

Supporting Information

Catalytic Asymmetric Allylation of Aldehydes with Alkenes through Allylic C(sp³)-H Functionalization Mediated by Organophotoredox and Chiral Chromium Hybrid Catalysis

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1. General Method

^1H NMR spectra were recorded on JEOL ECX500 (500 MHz for ^1H NMR and 125.65 MHz for ^{13}C NMR), and JEOL ECS400 (400 MHz for ^1H NMR and 100 MHz for ^{13}C NMR, and 370 MHz for ^{19}F NMR) spectrometer. For ^1H NMR and ^{13}C NMR, chemical shifts were reported in the scale relative to CDCl_3 ($\delta = 7.26$ for ^1H NMR and $\delta = 77.0$ for ^{13}C NMR) used as an internal reference. Electrospray ionization (ESI)-mass spectra were measured on a JEOL JMS-T100LC AccuTOF spectrometer for HRMS. DART-mass spectra were measured on a JEOL JMS-T100LC AccuTOF spectrometer. Infrared (IR) spectra were recorded on a JASCO FT/IR 410 Fourier transform infrared spectrophotometer. The enantiomeric excess (ee) was determined by HPLC analysis (JASCO HPLC systems; pump: PU-986; detector: UV-2075, measured at 210 nm or 254 nm; column: CHIRALPAK IA3, IBN3, IC3, IF3). Optical rotations were measured on a JASCO P-1010 polarimeter. Column chromatographies were performed with silica gel Merck 60 (230-400 mesh ASTM), Biotage Isolera One and Biotage SNAP Ultra, or Yamazen Smart Flash and Universal Column Premium. All non-commercially available compounds were prepared and characterized as described in Section 5 of this SI. Other reagents were purchased from Aldrich, Tokyo Chemical Industry Co., Ltd. (TCI), Kanto Chemical Co., Inc., and Wako Pure Chemical Industries, Ltd., and were used as received. A Valore VBP-L24-C2 with 38W LED lamp (VBL-SE150-BBB (430)) was used as the 430 nm light source.

2. Preparation of Catalysts and Substrates

Alkenes (**1a-1e**), aldehydes (**2**), Cr catalysts (CrCl_2 , $\text{CrCl}_3 \cdot 3\text{THF}$), ligand **L2**, photocatalysts (**10**, **11**) and additives (LiCl , LiI , LiBF_4 , LiClO_4 , NaClO_4 , $\text{Ca}(\text{ClO}_4)_2 \cdot x\text{H}_2\text{O}$, $\text{Mg}(\text{ClO}_4)_2$, NaOtBu) were commercially available and used as purchased. Chiral ligands (**L1**, **L3-L5**) were prepared according to the reported method.¹⁻⁴ MgPhPO_3 was synthesized according to related procedure for the preparation of metal phosphates.⁵

Preparation of MgPhPO_3

To a solution of $\text{Mg}(\text{OEt})_2$ (572 mg, 5 mmol) in EtOH (50 ml), phenylphosphonic acid (791 mg, 5 mmol) was added portion wise at 0 °C. The mixture was stirred for 15 min and concentrated under vacuo. The white solid was used for next reaction without further purification.

3. General Procedure for Catalytic Asymmetric Allylation of Aldehydes

Procedure for preparation of **8a-8i**, **8k**

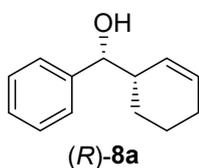


CrCl_2 (1.5 mg, 0.0125 mmol, 5 mol %) and **L5** (4.5 mg, 0.0125 mmol, 5 mol %) were dissolved in degassed

CH₂Cl₂ (2.5 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. Cyclohexene **1a** (507 μL, 5.0 mmol, 20 eq), Mg(ClO₄)₂ (55.8 mg, 0.25 mmol, 100 mol %), benzaldehyde **2a** (25.5 μL, 0.25 mmol, 1 eq) and photocatalyst **10** (2.5 mg, 0.00625 mmol, 2.5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was passed through a pad of silica gel with CH₂Cl₂ elution. After evaporation, diastereomeric ratio was determined by NMR analysis (68%, >20/1). The residue was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/2 to 2/1, v/v) to afford (*R*)-**8a** as colorless oil (25.8 mg, 55%).

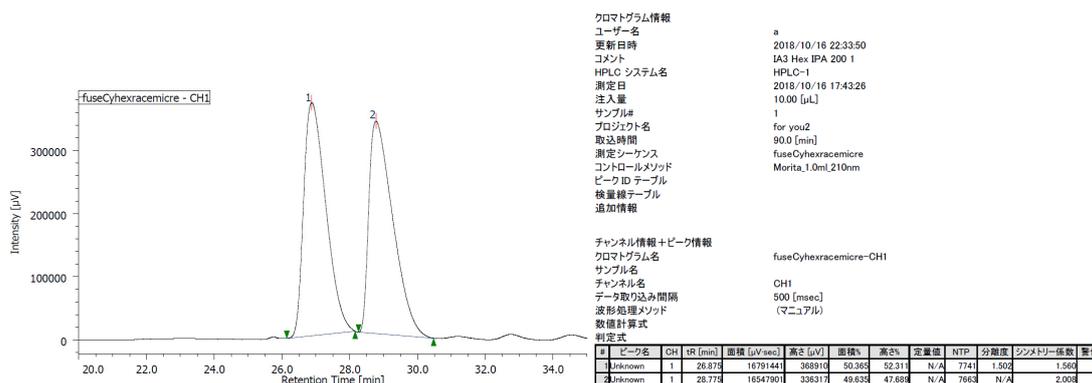
The amount of CrCl₂ and **L5** was increased to 10 mol% (CrCl₂: 3.0 mg, 0.025 mmol, **L5**: 9.0 mg, 0.025 mmol), (*R*)-**8a** was obtained in 65% (30.5 mg).

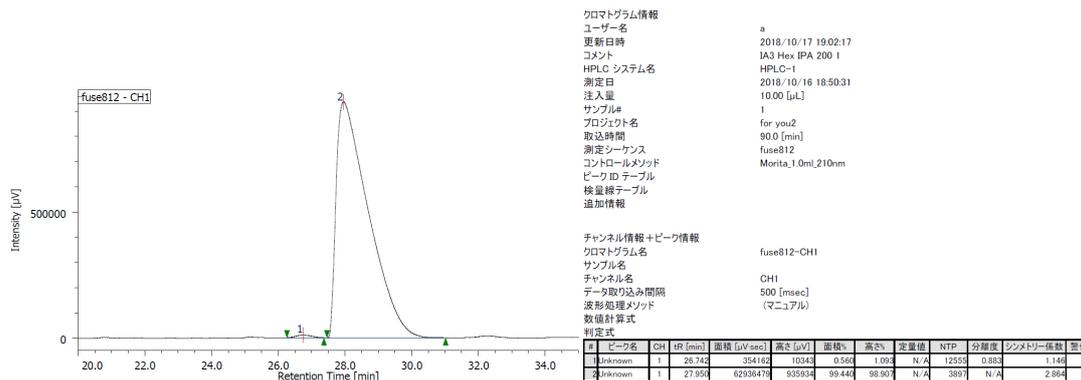
(*R*)-((*S*)-cyclohex-2-en-1-yl)(phenyl)methanol (8a**).**



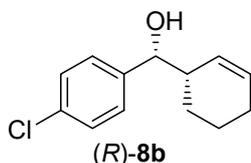
All the spectroscopic data matches with the previously reported data.⁶

¹H NMR (CDCl₃, 500 MHz) δ 1.38-1.61 (m, 2H), 1.62-1.83 (m, 2H), 1.84-2.10 (m, 3H), 2.44-2.56 (m, 1H), 4.58 (d, *J* = 6.4 Hz, 1H), 5.38 (dd, *J* = 1.8, 10.1 Hz, 1H), 5.77-5.87 (m, 1H), 7.25-7.39 (m, 5H); ¹³C NMR (CDCl₃, 125 MHz) δ 21.0, 23.8, 25.2, 43.0, 76.7, 126.5, 127.4, 128.0, 128.2, 130.4, 142.8; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 26.7 min (minor) and 28.0 min (major); [α]_D²⁰ = -4.8 (*c* = 0.76, CHCl₃, for 95% ee sample).



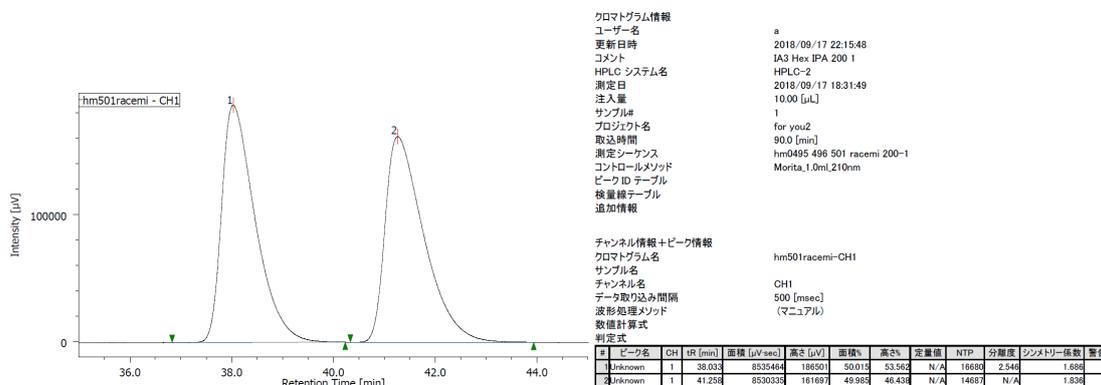


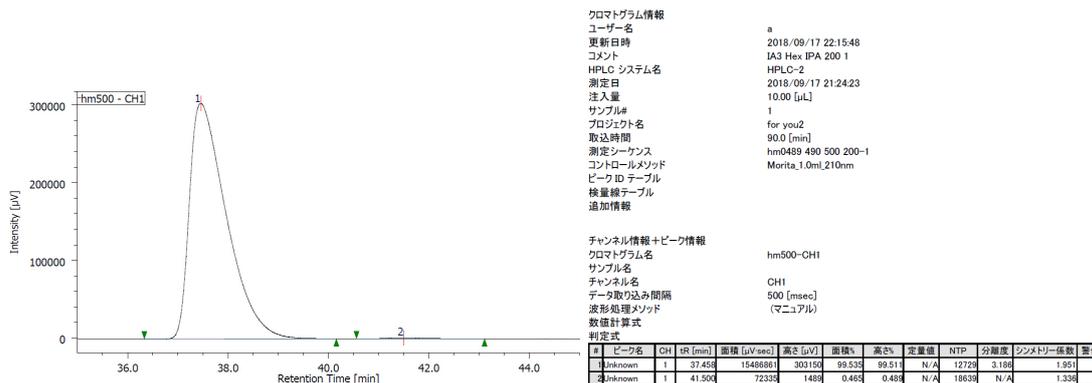
(R)-(4-chlorophenyl)((S)-cyclohex-2-en-1-yl)methanol (8b).



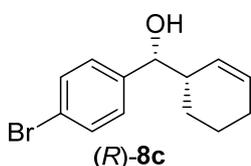
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (R)-8b as colorless oil (45.8 mg, 82%).

IR (neat) ν 3396, 3022, 2927, 2861, 1698, 1651, 1597, 1541, 1492, 1410, 1090, 1013, 837, 725 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 1.42-1.54 (m, 2 H), 1.58-1.68 (m, 1H), 1.69-1.78 (m, 1H), 1.86 (d, *J* = 2.3 Hz, 1H), 1.91-2.02 (m, 2H), 2.40-2.50 (m, 1H), 4.60 (dd, *J* = 2.3, 6.3 Hz, 1H), 5.39 (dd, *J* = 2.3, 10.3 Hz, 1H), 5.78-5.87 (m, 1H), 7.24-7.33 (m, 4H); ¹³C NMR (CDCl₃, 125 MHz) δ 21.0, 23.6, 25.1, 43.0, 76.6, 127.5, 127.8, 128.3, 130.9, 132.9, 141.2; LRMS (ESI): *m/z* 245 [M+Na⁺]; HRMS (ESI): *m/z* calculated for C₁₃H₁₅ClNaO⁺ [M+Na⁺]: 245.0704, found: 245.0701; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 37.5 min (major) and 41.5 min (minor); [α]_D²⁰ = -24.6 (*c* = 1.0, C₆H₆, for 99% ee sample).



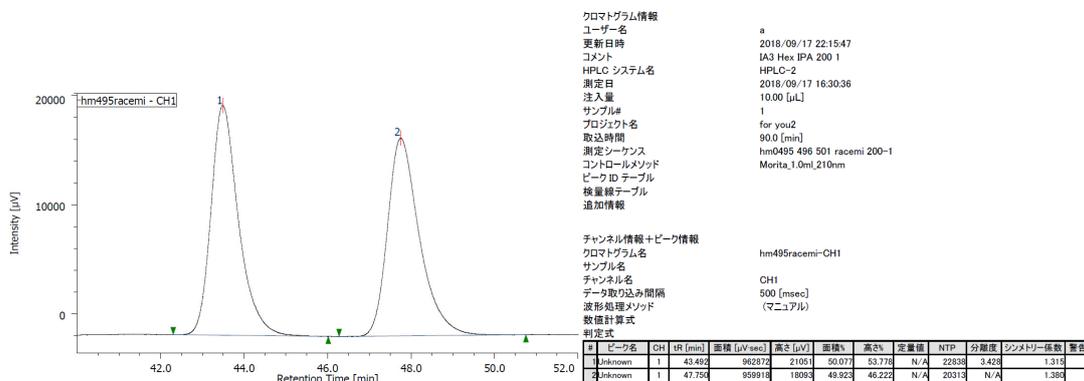


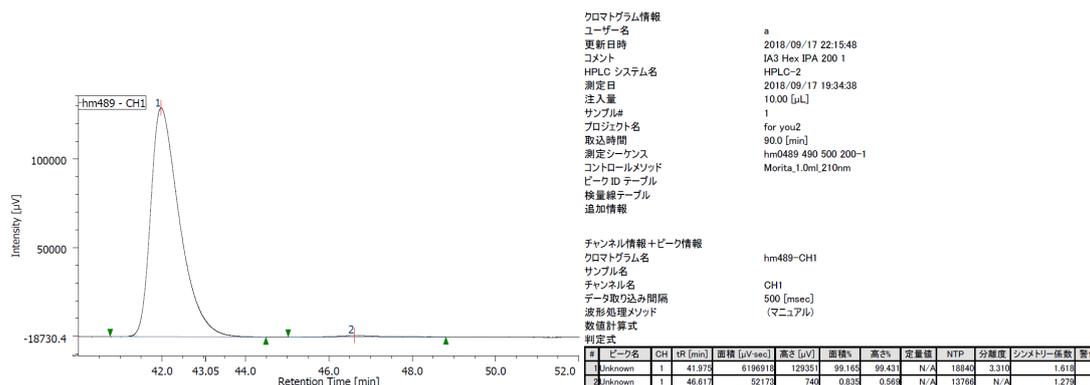
(R)-(4-bromophenyl)((S)-cyclohex-2-en-1-yl)methanol (8c).



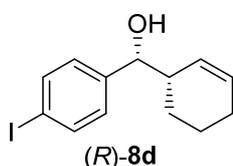
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (R)-8c as colorless oil (54 mg, 81%).

IR (neat) ν 3396, 3022, 2927, 2860, 1651, 1592, 1488, 1404, 1072, 1010, 812, 725 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 1.45-1.52 (m, 2 H), 1.57-1.68 (m, 1H), 1.70-1.78 (m, 1H), 1.85 (d, *J* = 2.9 Hz, 1H), 1.93-2.20 (m, 2H), 2.41-2.50 (m, 1H), 4.59 (dd, *J* = 2.3, 6.3 Hz, 1H), 5.39 (dd, *J* = 2.3, 10.3 Hz, 1H), 5.80-5.88 (m, 1H), 7.18-7.30 (m, 2H) , 7.43-7.49 (m, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 21.2, 23.8, 25.4, 43.1, 77.5, 121.3, 127.8, 128.4, 131.1, 131.4, 142.0; LRMS (ESI): *m/z* 289 [M+Na⁺]; HRMS (ESI): *m/z* calculated for C₁₃H₁₅BrNaO⁺ [M+Na⁺]: 289.0198, found: 289.0197; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 42.0 min (major) and 46.6 min (minor); [α]_D²⁰ = -5.8 (*c* = 0.66, CHCl₃, for 98% ee sample).



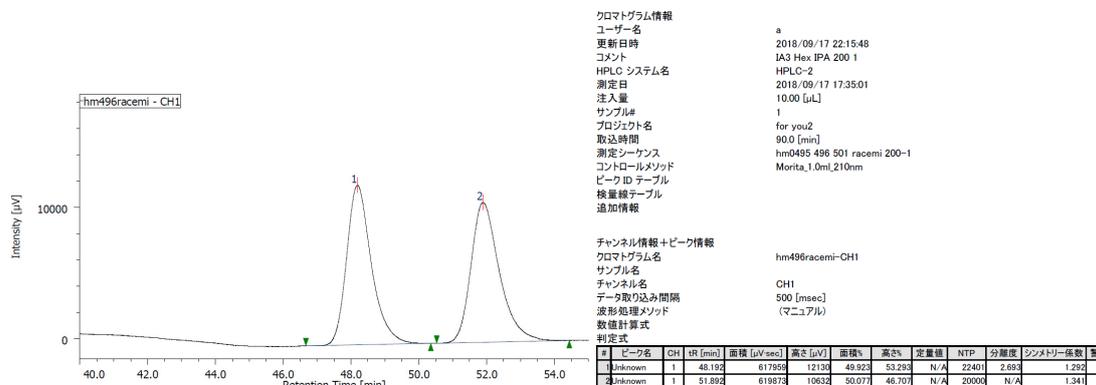


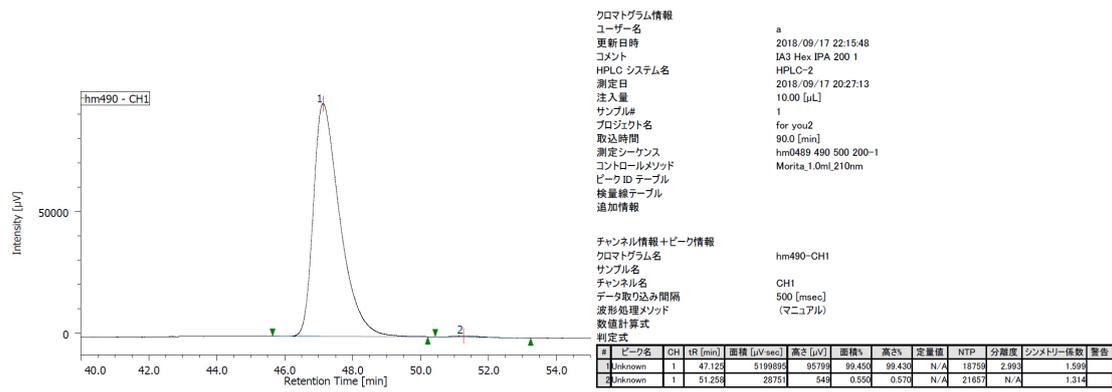
(R)-((S)-cyclohex-2-en-1-yl)(4-iodophenyl)methanol (8d).



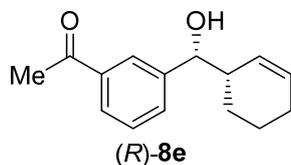
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (R)-**8d** as colorless oil (36.3 mg, 46%).

IR (neat) ν 3392, 3021, 2926, 2859, 1698, 1651, 1588, 1481, 1399, 1059, 1004, 811, 725 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 1.42-1.54 (m, 2 H), 1.59-1.67 (m, 1H), 1.68-1.79 (m, 1H), 1.86 (d, *J* = 1.7 Hz, 1H), 1.93-2.20 (m, 2H), 2.39-2.50 (m, 1H), 4.57 (dd, *J* = 1.7, 6.3 Hz, 1H), 5.39 (dd, *J* = 2.3, 10.3 Hz, 1H), 5.78-5.89 (m, 1H), 7.10 (d, *J* = 8.3 Hz, 2H), 7.67 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 21.0, 23.5, 25.1, 42.9, 76.6, 92.7, 127.5, 128.5, 130.9, 137.2, 142.4; LRMS (ESI): *m/z* 337 [M+Na⁺]; HRMS (ESI): *m/z* calculated for C₁₃H₁₅INaO⁺ [M+Na⁺]: 337.0060, found: 337.0073; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 47.1 min (major) and 51.3 min (minor); [α]_D²⁰ = -6.3 (*c* = 2.7, CHCl₃, for 99% ee sample).



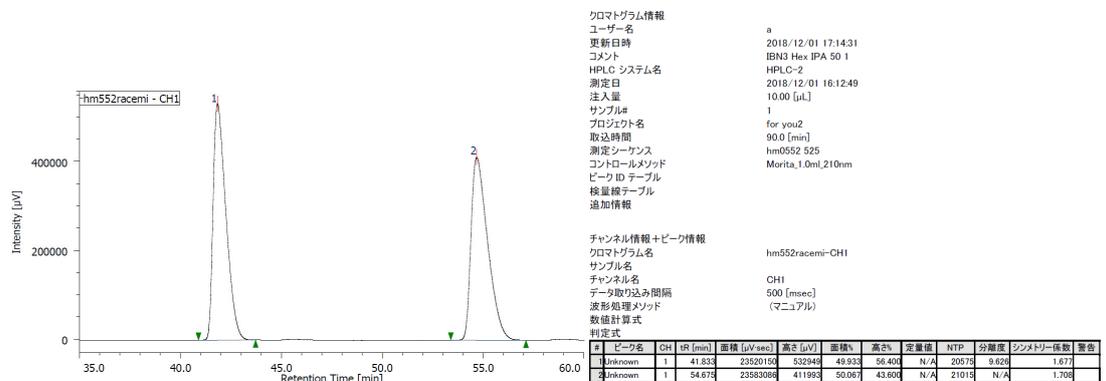


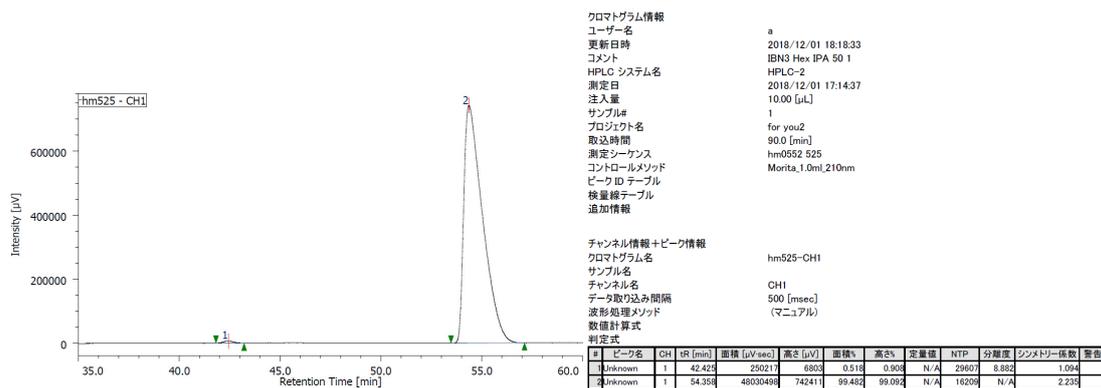
1-(3-((R)-((S)-cyclohex-2-en-1-yl)(hydroxy)methyl)phenyl)ethan-1-one (8e).



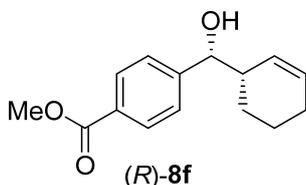
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (AcOEt/hexane = 1/10 to 1/1, v/v) to afford (R)-8e as colorless oil (26.7 mg, 46%).

IR (neat) ν 3444, 3021, 2927, 2860, 1683, 1601, 1584, 1433, 1360, 1276, 1184, 1025, 699 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 1.45-1.54 (m, 2 H), 1.58-1.67 (m, 1H), 1.71-1.81 (m, 1H), 1.92 (d, $J = 2.3$ Hz, 1H), 1.96-2.03 (m, 2H), 2.48-2.57 (m, 1H), 2.62 (s, 3H), 4.71 (dd, $J = 1.7, 5.7$ Hz, 1H), 5.41 (dd, $J = 1.7, 9.7$ Hz, 1H), 5.83-5.90 (m, 1H), 7.45 (dd, $J = 7.7, 7.7$ Hz, 1H), 7.57 (d, $J = 7.7$ Hz, 1H), 7.87 (d, $J = 7.7$ Hz, 1H), 7.93 (s, 1H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 21.0, 23.4, 25.1, 26.7, 43.0, 76.8, 126.3, 127.4, 127.5, 128.4, 131.0, 131.2, 137.0, 143.4, 198.3; LRMS (DART): m/z 231 [$\text{M}+\text{H}^+$]; HRMS (DART): m/z calculated for $\text{C}_{15}\text{H}_{19}\text{O}_2^+$ [$\text{M}+\text{H}^+$]: 231.1380, found: 231.1371; HPLC (chiral column: CHIRALPAK IBN3; solvent: hexane/2-propanol = 50/1; flow rate: 1.0 mL/min; detection: at 210 nm): $t_R = 42.4$ min (minor) and 54.4 min (major); $[\alpha]_D^{20} = +6.9$ ($c = 0.54$, CHCl_3 , for 99% ee sample).



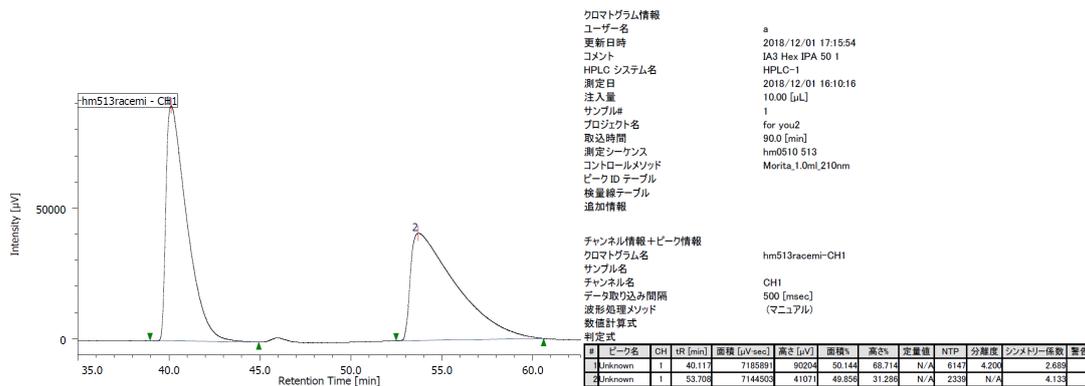


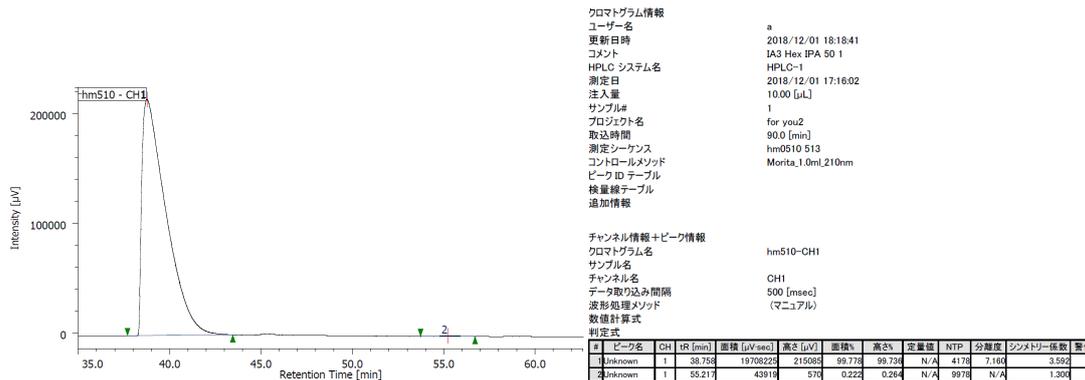
Methyl 4-((*R*)-((*S*)-cyclohex-2-en-1-yl)(hydroxy)methyl)benzoate (**8f**).



Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (AcOEt/hexane = 1/10 to 1/1, v/v) to afford (*R*)-**8f** as colorless oil (49.4 mg, 80%). All the spectroscopic data matches with the previously reported data.⁷

¹H NMR (CDCl₃, 500 MHz) δ 1.44-1.62 (m, 1H), 1.66-1.80 (m, 1H), 1.90-2.02 (m, 3H), 2.41-2.46 (m, 1H), 3.91 (s, 3H), 4.70 (dd, *J* = 2.3, 6.3 Hz, 1H), 5.42 (dd, *J* = 1.7, 10.3 Hz, 1H), 5.84-5.90 (m, 1H), 7.42 (d, *J* = 8.3 Hz, 2H), 8.02 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 21.0, 23.3, 25.1, 42.9, 52.0, 76.6, 126.4, 127.5, 129.0, 129.4, 131.0, 148.1, 167.0; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 50/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 38.8 min (major) and 55.2 min (minor); [α]_D^{20.9} = +5.1 (*c* = 0.97, CHCl₃, for 99% ee sample).

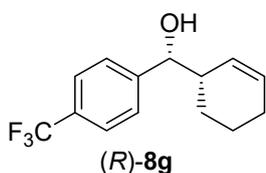




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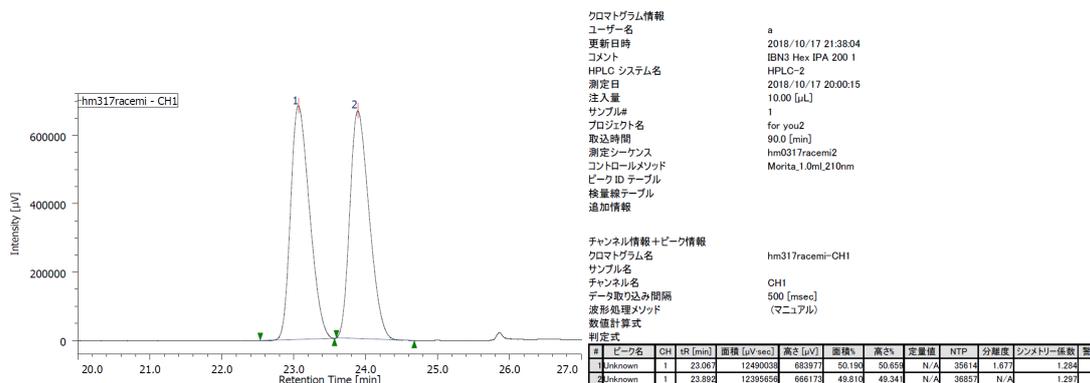
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(R)-((S)-cyclohex-2-en-1-yl)(4-(trifluoromethyl)phenyl)methanol (8g).



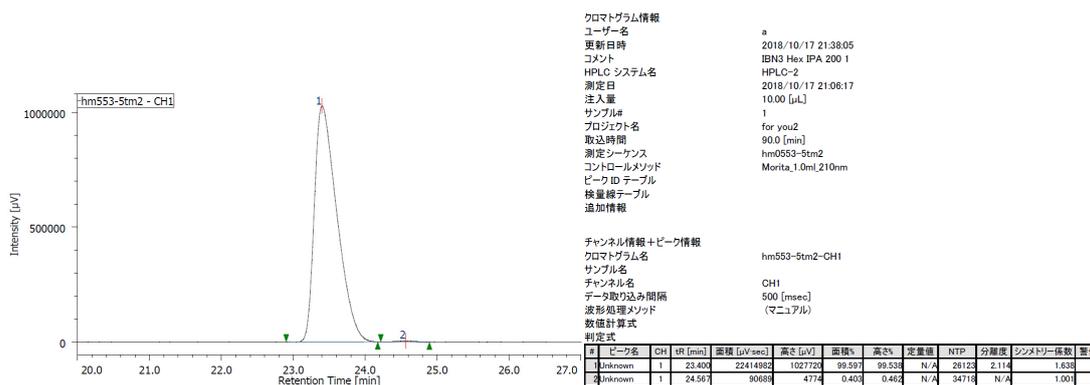
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (R)-8g as colorless oil (40.2 mg, 63%). All the spectroscopic data matches with the previously reported data.⁸

¹H NMR (CDCl₃, 500 MHz) δ 1.45-1.62 (m, 2H), 1.70-1.81 (m, 1H), 1.91 (d, *J* = 2.3 Hz, 1H), 1.97-2.03 (m, 2H), 2.47-2.56 (m, 1H), 4.71 (d, *J* = 8.0 Hz, 2H), 5.42 (dd, *J* = 2.3, 10.3 Hz, 1H), 5.85-5.92 (m, 1H), 7.47 (d, *J* = 8.0 Hz, 2H), 7.60 (d, *J* = 8.3 Hz, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 21.0, 23.2, 25.1, 43.0, 76.5, 124.2 (q, *J* = 270.7 Hz), 125.1 (q, *J* = 4.8 Hz), 126.7, 127.3, 129.5 (q, *J* = 32.2 Hz), 131.3, 146.7; HPLC (chiral column: CHIRALPAK IBN3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 23.4 min (major) and 24.6 min (minor); [*α*]_D²⁰ = +8.6 (*c* = 1.1, CHCl₃, for 99% ee sample).

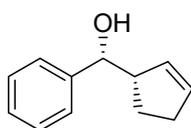


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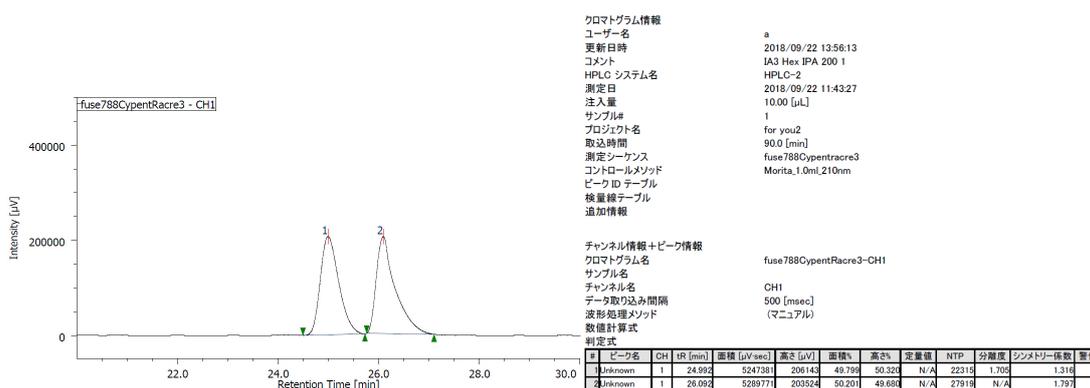
(R)-((S)-cyclopent-2-en-1-yl)(phenyl)methanol (8h).

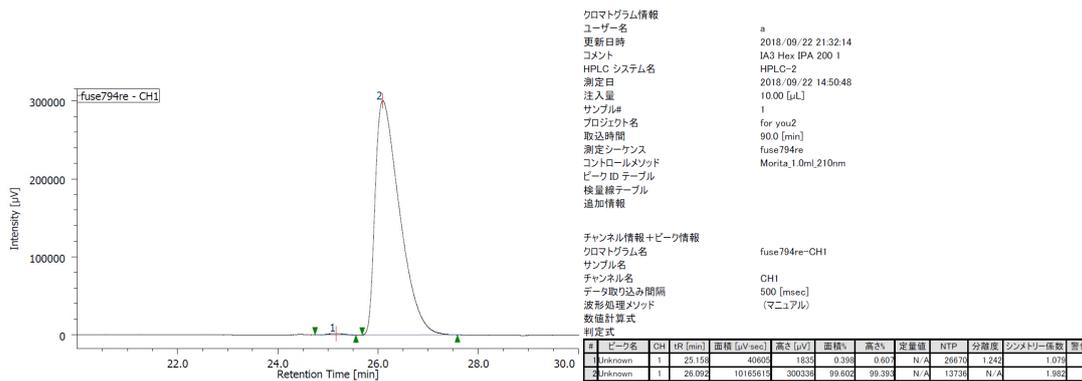


(R)-8h

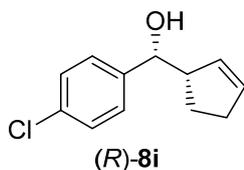
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/2 to 2/1, v/v) to afford (R)-**8h** as colorless oil (25.5 mg, 59%). All the spectroscopic data matches with the previously reported data.⁶

¹H NMR (CDCl₃, 500 MHz) δ 1.78-2.00 (m, 3H), 2.24-2.45 (m, 2H), 3.05-3.17 (m, 1H), 4.58 (d, *J* = 6.4 Hz, 1H), 5.38-5.44 (m, 1H), 5.83-5.92 (m, 1H), 7.22-7.42 (m, 5H); ¹³C NMR (CDCl₃, 125 MHz) δ 25.0, 32.2, 53.9, 76.7, 126.2, 127.3, 128.2, 131.2, 133.7, 143.4; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 25.2 min (minor) and 26.1 min (major); [α]_D²⁰ = -67 (*c* = 1.56, CHCl₃, for 99% ee sample).



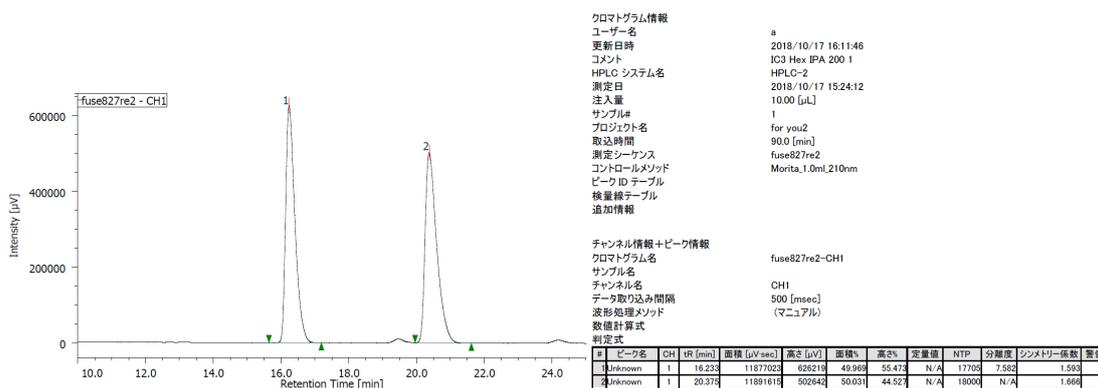


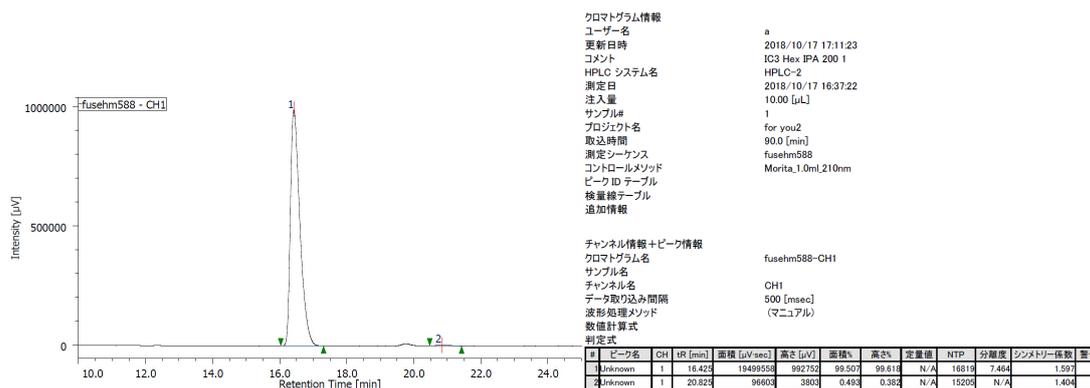
(R)-(4-chlorophenyl)((S)-cyclopent-2-en-1-yl)methanol (8i).



Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/2 to 2/1, v/v) to afford (R)-8i as colorless solid (28.5 mg, 55%).

IR (neat) ν 3390, 3053, 2929, 2850, 1902, 1716, 1597, 1489, 1410, 1293, 1089, 1013, 827, 726 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 1.72-1.97 (m, 3H), 2.23-2.43 (m, 2H), 3.01-3.12 (m, 1H), 4.58 (d, *J* = 6.4 Hz, 1H), 5.37-5.45 (m, 1H), 5.84-5.93 (m, 1H), 7.24-7.34 (m, 4H); ¹³C NMR (CDCl₃, 125 MHz) δ 24.6, 32.2, 53.9, 76.0, 127.6, 128.3, 130.8, 132.9, 134.2, 141.8; LRMS (ESI): *m/z* 231 [M+Na⁺]; HRMS (ESI): *m/z* calculated for C₁₂H₁₃ClNaO⁺ [M+Na⁺]: 231.0547, found: 231.0550; HPLC (chiral column: CHIRALPAK IC3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 16.4 min (major) and 20.8 min (minor); [α]_D²⁰ = -60 (*c* = 0.98, CHCl₃, for 99% ee sample).

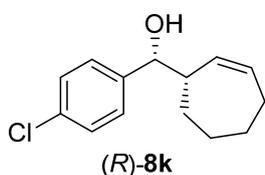




クロマトグラム情報
 ユーザー名 a
 更新日時 2018/10/17 17:11:23
 コメント IC3 Hex IPA 200 1
 HPLC システム名 HPLC-2
 測定日 2018/10/17 16:37:22
 注入量 10.00 [μL]
 サンプル# 1
 プロジェクト名 for you2
 取込時間 90.0 [min]
 測定シーケンス fusehm588
 コントロールメソッド Morita_1.0ml_210nm
 ピークIDテーブル
 検量線テーブル
 追加情報

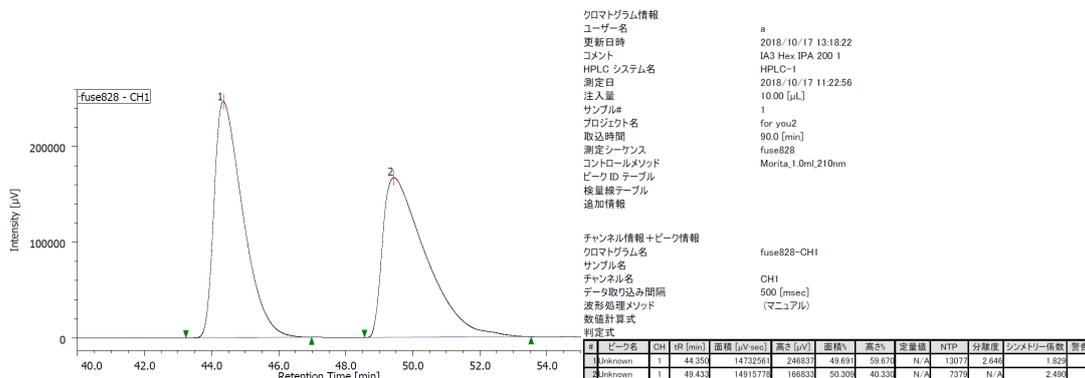
チャンネル情報+ピーク情報
 クロマトグラム名 fusehm588-CH1
 サンプル名
 チャンネル名 CH1
 データ取り込み間隔 500 [msec]
 波形処理メソッド (マニュアル)
 数値計算式
 判定式

(R)-(4-chlorophenyl)((S)-cyclohept-2-en-1-yl)methanol (8k).



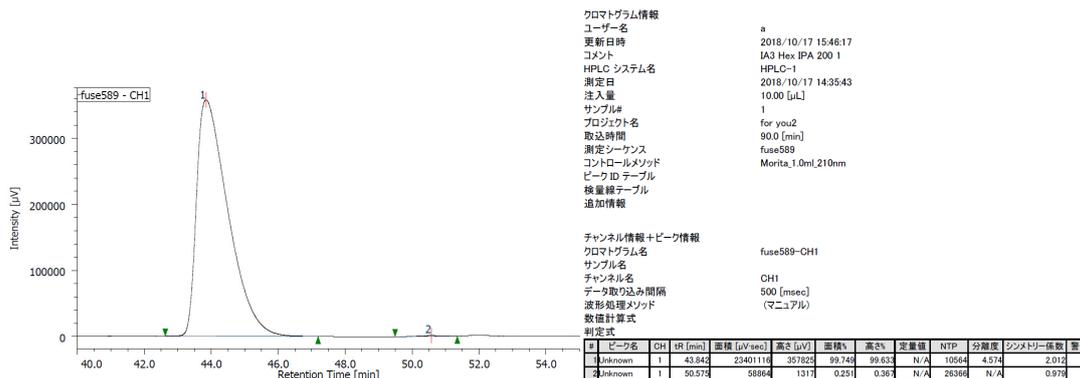
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/2 to 2/1, v/v) to afford (R)-8k as colorless solid (28.1 mg, 47%).

IR (neat) ν 3389, 3020, 2923, 2852, 1901, 1721, 1650, 1597, 1490, 1444, 1090, 1013, 832, 687 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 1.23-1.41 (m, 2H), 1.47-1.72 (m, 2H), 1.77-2.21 (m, 5H), 2.56-2.65 (m, 1H), 4.65 (d, *J* = 6.3 Hz, 1H), 5.46-5.54 (m, 1H), 5.75-5.84 (m, 1H), 7.21-7.37 (m, 4H); ¹³C NMR (CDCl₃, 125 MHz) δ 26.7, 27.9, 28.5, 29.8, 47.0, 76.7, 128.0, 128.4, 132.8, 133.1, 133.1, 141.6; LRMS (ESI): *m/z* 259[M+Na⁺]; HRMS (ESI): *m/z* calculated for C₁₄H₁₇ClNaO⁺ [M+Na⁺]: 259.0860, found: 259.0854; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 43.8 min (major) and 50.6 min (minor); [α]_D^{20.9} = +63 (*c* = 0.92, CHCl₃, for 99% ee sample).

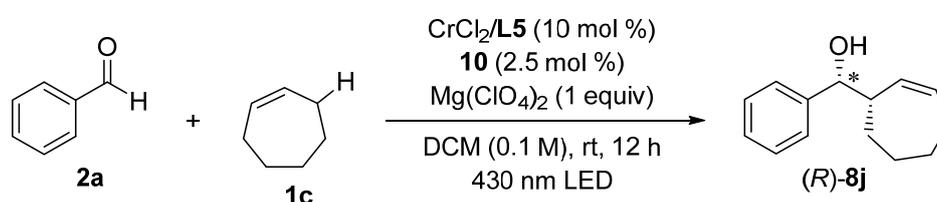


クロマトグラム情報
 ユーザー名 a
 更新日時 2018/10/17 13:18:22
 コメント IA3 Hex IPA 200 1
 HPLC システム名 HPLC-1
 測定日 2018/10/17 11:22:56
 注入量 10.00 [μL]
 サンプル# 1
 プロジェクト名 for you2
 取込時間 90.0 [min]
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 コントロールメソッド Morita_1.0ml_210nm
 ピークIDテーブル
 検量線テーブル
 追加情報

チャンネル情報+ピーク情報
 クロマトグラム名 fuse828-CH1
 サンプル名
 チャンネル名 CH1
 データ取り込み間隔 500 [msec]
 波形処理メソッド (マニュアル)
 数値計算式
 判定式

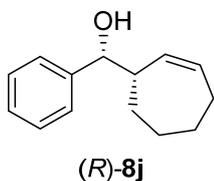


Procedure for preparation of **8j**



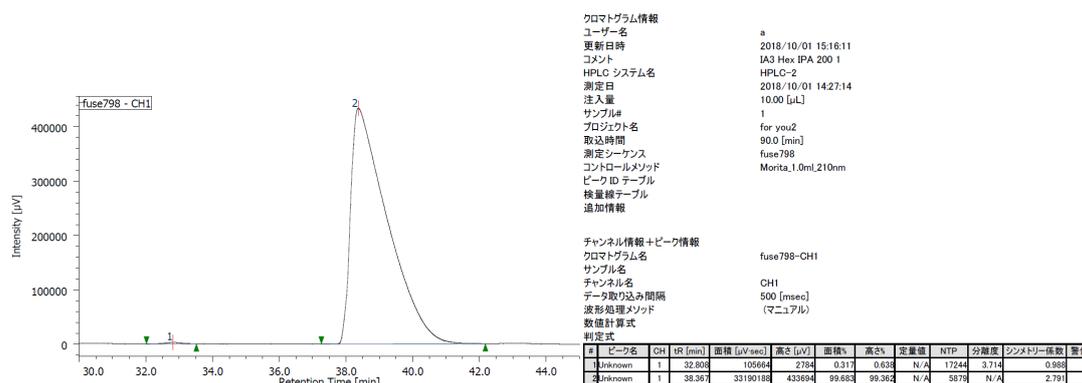
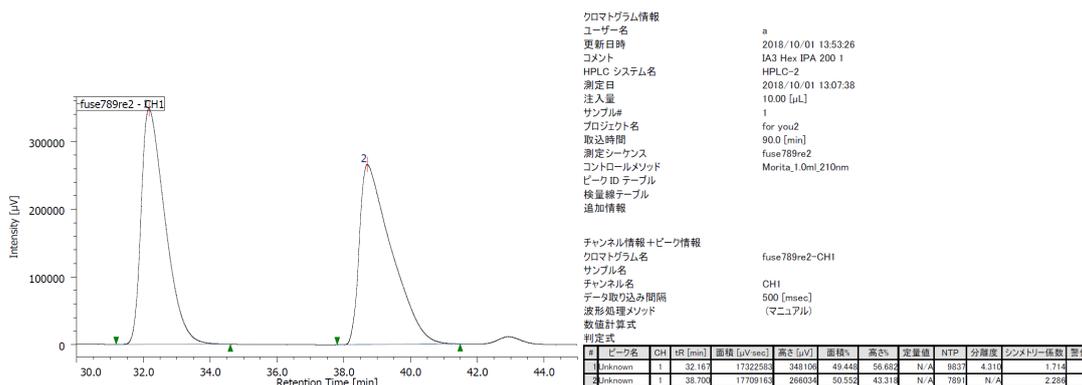
CrCl_2 (3.0 mg, 0.025 mmol, 10 mol %) and **L5** (9.0 mg, 0.025 mmol, 10 mol %) were dissolved in degassed CH_2Cl_2 (2.5 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. Cyclohexene **1c** (584 μL, 5.0 mmol, 20 eq), $\text{Mg}(\text{ClO}_4)_2$ (55.8 mg, 0.25 mmol, 100 mol %), benzaldehyde **2a** (25.5 μL, 0.25 mmol, 1 eq) and photocatalyst **10** (2.5 mg, 0.00625 mmol, 2.5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was passed through a pad of silica gel with CH_2Cl_2 elution. After evaporation, diastereomeric ratio was determined by NMR analysis (68%, >20/1). The residue was purified by silica gel flash column chromatography (CH_2Cl_2 /hexane = 1/2 to 2/1, v/v) to afford (*R*)-**8a** as colorless oil (28.4 mg, 56%).

(*R*)-((*S*)-cyclohept-2-en-1-yl)(phenyl)methanol (**8j**).

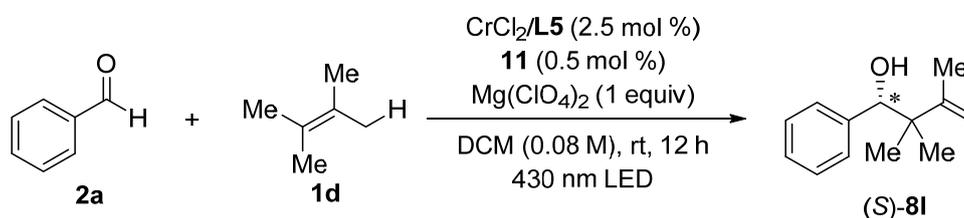


All the spectroscopic data matches with the previously reported data.⁹

^1H NMR (CDCl_3 , 500 MHz) δ 1.21-1.40 (m, 2H), 1.48-1.75 (m, 2H), 1.85-2.24 (m, 5H), 2.62-2.71 (m, 1H), 4.66 (d, $J = 7.3$ Hz, 1H), 5.53 (dd, $J = 3.2, 14.5$ Hz, 1H), 5.73-5.83 (m, 1H), 7.25-7.39 (m, 5H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 26.7, 28.1, 28.6, 30.0, 46.9, 77.4, 126.7, 127.5, 128.3, 132.4, 133.5, 143.2; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): $t_R = 32.8$ min (minor) and 38.4 min (major); $[\alpha]_D^{20} = +84$ ($c = 0.77$, CHCl_3 , for 99% ee sample).

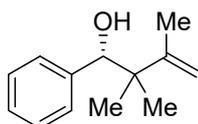


Procedure for preparation of **8l** and **8v**



CrCl_2 (1.5 mg, 0.0125 mmol, 2.5 mol %) and **L5** (4.5 mg, 0.0125 mmol, 2.5 mol %) were dissolved in degassed CH_2Cl_2 (4 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1d** (106 μL , 1.0 mmol, 2 eq), $\text{Mg}(\text{ClO}_4)_2$ (112 mg, 0.5 mmol, 100 mol %), benzaldehyde **2a** (51 μL , 0.5 mmol, 1 eq) and photocatalyst **11** (1.0 mg, 0.0025 mmol, 0.5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was directly loaded to silica gel and purified by flash column chromatography ($\text{CH}_2\text{Cl}_2/\text{hexane} = 1/10$ to 1/0, v/v) to afford **(R)-8l** as colorless oil (82.1 mg, 87%).

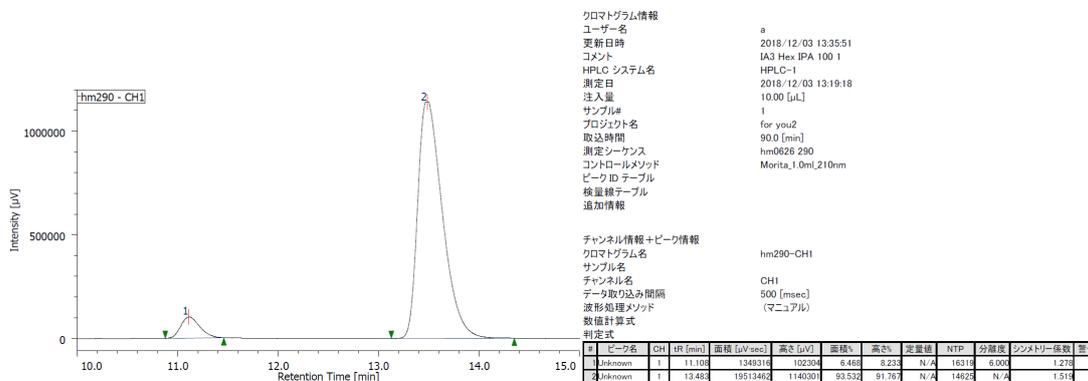
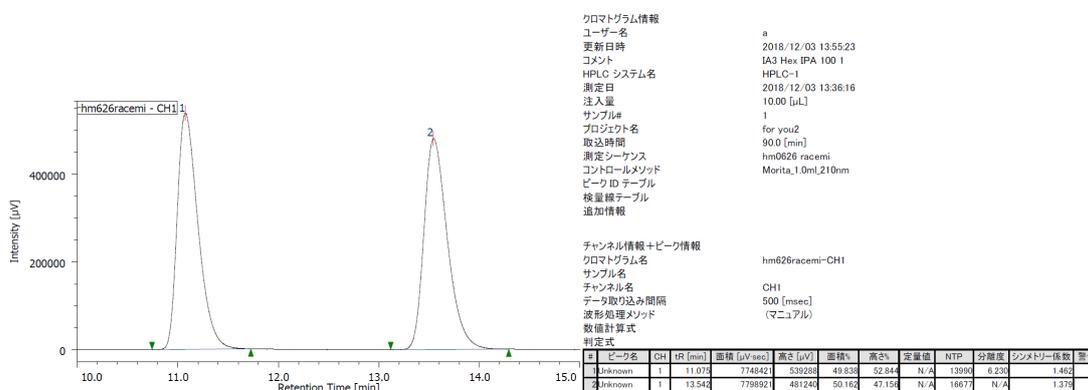
(S)-2,2,3-trimethyl-1-phenylbut-3-en-1-ol (8l).



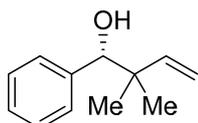
(S)-8I

All the spectroscopic data matches with the previously reported data.¹⁰

¹H NMR (CDCl₃, 500 MHz) δ 0.94 (s, 3H), 0.99 (s, 3H), 1.87 (s, 3H), 2.08 (d, *J* = 1.7 Hz, 1H), 4.63 (s, 1H), 4.97 (s, 1H), 5.04 (s, 1H), 7.18-7.36 (m, 5H); ¹³C NMR (CDCl₃, 125 MHz) δ 19.8, 20.1, 24.2, 44.5, 77.4, 113.2, 127.2, 127.4, 127.9, 140.2, 150.6; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 100/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 11.1 min (minor) and 13.5 min (major); [α]_D²⁰ = +99 (*c* = 0.24, CHCl₃, for 88% ee sample).



(R)-2,2-dimethyl-1-phenylbut-3-en-1-ol (8v).

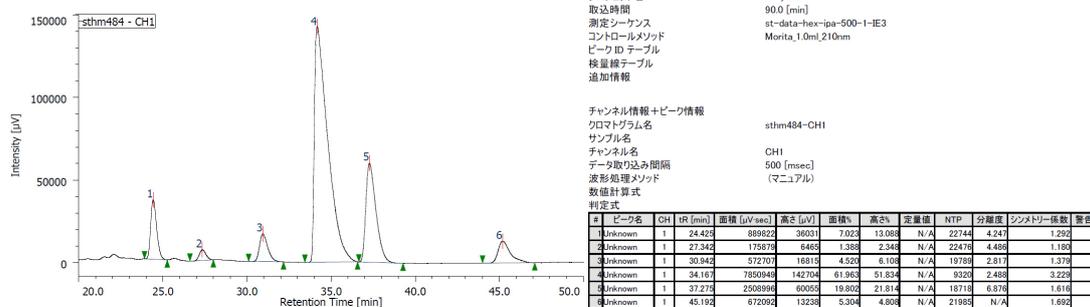
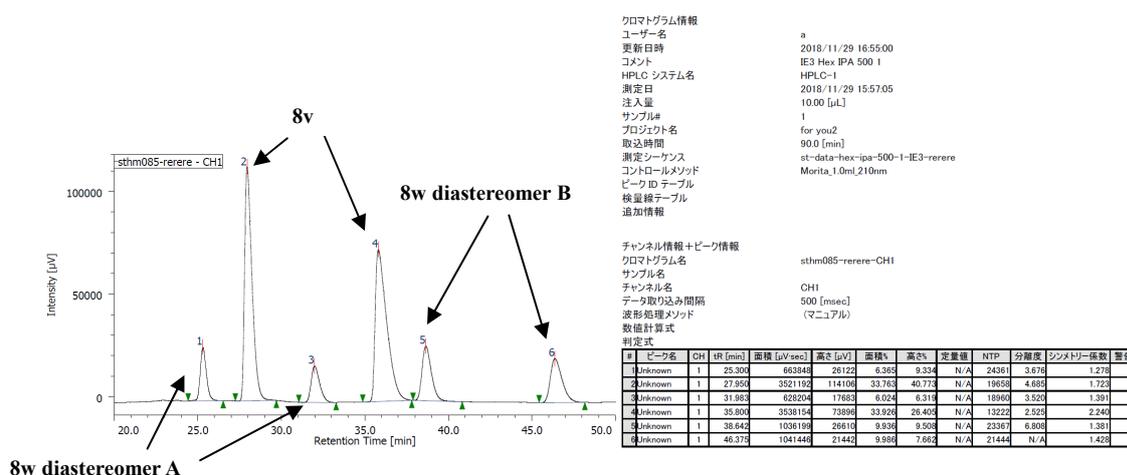


(R)-8v

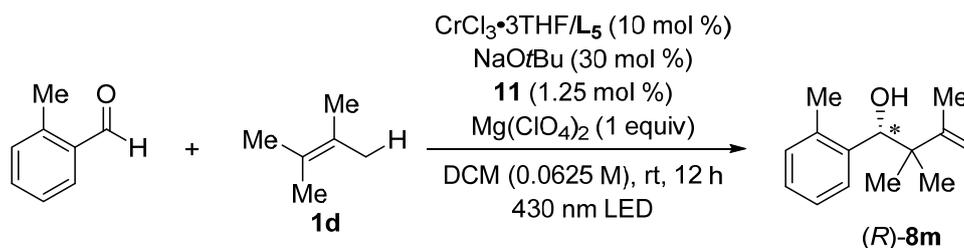
Prepared according to the general procedure, then the reaction mixture was passed through a pad of silica gel with CH₂Cl₂. After evaporation, regioisomeric ratio was determined by NMR analysis (1.9/1 rr). The residue was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (R)-8v

as colorless oil along with the inseparable mixture of stereoisomers (**8w**) (42.8 mg, 97%). All the spectroscopic data matches with the previously reported data.¹¹ The following spectroscopic data were collected with a pure sample obtained by preparative TLC.

¹H NMR (CDCl₃, 500 MHz) δ 0.88 (s, 3H), 0.93 (s, 3H), 2.01 (s, 1H), 4.33 (s, 1H), 4.99 (dd, *J* = 1.4, 17.9 Hz, 1H), 5.05 (dd, *J* = 1.4, 11.0 Hz, 1H), 5.83 (dd, *J* = 11.0, 17.9 Hz, 1H), 7.12-7.26 (m, 5H); ¹³C NMR (CDCl₃, 125 MHz) δ 20.1, 24.4, 42.2, 80.6, 113.8, 127.4, 127.4, 127.7, 140.7, 145.0; HPLC (chiral column: CHIRALPAK IE3; solvent: hexane/2-propanol = 500/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 27.3 min (minor) and 34.2 min (major); [α]_D²⁰ = +111.5 (*c* = 1.23, CHCl₃, for 83% ee sample).



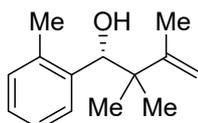
Procedure for preparation of **8m** and **8p**



CrCl₃·3THF (9.4 mg, 0.025 mmol, 10 mol %), NaOtBu (7.2 mg, 0.075 mmol, 30 mol %) and **L5** (9.0 mg, 0.025 mmol, 10 mol %) were dissolved in degassed CH₂Cl₂ (4 mL) in a screw-capped vial under argon

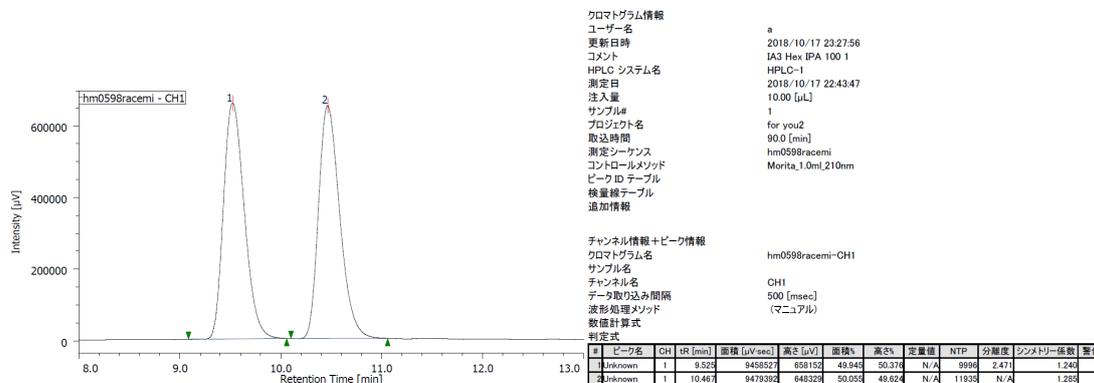
atmosphere and the mixture was stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1d** (540 μL , 5.0 mmol, 20 eq), $\text{Mg}(\text{ClO}_4)_2$ (55.8 mg, 0.25 mmol, 100 mol %), *o*-tolaldehyde (29 μL , 0.25 mmol, 1 eq) and photocatalyst **11** (1.3 mg, 0.03125 mmol, 1.25 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 $^\circ\text{C}$). Then, the reaction mixture was directly loaded to silica gel and purified by flash column chromatography ($\text{CH}_2\text{Cl}_2/\text{hexane} = 1/10$ to $1/0$, v/v) to afford (*R*)-**8m** as colorless oil (51.1 mg, 50%).

(R)-2,2,3-trimethyl-1-(*o*-tolyl)but-3-en-1-ol (8m).

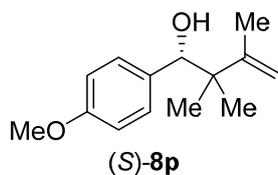


(*R*)-**8m**

IR (neat) ν 3466, 3090, 2972, 1633, 1607, 1463, 1377, 1150, 1049, 892, 790, 734 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 0.93 (s, 3H), 1.10 (s, 3H), 1.90 (s, 3H), 2.00 (d, $J = 1.7$ Hz, 1H), 2.38 (s, 3H), 4.90–5.10 (m, 2H), 5.05 (s, 1H), 7.10–7.25 (m, 3H), 7.50 (dd, $J = 1.7, 7.4$ Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 20.1, 20.4, 20.5, 23.8, 45.8, 72.2, 113.3, 125.3, 127.1, 128.3, 130.1, 136.1, 138.6, 150.9; LRMS (ESI): m/z 227 [$\text{M}+\text{Na}^+$]; HRMS (ESI): m/z calculated for $\text{C}_{14}\text{H}_{20}\text{NaO}^+$ [$\text{M}+\text{Na}^+$]: 227.1406, found: 227.1395; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 100/1; flow rate: 1.0 mL/min; detection: at 210 nm): $t_R = 9.7$ min (major) and 10.7 min (minor); $[\alpha]_D^{20} = +110$ ($c = 1.05$, CHCl_3 , for 96% ee sample).

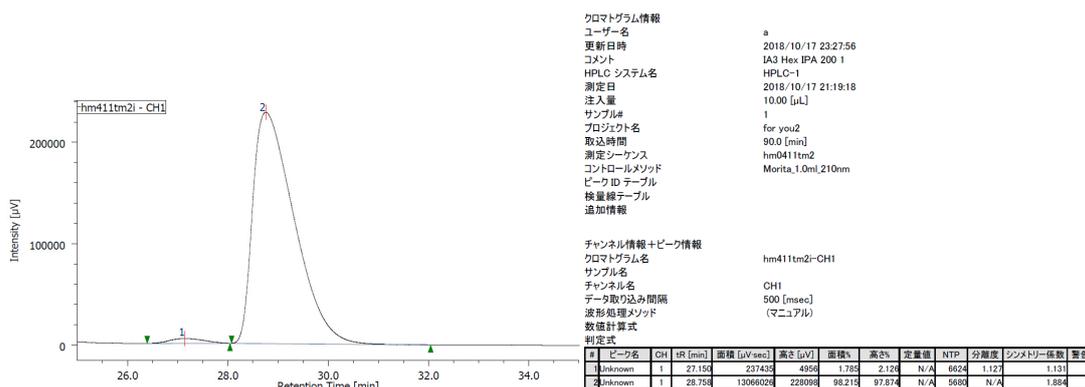
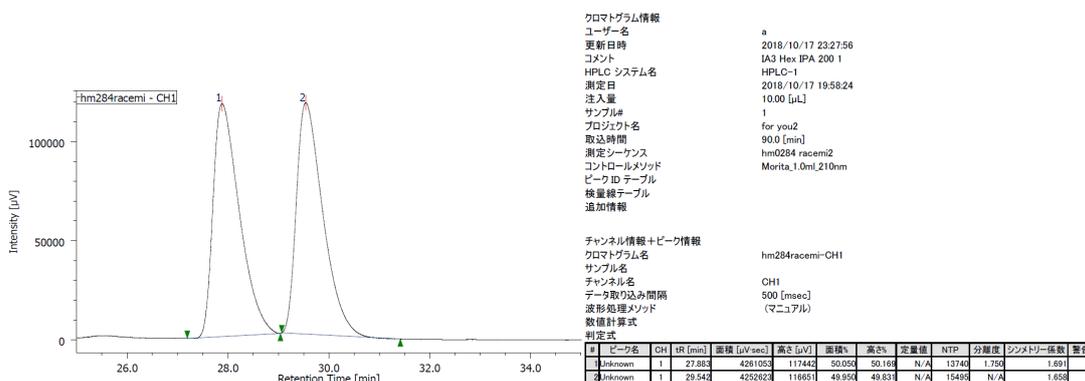


(S)-1-(4-methoxyphenyl)-2,2,3-trimethylbut-3-en-1-ol (8p).

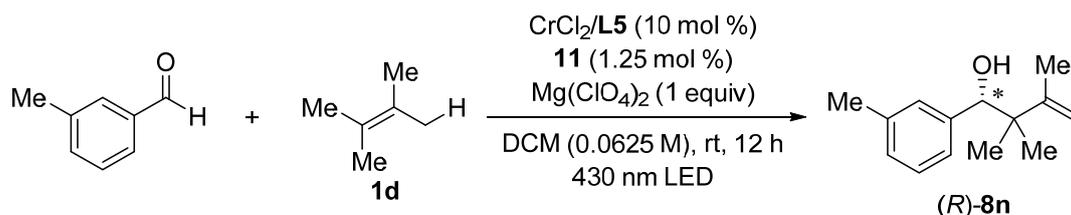


Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (AcOEt/hexane = 1/10 to 1/1, v/v) to afford (S)-8p as colorless oil (21.5 mg, 39%). All the spectroscopic data matches with the previously reported data.¹⁰

¹H NMR (CDCl₃, 500 MHz) δ 0.92 (s, 3H), 0.98 (s, 3H), 1.86 (s, 3H), 2.03 (d, *J* = 1.7 Hz, 1H), 3.81 (s, 3H), 4.59 (s, 1H), 4.96 (s, 1H), 5.02 (s, 1H), 6.83-6.88 (m, 2H), 7.23-7.27 (m, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 19.8, 20.0, 24.2, 44.6, 55.2, 77.1, 112.8, 113.1, 129.0, 132.3, 150.7, 158.8; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 27.2 min (minor) and 28.8 min (major); [α]_D²⁰ = +138 (*c* = 0.49, CHCl₃, for 96% ee sample).

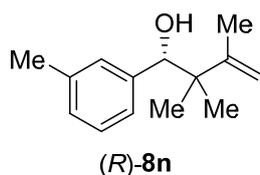


Procedure for preparation of 8n and 8o

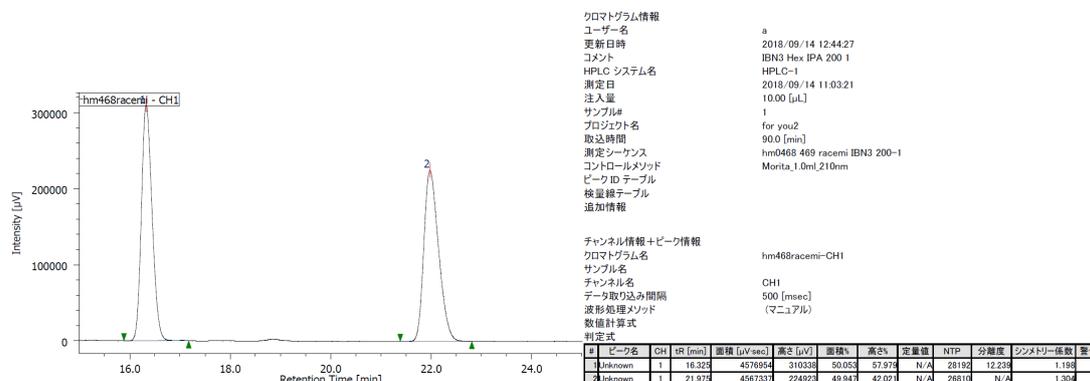


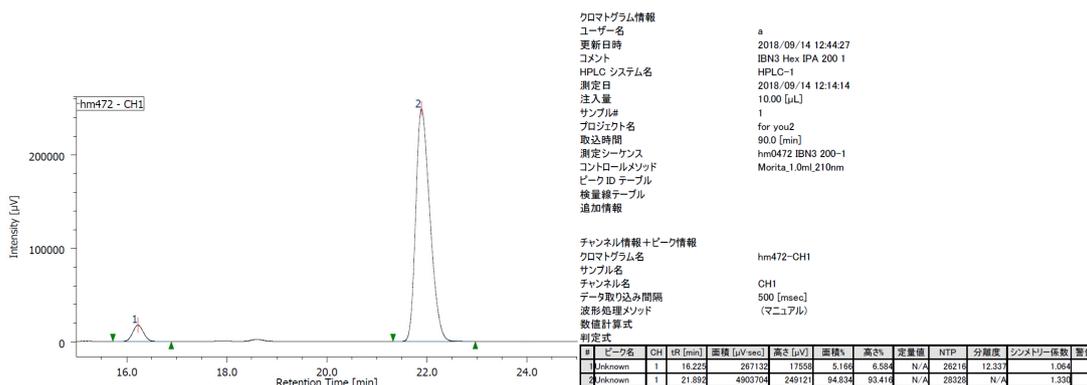
CrCl_2 (3.0 mg, 0.025 mmol, 10 mol %) and **L5** (9.0 mg, 0.025 mmol, 10 mol %) were dissolved in degassed CH_2Cl_2 (4 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1d** (148 μL , 1.25 mmol, 5 eq), $\text{Mg}(\text{ClO}_4)_2$ (55.8 mg, 0.25 mmol, 100 mol %), *m*-tolaldehyde (29 μL , 0.25 mmol, 1 eq) and photocatalyst **11** (1.3 mg, 0.003125 mmol, 1.25 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was directly loaded to silica gel and purified by flash column chromatography (CH_2Cl_2 /hexane = 1/10 to 1/0, v/v) to afford (*S*)-**8n** as colorless oil (43.2 mg, 85%).

(S)-2,2,3-trimethyl-1-(*m*-tolyl)but-3-en-1-ol (8n).

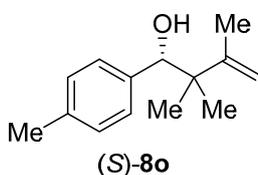


IR (neat) ν 3468, 2974, 1715, 1632, 1461, 1378, 1181, 1039, 1007, 893, 764, 738 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 0.94 (s, 3H), 0.99 (s, 3H), 1.87 (s, 3H), 2.05 (s, 1H), 2.36 (s, 3H), 4.60 (s, 1H), 4.97 (d, J = 1.1 Hz, 1H), 5.03 (d, J = 1.1 Hz, 1H), 7.08 (d, J = 7.4 Hz, 1H), 7.12 (d, J = 7.4 Hz, 1H), 7.16 (s, 1H), 7.20 (dd, J = 7.4, 7.4 Hz, 1H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 19.8, 20.0, 21.5, 24.3, 44.4, 77.4, 113.1, 125.1, 127.2, 128.0, 128.6, 136.8, 140.2, 150.7; LRMS (ESI): m/z 227 [$\text{M}+\text{Na}^+$]; HRMS (ESI): m/z calculated for $\text{C}_{14}\text{H}_{20}\text{NaO}^+$ [$\text{M}+\text{Na}^+$]: 227.1406, found: 227.1399; HPLC (chiral column: CHIRALPAK IBN3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): t_R = 16.2 min (minor) and 21.9 min (major); $[\alpha]_D^{20}$ = +103 (c = 0.9, CHCl_3 , for 90% ee sample).



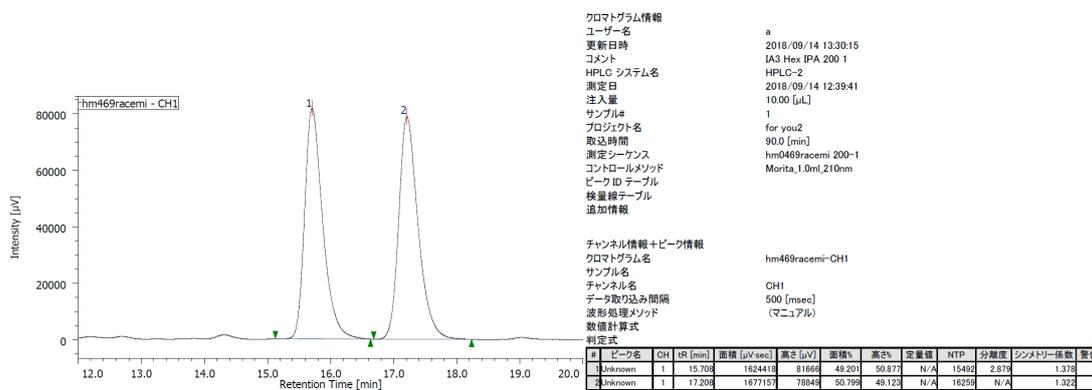


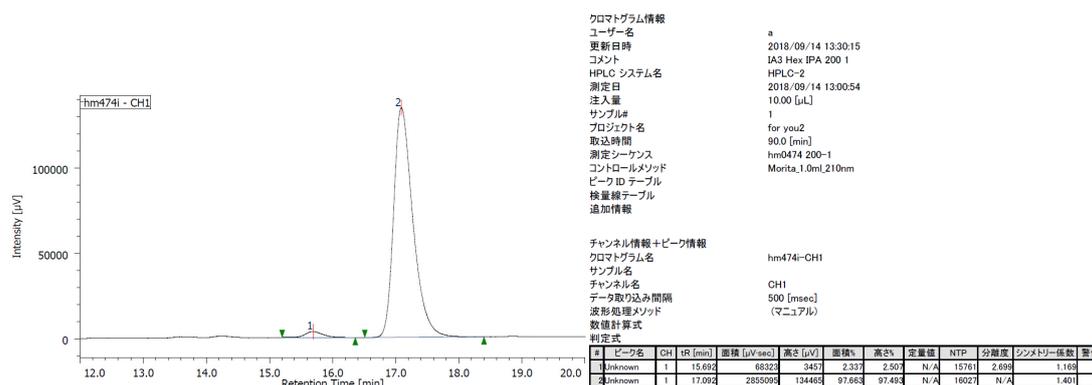
(S)-2,2,3-trimethyl-1-(p-tolyl)but-3-en-1-ol (8o).



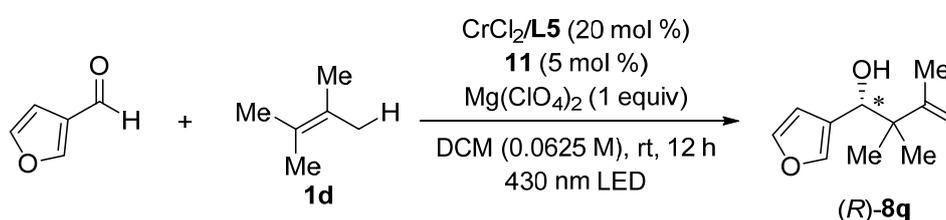
Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (S)-8o as colorless oil (49.4 mg, 97%). All the spectroscopic data matches with the previously reported data.¹²

¹H NMR (CDCl₃, 500 MHz) δ 0.93 (s, 3H), 0.98 (s, 3H), 1.86 (s, 3H), 2.03 (d, *J* = 1.7 Hz, 1H), 2.34 (s, 3H), 4.60 (s, 1H), 4.96 (d, *J* = 1.1 Hz, 1H), 5.02 (d, *J* = 1.1 Hz, 1H), 7.12 (d, *J* = 8.0 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 19.8, 20.1, 21.1, 24.2, 44.4, 77.3, 113.0, 127.8, 128.1, 136.8, 137.3, 150.7; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 15.7 min (minor) and 17.1 min (major); [α]_D²⁰ = +134 (*c* = 1.76, CHCl₃, for 95% ee sample).



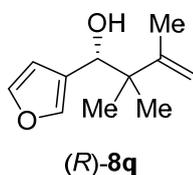


Procedure for preparation of **8q**

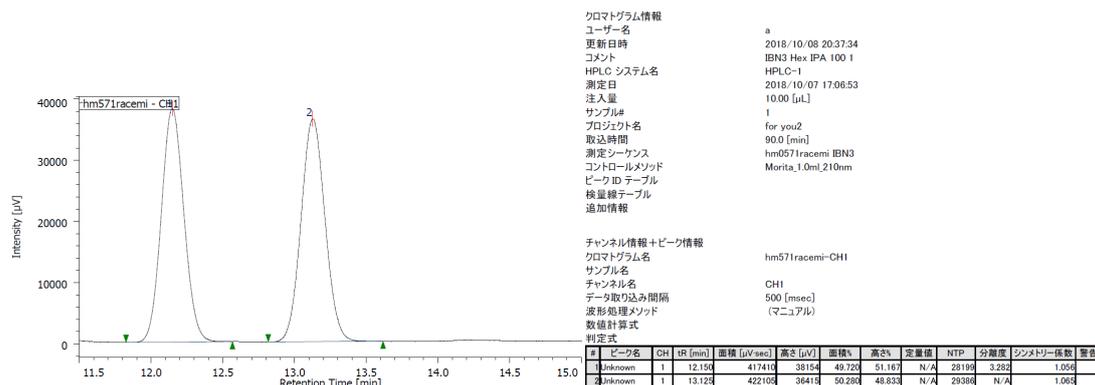


CrCl_2 (6.1 mg, 0.05 mmol, 20 mol %) and **L5** (17.9 mg, 0.05 mmol, 20 mol %) were dissolved in degassed CH_2Cl_2 (4 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1d** (593 μL , 5.0 mmol, 20 eq), $\text{Mg}(\text{ClO}_4)_2$ (55.8 mg, 0.25 mmol, 100 mol %), 3-furaldehyde (21 μL , 0.25 mmol, 1 eq) and photocatalyst **11** (5.2 mg, 0.00125 mmol, 5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was directly loaded to silica gel and purified by flash column chromatography ($\text{CH}_2\text{Cl}_2/\text{hexane} = 1/10$ to $1/0$, v/v) to afford **(R)-8q** as colorless oil (14.8 mg, 33%).

(R)-1-(furan-3-yl)-2,2,3-trimethylbut-3-en-1-ol (8q).

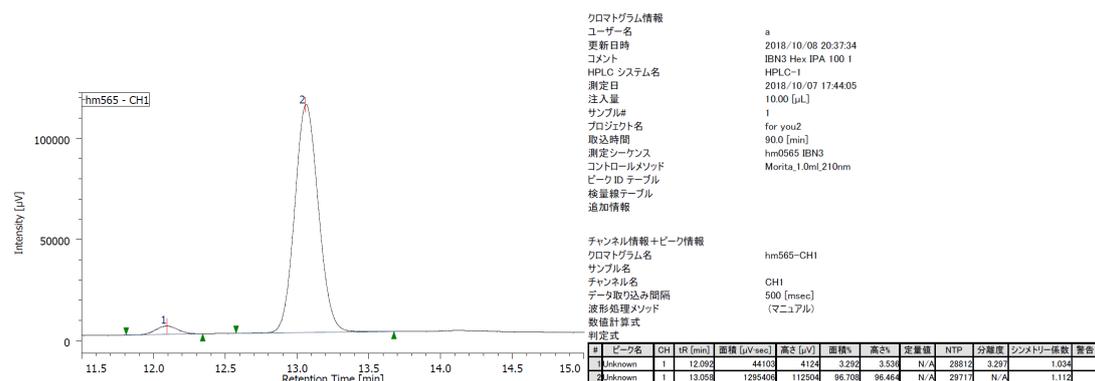


IR (neat) ν 3464, 3091, 2971, 1716, 1635, 1501, 1460, 1377, 1261, 1163, 1023, 875, 798, 736 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 0.98 (s, 3H), 1.05 (s, 3H), 1.83 (s, 3H), 1.93 (s, 1H), 4.61 (s, 1H), 4.95 (s, 1H), 5.01 (s, 1H), 6.39 (s, 1H), 7.35–7.39 (m, 2H); ^{13}C NMR (CDCl_3 , 125 MHz) δ 19.8, 20.5, 23.7, 44.2, 71.3, 110.3, 113.1, 124.9, 140.3, 142.3, 150.3; LRMS (ESI): m/z 203 [$\text{M}+\text{Na}^+$]; HRMS (ESI): m/z calculated for $\text{C}_{11}\text{H}_{16}\text{NaO}_2^+$ [$\text{M}+\text{Na}^+$]: 203.1043, found: 203.1040; HPLC (chiral column: CHIRALPAK IBN3; solvent: hexane/2-propanol = 100/1; flow rate: 1.0 mL/min; detection: at 210 nm): $t_R = 12.1$ min (major) and 13.1 min (minor); $[\alpha]_D^{20} = +56$ ($c = 0.54$, CHCl_3 , for 93% ee sample).



クロマトグラム情報
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 更新日時 2018/10/08 20:37:34
 コメント IBN3 Hex IPA 100 1
 HPLC システム名 HPLC-1
 測定日 2018/10/07 17:06:53
 注入量 10.00 [μL]
 サンプル# 1
 プロジェクト名 for you2
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 コントロールメソッド Morita_1.0ml_210nm
 ピークID テーブル
 検量線テーブル
 追加情報

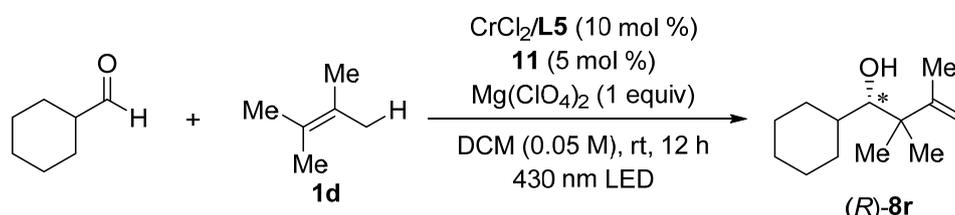
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 サンプル名
 チャンネル名 CH1
 データ取り込み間隔 500 [msec]
 波形処理メソッド (マニュアル)
 数値計算式



クロマトグラム情報
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 追加情報

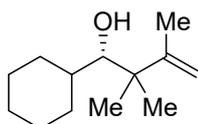
チャンネル情報+ピーク情報
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 数値計算式

Procedure for preparation of **8r**



CrCl_2 (2.4 mg, 0.02 mmol, 20 mol %) and **L5** (7.2 mg, 0.02 mmol, 20 mol %) were dissolved in degassed 1,2-dichloroethane (4 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1a** (119 μL , 1.0 mmol, 5 eq), $\text{Mg}(\text{ClO}_4)_2$ (44.6 mg, 0.2 mmol, 100 mol %), cyclohexanecarbaldehyde (24 μL , 0.2 mmol, 1 eq) and photocatalyst **11** (4.1 mg, 0.01 mmol, 5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was directly loaded to silica gel and purified by flash column chromatography (CH_2Cl_2 /hexane = 1/10 to 1/0, v/v) to afford (*S*)-**8r** as colorless oil (35.5 mg, 90%).

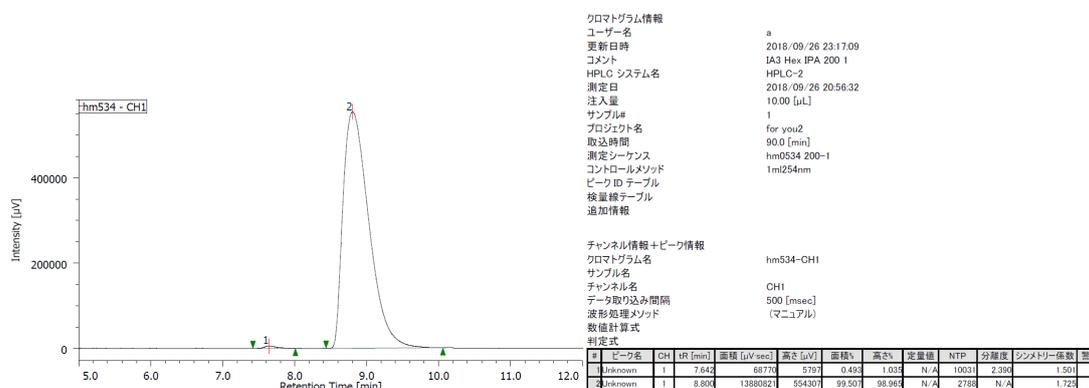
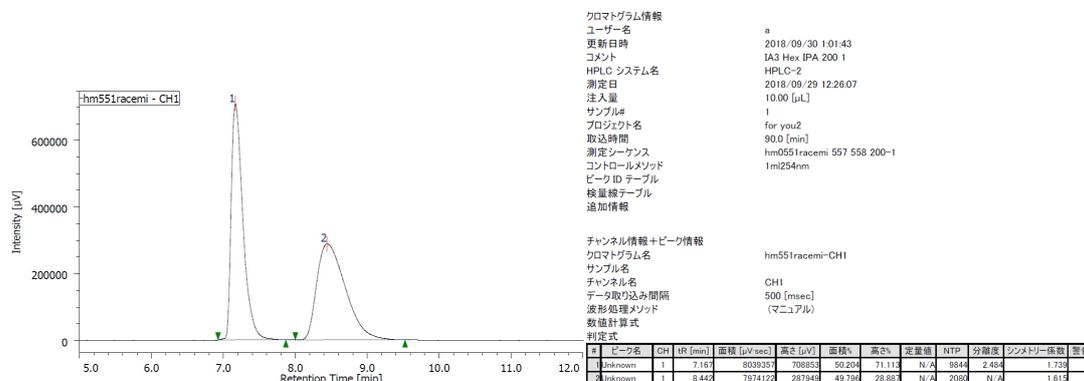
(S)-1-cyclohexyl-2,2,3-trimethylbut-3-en-1-ol (8r).



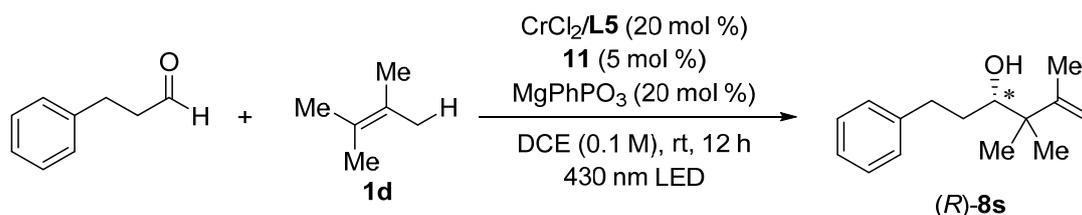
(S)-**8r**

All the spectroscopic data matches with the previously reported data.¹⁰ Enantiomeric excess was determined after protection of **8r** by *p*-NO₂-benzoyl chloride.

¹H NMR (CDCl₃, 500 MHz) δ 1.06 (s, 3H), 1.08 (s, 3H), 1.19-1.37 (m, 6H), 1.43 (d, *J* = 4.6 Hz, 1H), 1.45-1.66 (m, 2H), 1.68-1.80 (m, 6H), 3.28 (dd, *J* = 2.9, 4.0 Hz, 1H), 4.83 (s, 1H), 4.87 (s, 1H); ¹³C NMR (CDCl₃, 125 MHz) δ 20.0, 22.9, 23.2, 26.4, 26.4, 26.9, 28.0, 34.0, 39.1, 44.6, 79.5, 111.7, 151.7; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 254 nm): *t*_R = 7.6 min (minor) and 8.8 min (major); [α]_D²⁰ = -49 (*c* = 1.19, CHCl₃, for 99% ee sample).



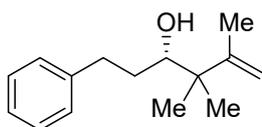
Procedure for preparation of **8s-8u**



CrCl₂ (6.1 mg, 0.05 mmol, 20 mol %) and **L5** (17.9 mg, 0.05 mmol, 20 mol %) were dissolved in degassed

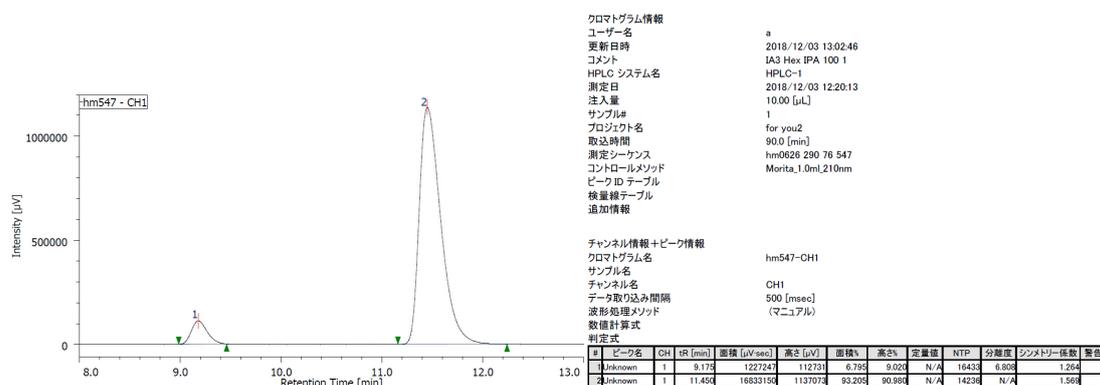
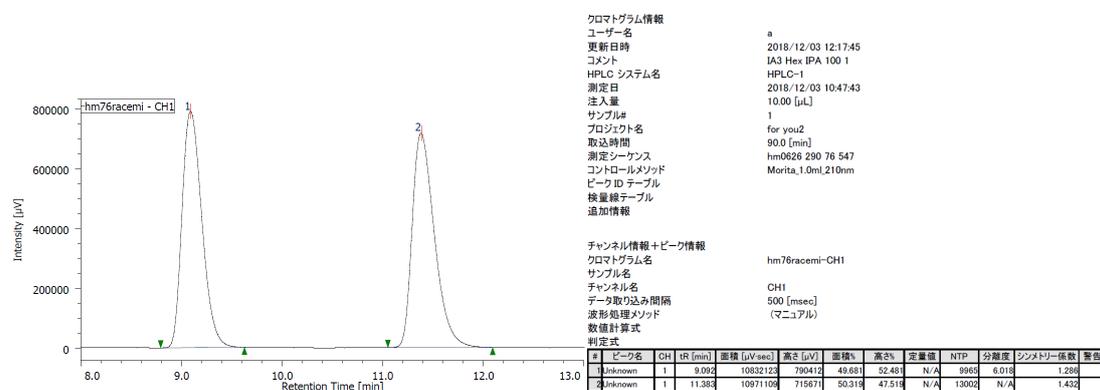
1,2-dichloroethane (2.5 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1d** (593 μ L, 5.0 mmol, 20 eq), MgPhPO₃ (9.0 mg, 0.05 mmol, 20 mol %), hydrocinnamaldehyde (33 μ L, 0.25 mmol, 1 eq) and photocatalyst **11** (5.2 mg, 0.0125 mmol, 5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 48 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was directly loaded to silica gel and purified by flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (*S*)-**8s** as colorless oil (33.8 mg, 69%).

(*S*)-4,4,5-trimethyl-1-phenylhex-5-en-3-ol (8s).

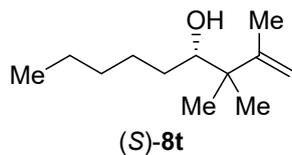


(*S*)-8s

IR (neat) ν 3479, 3087, 3027, 2970, 1634, 1603, 1496, 1455, 1377, 1302, 1067, 894, 748, 699 cm^{-1} ; ¹H NMR (CDCl₃, 500 MHz) δ 0.99 (s, 3H), 1.04 (s, 3H), 1.50-1.81 (m, 6H), 2.55-2.66 (m, 1H), 2.88-2.99 (m, 1H), 3.50 (d, *J* = 10.3 Hz, 1H), 4.84 (s, 1H), 4.90 (s, 1H), 7.15-7.32 (m, 5H); ¹³C NMR (CDCl₃, 125 MHz) δ 19.6, 21.5, 22.4, 33.0, 33.6, 43.7, 74.8, 112.1, 125.7, 128.3, 128.5, 142.5, 150.8; LRMS (DART): *m/z* 219 [M+H⁺]; HRMS (DART): *m/z* calculated for C₁₅H₂₃O⁺ [M+H⁺]: 219.1743, found: 219.1753; HPLC (chiral column: CHIRALPAK IA3; solvent: hexane/2-propanol = 100/1; flow rate: 1.0 mL/min; detection: at 210 nm): *t*_R = 9.2 min (minor) and 10.5 min (major); [α]_D²⁰ = +98 (*c* = 1.38, CHCl₃, for 86% ee sample).

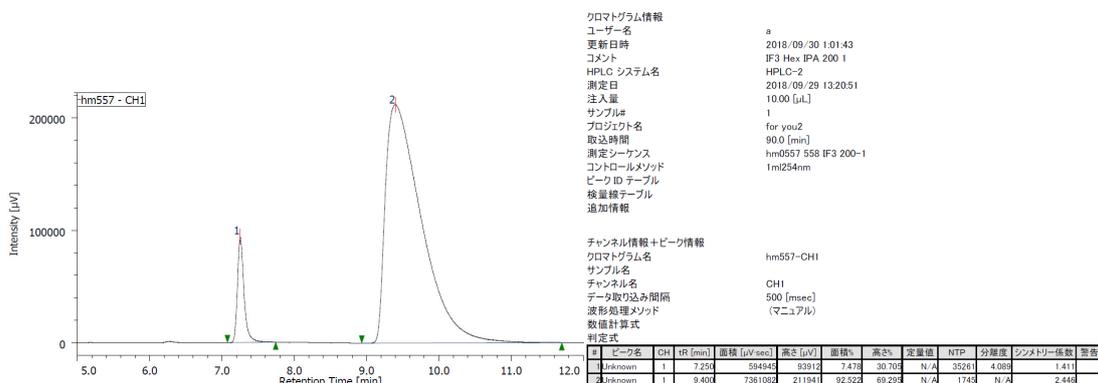
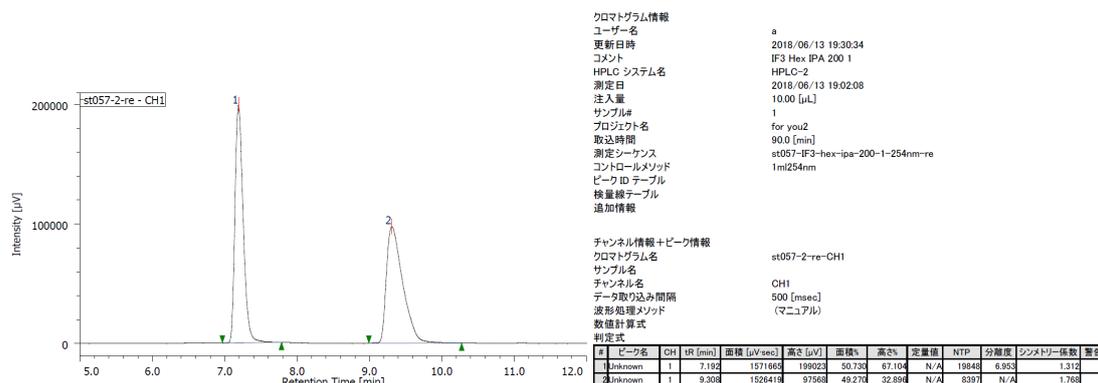


(S)-2,3,3-trimethylnon-1-en-4-ol (8t).

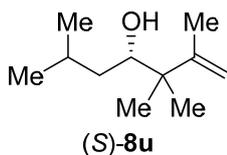


Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (S)-8t as colorless oil (43.6 mg, 95%). Enantiomeric excess was determined after protection of 8t by *p*-NO₂-benzoyl chloride.

IR (neat) ν 3446, 2928, 2856, 1732, 1635, 1457, 1377, 1289, 1120, 1073, 934, 892 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 0.89 (t, *J* = 6.9 Hz, 3H), 1.00 (s, 3H), 1.04 (s, 3H), 1.18-1.37 (m, 6H), 1.38-1.47 (m, 1H), 1.47-1.63 (m, 2H), 1.75 (d, *J* = 1.1 Hz, 3H), 3.46 (d, *J* = 10.3 Hz, 1H), 4.84 (s, 1H), 4.91 (s, 1H); ¹³C NMR (CDCl₃, 125 MHz) δ 14.1, 19.7, 21.4, 22.6, 22.7, 27.1, 31.0, 32.0, 43.7, 75.5, 111.9, 151.1; LRMS (DART): *m/z* 633 [M+H⁺]; HRMS (DART): *m/z* calculated for C₁₂H₂₅O⁺ [M+H⁺]: 185.1900, found: 185.1894; HPLC (chiral column: CHIRALPAK IF3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 254 nm): *t*_R = 7.3 min (minor) and 9.4 min (major); [α]_D²⁰ = +70 (*c* = 0.26, CHCl₃, for 85% ee sample).

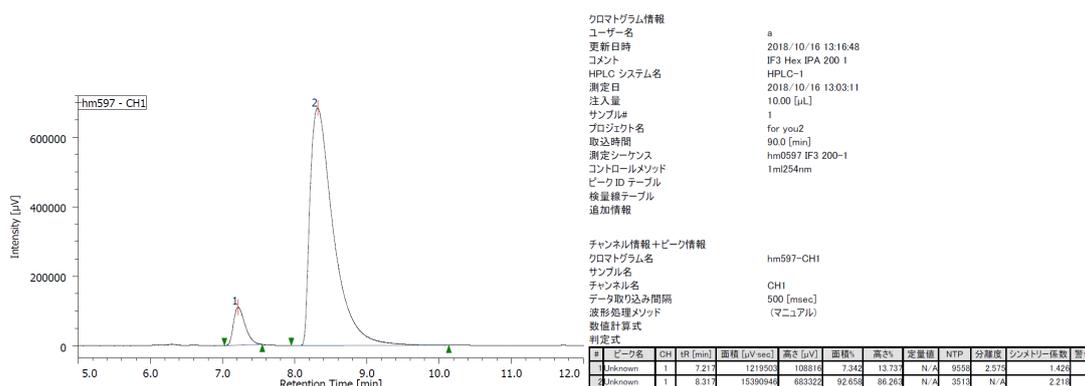
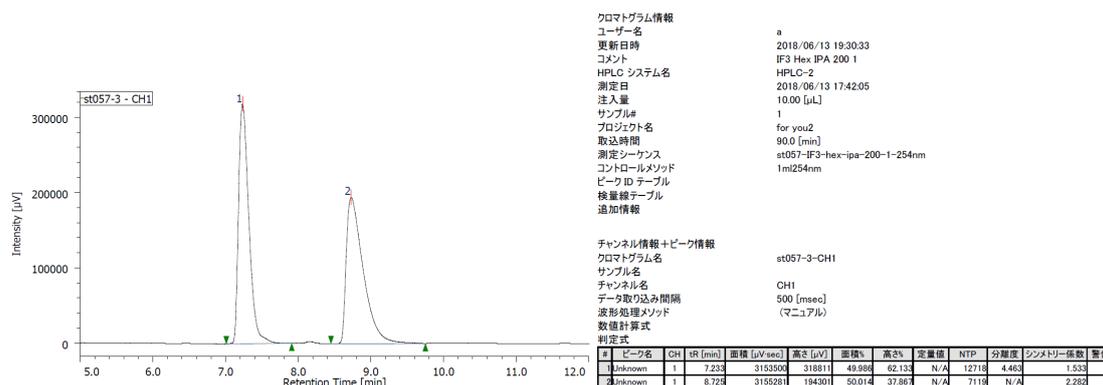


(S)-2,3,3,6-tetramethylhept-1-en-4-ol (8u).



Prepared according to the general procedure, then the crude material was purified by silica gel flash column chromatography (CH₂Cl₂/hexane = 1/10 to 1/0, v/v) to afford (*R*)-**8u** as colorless oil (33.3 mg, 78%). Enantiomeric excess was determined after protection of **8u** by *p*-NO₂-benzoylchloride.

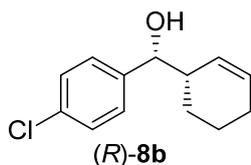
IR (neat) ν 3408, 3091, 2957, 2970, 1635, 1468, 1377, 1296, 1069, 983, 893 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 0.90 (t, *J* = 6.3 Hz, 3H), 0.94 (t, *J* = 6.9 Hz, 3H), 0.99 (s, 3H), 1.04 (s, 3H), 1.10-1.18 (m, 1H), 1.22-1.32 (m, 1H), 1.46 (d, *J* = 2.2 Hz, 1H), 1.73-1.85 (m, 4H), 3.56 (dd, *J* = 2.2, 10.3 Hz, 1H), 4.84 (s, 1H), 4.90 (s, 1H); ¹³C NMR (CDCl₃, 125 MHz) δ 14.1, 19.7, 21.4, 22.6, 22.7, 27.1, 31.0, 32.0, 43.7, 75.5, 111.9, 151.1; LRMS (ESI): *m/z* 193 [M+Na⁺]; HRMS (ESI): *m/z* calculated for C₁₁H₂₂NaO⁺ [M+Na⁺]: 193.1563, found: 193.1570; HPLC (chiral column: CHIRALPAK IF3; solvent: hexane/2-propanol = 200/1; flow rate: 1.0 mL/min; detection: at 254 nm): *t*_R = 7.2 min (minor) and 8.3 min (major); [α]_D²⁰ = +116 (*c* = 0.84, CHCl₃, for 85% ee sample).



Determination of absolute configuration

The absolute configurations of **8** were determined as follows:

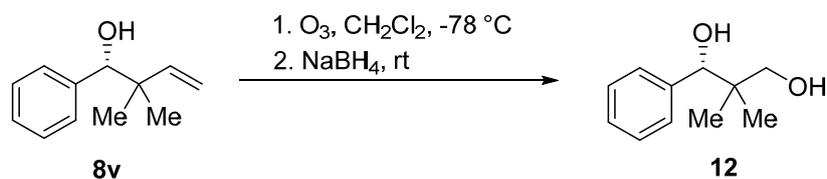
8a-8k: The absolute configuration was determined by optical rotation value of **8b**. Those of others were assigned by analogy.



This time: $[\alpha]_D^{20} = -24.6$ ($c = 1.0$, C_6H_6 , for 99% ee sample)

Reported value¹³: $[\alpha]_D^{20} = -15.0$ ($c = 1.0$, C_6H_6 , for 64% ee sample)

8l-8v: Following the following reaction scheme, diol **12** was obtained. The absolute configuration was determined by optical rotation value of diol **12**. Those of others were assigned by analogy.

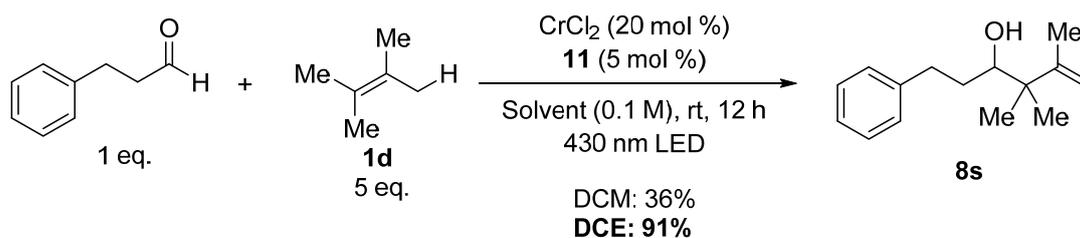


8v (38 mg, 0.216 mmol, 1 eq, 83% ee) were dissolved in CH_2Cl_2 (2 mL) in a round bottom flask. Then the flask was connected to an oxygen tank with a bubbler. The reaction mixture was cooled to $-78\text{ }^\circ\text{C}$. The ozone generator was switched on and the ozone went into the reaction flask. The reaction mixture was allowed to stir until the color changed to blue. $NaBH_4$ (16.8 mg, 0.431 mmol, 2 eq) and MeOH (1 ml) were added and the mixture was warmed up to room temperature. The reaction mixture was stirred for 15 h, and then quenched with saturated *aq.* NH_4Cl . The organic layer was separated, and the aqueous layer was further extracted three times with AcOEt. The combined organic layer was washed with water and brine and dried over Na_2SO_4 . After the evaporation under reduced pressure, the residue was purified by silica gel flash column chromatography (hexane/AcOEt = 3/1 to 1/1, v/v) to give the desired diol **9** (30.2 mg, 78% yield) as a colorless oil. $[\alpha]_D^{20} = +65.9$ ($c = 2.5$, $CHCl_3$, for 83% ee sample)

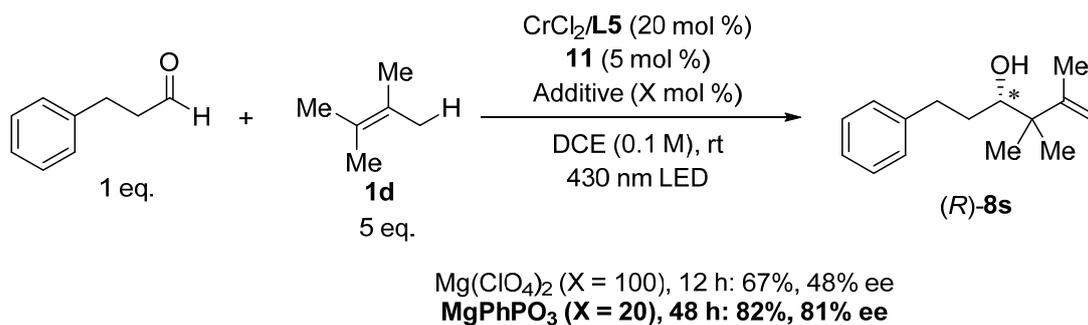
Reported optical rotation value¹⁴: $[\alpha]_D^{30} = +44.7$ ($c = 1.0$, $CHCl_3$, for 99% ee sample)

4. Optimization Study for Aliphatic Aldehydes

During the optimization of racemic reaction, DCE solvent was found to be better solvent than DCM in terms of reactivity.



Addition of $MgPhPO_3$ was effective for improving enantioselectivity, while the reaction rate was slower than in the presence of $Mg(ClO_4)_2$. 48 hours was required for the completion of the reaction.



5. Mechanistic Study

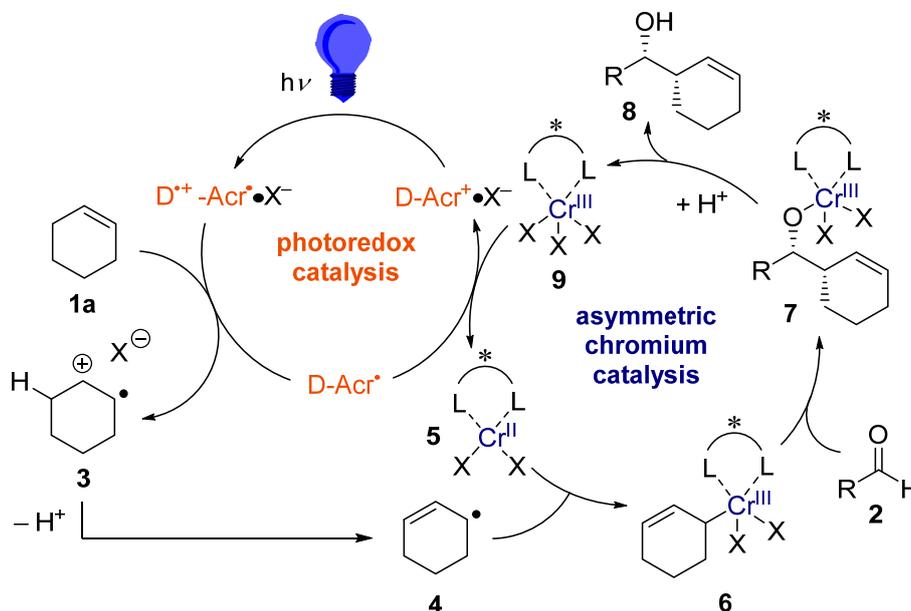
5.1 Redox potentials of intermediates in catalytic cycle

We assumed that the proposed catalytic cycle (Figure 2 in the text) is feasible based on the following redox potentials of intermediates. Although oxidation potential of cyclohexene is higher than that of excited state of photocatalyst **10**, one electron transfer would be possible based on the related precedent (cyclopentene and photocatalyst **11**).¹⁵ In addition, addition of Mg(ClO₄)₂ might facilitate this step by stabilizing ion pair **3** (see ref 20 in the text).

Cyclohexene **1a**: [**1a**/**3**] = +2.37 V versus SCE¹⁶

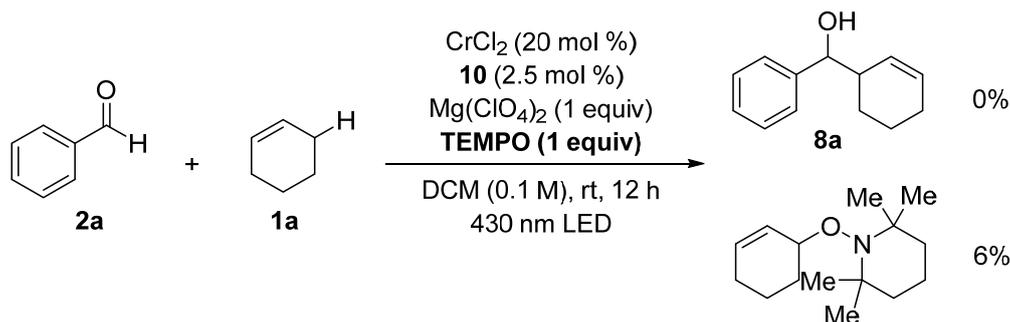
Photocatalyst **10**: [D-Acr[•]/D⁺-Acr⁺] = +2.24 V versus SCE, [D-Acr⁺/D-Acr[•]] = -0.46 V versus SCE¹⁷

Cr catalyst: [Cr(III)/Cr(II)] = -0.41 V versus SCE¹⁸



5.2 TEMPO trapping experiment

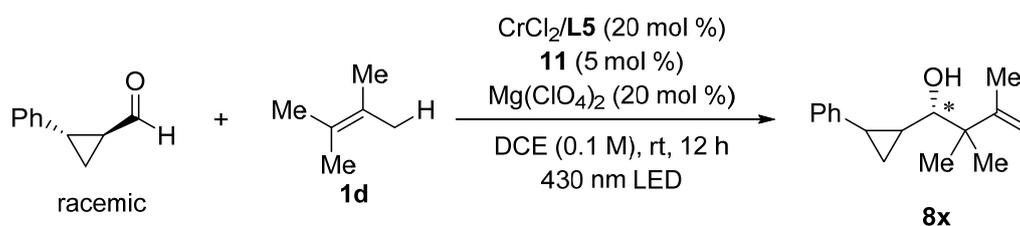
To confirm the generation of an allyl radical, a radical trapping experiment by the addition of TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy, free radical) was conducted.



CrCl_2 (6.1 mg, 0.05 mmol, 20 mol %), cyclohexene **1a** (507 μL , 5.0 mmol, 20 eq), benzaldehyde **2a** (25.5 μL , 0.25 mmol, 1 eq), $\text{Mg}(\text{ClO}_4)_2$ (55.8 mg, 0.25 mmol, 100 mol %) and photocatalyst **10** (2.5 mg, 0.00625 mmol, 2.5 mol %) were dissolved in degassed CH_2Cl_2 (2.5 mL) in a screw-capped vial under argon atmosphere at room temperature. Then, TEMPO (39.1 mg, 0.25 mmol, 1 eq) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 $^\circ\text{C}$). Then, the reaction mixture was passed through a pad of silica gel with CH_2Cl_2 . After evaporation, the crude mixture was analyzed by NMR spectroscopy using 1,1,2,2-tetrachloroethane as internal standard. No desired product **8a** was formed, and TEMPO adduct¹⁹ was detected in 6% NMR yield. Therefore, we assume allyl radical is formed as an intermediate of the reaction.

5.3 Radical clock experiment

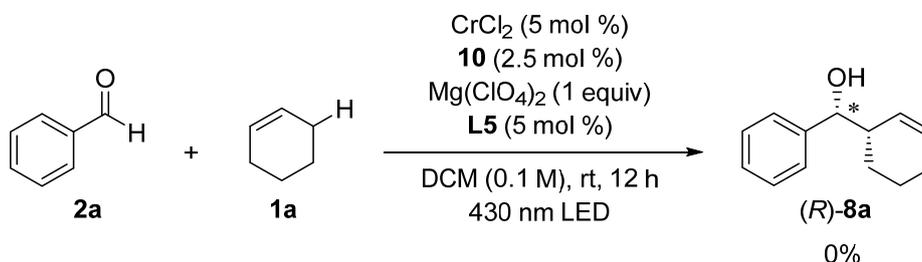
To deny the involvement of ketyl radicals, a radical clock experiment was performed using 2-phenylcyclopropylcarbaldehyde as electrophile.



CrCl_2 (6.1 mg, 0.05 mmol, 20 mol %) and **L5** (17.9 mg, 0.05 mmol, 20 mol %) were dissolved in degassed 1,2-dichloroethane (2.5 mL) in a screw-capped vial under argon atmosphere and stirred for 1 hour at room temperature. 2,3-Dimethyl-2-butene **1d** (119 μL , 1.0 mmol, 5 eq), $\text{Mg}(\text{ClO}_4)_2$ (11.2 mg, 0.05 mmol, 20 mol %), 2-phenylcyclopropylcarbaldehyde²⁰ (33 μL , 0.25 mmol, 1 eq) and photocatalyst **11** (5.2 mg, 0.0125 mmol, 5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 $^\circ\text{C}$). Then, the reaction mixture was passed through a pad of silica gel with a CH_2Cl_2 eluent. After evaporation, the crude mixture was analyzed by NMR spectroscopy using 1,1,2,2-tetrachloroethane as internal standard. Desired product **8x** was formed (77%, 1.1/1 dr), and no ring opened product was observed. Therefore, we assume that ketyl radicals are not formed in the reaction.

5.4 Alternative pathway of chirality induction

We studied a feasibility of an alternative asymmetric induction mechanism involving in situ-generated $\text{Mg}(\text{ClO}_4)_2/\mathbf{L5}$ as chiral Lewis acid by premixing $\text{Mg}(\text{ClO}_4)_2$ and $\mathbf{L5}$. However, the desired reaction did not proceed at all, indicating that $\mathbf{L5}$ remains coordinating to the chromium atom.



$\text{Mg}(\text{ClO}_4)_2$ (55.8 mg, 0.25 mmol, 100 mol %) and $\mathbf{L5}$ (4.5 mg, 0.0125 mmol, 5 mol %) were dissolved in degassed CH_2Cl_2 (2.5 mL) in a screw-capped vial under argon atmosphere and the mixture was stirred for 1 hour at room temperature. Cyclohexene **1a** (507 μL , 5.0 mmol, 20 eq), CrCl_2 (1.5 mg, 0.0125 mmol, 5 mol %), benzaldehyde **2a** (25.5 μL , 0.25 mmol, 1 eq) and photocatalyst **10** (2.5 mg, 0.00625 mmol, 2.5 mol %) were added to the reaction mixture. The vial was subjected to 430 nm LED irradiation for 12 hours under temperature control (ca. 27–29 °C). Then, the reaction mixture was passed through a pad of silica gel with CH_2Cl_2 . After evaporation, the crude mixture was analyzed by NMR. However, no desired product **8a** was observed. Therefore, we assume that the asymmetric induction is not due to $\text{Mg}(\text{ClO}_4)_2/\mathbf{L5}$, but due to $\text{CrCl}_2/\mathbf{L5}$.

5.5 Transient absorption analysis

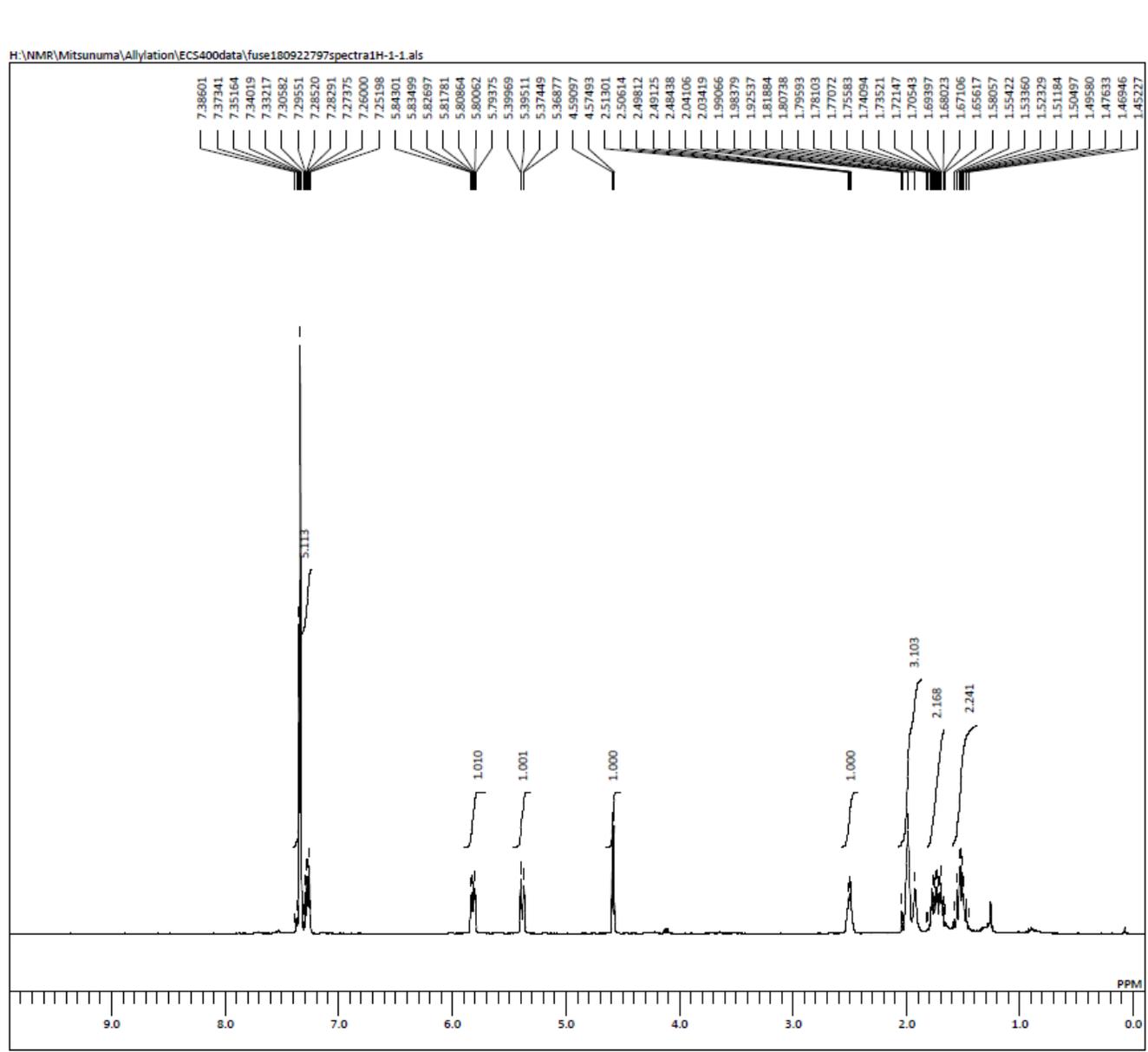
Nanosecond time-resolved transient absorption measurements were carried out using the laser system provided by UNISOKU Co., Ltd. Measurements of nanosecond transient absorption spectrum were performed according to the following procedure. A deaerated solution containing a sample in a quartz cell (1 cm \times 1 cm) was excited by a Nd:YAG laser (Continuum SLII-10, 4-6 ns fwhm, $\lambda_{\text{ex}} = 355$ nm, 5mJ/pulse). The photodynamics were monitored by continuous exposure to a xenon lamp (150 W) as a probe light and a photomultiplier tube (Hamamatsu 2949) as a detector. The solution in a quartz cuvette was deoxygenated by nitrogen purging for 10 min prior to measurements. In following experiments, the sample containing $\text{Mg}(\text{ClO}_4)_2$ was prepared using saturated $\text{Mg}(\text{ClO}_4)_2$ solution in DCM. ($\text{Mg}(\text{ClO}_4)_2$ (2 mg) was added to DCM (10 ml) and sonicated 30 min at room temperature. The supernatant was used as saturated $\text{Mg}(\text{ClO}_4)_2$ solution in DCM.)

6. References

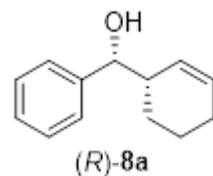
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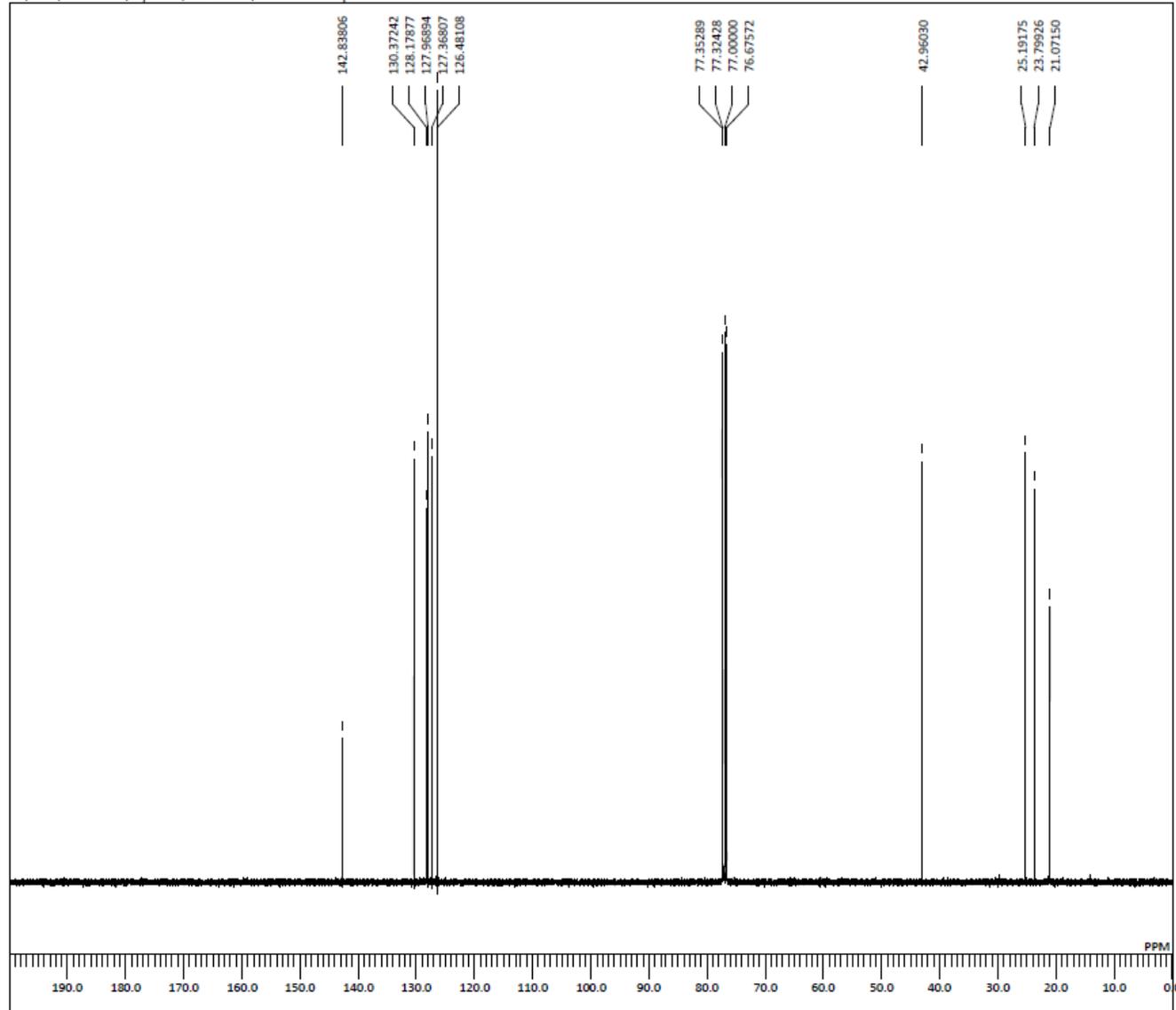
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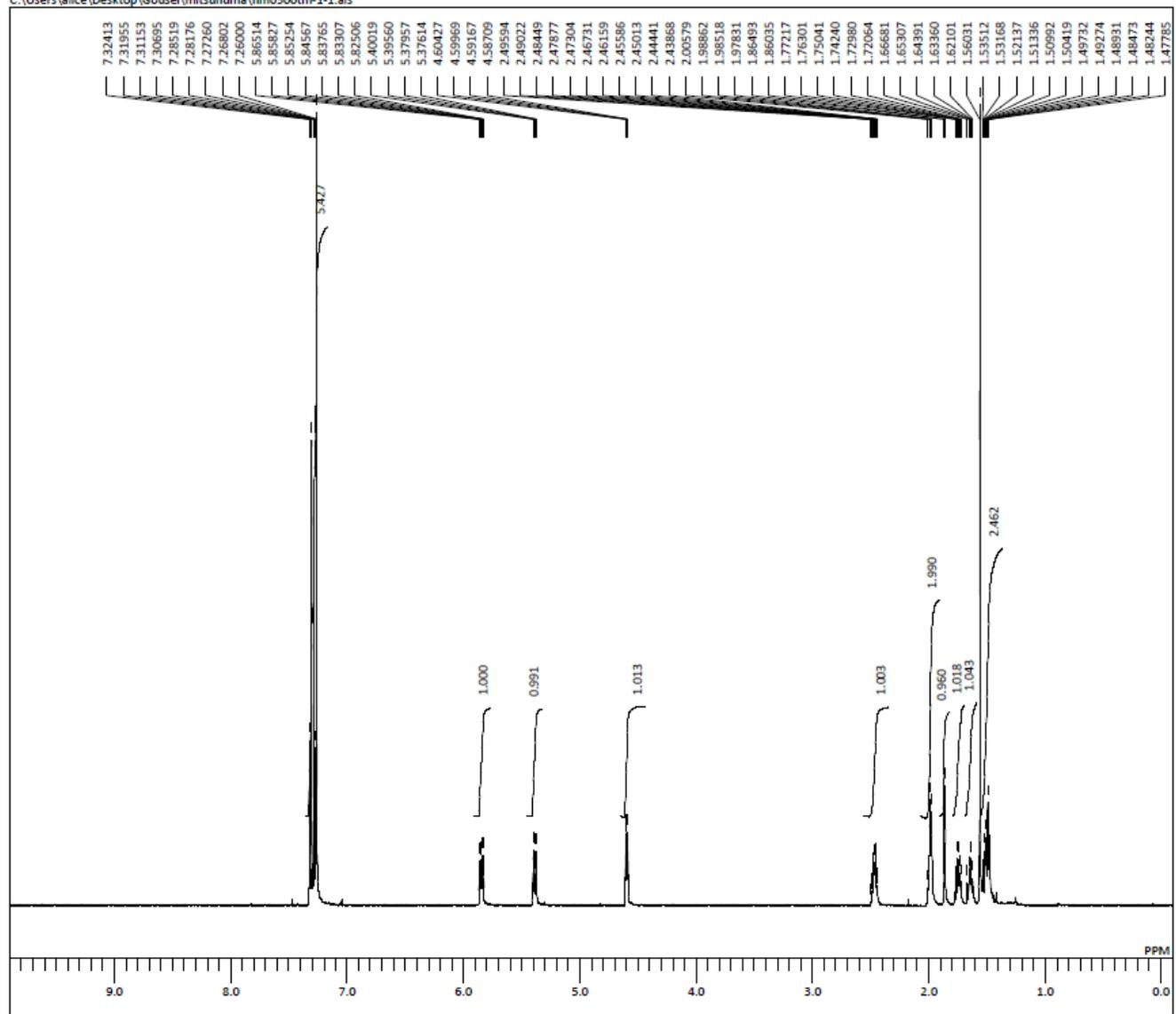


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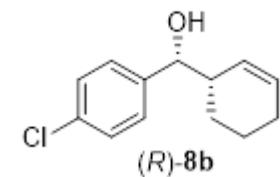


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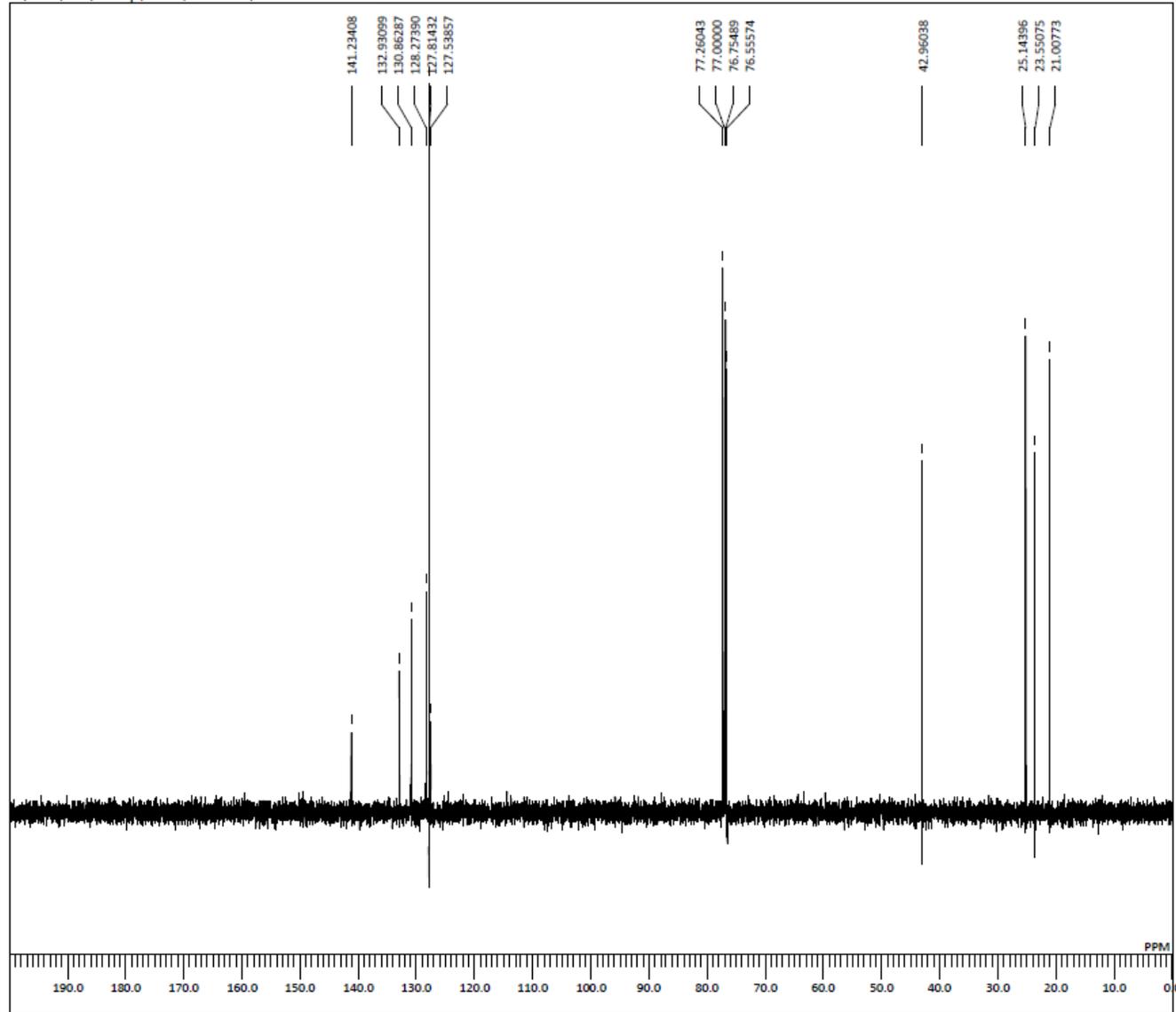
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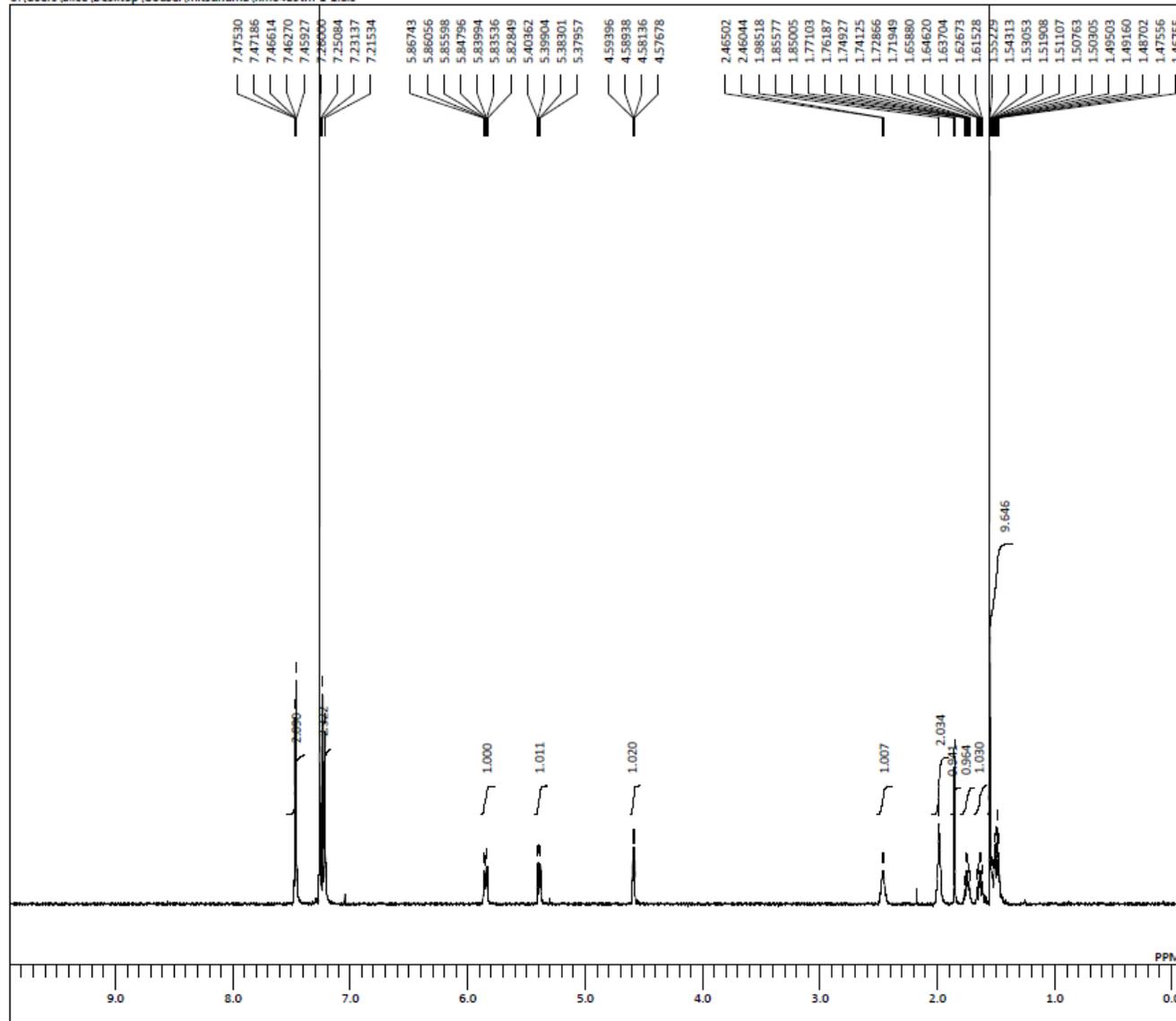


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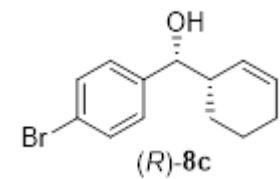


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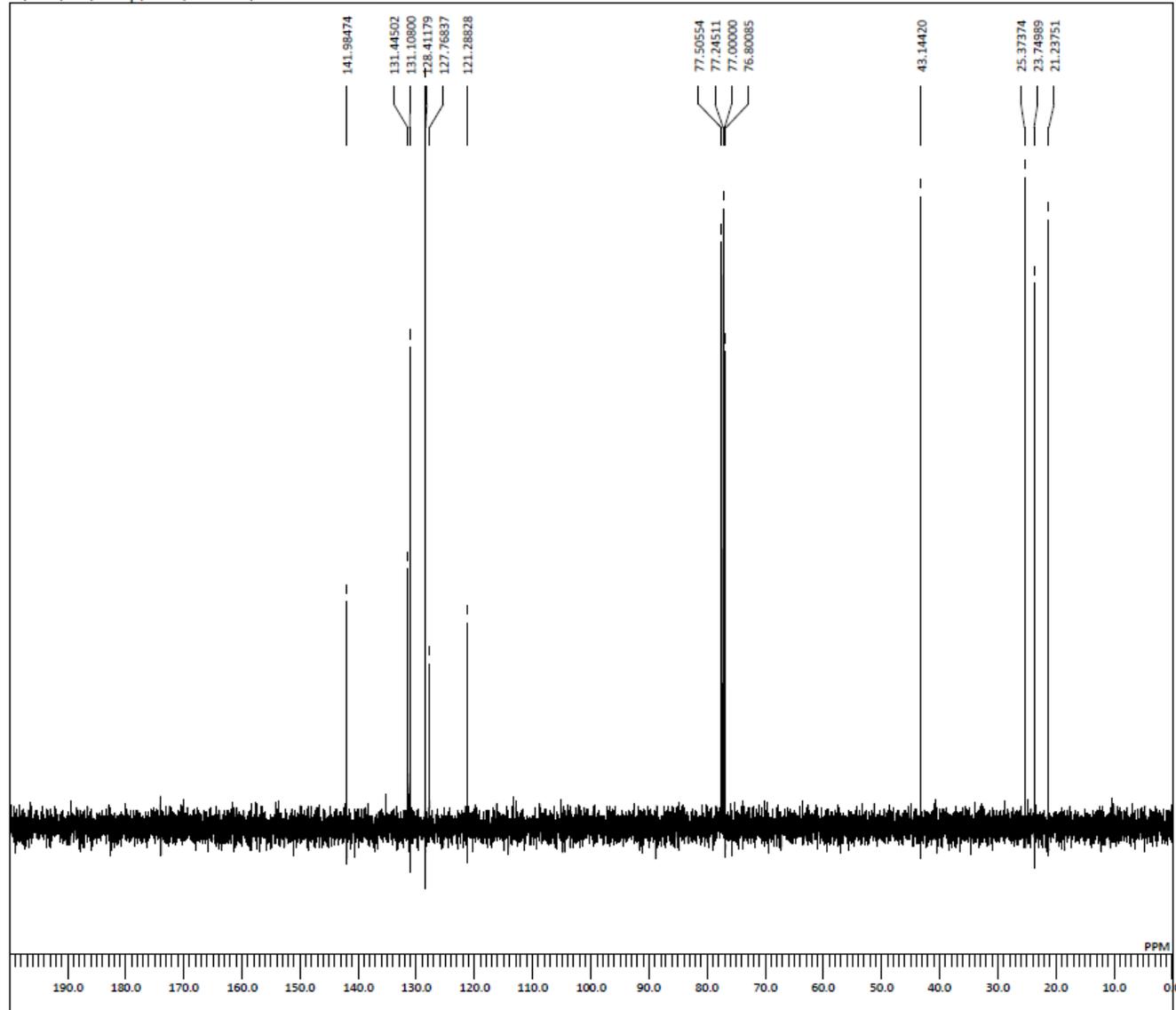
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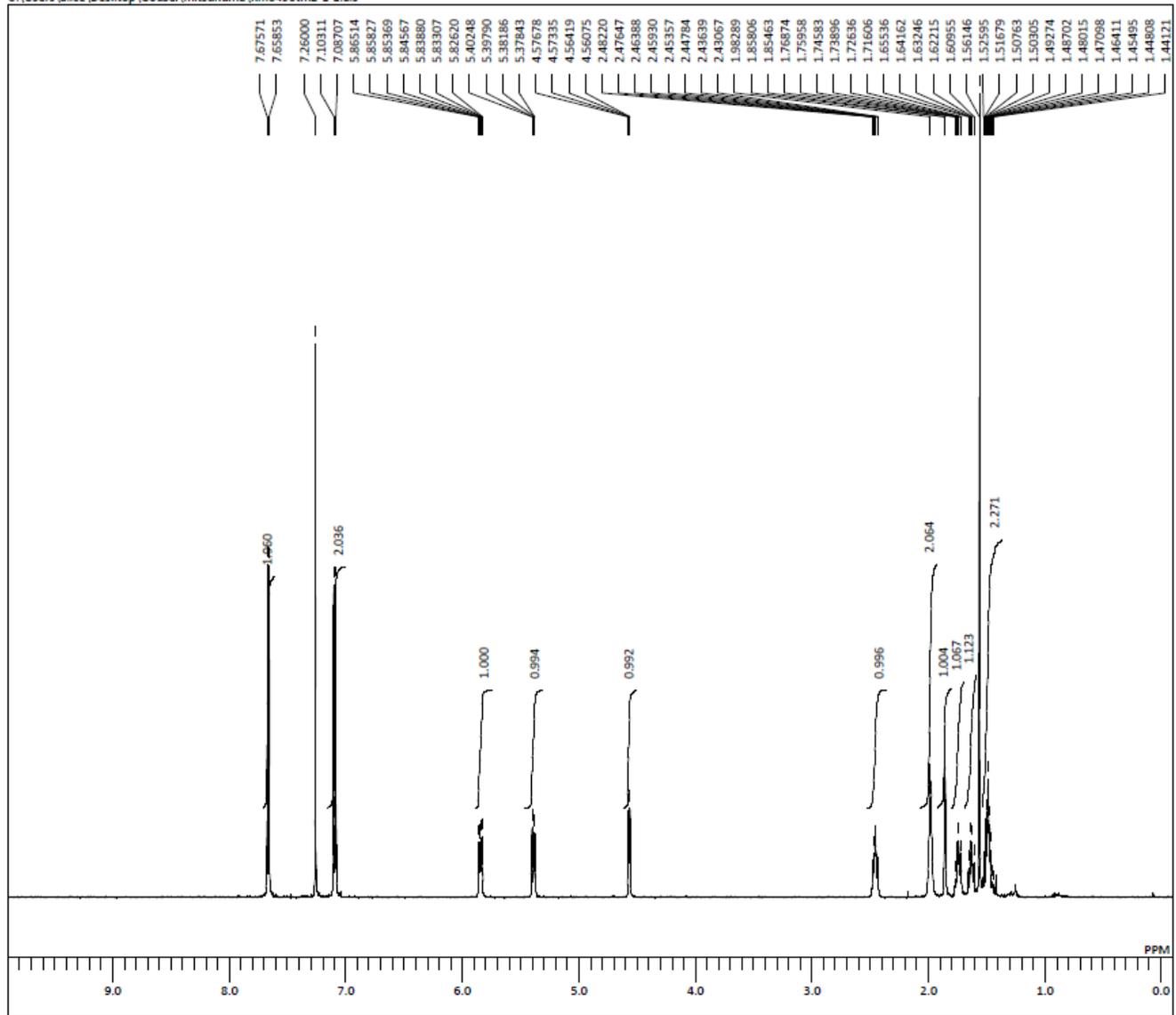


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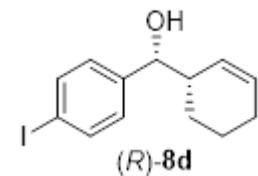


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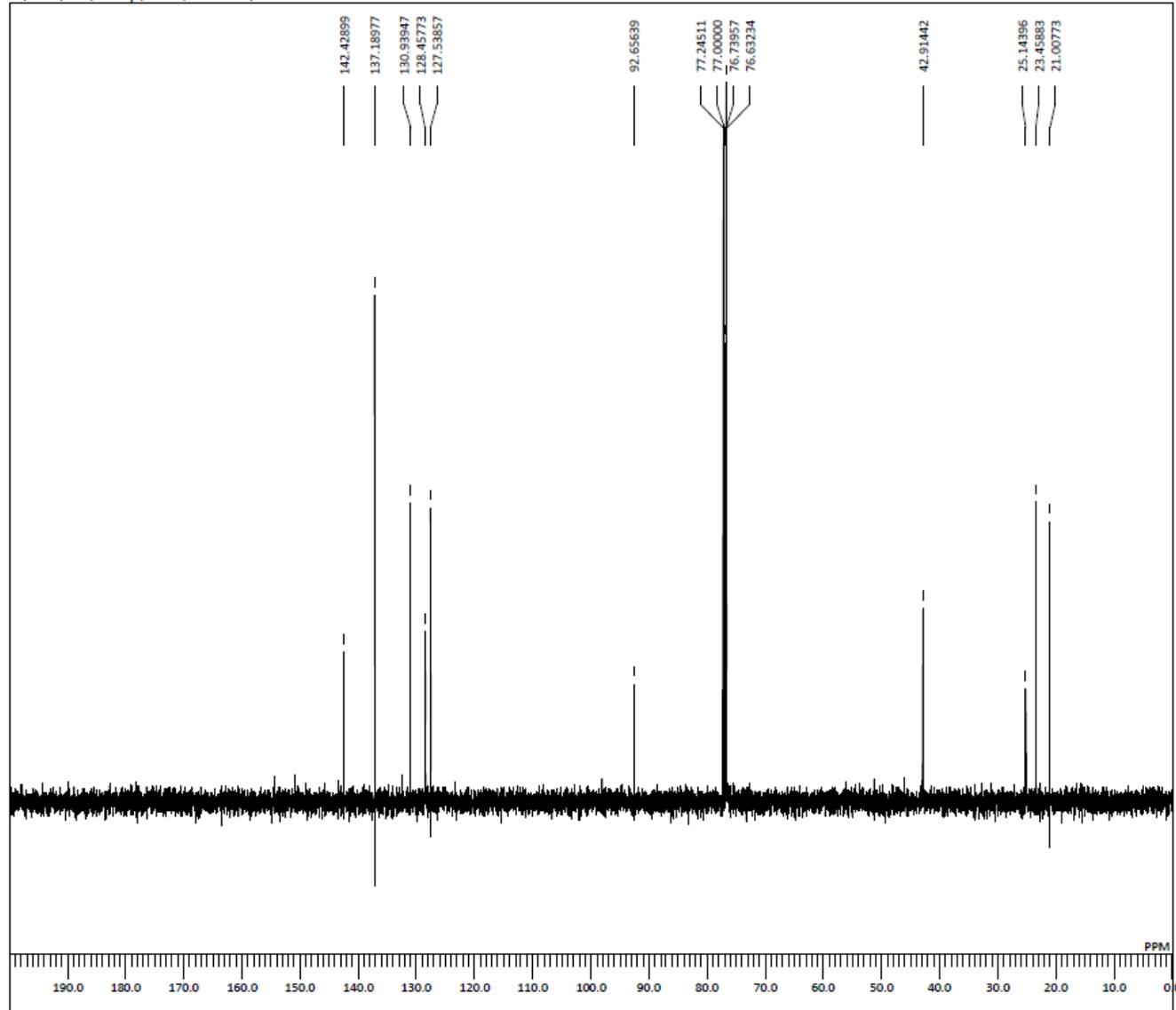
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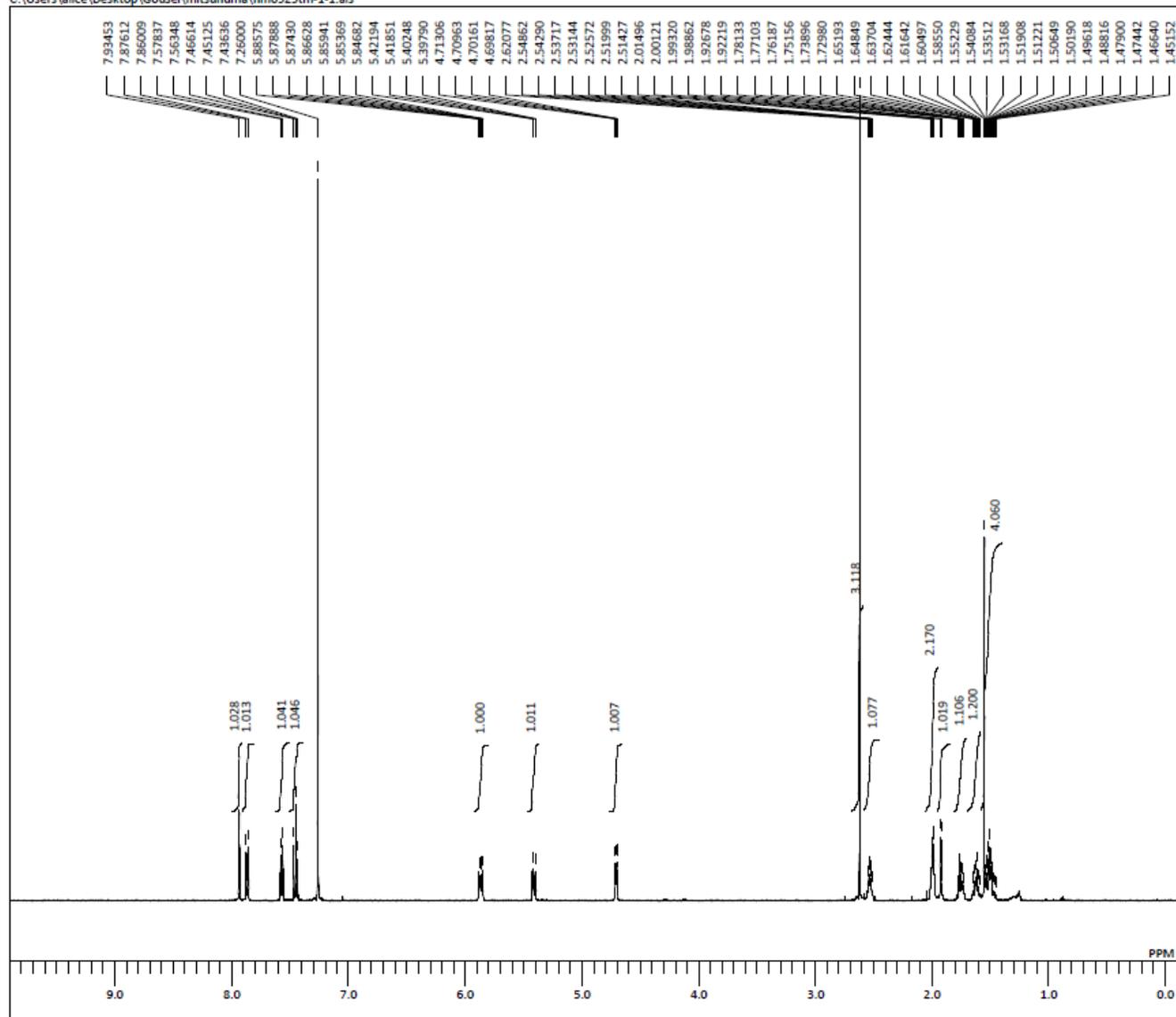


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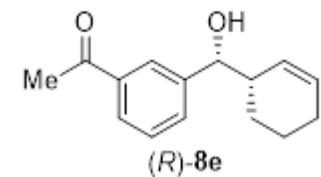


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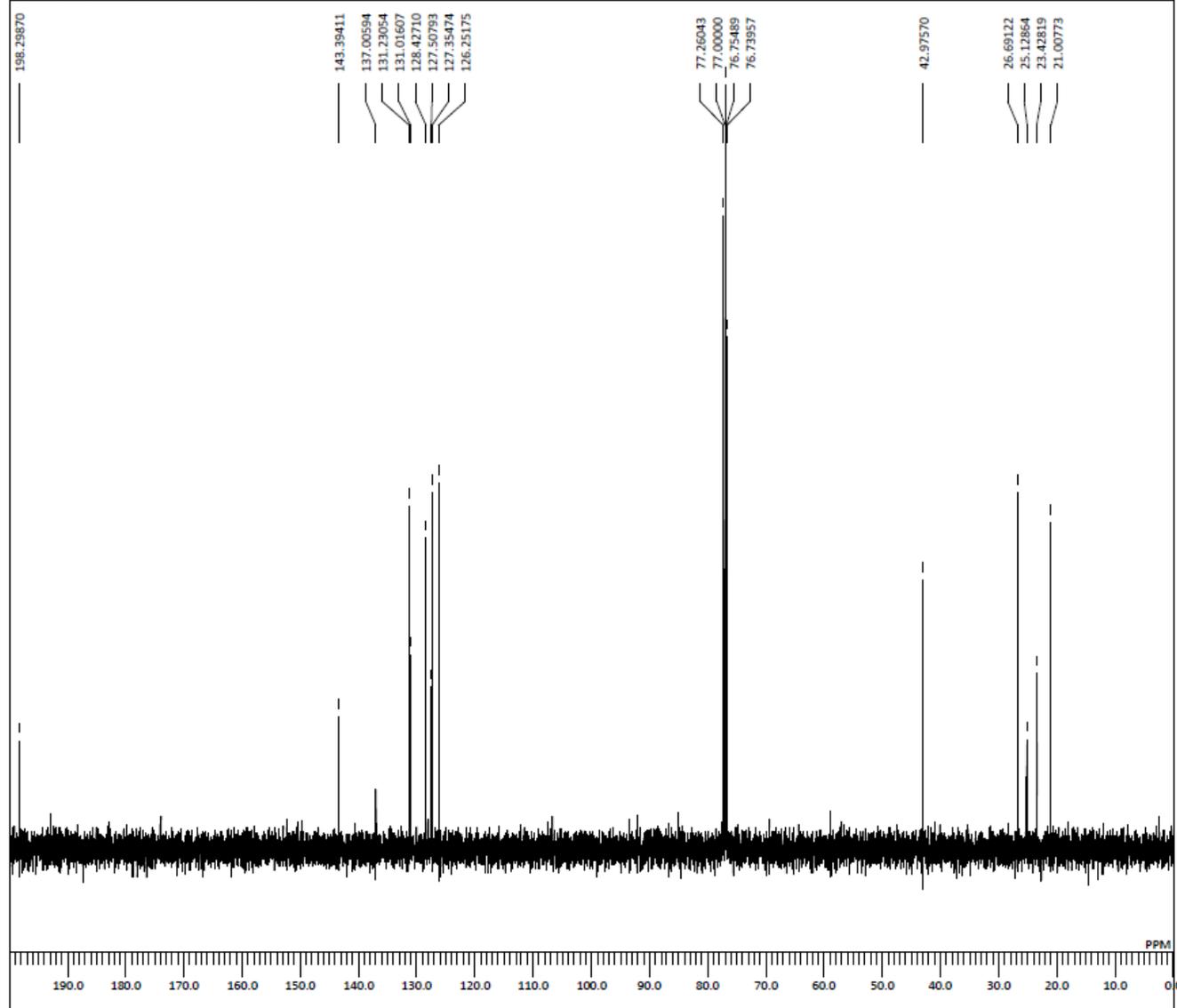
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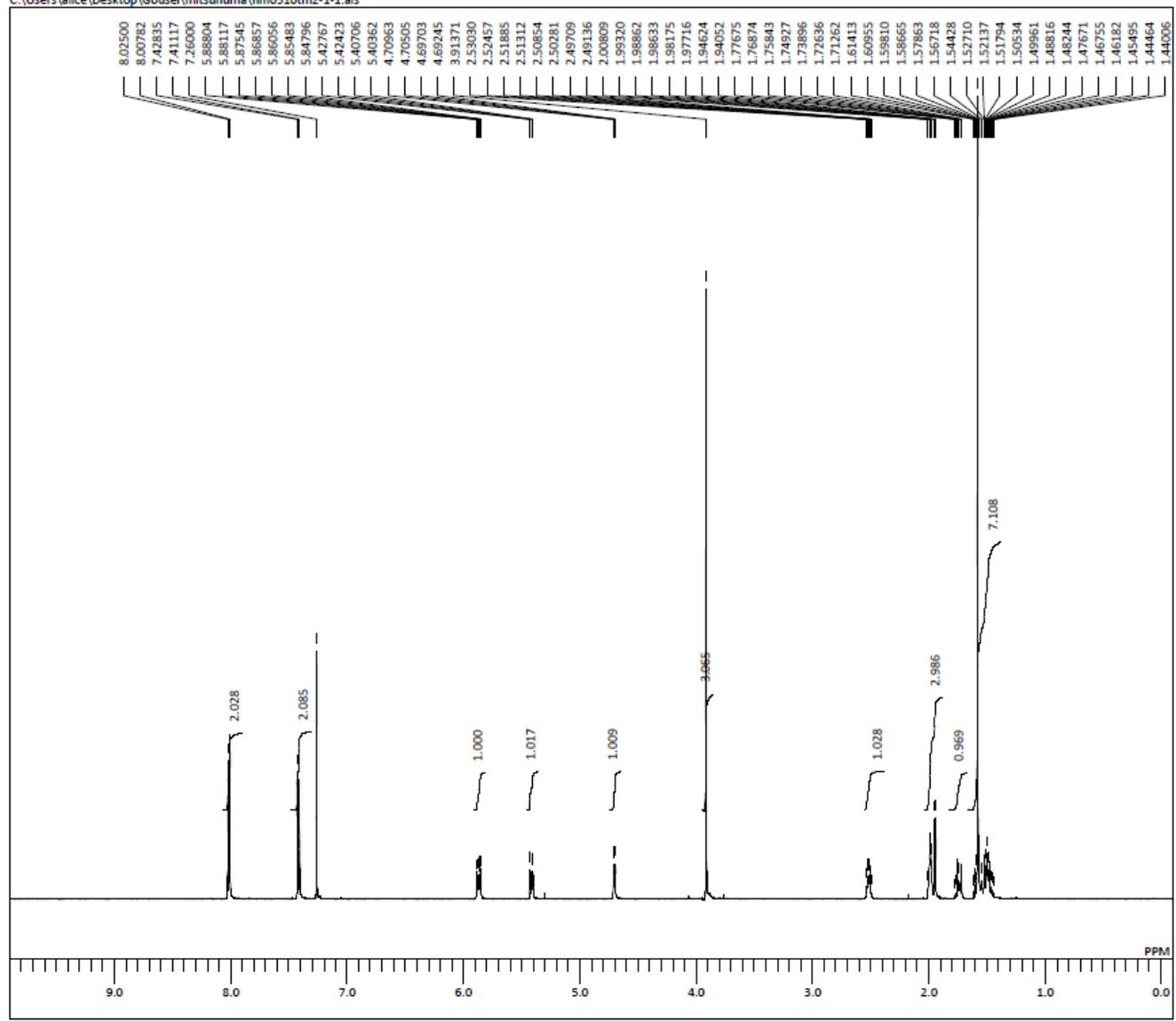


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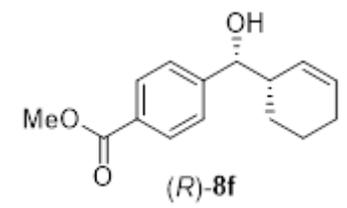


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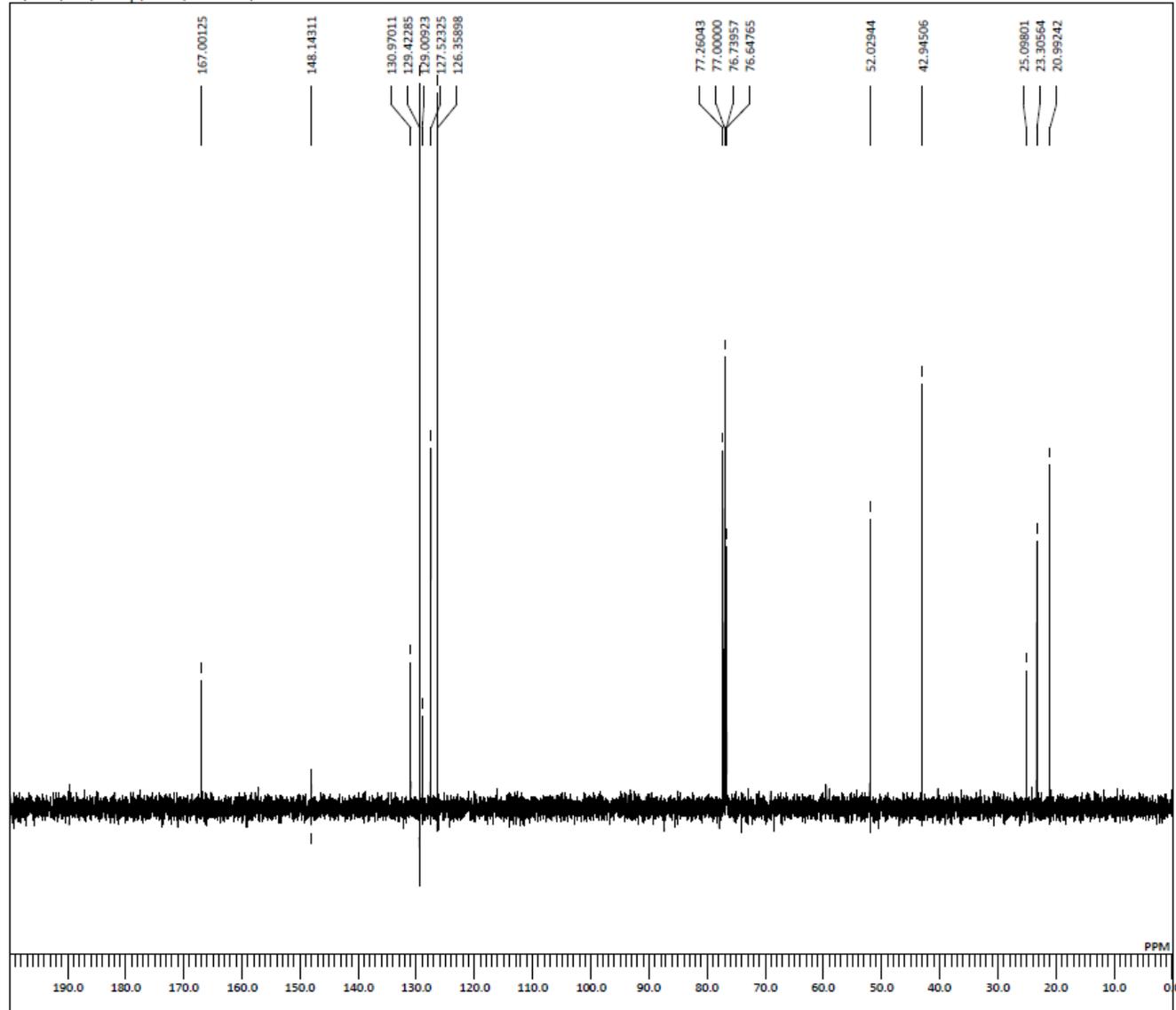
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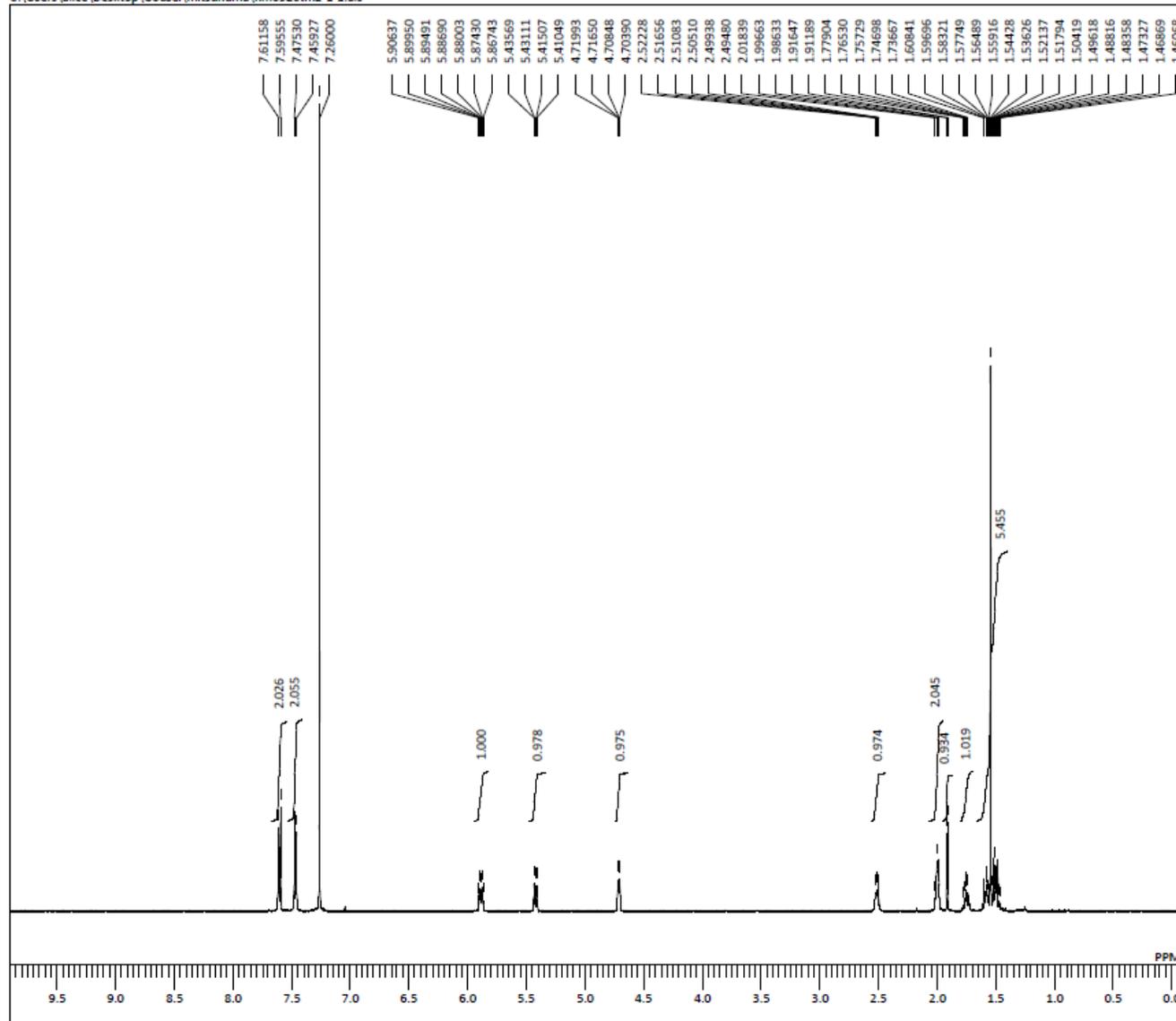


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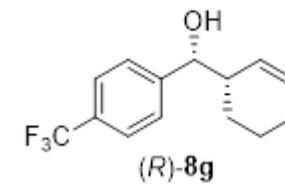


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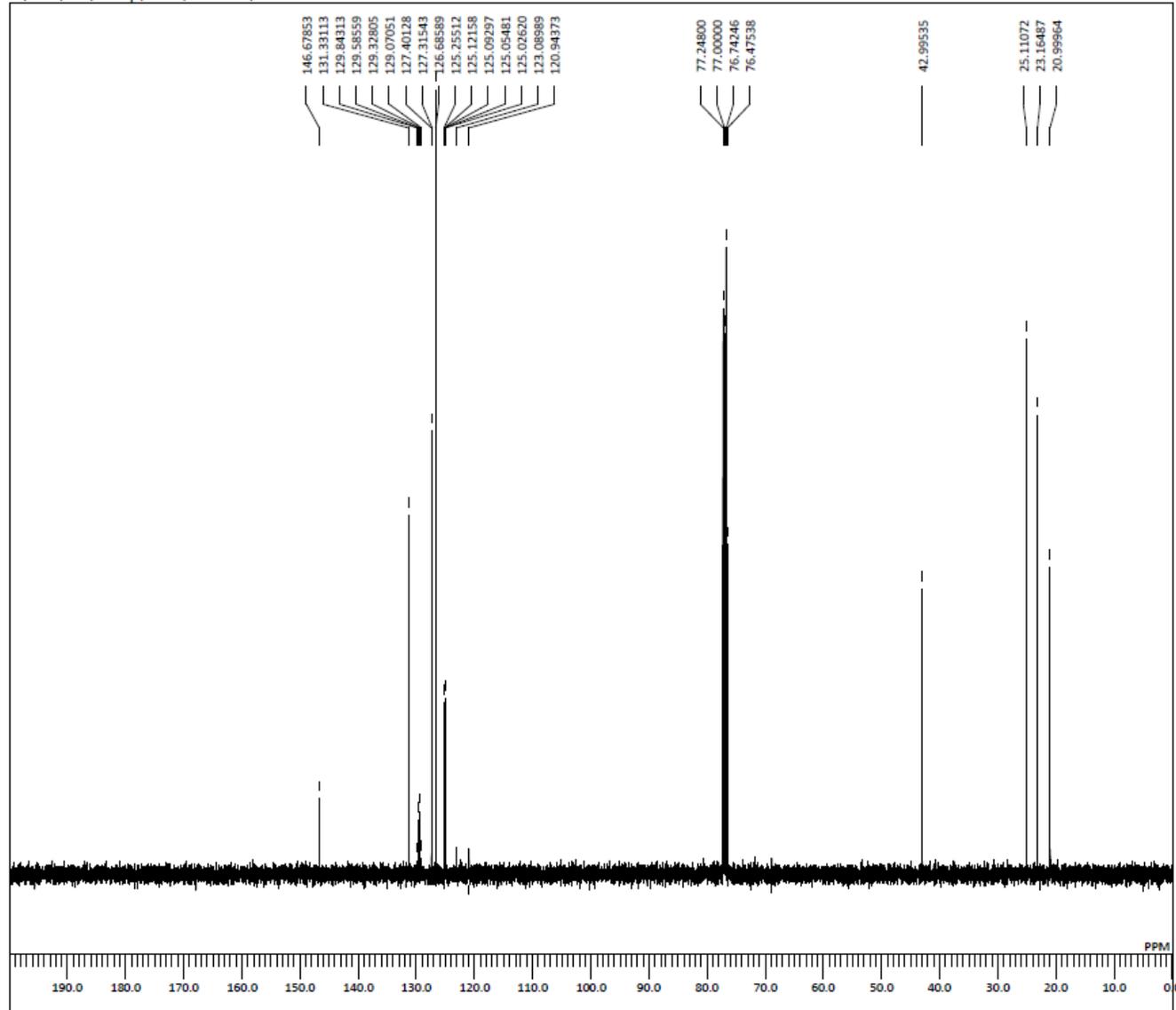
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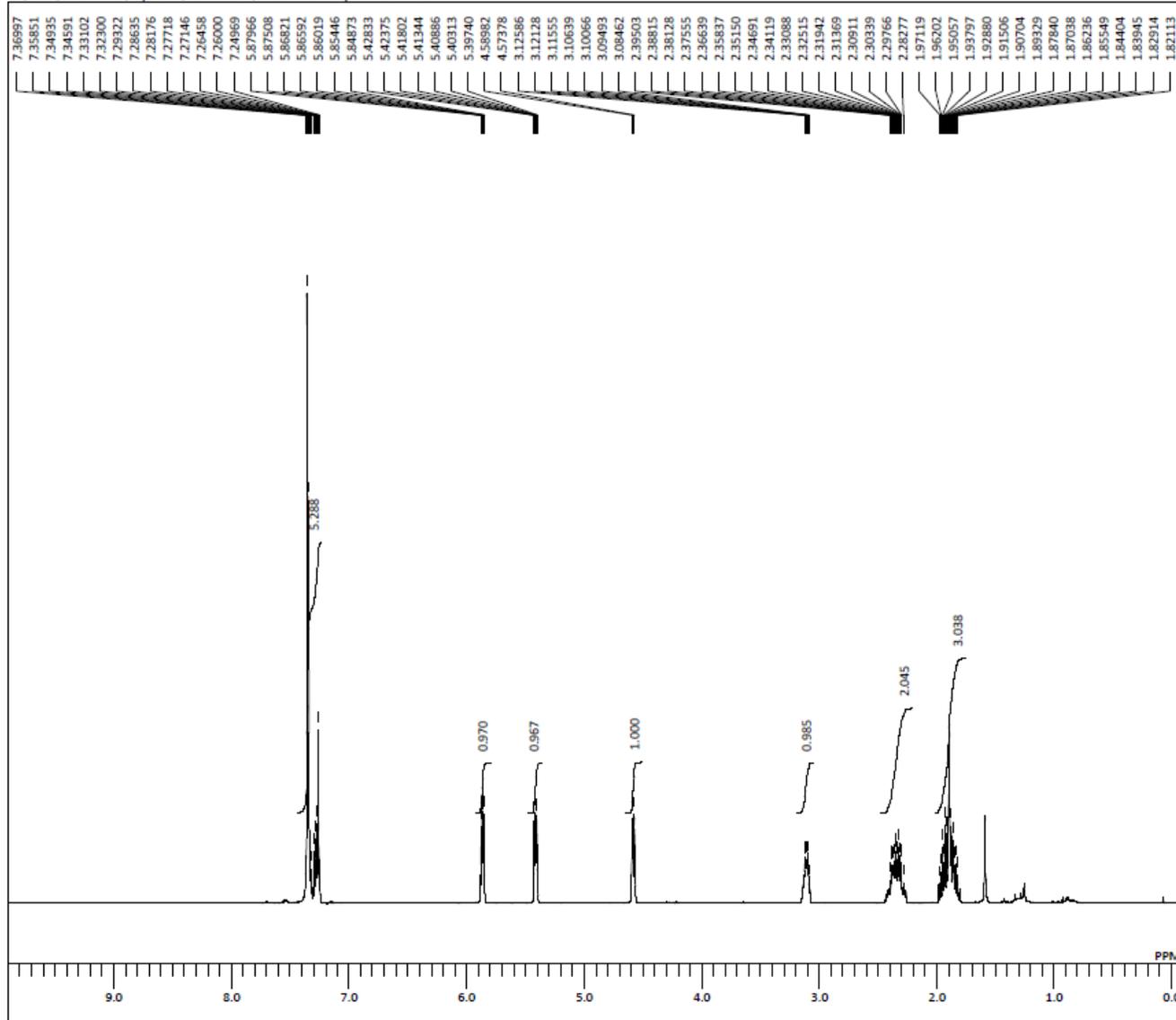


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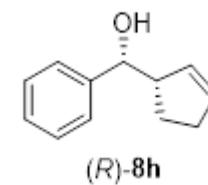


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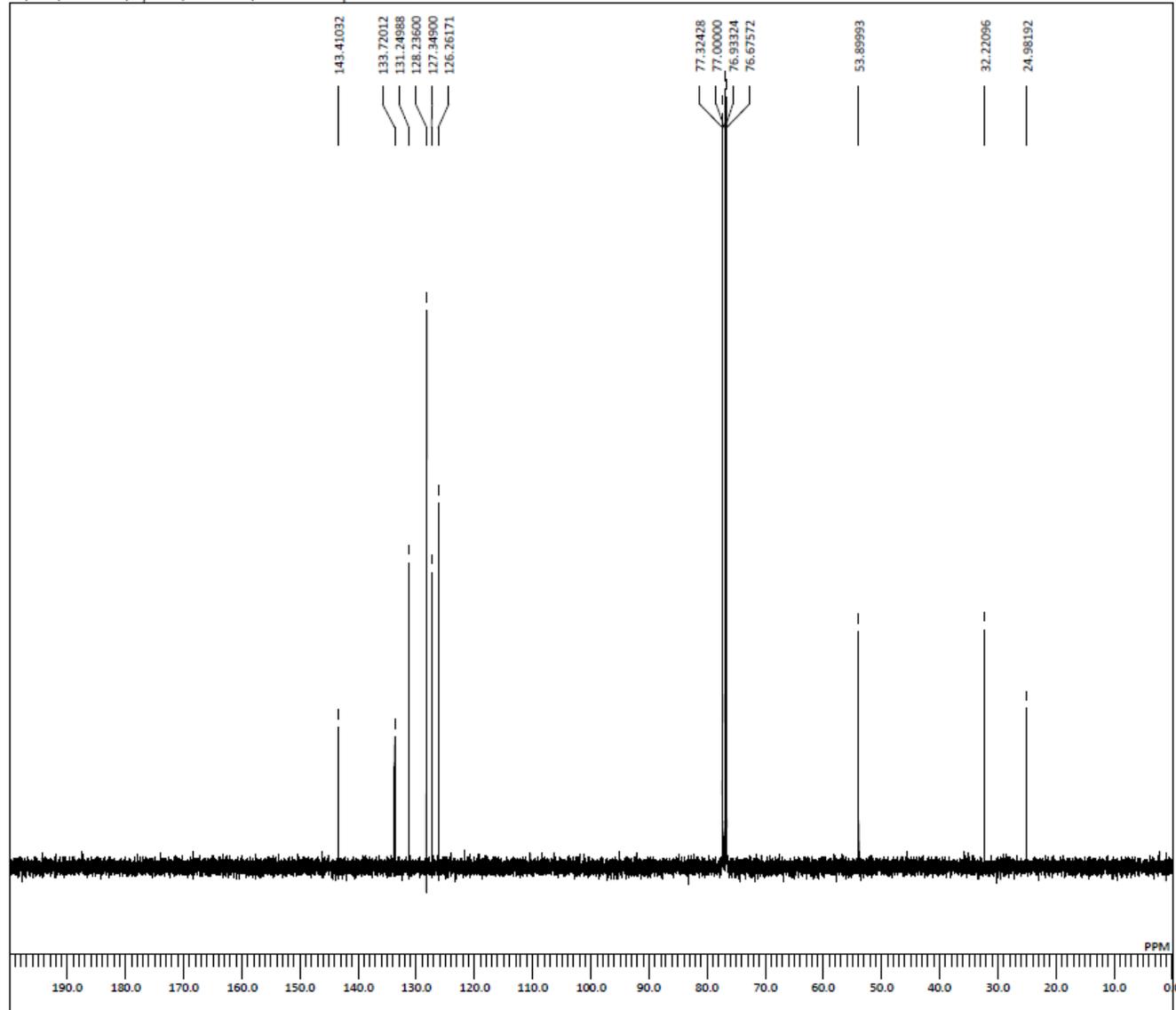
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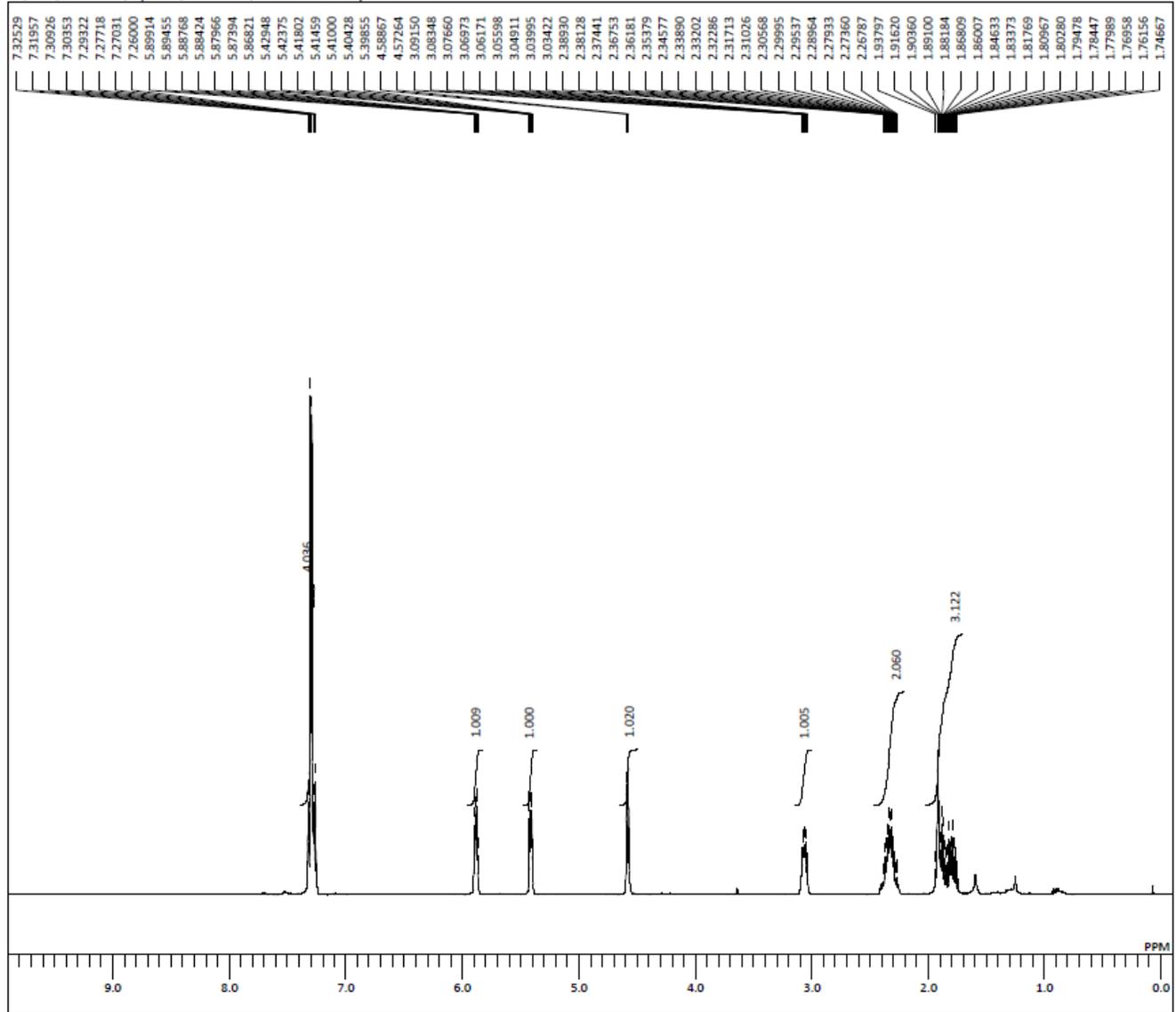


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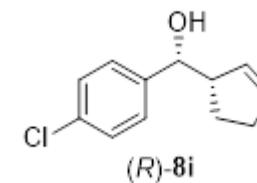


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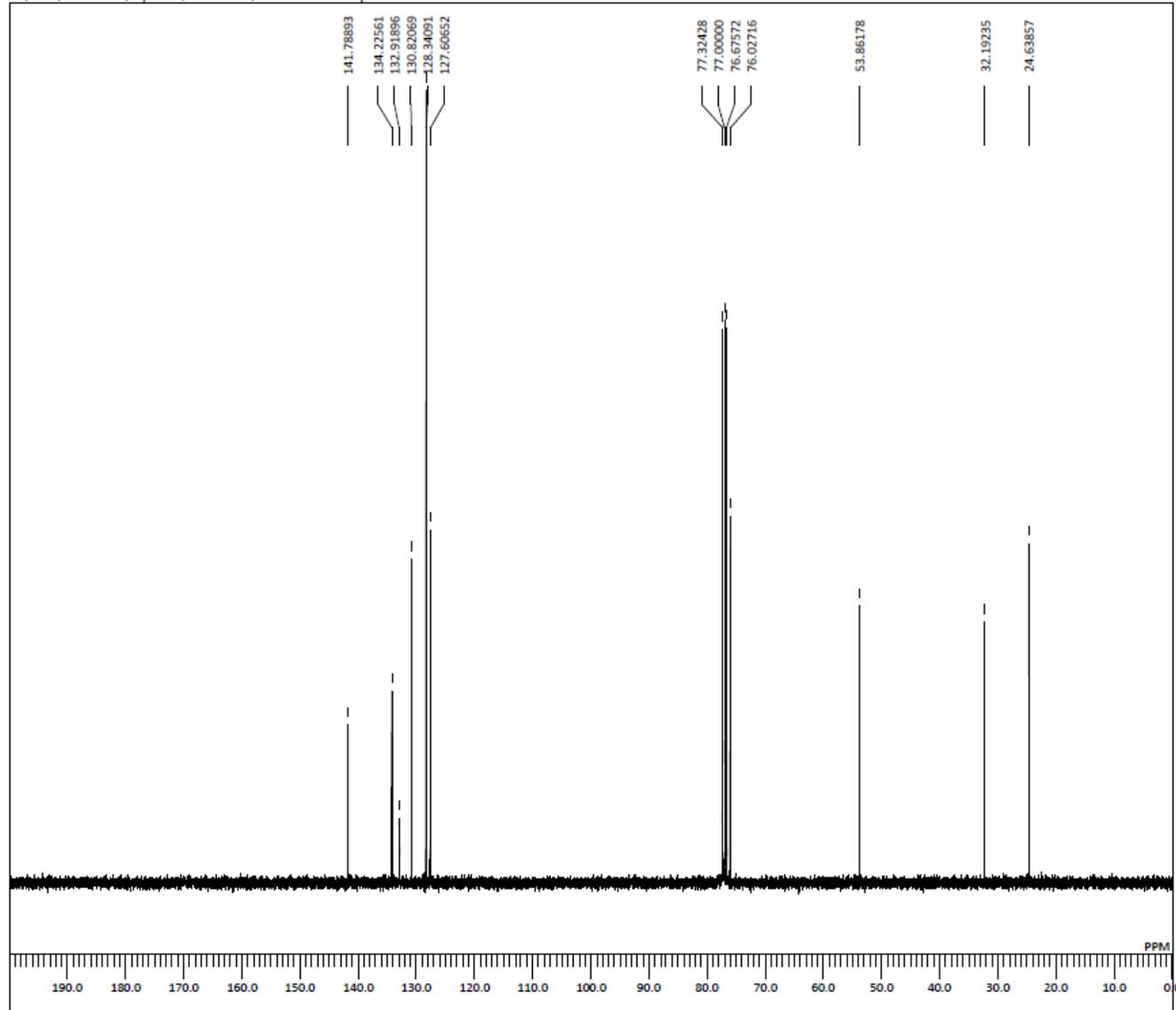
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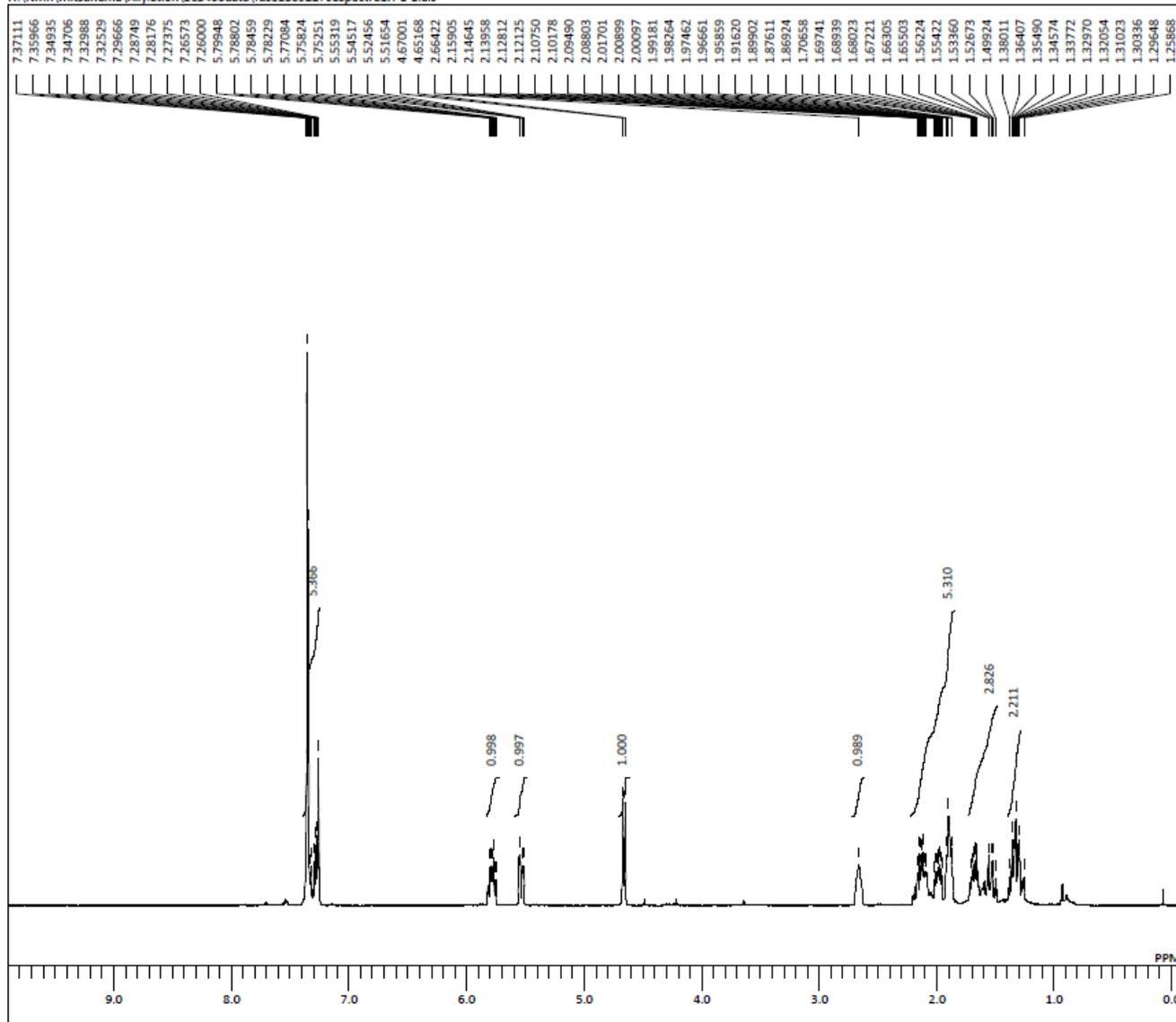


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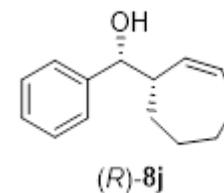


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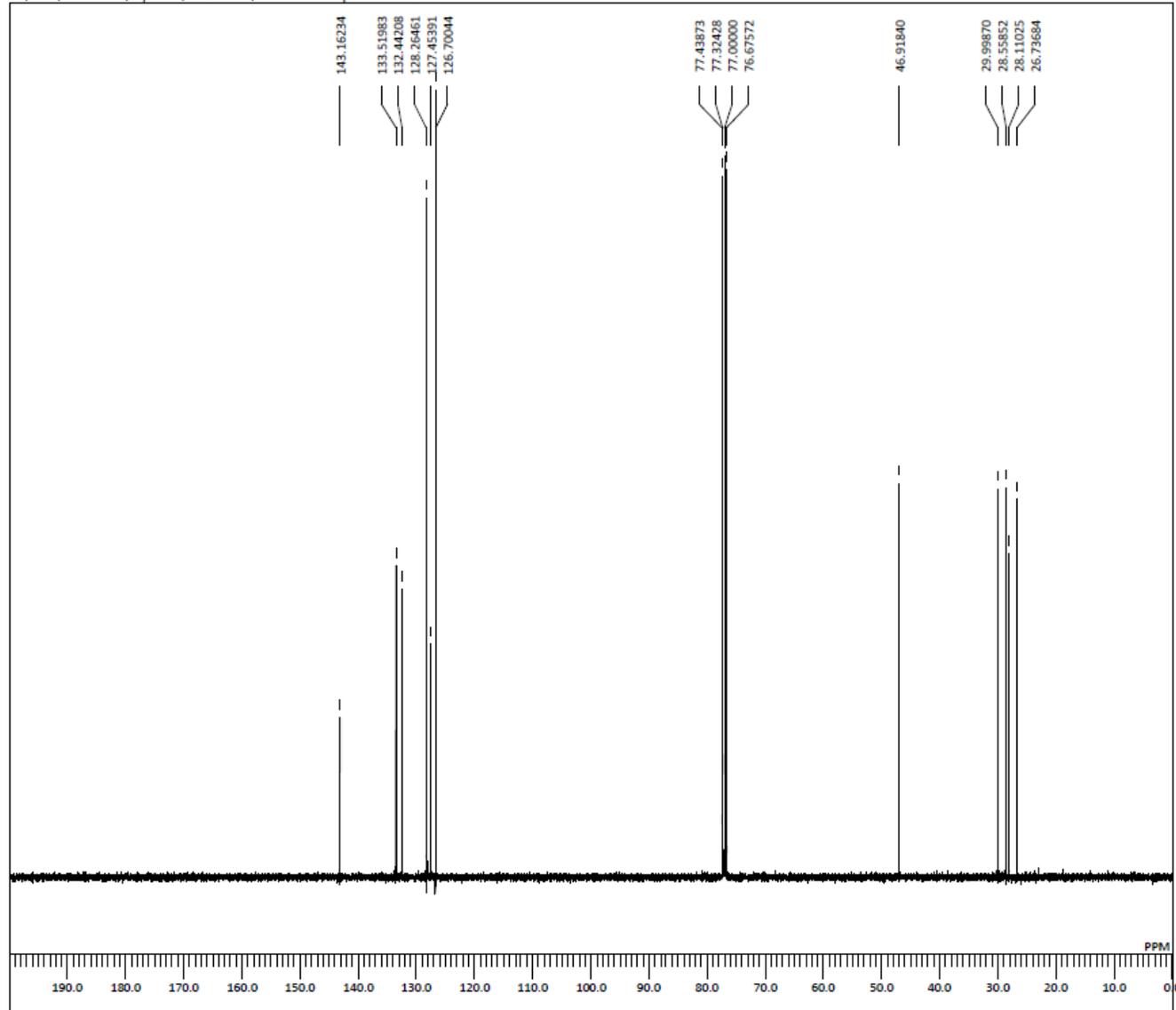
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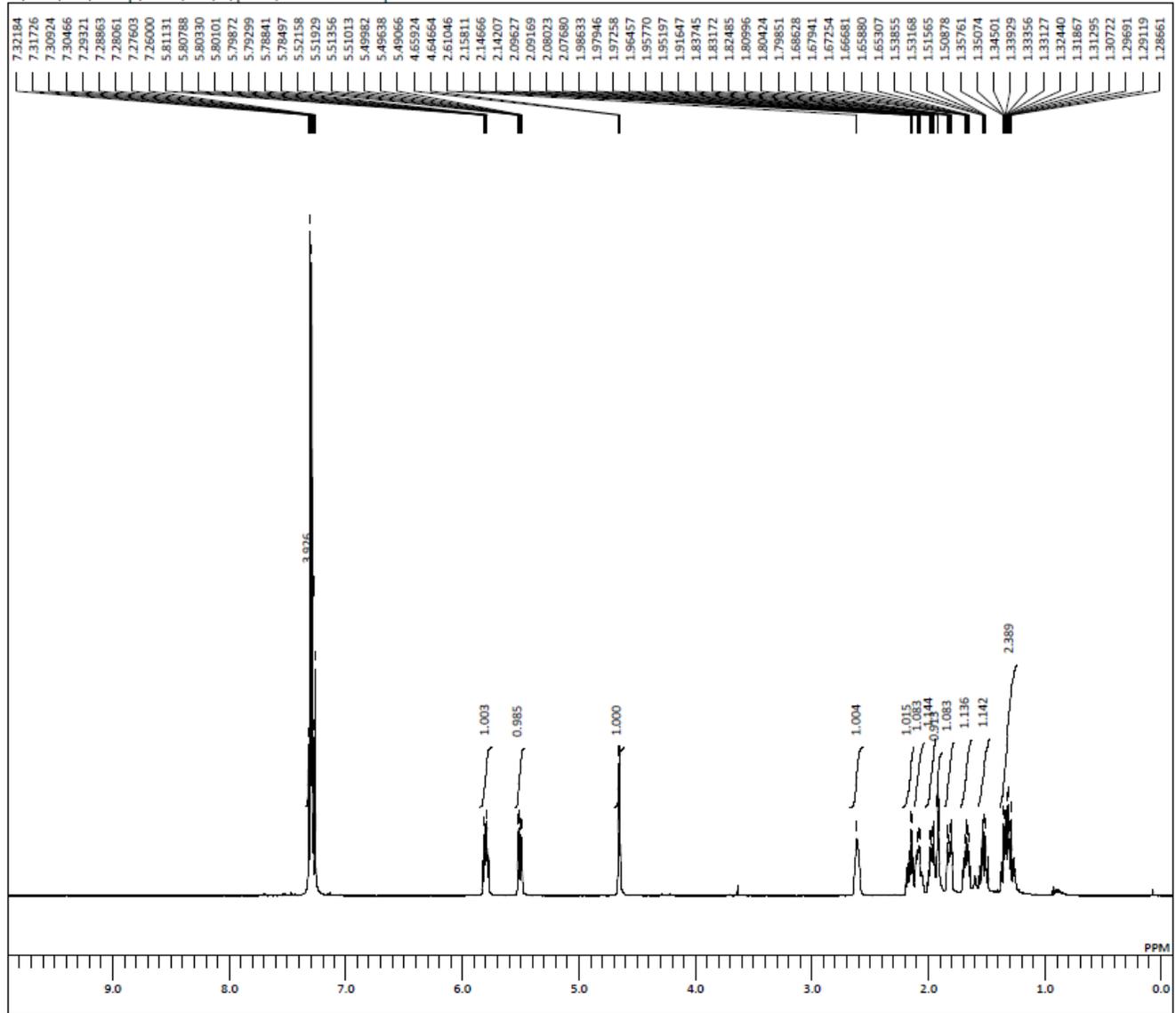


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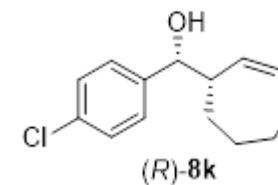


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OBFRQ 98.52 MHz
OBSET 4.64 kHz
OBFIN 8.74 Hz
POINT 26214
FREQU 24630.54 Hz
SCANS 1774
ACQTM 1.0643 sec
PD 2.0000 sec
PW1 3.12 usec
IRNUC 1H
CTEMP 20.3 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

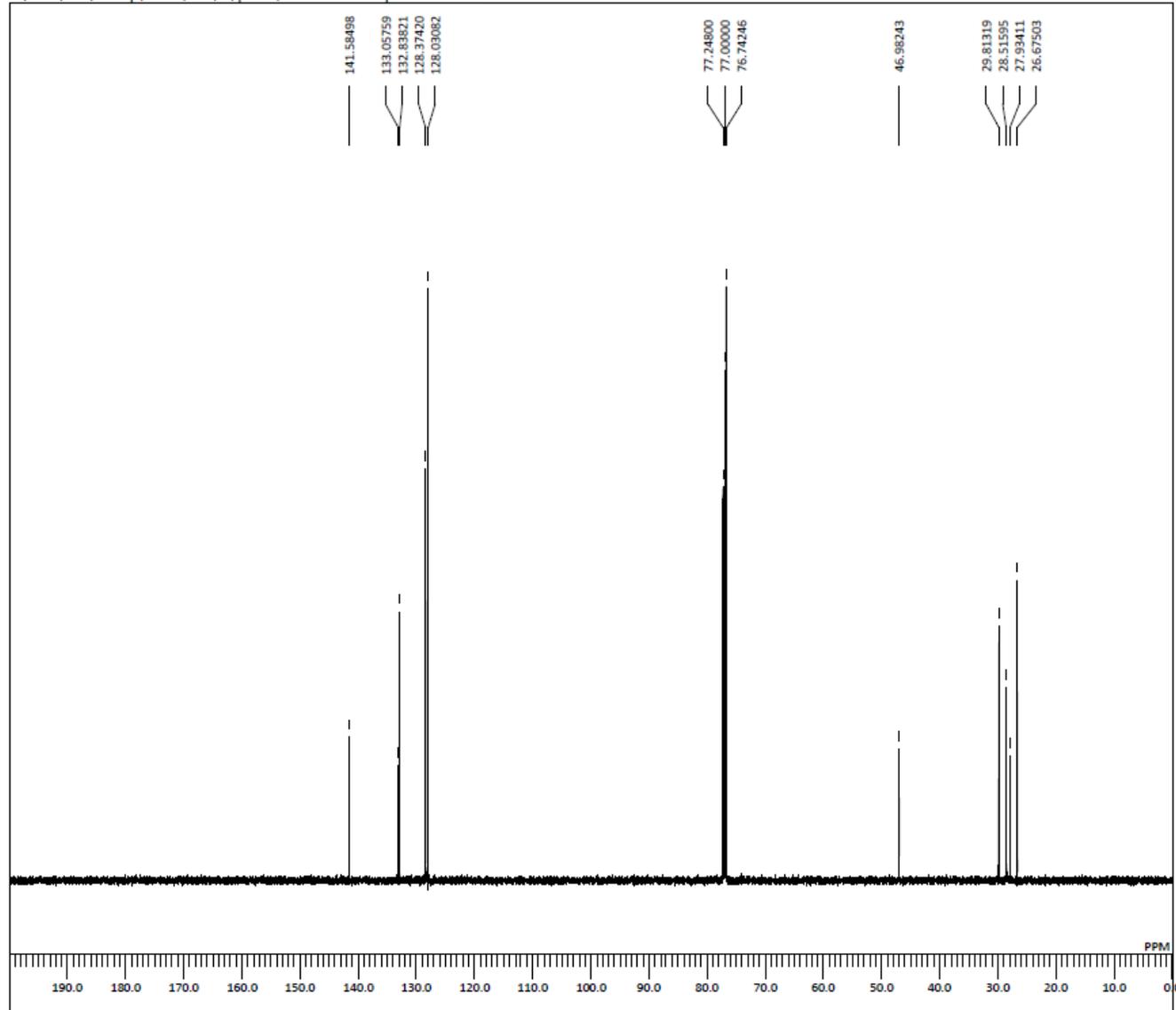
C:\Users\alice\Desktop\Gousei\Fuse\Cr\spectra\fuse181016hm589spectra1H-1-1.als



DFILE fuse181016hm589spectra1H-1-1.als
 COMMT
 DATIM 2018-10-16 21:52:32
 OBNUC 1H
 EXMOD proton.jxp
 OBFRO 500.16 MHz
 OBSET 2.41 KHz
 OBFIN 6.01 Hz
 POINT 13107
 FREQU 7507.51 Hz
 SCANS 8
 ACQTM 1.7459 sec
 PD 5.0000 sec
 PW1 5.55 usec
 IRNUC 1H
 CTEMP 21.8 c
 SLVNT CDCL3
 EXREF 7.26 ppm
 BF 0.12 Hz
 RGAIN 30

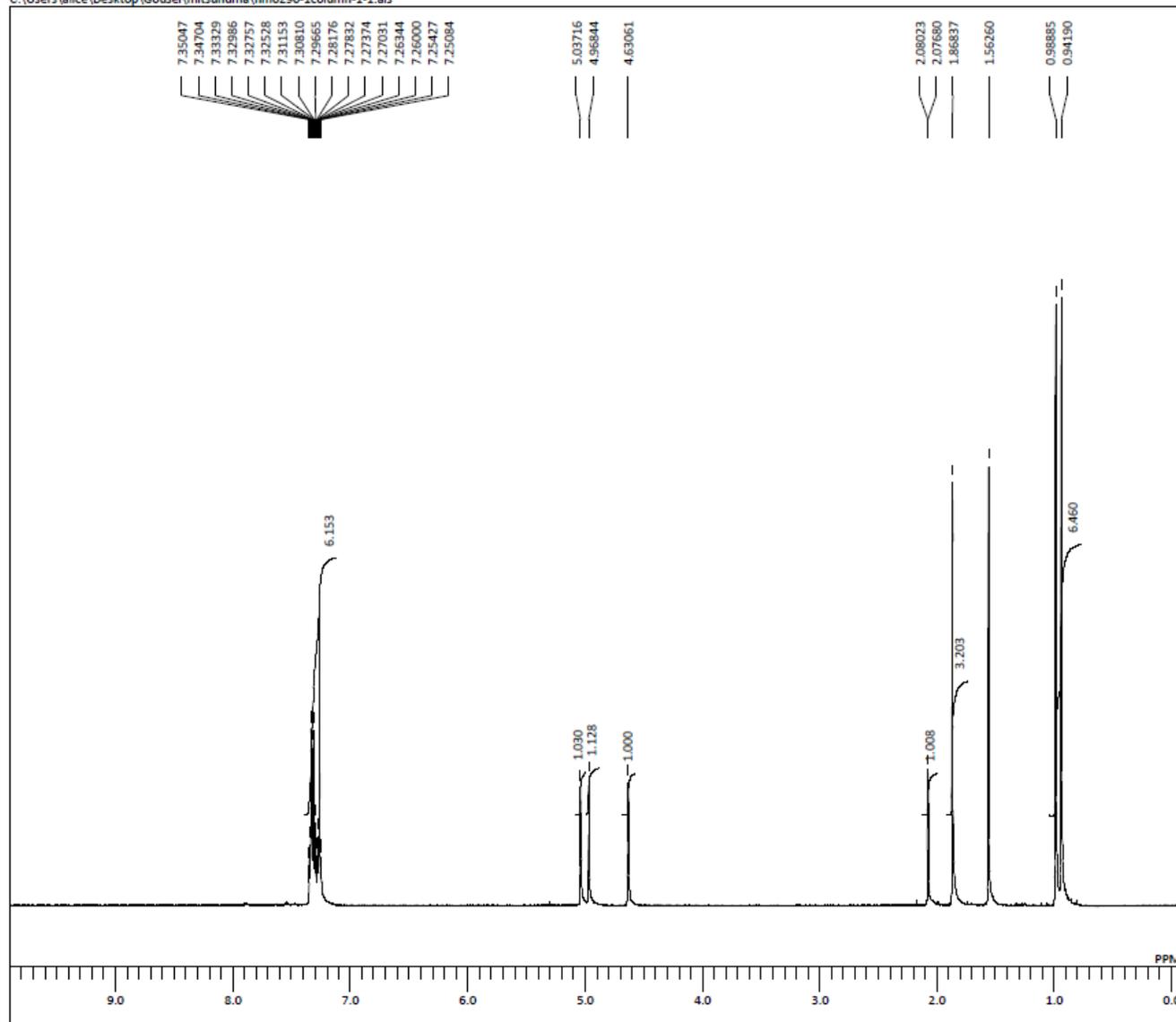


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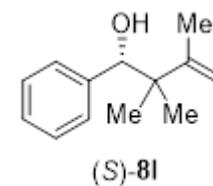


DFILE fuse181016hm589\spectra13C-1-1.als
COMMT
DATIM 2018-10-16 21:54:07
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 7.87 kHz
OBFIN 4.21 Hz
POINT 26214
FREQU 31446.54 Hz
SCANS 697
ACQTM 0.8336 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 22.2 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

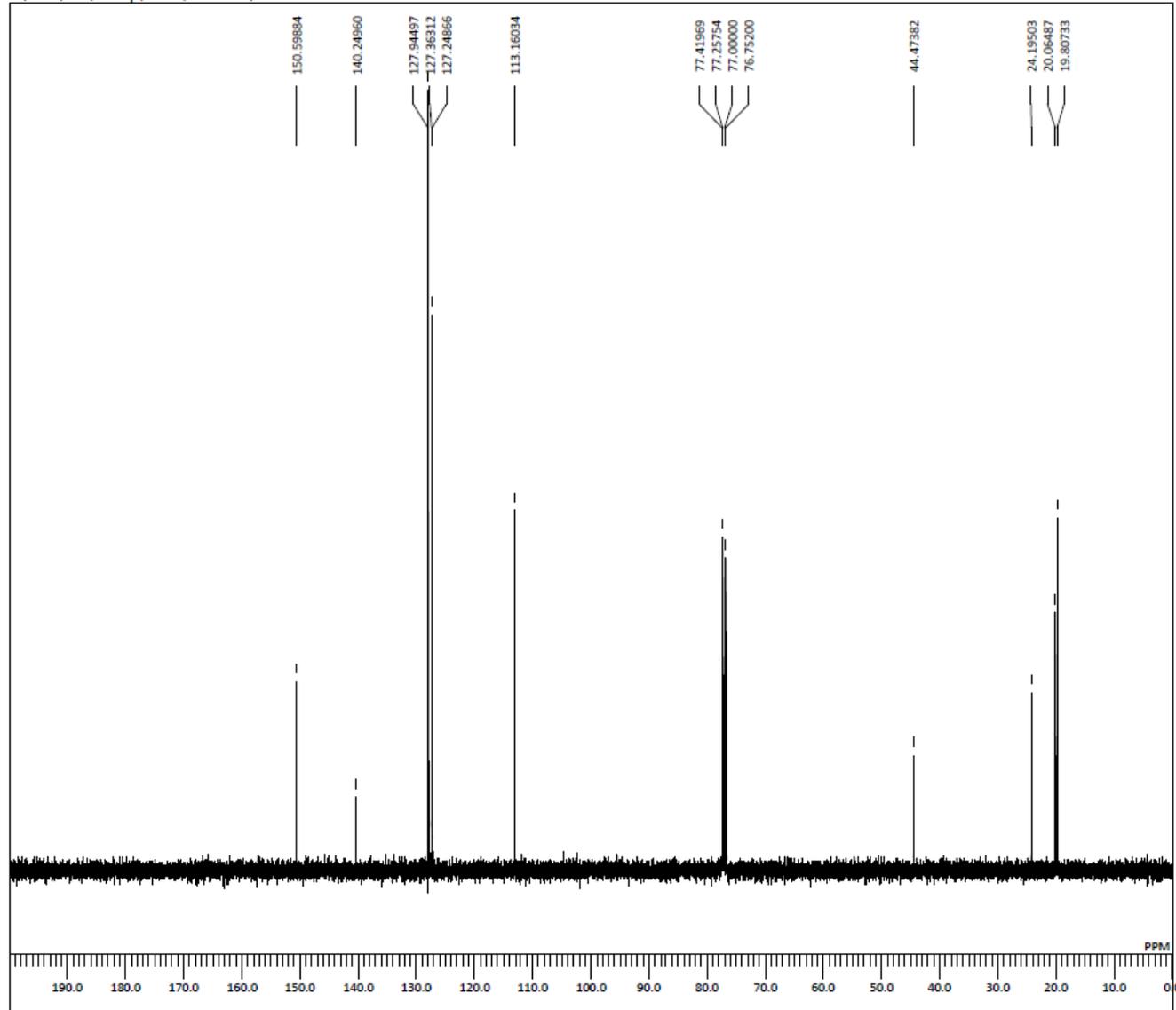
C:\Users\alice\Desktop\Gousei\mitsunuma\hm0290-1column-1-1.als



DFILE hm0290-1column-1-1.als
COMNT
DATIM 2018-06-29 18:17:57
OBNUC 1H
EXMOD proton.jpg
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.6 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

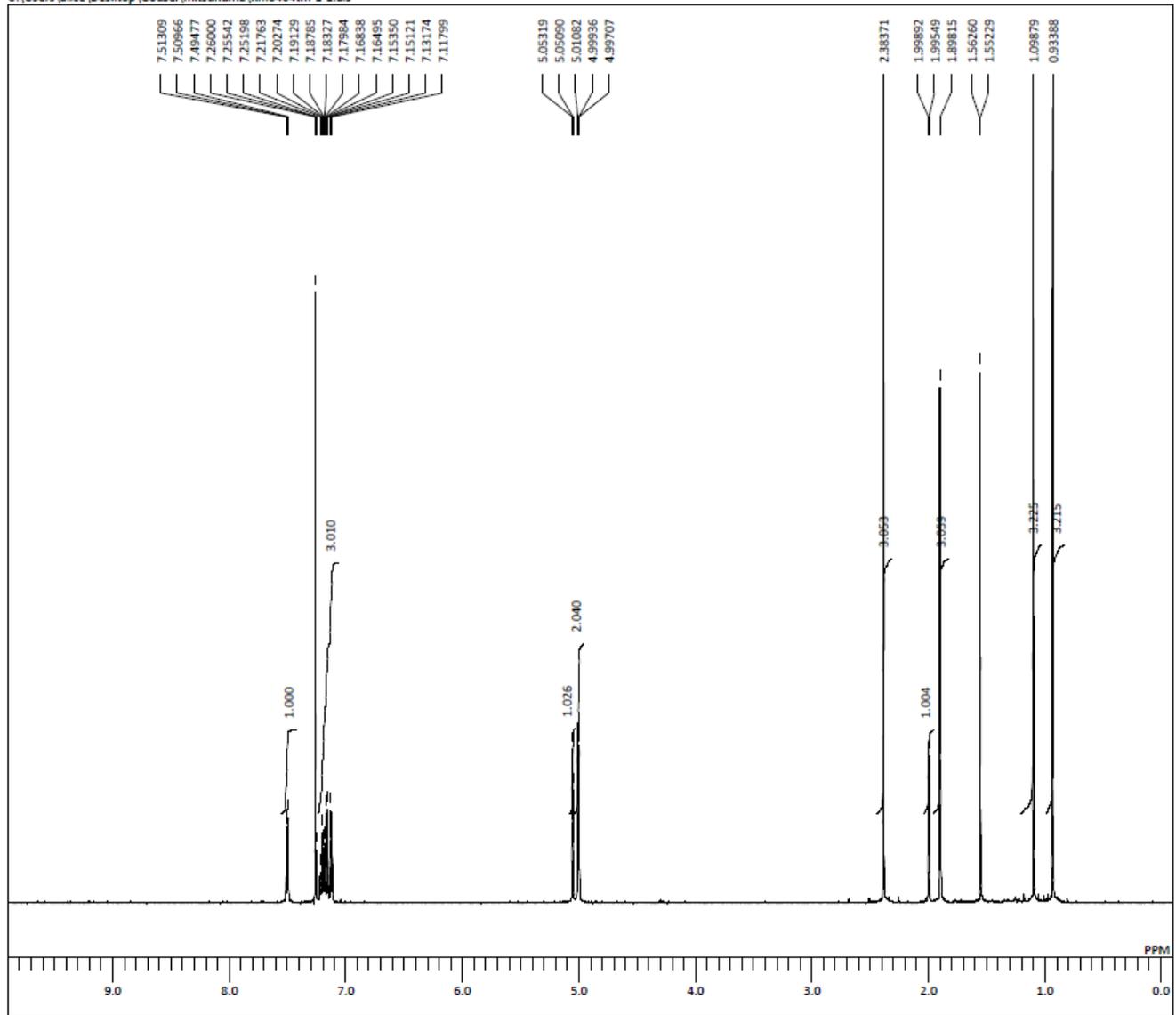


C:\Users\alice\Desktop\Gousei\mitsunuma\hm0290-1column 13C-1-1.als

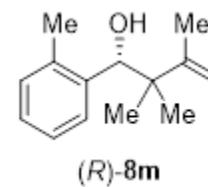


DFILE hm0290-1column 13C-1-1.als
COMMT
DATIM 2018-06-29 18:32:36
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 7.87 kHz
OBFIN 4.21 Hz
POINT 26214
FREQU 31446.54 Hz
SCANS 48
ACQTM 0.8336 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.5 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

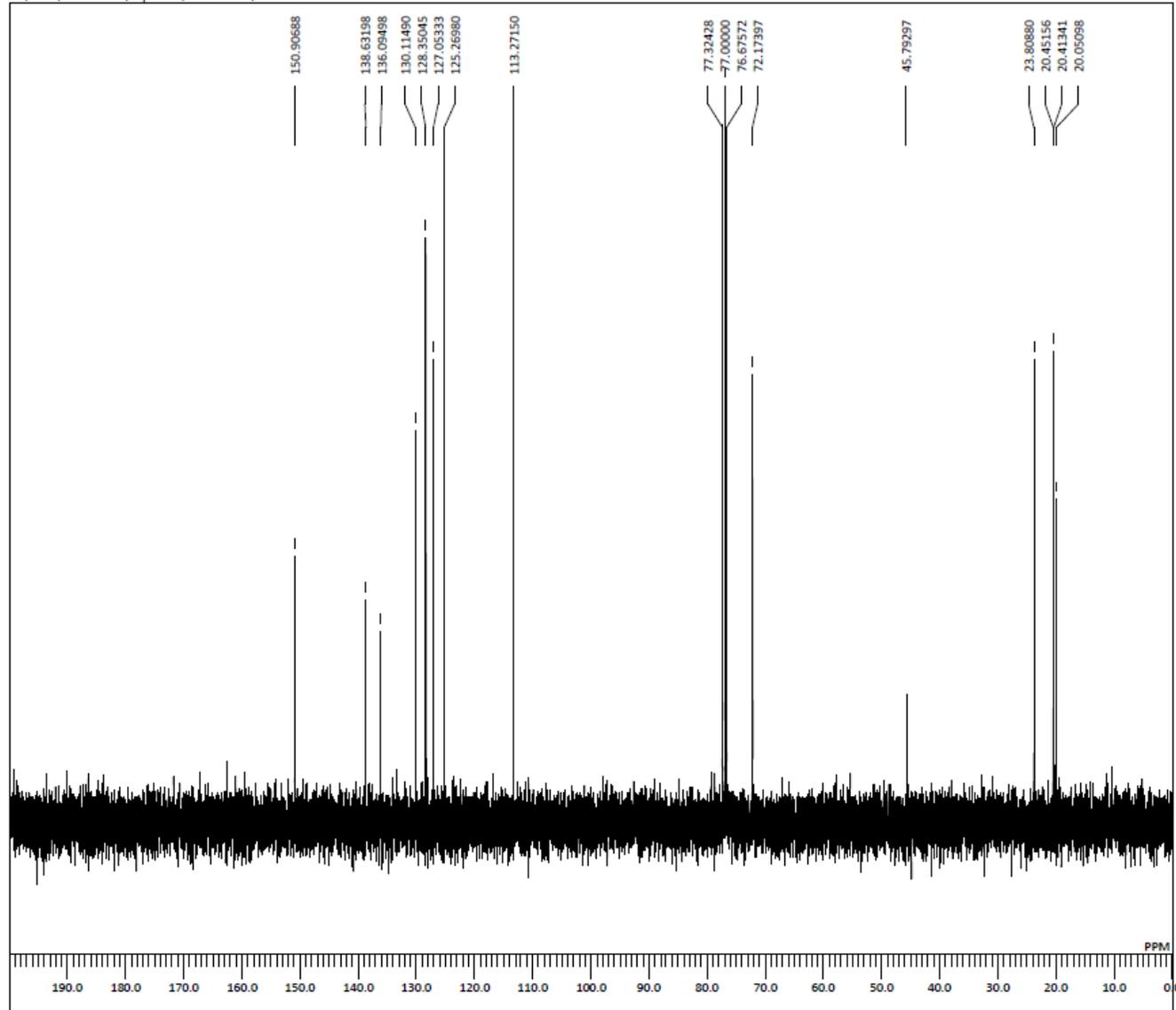
C:\Users\alice\Desktop\Gousei\mitsunuma\hm0464tm-1-1.als



DFILE hm0464tm-1-1.als
COMMT
DATIM 2018-09-11 20:58:10
OBNUC 1H
EXMOD proton.jpg
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.8 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

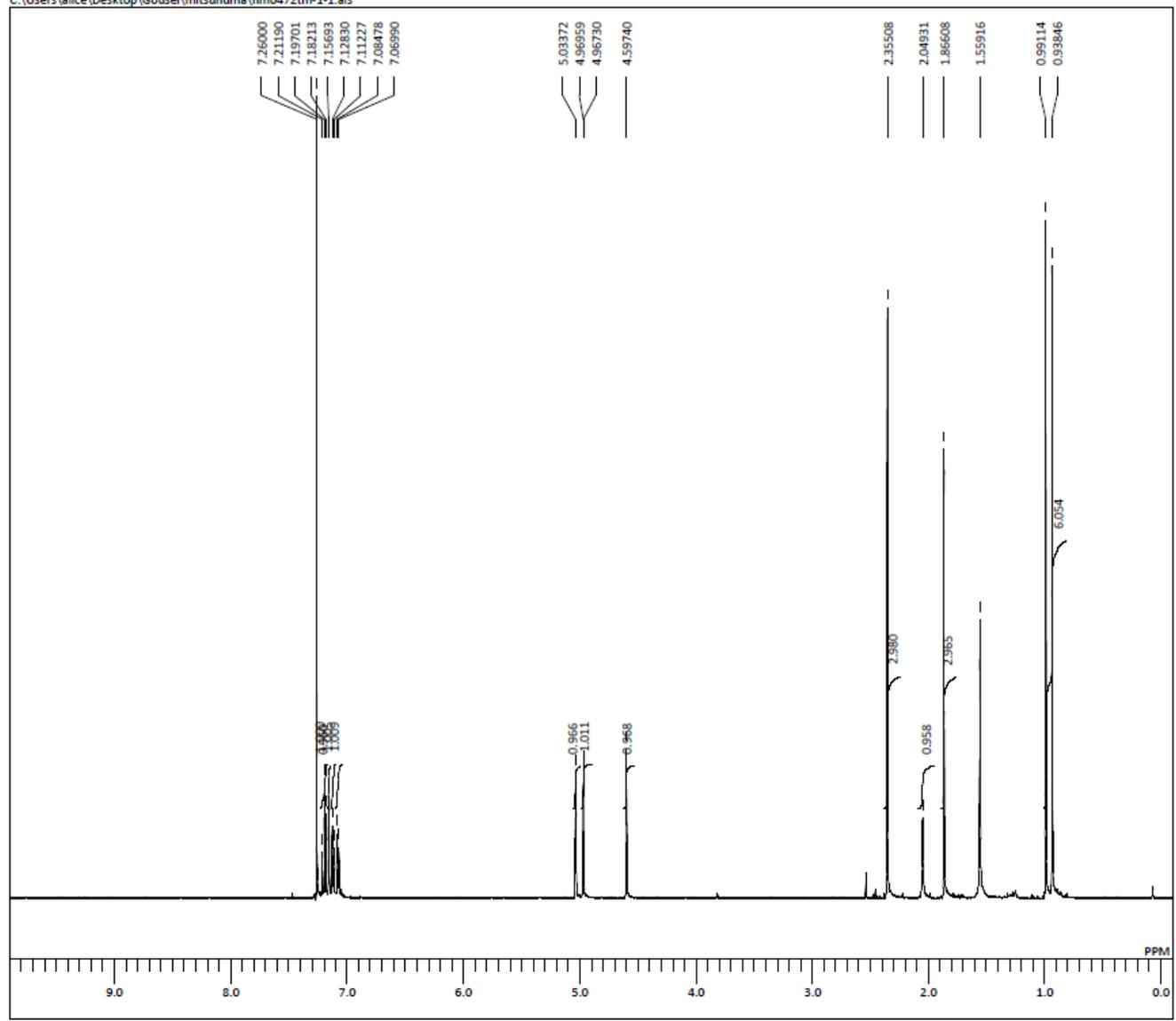


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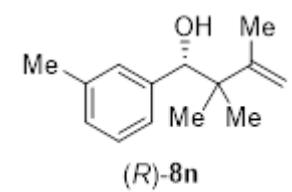


DFILE hm0464tm13c-1-1.als
COMMT
DATIM 11-09-2018 22:50:17
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 98.52 MHz
OBSET 4.64 kHz
OBFIN 8.74 Hz
POINT 26214
FREQU 24630.54 Hz
SCANS 20
ACQTM 1.0643 sec
PD 2.0000 sec
PW1 3.12 usec
IRNUC 1H
CTEMP 20.5 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

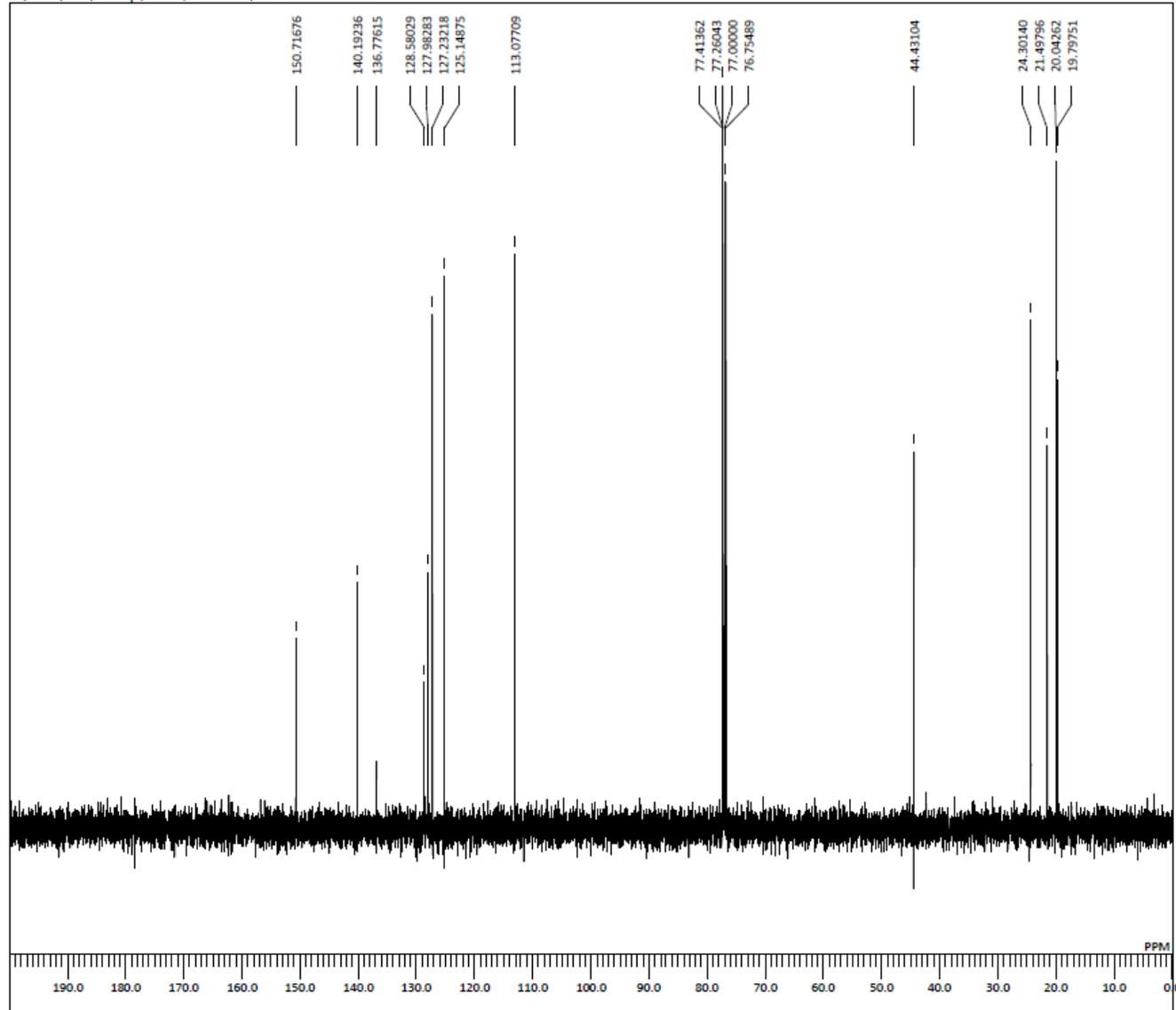
C:\Users\alice\Desktop\Gousei\mitsunuma\hm0472tm-1-1.als



DFILE hm0472tm-1-1.als
COMMT
DATIM 2018-09-13 21:59:42
OBNUC 1H
EXMOD proton.jpg
OBFREQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.4 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

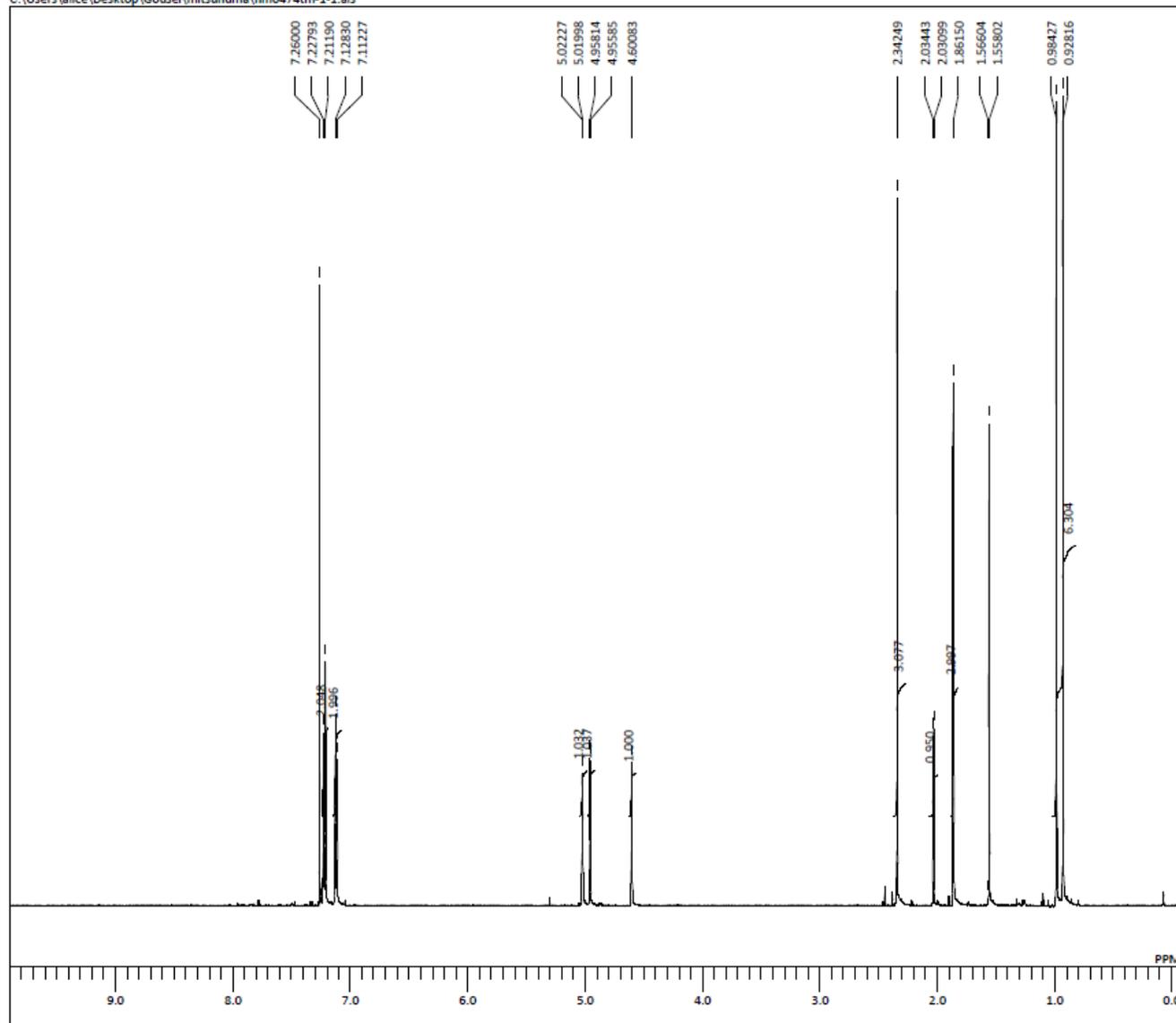


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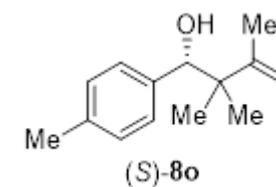


DFILE hm0472tm13C-1-1.als
COMMT
DATIM 2018-09-13 22:16:04
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 1.58 kHz
OBFIN 5.95 Hz
POINT 26214
FREQU 50505.05 Hz
SCANS 24
ACQTM 0.5190 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.3 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 58

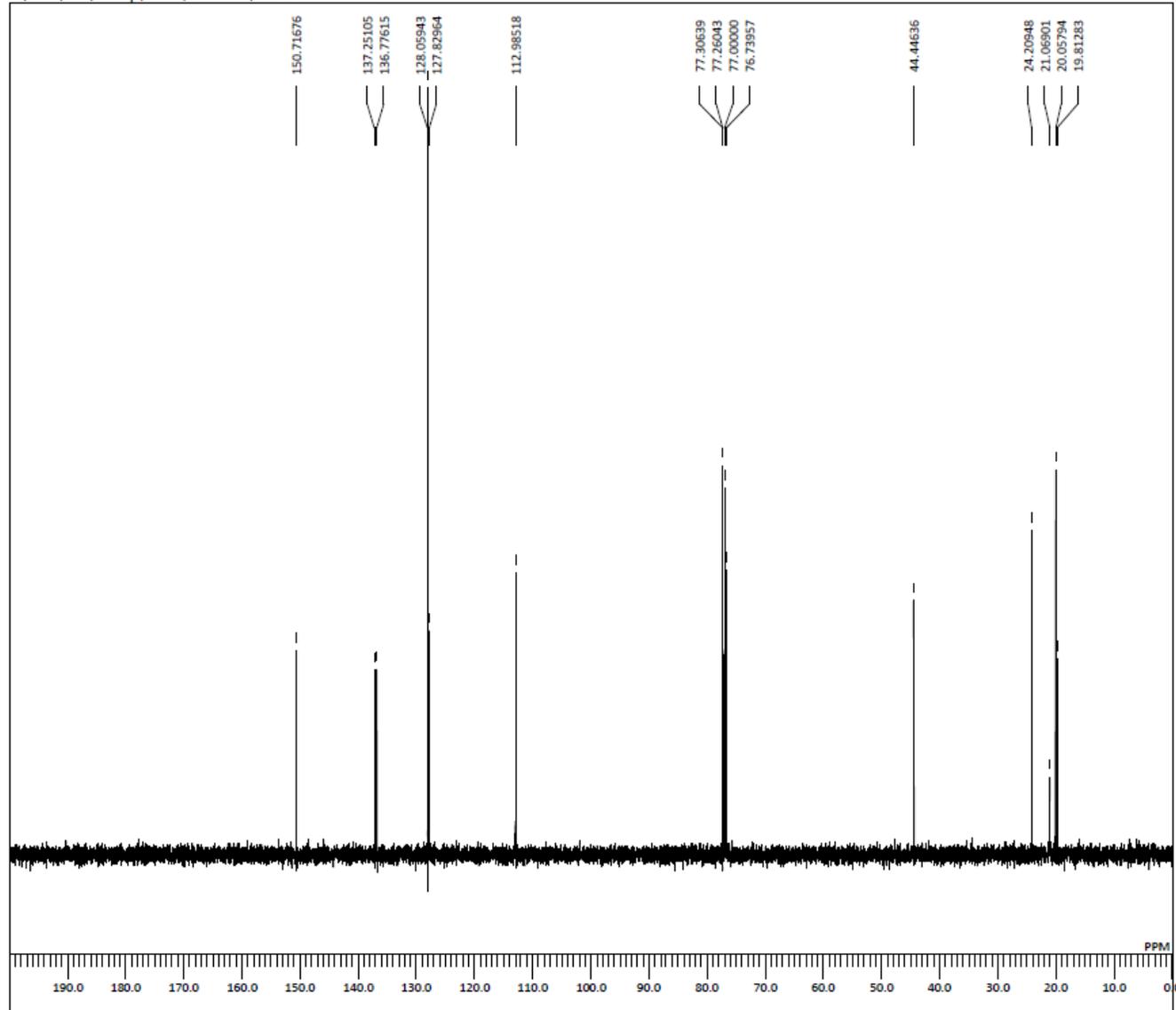
C:\Users\alice\Desktop\Gousei\mitsunuma\hm0474tm-1-1.als



DFILE hm0474tm-1-1.als
COMMT
DATIM 2018-09-13 22:03:22
OBNUC 1H
EXMOD proton.jpg
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.4 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 34

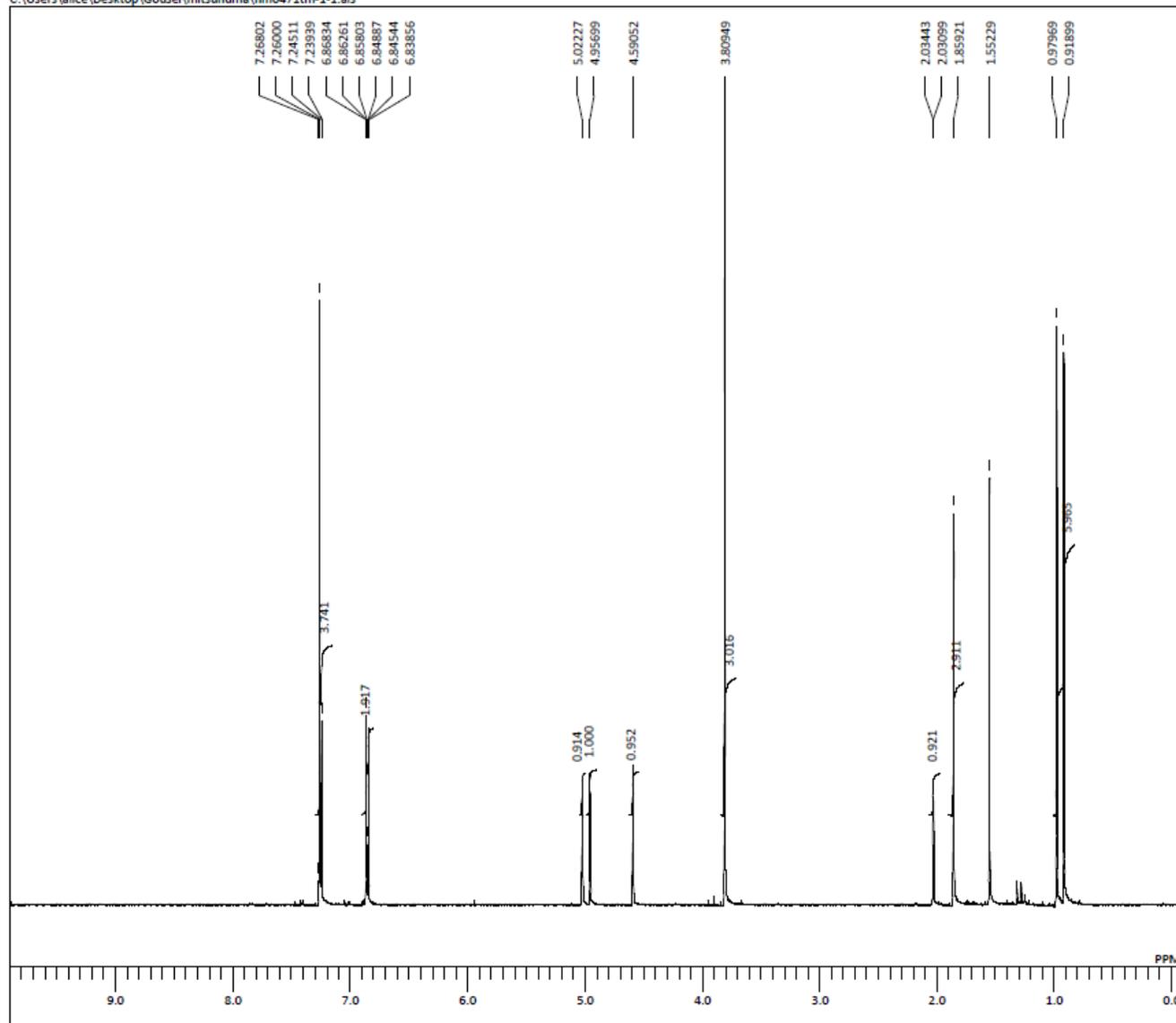


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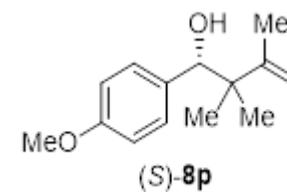


DFILE hm0474tm13C-1-1.als
COMNT
DATIM 2018-09-13 22:19:23
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 1.58 kHz
OBFIN 5.95 Hz
POINT 26214
FREQU 50505.05 Hz
SCANS 25
ACQTM 0.5190 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.5 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 58

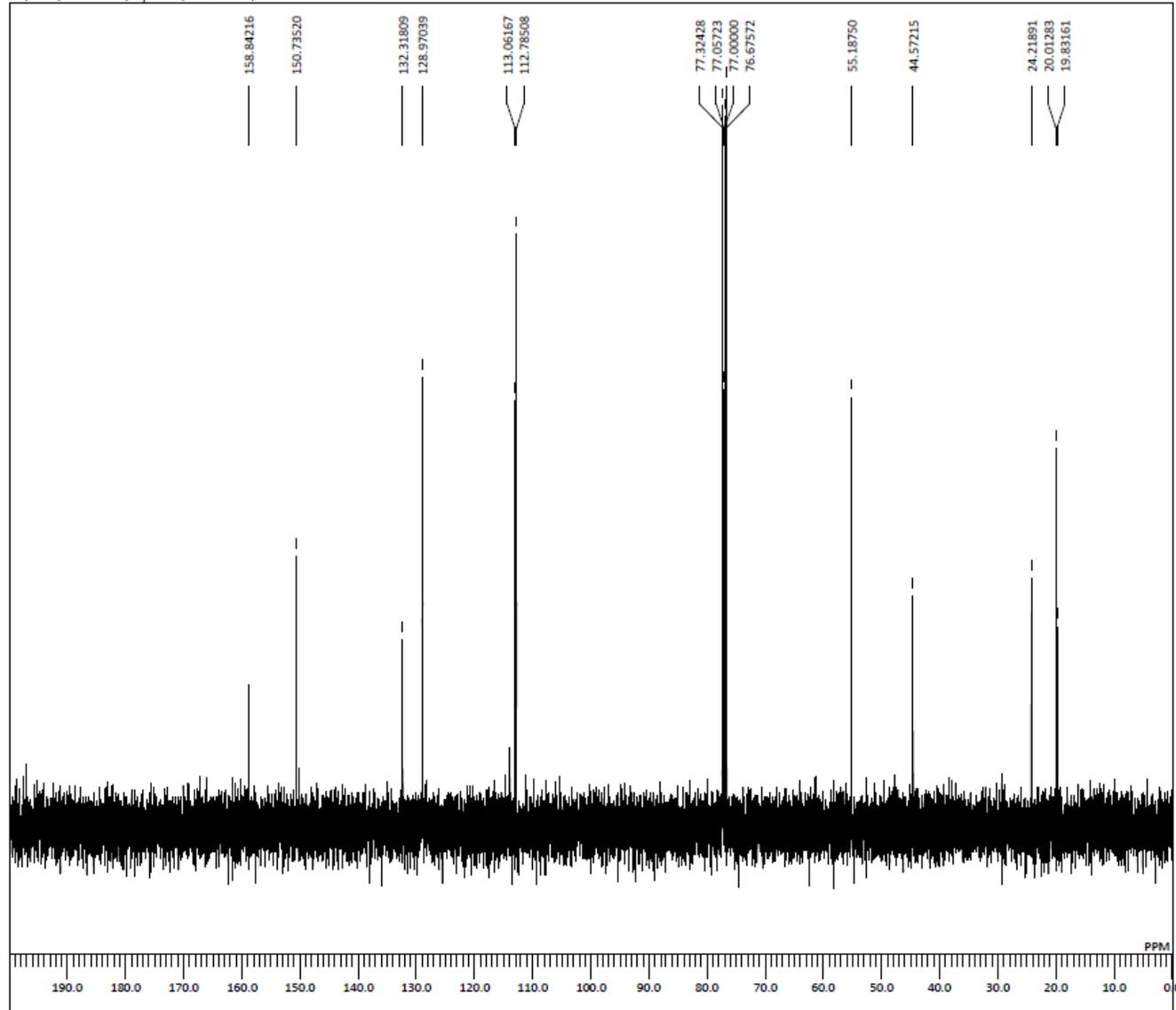
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DFILE hm0471tm-1-1.als
COMMT
DATIM 2018-09-11 21:09:39
OBNUC 1H
EXMOD proton.jpg
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.6 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

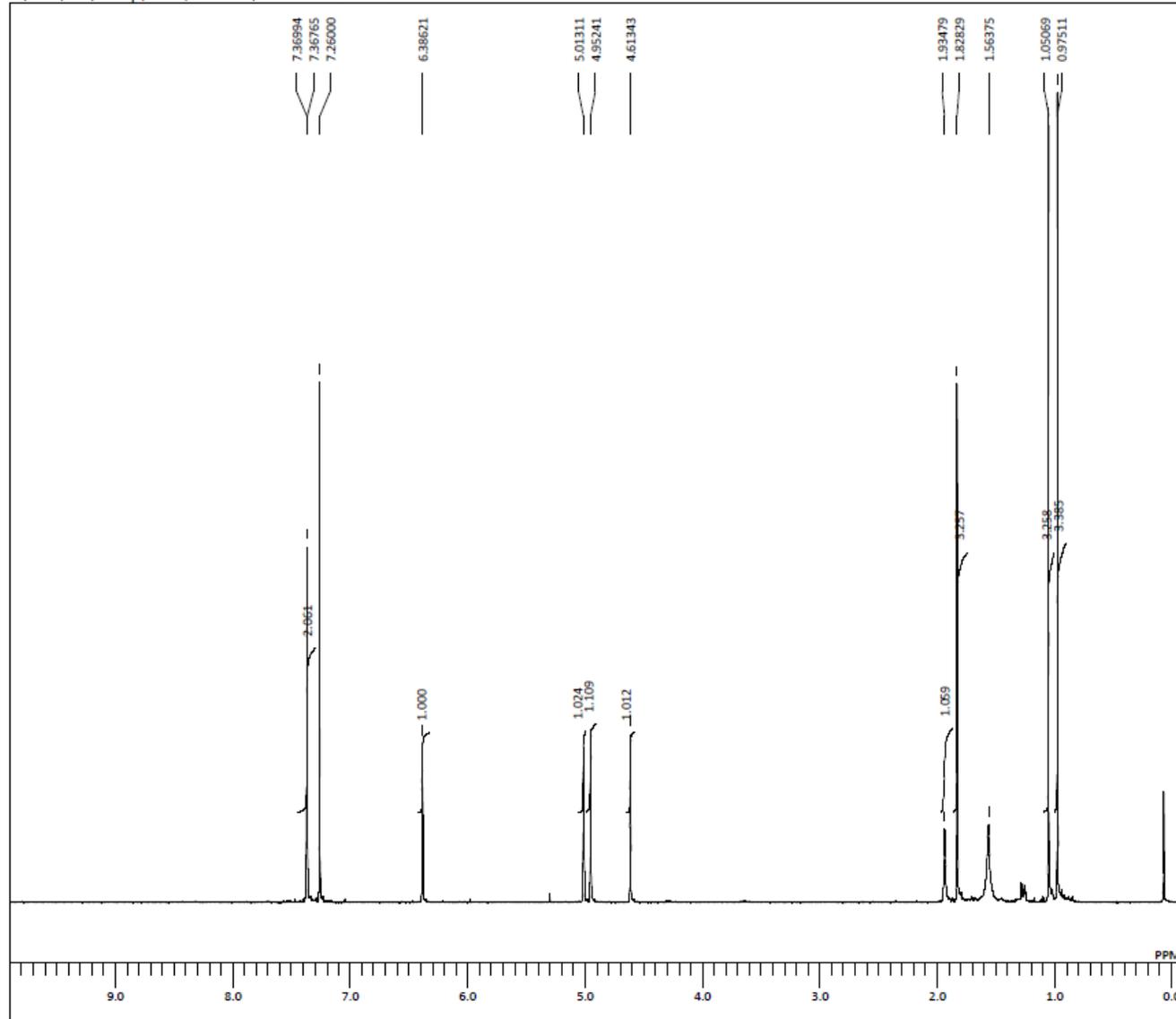


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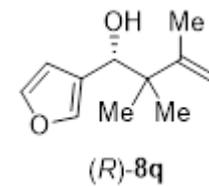


DFILE hm0471tm13c-1-1.als
COMMT
DATIM 11-09-2018 22:58:34
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 98.52 MHz
OBSET 4.64 kHz
OBFIN 8.74 Hz
POINT 26214
FREQU 24630.54 Hz
SCANS 32
ACQTM 1.0643 sec
PD 2.0000 sec
PW1 3.12 usec
IRNUC 1H
CTEMP 20.5 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

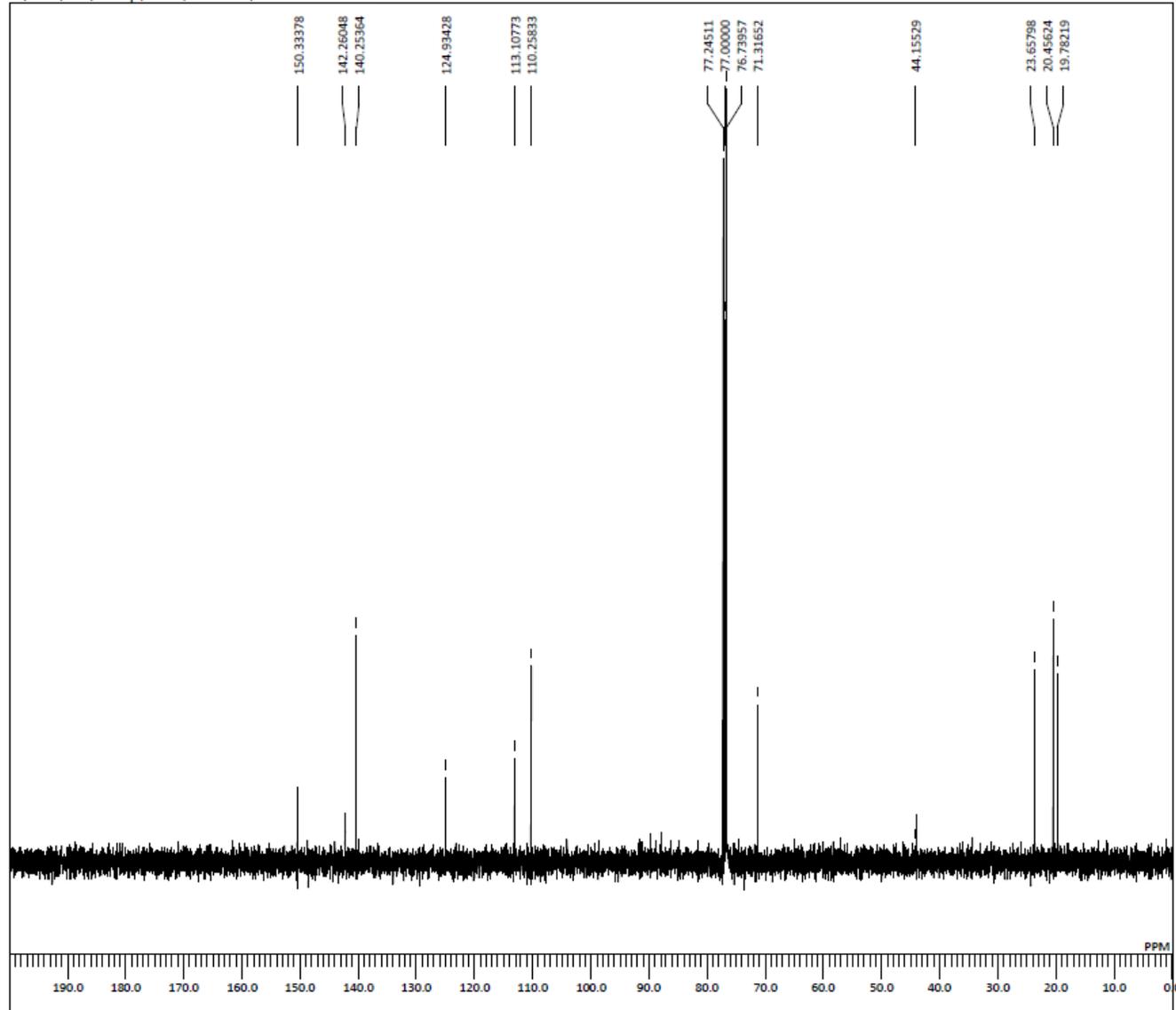
C:\Users\alice\Desktop\Gousei\mitsunuma\hm0565column-1-1.als



DFILE hm0565column-1-1.als
COMNT
DATIM 2018-10-05 22:27:10
OBNUC 1H
EXMOD proton.jpg
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.6 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 34

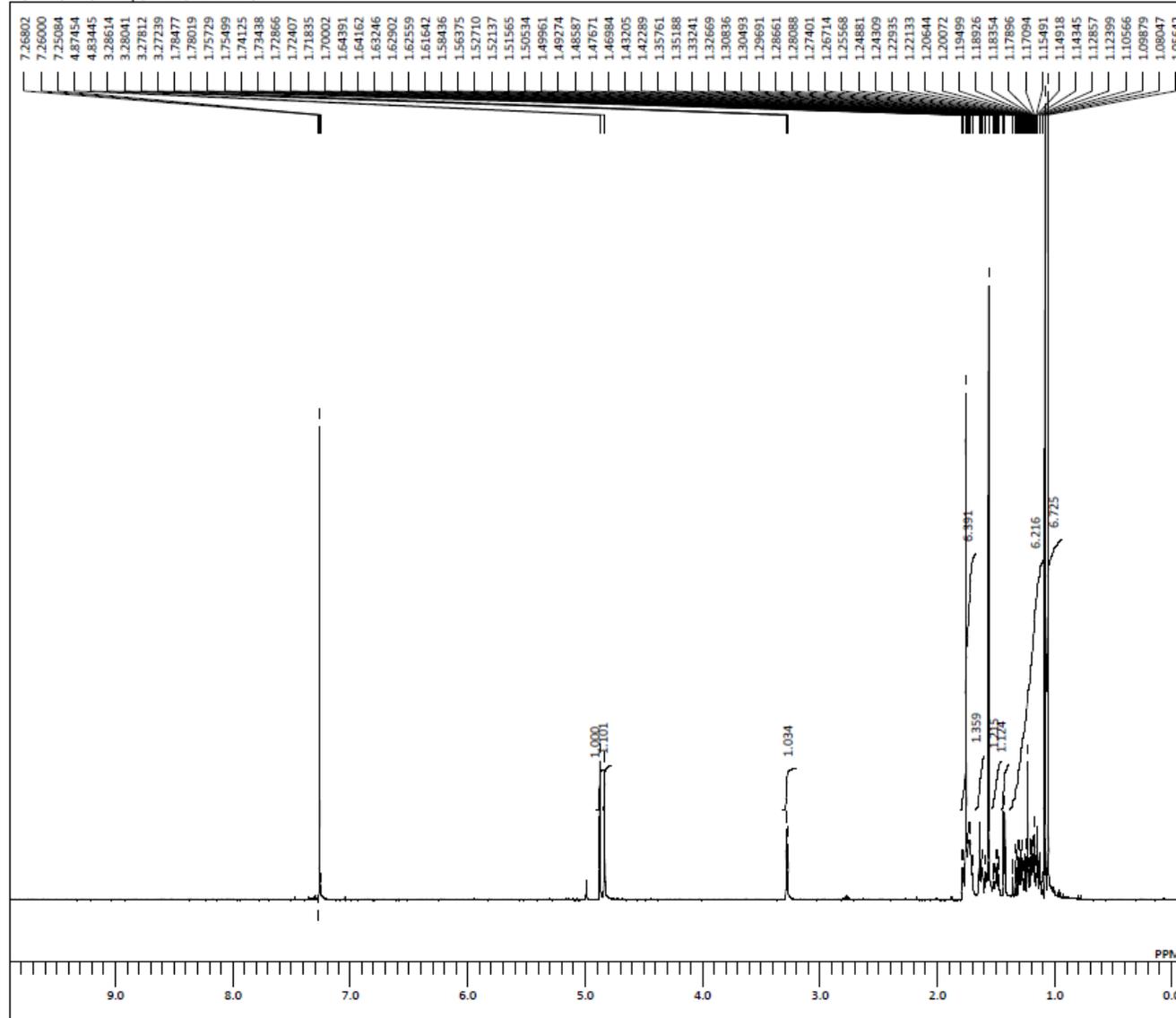


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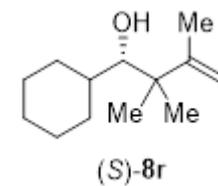


DFILE hm0565tm 13C-1-1.als
COMMT
DATIM 2018-10-05 23:09:42
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 1.58 kHz
OBFIN 5.95 Hz
POINT 26214
FREQU 50505.05 Hz
SCANS 60
ACQTM 0.5190 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.7 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 58

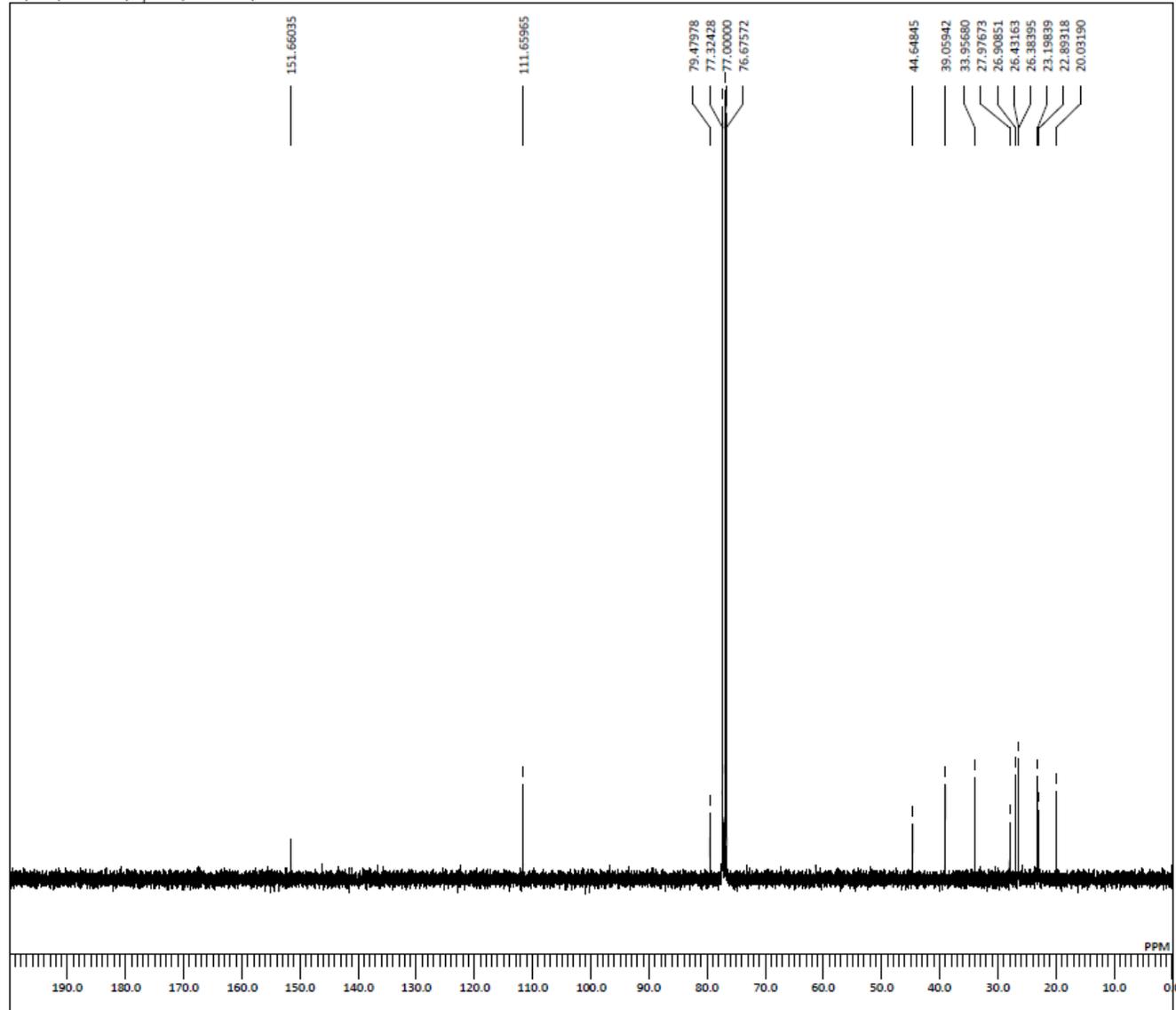
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DFILE hm0494tm-1-1.als
COMNT
DATIM 2018-09-16 23:46:03
OBNUC 1H
EXMOD proton.jxp
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.7 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

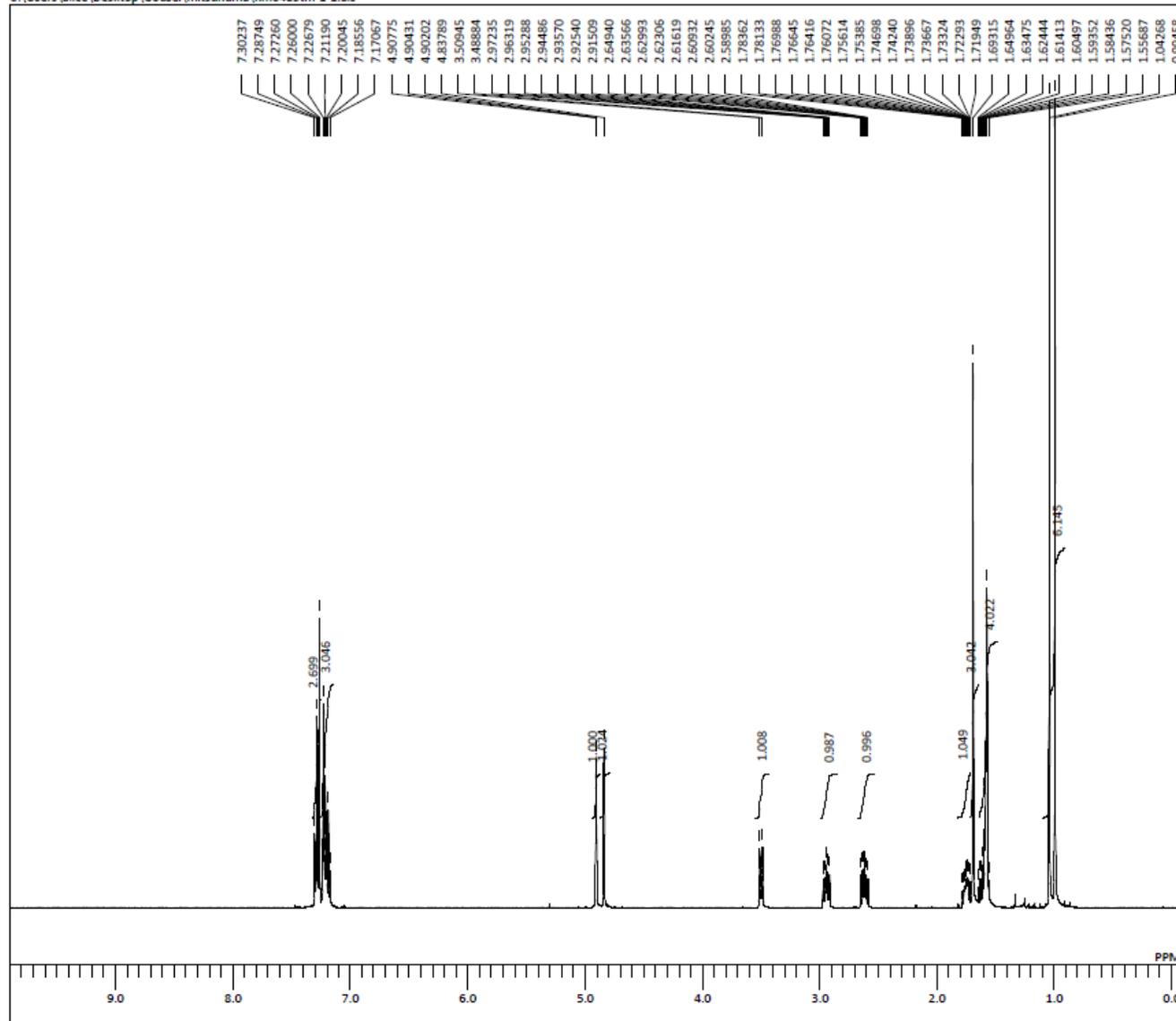


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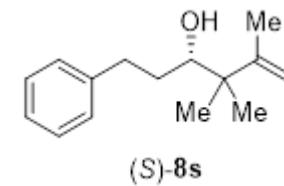


DFILE hm0494tm 13C-1-1.als
COMMT
DATIM 17-09-2018 19:04:55
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 98.52 MHz
OBSET 4.64 kHz
OBFIN 8.74 Hz
POINT 26214
FREQU 24630.54 Hz
SCANS 640
ACQTM 1.0643 sec
PD 2.0000 sec
PW1 3.12 usec
IRNUC 1H
CTEMP 20.5 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

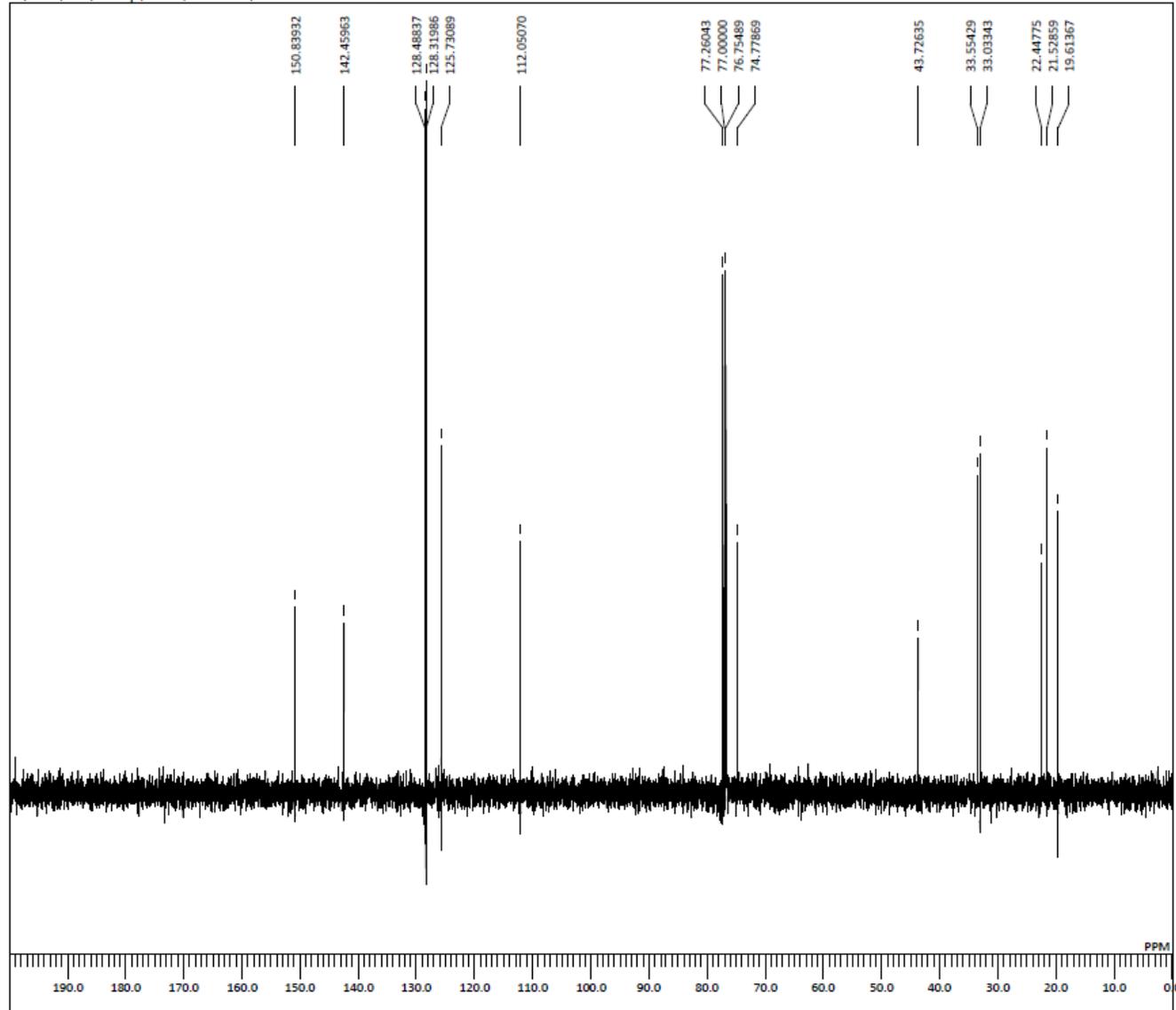
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DFILE hm0485tm-1-1.als
COMMT
DATIM 2018-09-18 10:24:20
OBNUC 1H
EXMOD proton.jxp
OBFREQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.9 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

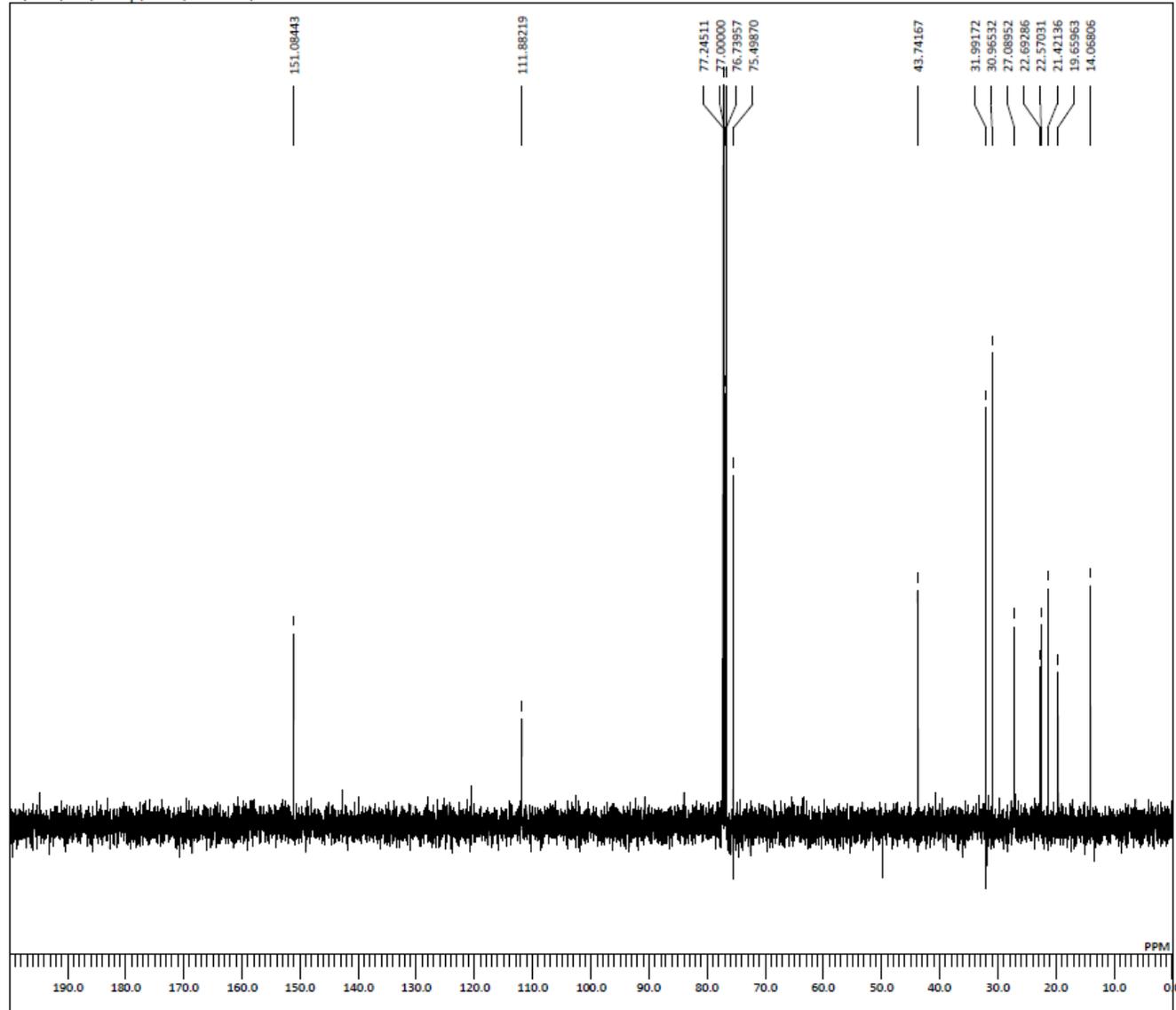


C:\Users\alice\Desktop\Gousei\mitsunuma\hm0485tm 13C-1-1.als



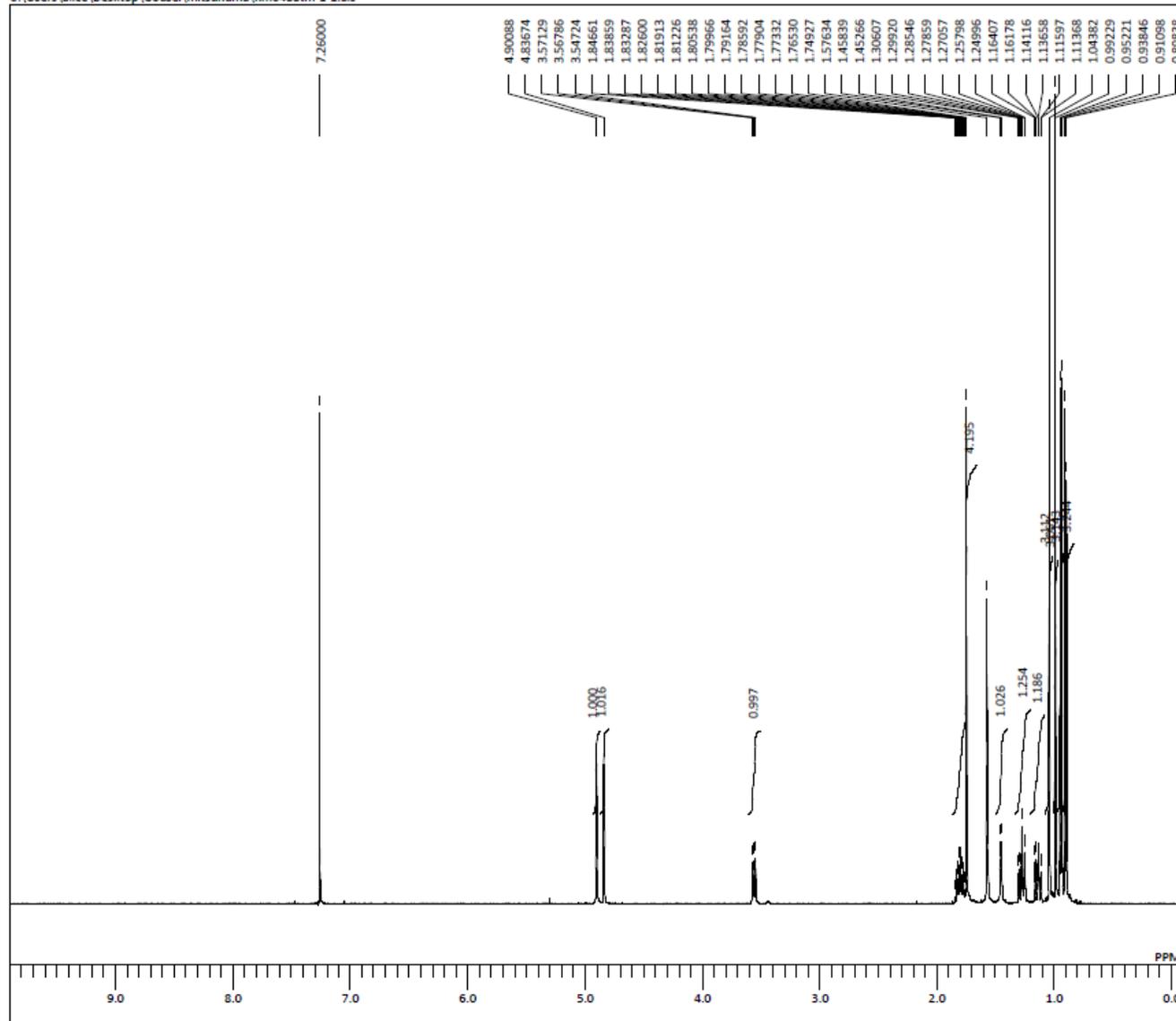
DFILE hm0485tm 13C-1-1.als
COMMT
DATIM 2018-09-18 10:49:26
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 1.58 kHz
OBFIN 5.95 Hz
POINT 26214
FREQU 50505.05 Hz
SCANS 20
ACQTM 0.5190 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.8 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 58

C:\Users\alice\Desktop\Gousei\mitsunuma\hm0487tm 13C-1-1.als

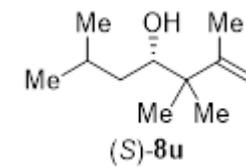


DFILE hm0487tm 13C-1-1.als
COMMT
DATIM 2018-09-18 10:55:58
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 1.58 kHz
OBFIN 5.95 Hz
POINT 26214
FREQU 50505.05 Hz
SCANS 25
ACQTM 0.5190 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.6 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

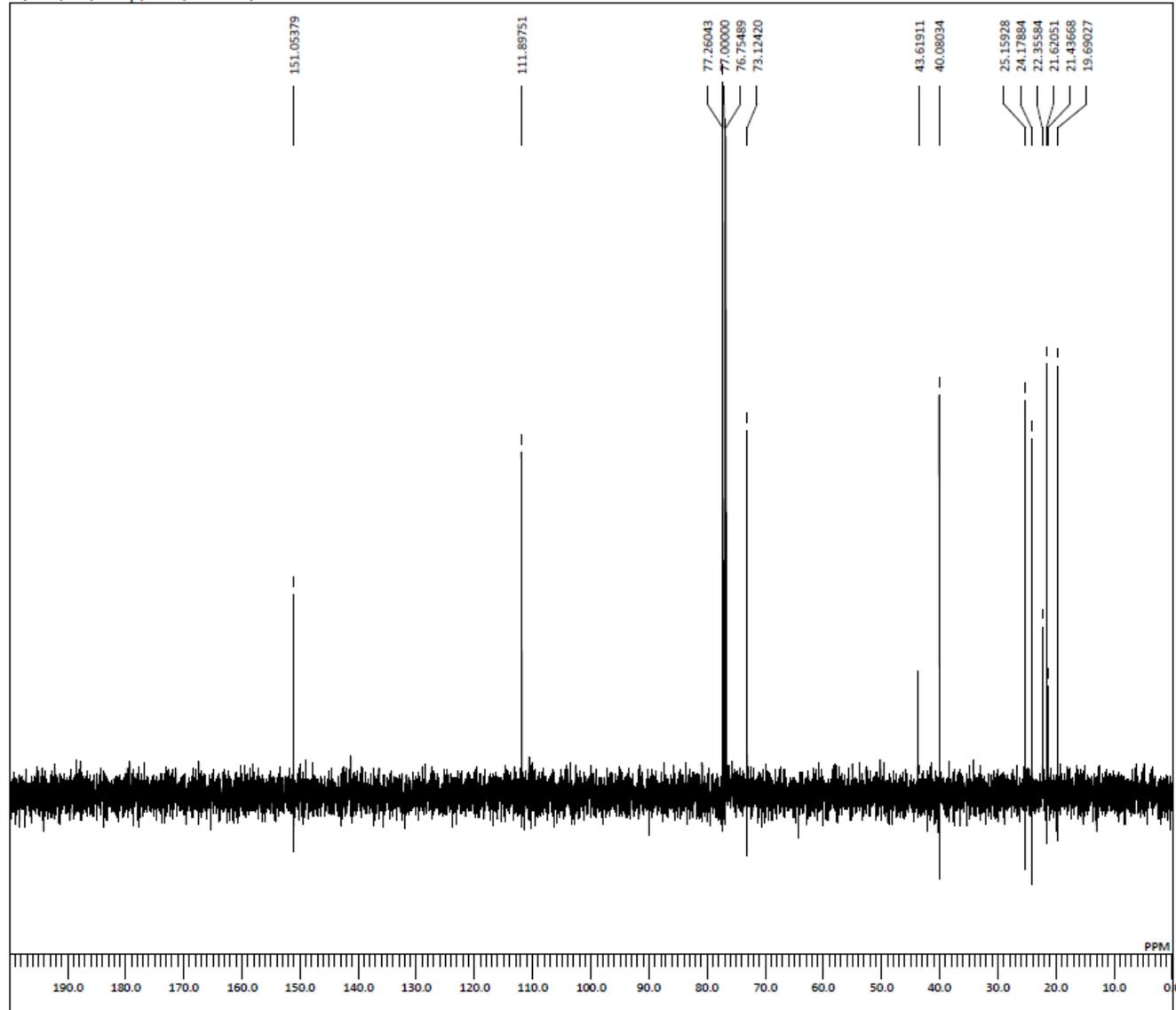
C:\Users\alice\Desktop\Gousei\mitsunuma\hm0488tm-1-1.als



DFILE hm0488tm-1-1.als
COMMT
DATIM 2018-09-18 10:34:60
OBNUC 1H
EXMOD proton.jpg
OBFRQ 500.16 MHz
OBSET 2.41 KHz
OBFIN 6.01 Hz
POINT 13107
FREQU 7507.51 Hz
SCANS 8
ACQTM 1.7459 sec
PD 5.0000 sec
PW1 5.55 usec
IRNUC 1H
CTEMP 21.6 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 32

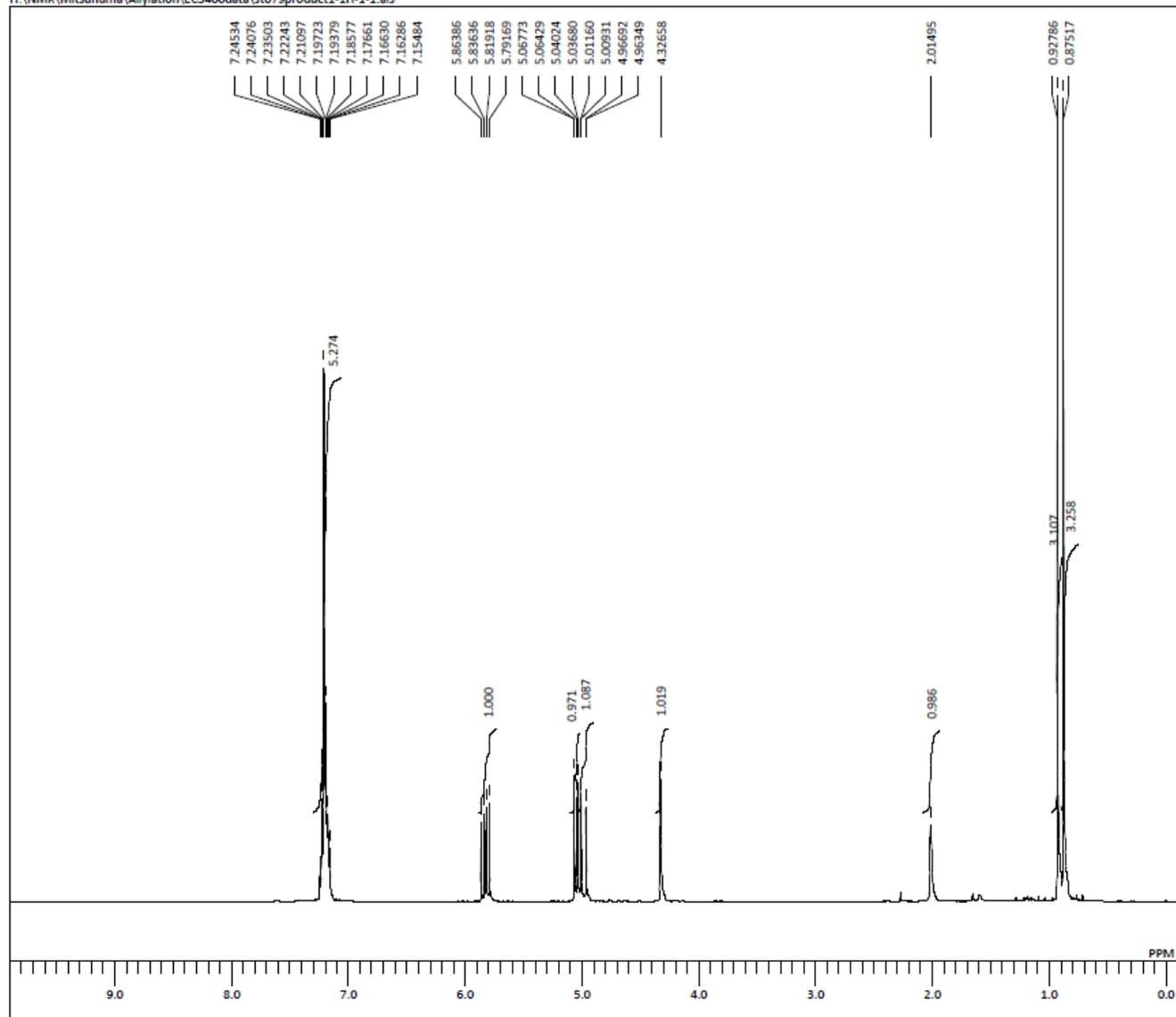


C:\Users\alice\Desktop\Gousei\mitsunuma\hm0488tm 13C-1-1.als

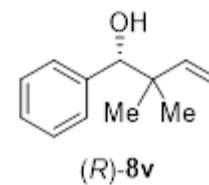


DFILE hm0488tm 13C-1-1.als
COMMT
DATIM 2018-09-18 10:58:48
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 125.77 MHz
OBSET 1.58 kHz
OBFIN 5.95 Hz
POINT 26214
FREQU 50505.05 Hz
SCANS 20
ACQTM 0.5190 sec
PD 2.0000 sec
PW1 3.40 usec
IRNUC 1H
CTEMP 21.3 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

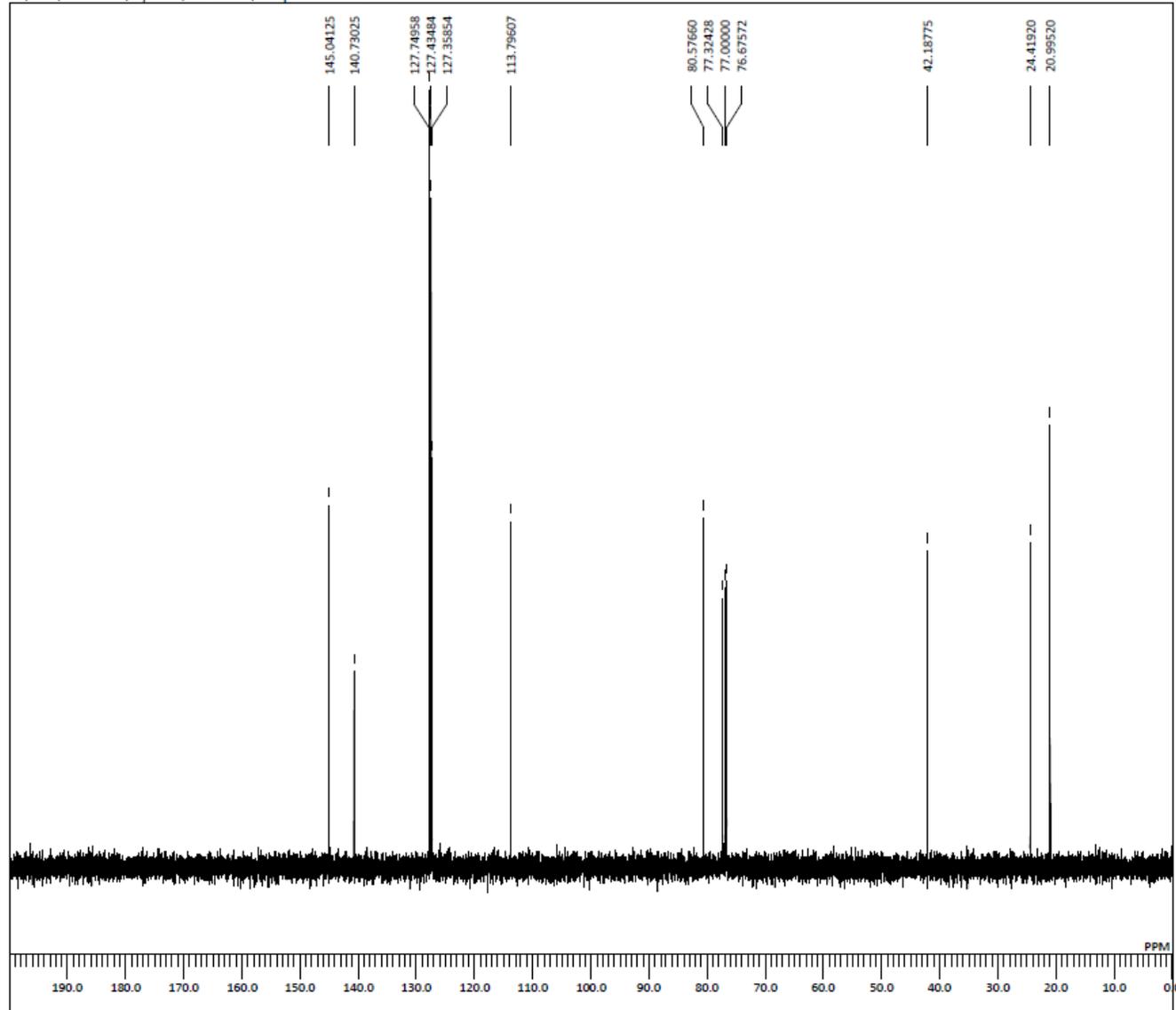
H:\NMR\Mitsunuma\Allylation\ECS400data\st079product1-1H-1-1.als



DFILE st079product1-1H-1-1.als
COMNT
DATIM 01-10-2018 21:37:08
OBNUC 1H
EXMOD proton.jxp
OBFRQ 391.78 MHz
OBSET 8.51 kHz
OBFIN 3.34 Hz
POINT 13107
FREQU 5882.35 Hz
SCANS 8
ACQTM 2.2282 sec
PD 5.0000 sec
PW1 5.22 usec
IRNUC 1H
CTEMP 21.1 c
SLVNT CDCL3
EXREF 0.00 ppm
BF 0.12 Hz
RGAIN 22



H:\NMR\Mitsunuma\Allylation\ECS400data\st079product1-13C-1-1.als



DFILE st079product1-13C-1-1.als
COMMT
DATIM 01-10-2018 21:33:30
OBNUC 13C
EXMOD carbon.jpg
OBFRQ 98.52 MHz
OBSET 4.64 kHz
OBFIN 8.74 Hz
POINT 26214
FREQU 24630.54 Hz
SCANS 32
ACQTM 1.0643 sec
PD 2.0000 sec
PW1 3.12 usec
IRNUC 1H
CTEMP 21.1 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60