Reviewer Report

Title: Single-cell RNA-seq reveals dynamic transcriptome profiling in human early neural differentiation

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Reviewer Comments to Author:

The present manuscript by Shang et al. performed parallel single cell transcriptome and bulk chromatin landscape profiling during six consecutive stages during neural differentiation of fibroblast-based iPSCs. Based on a comprehensive amount of data and detailed analysis, this well-written and well-visualized paper provides many specific and novel insights into gene expression changes specific to neural differentiation. While I feel that this manuscript is a very interesting read as well as a valuable resource for the field, functional validation of cellular heterogeneity, as well as of the putative novel hub TFs would substantially improve the current study. Below please find my specific points that I feel should be addressed prior to publication.

Major points:

1. Heterogeneity

Definition of the subclasses at the different stages is done based on single-cell RNAseq. Based on the presented data, it is not clear if the observed heterogeneity...

i) ...stems from the fact that the harvested cells at each stage contain lagging and leading cells (slow and fast differentiating cells).

ii) ... represent temporal transcriptional states (e.g. during cell cycle or circadian rhythm).

iii) ... represent distinct cellular subpopulation that occur in parallel, but that have different fates (e.g. dorsal, ventral, neural crest, ...).

While the Monocle analysis appears to suggest iii), the data is not able to convincingly draw a conclusion in this regard. I thus suggest that the authors should mention this as a caveat more prominently. Alternatively, single-cell ATACseq might help to gain insights if the observed subclasses are also distinguishable epigenetically, but the effort seems immense.

2. Validation of heterogeneity

For EBs, Ros-E and Ros-L, the authors find strikingly different subclasses of cells. Based on a few selected 'novel' markers, it would be very interesting to see if this heterogeneity can be confirmed by immunostaining with reasonable effort.

3. Fibroblasts

The study analyzed fibroblasts as well, but in most of the presented the analyses they are not included. It would be interesting to see differences relative to fibroblasts also in the Monocle and ATACseq analysis. Also, why are some fibroblasts clustering with EBs in t-SNE (Fig 2a), and how would a PCA of these data look like?

4. Universal validity / N of genetic background

As that the authors apparently only used one fibroblast line/genetic background for this study, the cogency of the study is limited. While I agree that repeating all experiments with a second line would be very time and money consuming, this caveat leaves the possibility for each finding to be an artifact of this one cell line/genetic background. The authors should try to at least validate some of their key findings/TFs on one another genetic background.

5. Functional validation of new TFs

Based on the identification of 'novel' TFs involved in neural differentiation, it would be interesting if overexpression or knockdown of these factors boost/impair neuronal differentiation of iPSCs.

6. Chromatin closing

For the ATACseq data, in addition to reporting the % novel peaks for each stage, I would be curious to know the % change in peak diversity between each time, because this would take into account both regions of the genome opening up, and regions of the genome closing up, instead of just opening.

7. Neurons

It would be interesting to see differentiated neurons included in this already very interesting paper!

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