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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see Authors & Referees and the Editorial Policy Checklist.

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Statistics	
For all statistical analy	yses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a Confirmed	
☐ ☐ The exact sa	mple size (n) for each experimental group/condition, given as a discrete number and unit of measurement
A statement	on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
The statistics	al test(s) used AND whether they are one- or two-sided tests should be described solely by name; describe more complex techniques in the Methods section.
A description	n of all covariates tested
A description	n of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
A full description AND variation	otion of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) on (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
For null hypo	othesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted as exact values whenever suitable.
For Bayesian	analysis, information on the choice of priors and Markov chain Monte Carlo settings
For hierarch	ical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
Estimates of	effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
1	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.
Software and	code
Policy information ab	out <u>availability of computer code</u>
Data collection	Image Lab, Stepone Software v2.3, cellSens Dimension.
Data analysis	Microsoft Excel, ImageJ (JACoP v.2.0), SPSS software (version 22.0, SPSS Inc), Adobe Photoshop CC2015, Adobe Ilustrator CC 2015, Microsoft Powerpoint.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The data that support the findings of this study are available within the article and its Supplementary Information or from the corresponding authors on reasonable request. The source data underlying Figs 4a, 4e, 5d, 5i, 6c, and 6i and Supplementary Figs 5b, 5d, 6b and 7a are provided as a Source Data file

Field-spe	cific reporting	
Please select the or	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.	
∠ Life sciences	Behavioural & social sciences Ecological, evolutionary & environmental sciences	
For a reference copy of t	he document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>	
Life scier	ices study design	
All studies must dis	close on these points even when the disclosure is negative.	
Sample size	The sample size for each experiment, n, is included in the results section and the associated figure legend.	
Data exclusions	No data were excluded from the analyses.	
Replication	Data are representatives of at least three independent experiments with similar results and plotted as mean ± SD	
Randomization	Mice were random allocated into experimental group	
Blinding	The investigators were blinded to group allocation during data collection and analysis.	
Reportin	g for specific materials, systems and methods	
	on from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, ed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.	
	perimental systems Methods	
n/a Involved in th	<u> </u>	
Antibodies	ChIP-seq	
☐ Eukaryotic	cell lines	
Palaeontol	pgy MRI-based neuroimaging	
	d other organisms	
	earch participants	
Clinical dat	a	
Antibodies		
Antibodies used Antibody against PAR (Cat#4335-MC-100; 1:1000 for WB; 1:100 for IF) were purchased from Trevigen (MD, USA). A against PARP1 (Cat#9532; 1:1000), Runx2 (Cat#12556; 1:1000 for WB; 1:100 for IF), pSTAT3 (Cat#9145; 1:1000), ST (Cat#12640; 1:1000), pJAK2 (Cat#3776; 1:1000), JAK2 (Cat#3230; 1:1000), pSRC (Cat#6943; 1:1000) and SRC (Cat#2 were obtained from Cell Signaling Technology (MA, USA). Antibodies against SMA (Cat#ab8226; 1:2000 for WB; 1:1 osteopotin (Cat#ab8448; 1:500), OC (Cat#ab13420; 1:500), osterix (Cat#ab209484; 1:1000), MSX2 (Cat#ab223692; (Cat#ab185966; 1:1000), GAPDH (Cat#ab8245; 1:2000) and Argonaute-2 (Cat#ab32381;1:100) were purchased from (Cambridge, UK). Antibodies against SM22 (Cat#10493-1-AP; 1:1000) and SMMHC (Cat#21404-1-AP; 1:1000) were Proteintech (IL, USA). Antibodies against Pit1 (Cat#a4117; 1:500) and Pit2 (Cat#a6739; 1:500) were purchased from (Wuhan, China). The Col1A1 (Cat#NB600-408; 1:500) antibody was purchased from — Novus (CO, USA). TRPM3 (Cat 1:500) antibody was purchased from GeneTex (CA, USA).		
Validation	All antibodies are commercial available and validated.	
Fukaryotic o	all lines	
Eukaryotic cell lines Policy information about cell lines		
Cell line source(s)		

Cell line source(s)

A7r5 cell is from the aorta of rat. HASMC cell is from the aorta of human.

Authentication

The A7r5 and HASMC cell lines were obtained from ATCC.

Mycoplasma contamination

All cell lines are negative for mycoplasma contamination

Commonly misidentified	line
(See ICLAC register)	

Field-collected samples

N/A			
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Animals and other organisms

N/A

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research		
Laboratory animals	eight-week-old male Wistar rats	

Wild animals N/A

Ethics oversight

All experimental protocols were approved by the Ethics Committee of Tongji Medical College, Huazhong University of Science and Technology, and were performed in accordance with relevant institutional and national guidelines and regulations.

Note that full information on the approval of the study protocol must also be provided in the manuscript.