iRGD synergizes with PD-1 knockout immunotherapy by enhancing lymphocyte infiltration in gastric cancer

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Supplementary Figure 1 Cell surface modification with DSPE-PEG-iRGD. a. ¹H NMR spectrum of iRGD, DSPE-PEG-Mal and DSPE-PEG-iRGD. b. Mean fluorescence intensity of T cells alone (grey) and the cells incubated with iRGD-FAM (blue) and DSPE-PEG-iRGD-FAM (red). Student's *t* test. ***p < 0.001. c. Analysis of mean fluorescence intensity of cells treated with different ratios of DSPE-PEG-iRGD. Data represent mean \pm s.e.m.; n = 3.



Supplemeta PEG-based modification on T cells in vitro. a. No r n in T, T+iRGD and T-iRGD group. In vitro sed to iRGD or DSPE-PEG-iRGD solutions cultured activ at the concen irs, the morphological change was analyzed by optical microscopy. Scale bar, 100 µm. **b**. Vitality of cells after exposure to iRGD or DSPE-PEG-i 143 **1**44 t concentrations. c-e. Cytotoxicity, cellular l response and specific CTLs of different format. LMP2Aspecific CTL g of HLA-A24 positive human PBMC with HLA-A24-restricted LMP2A peptide (TYGPVFMCL) loaded DCs in the presence of IL-2, IL-7 and IL-15 for 14 days. And then, EBV-LMP2A-CTLs were incubated with iRGD or DSPE-PEG-iRGD solutions at the concentration of 5 µg ml⁻¹ for 30 minutes and analyzed. LMP2A-specific CTLs of different format were incubated with CFSE labeled HLA-A24 positive SNU719 cells at effector-to-target ratio (E:T) of 0:1, 5:1, 10:1, 20:1 and 40:1 respectively, PI was added 6 hours after incubation and the percentage of dead cells was analyzed by flow cytometry (c). CTLs were stimulated with LAM2A loaded autologous DCs for 20 hours, culture supernatant was collected and the cytokine level was analyzed by CBA Human IFN- γ kit (d). The phenotype of the in vitro cultured T cells of different format were analyzed by flow cytometry (e). Data represent mean \pm s.e.m.; n = 3. Student's *t* test.



Supplementary Figure 3 Cell expression of $\alpha\nu\beta3$, $\alpha\nu\beta5$ and NRP-1. $\alpha\nu\beta3$, anti- $\alpha\nu\beta5$ and NRP-1 expression in tumor cells was determined by flow cytometry. Integrin $\alpha\nu\beta3$ was detected using FITC-conjugated mouse anti-human $\alpha\nu\beta3$ monoclonal antibody, and integrin $\alpha\nu\beta5$ was detected using FITC-conjugated mouse anti-human $\alpha\nu\beta5$ monoclonal antibody. The matched isotype control was FITC-conjugated mouse IgG1 κ . NRP-1 was detected using PE-conjugated mouse anti-human NRP-1 monoclonal antibody and an isotype control.



Supplementary Figure 4 The establishment of peritoneal metastasis tumor model. 10⁶ MKN45 cells were injected intraperitoneally and 20 days later, the formation of tumor nodules were evaluated by both dissection (**a**) and T2-weighted MR imaging (**b**, **c**). The white arrow and the red circle indicate the tumor nodules.



Supplementary Figure 5 iRGD-mediated *in vivo* **tumor penetration of T cells in intraperitoneal tumor model.** Mice bearing intraperitoneal tumor were treated with 50ug anti-NRP-1 antibody or control sheep anti-IgG 15 min prior to T cells injection, tumor nodules were analyzed 3 h post T cells injection.



Supplementary Figure 6 Enhanced tumor specific delivery of lymphocytes 24 h post intraperitoneal injection. a. *Ex vivo* images of tumor, liver and spleen at 24 h post intraperitoneal T cells injection. top layer: small tumor nodules (diameter < 3mm); second layer: big tumor nodules (diameter > 3mm). White dashed lines, peritoneal tumors. b. Quantification of the fluorescent signal in tumors and control organs at 24 h post lymphocytes injection. Data are represented as mean \pm s.e.m.; n = 3. Student's *t* test. **p < 0.05, *p < 0.01. c. Immunohistochemistry analysis of resected tumor nodules at 24 h post intraperitoneal injection. T cells were stained with anti-CD3. Scale bar, 100 µm.



Supplementary Figure 7 Cytotoxicity of iRGD modified T cell on MKN45 in vitro. Activated T cells of different format were incubated with CFSE labeled MKN45 cells at effector-to-target ratio (E:T) of 0:1, 5:1,10:1,20:1 and 40:1 respectively, PI was added 6 hours after incubation and the percentage of dead cells was analyzed by flow cytometry.



Supplementary Figure 8 Safety profile evaluation of intraperitoneal T cells injection. a. Body weight change of mice treated by intraperitoneal injections of PBS, T, T+iRGD or T-iRGD. b. Main organs were dissected for H&E staining. Scale bar, 100 μ m. c. Assessment of liver and kidney function at the end of treatment. Data are represented as mean \pm s.e.m.; n = 8. Student's *t* test. He, Heart; Li, Liver; Sp, Spleen; Lu, Lung; Ki, Kidney; AST, Aspartate aminotransferase; ALT, Alanine aminotransferase; BUN, Blood urea nitrogen; Cr, Creatinine.



Supplementary Figure 9 Tumor target infiltration of iRGD modified lymphocytes in a SNU719 subcutaneous tumor model. a. *In vivo* imaging of SNU719 tumorbearing mice at 6, 24, 72 and 144 h after intravenous injection of DiR labeled T cells. White dashed lines, subcutaneous tumors. b. *Ex vivo* imaging of tumors and other organs at 144 h after T cells injection. c. Semiquantification of T cells biodistribution in mice organs collected at 144 h post T cells injection. Results are expressed as mean fluorescence intensity \pm s.e.m.; n = 5. Student's *t* test. *p < 0.05, ***p < 0.001. d. Confocal images of frozen tumor sections at 24 h post T cells transfusion. T cells were labeled with CFSE before injection. Tumor blood vessels were labeled with CD31.T cells, green; vessel, red; nucleus, blue. Scale bar, 100 µm.



Supplementary Figure 10 Body weight change of SNU719 tumor-bearing mice that received the indicated treatments. Data are represented as mean \pm s.e.m.; n = 7.



Supplementary Figure 11 Representative Sequential Gating Strategies for Flow Cytometry Analysis. a. Gating strategy to identify the phenotype of the in vitro cultured T cells presented in Supplementary Figure 2e. The gating strategy depicted here is also applied to experiments from which data is shown in Figure 1c-e, Supplementary Figure 2b and Supplementary Figure 3. b. Gating strategy to determine the percentage of dead cell presented in Supplementary 2c and Supplementary 7. c. Gating strategy to determine the IFN- γ level presented in Supplementary 2d.