Investigating the promiscuity of the chloramphenicol nitroreductase from *Haemophilus influenzae* **towards the reduction of 4-nitrobenzene derivatives**

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Materials and instrumentation for chemistry and biological studies. 4-Chloro-2 fluorobenzylalcohol was purchased form Matrix Scientific (Columbia, SC, USA). All other chemicals for the synthesis of 4-nitrobenzyl ethers, esters, and thioethers, as well as other nitrocontaining molecules were purchased from Sigma-Aldrich (St. Louis, MO, USA) and used without any further purification. Chemical reactions were monitored by thin-layer chromatography (TLC) using Merck, Silica gel 60 F_{250} plates. All ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra were recorded on a Varian 400 MHz spectrometer. Chemical shifts (δ) and coupling constant (J) are provided in parts per million (ppm) and Hertz (Hz), respectively. Conventional abbreviations used for signal shape are as follows: br s = broad singlet; $s =$ singlet; $d =$ doublet; $t =$ triplet; $q =$ quartet; $p =$ pentet; sx = sextet; sp = septet; m = multiplet. Monitoring of nitroreduction reactions was performed on a multimode SpectraMax M5 plate reader. Reversed-phase high-pressure liquid chromatography (RP-HPLC) assays were performed on an Agilent Technologies 1260 Infinity HPLC system using the following general method 1: Flow rate = 1 mL/min; $\lambda = 254$ nm; column $=$ Vydac 201SP C18, 250 mm \times 4.6 mm, 90 Å, 5 µm. Eluents: A = H₂O + 0.1% TFA; B = MeCN. Gradient profile: starting from 5% B, increasing to 100% B over 10 min, holding at 100% B from 10 to 16 min, decreasing from 100% B to 5% B from 16 to 20 min. Prior to each injection the HPLC column was equilibrated for 15 min with 5% B. Liquid chromatography-mass spectrometry (LCMS) experiments were performed on an Agilent 1200 Series Quaternary LC system and with an Eclipse XDB-C18 column (150 mm \times 4.6 mm, 80 Å, 5 µm) equipped with an Agilent 6120 Quadropole MSD mass spectrometer (Agilent Technologies, Santa Clara, CA). All compounds were at least 95% pure.

CHEMISTRY

General procedure for the synthesis of 4-nitrobenzyl ethers (26a,b,d-k,o,q). 4- Nitrobenzylbromide (216 mg, 1 mmol) was added to a mixture of primary or secondary alcohol (1.1 mmol) and Ag₂O (340 mg, 1.5 mmol) in dry Et₂O (10 mL). The reaction mixture was heated at reflux temperature for 12 h. After cooling to room temperature, the solid was filtered off and washed with Et₂O (2×25 mL). The organic layer was collected, concentrated under reduced pressure, and purified by column chromatography ($SiO₂$, hexanes: $CH₂Cl₂/1:1$). 4-Nitrobenzylethyl ether $(26a)$,¹ 4-nitrobenzylpropyl ether $(26b)$,² 4-nitrobenzylbenzyl ether $(26g)$,³ 4-bromobenzyl 4-nitrobenzyl ether (26h),⁴ di(4-nitrobenzyl) ether (26j),⁵ and 4-nitrobenzylphenyl ether (26q),^{3,6}

have previously been reported. The physical appearance and spectral data of all ethers are listed below:

4-Nitrobenzylethyl ether (26a). Yellow solid (91 mg, 50%). ¹H NMR (CDCl₃, Fig. S1) δ 8.14 (d, *J* = 8.4 Hz, 2H), 7.47 (d, *J* = 8.4 Hz, 2H), 4.57 (s, 2H), 3.56 (q, *J* = 7.6 Hz, 2H), 1.25 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (CDCl₃, Fig. S2) δ 146.4, 127.7, 127.6, 123.5, 71.3, 66.4, 15.1.

4-Nitrobenzylpropyl ether (26b). Off-white solid $(51 \text{ mg}, 26\%)$. ¹H NMR (CDCl₃, Fig. S3) δ 8.17 $(d, J = 8.0 \text{ Hz}, 2\text{H})$, 7.48 $(d, J = 8.0 \text{ Hz}, 2\text{H})$, 4.58 (s, 2H), 3.47 (t, $J = 6.8 \text{ Hz}, 2\text{H}$), 1.65 (q, $J = 7.2$ Hz, 2H), 0.95 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, Fig. S4) δ 146.5, 130.9, 127.6, 123.5, 72.7, 71.5, 22.9, 10.6.

1-*[(Cyclobutylmethoxy)methyl]-4-nitrobenzene (26d)*. Colorless oil (70 mg, 31%). ¹H NMR $(CDCl₃, Fig. S5)$ δ 8.18 (d, *J* = 8.2 Hz, 2H), 7.48 (d, *J* = 8.2 Hz, 2H), 4.58 (s, 2H), 3.46 (d, *J* = 6.8 Hz, 2H), 2.61 (p, $J = 7.2$ Hz, 1H), 2.07-2.03 (m, 2H), 1.92-1.72 (m, 4H); ¹³C NMR (CDCl₃, Fig. S6) d 147.2, 146.5, 127.6, 123.5, 75.5, 71.6, 35.0, 25.0, 18.6.

1-*[(Cyclopentylmethoxy)methyl]-4-nitrobenzene (26e)*. Colorless oil (83 mg, 35%). ¹H NMR $(CDCl_3$, Fig. S7) δ 8.17 (d, $J = 7.2$ Hz, 2H), 7.49 (d, $J = 7.2$ Hz, 2H), 4.59 (s, 2H), 3.37 (d, $J = 5.6$ Hz, 2H), 2.21 (sp, *J* = 6.0 Hz, 1H), 1.76 (m, 2H), 1.52 (m, 4H), 1.26 (m, 2H); 13C NMR (CDCl3, Fig. S8) d 147.2, 146.6, 127.6, 123.5, 75.6, 71.6, 39.5, 29.6, 25.4.

1-[(Cyclohexylmethoxy)methyl]-4-nitrobenzene (26f). Colorless oil (178 mg, 71%). ¹H NMR $(CDCl₃, Fig. S9)$ δ 8.15 (d, *J* = 8.8 Hz, 2H), 7.46 (d, *J* = 8.8 Hz, 2H), 4.56 (s, 2H), 3.29 (d, *J* = 6.4 Hz, 2H), 1.78-1.60 (m, 6H), 1.21-1.11 (m, 3H), 0.95 (m, 2H); ¹³C NMR (CDCl₃, Fig. S10) δ 147.2, 146.6, 127.5, 123.5, 76.9, 71.6, 38.1, 30.0, 26.5, 25.8.

4-Nitrobenzylbenzyl ether (26g). Note: For the preparation of compound **1f**, 4-nitrobenzyl alcohol and benzyl bromide were used instead of 4-nitrobenzyl bromide and benzyl alcohol. Pale yellow solid (122 mg, 50%). ¹H NMR (CDCl₃, Fig. S11) δ 8.19 (d, *J* = 8.8 Hz, 2H), 7.52 (d, *J* = 8.8 Hz, 2H), 7.33 (m, 5H), 4.65 (s, 2H), 4.62 (s, 2H); 13C NMR (CDCl3, Fig. S12) d 147.3, 146.0, 137.6, 128.6, 128.0, 127.80, 127.76, 123.6, 72.8, 70.8.

4-bromobenzyl 4-nitrobenzyl ether (26h). White solid (315 mg, 50%). ¹H NMR (CDCl₃, Fig. S13) d 8.20 (d, *J* = 8.0 Hz, 2H), 7.50 (d, *J* = 8.0 Hz, 2H), 7.48 (d, *J* = 8.4 Hz, 2H), 7.23 (d, *J* = 8.4 Hz, 2H), 4.62 (s, 2H), 4.55 (s, 2H); 13C NMR (CDCl3, Fig. S14) d 147.4, 145.6, 136.5, 131.7 (2 carbons), 129.3 (2 carbons), 127.7 (2 carbons), 123.7 (2 carbons), 121.9, 72.0, 70.9.

4-Chlorobenzyl 4-nitrobenzyl ether (26i). Yellow oil (121 mg, 44%). ¹H NMR (CDCl₃, Fig. S15) d 8.20 (d, *J* = 8.8 Hz, 2H), 7.50 (d, *J* = 8.4 Hz, 2H), 7.33 (d, *J* = 8.8 Hz, 2H), 7.29 (d, *J* = 8.8 Hz, 2H), 4.62 (s, 2H), 4.56 (s, 2H); 13C NMR (CDCl3, Fig. S16) d 147.4, 145.6, 136.0, 133.7, 129.0 (2 carbons), 128.7 (2 carbons), 127.7 (2 carbons), 123.7 (2 carbons), 72.0, 70.9.

 $Di(4-nitrobenzyl)$ ether (26j). Off-white solid (130 mg, 45%). ¹H NMR (CDCl₃, Fig. S17) δ 8.22 (d, $J = 8.4$ Hz, 4H), 7.54 (d, $J = 8.4$ Hz, 4H), 4.72 (s, 4H); ¹³C NMR (CDCl₃, Fig. S18) δ 147.5, 145.1, 127.8, 123.7, 71.5.

4-Chloro-2-fluorobenzyl 4-nitrobenzyl ether (26k). White solid $(8 \text{ mg}, 3\%)$. ¹H NMR (CDCl₃, Fig. S19) d 8.20 (d, *J* = 8.4 Hz, 2H), 7.50 (d, *J* = 8.4 Hz, 2H), 7.37 (m, 1H), 7.12 (m, 2H), 4.66 (s, 2H), 4.61 (s, 2H); 13C NMR (CDCl3, Fig. S20) d 161.73-159.24 (1 *JC-F* = 249.7 Hz), 147.5, 145.4, 134.73-134.64 (${}^{3}J_{C-F}$ = 9.4 Hz), 130.81-130.75 (${}^{3}J_{C-F}$ = 5.3 Hz), 127.7 (2 carbons), 124.67-124.64 $(^{4}J_{C\text{-}F} = 3.8 \text{ Hz})$, 123.7 (2 carbons), 123.44-123.29 ($^3J_{C\text{-}F} = 14.4 \text{ Hz}$), 116.34-116.10 ($^2J_{C\text{-}F} = 29.4$ Hz), 71.22 , $65.85-65.81$ $(^3J_{C-F} = 3.8$ Hz).

1-*[(Cyclopentyloxy)methyl]-4-nitrobenzene (260)*. Colorless oil (70 mg, 31%). ¹H NMR (CDCl₃, Fig. S21) d 8.18 (d, *J* = 8.0 Hz, 2H), 7.49 (d, *J* = 8.0 Hz, 2H), 4.55 (s, 2H), 4.01 (s, 1H), 1.73 (m, 6H), 1.55 (m, 2H); ¹³C NMR (CDCl₃, Fig. S22) δ 147.2, 146.9, 127.5, 123.5, 81.7, 69.5, 32.3, 23.5.

4-*Nitrobenzylphenyl ether (26q)*. White solid (141 mg, 62%). ¹H NMR (CDCl₃, Fig. S23) δ 8.22 (d, *J* = 8.4 Hz, 2H), 7.60 (d, *J* = 8.4 Hz, 2H), 7.33 (t, *J* = 7.8 Hz, 2H), 7.01 (m, 3H), 5.17 (s, 2H); ¹³C NMR (CDCl₃, Fig. S24) δ 158.1, 147.5, 144.7, 129.7, 127.6, 123.8, 121.5, 114.8, 68.5.

General procedure for the synthesis of 4-nitrobenzyl esters (27c-i,k-n). The carboxylic acid (1.1 mmol) and K_2CO_3 (150 mg, 1.1 mmol) were added to acetone (15 mL). The reaction mixture was stirred for 10 min, and then the 4-nitrobenzylbromide (216 mg, 1 mmol) was added portionwise. The reaction mixture was continuously stirred at 50 \degree C for 6 h, and then cooled down to room temperature. The inorganic salts were removed by filtration. The organic materials were collected and concentrated under vacuum, and then partitioned between EtOAc (30 mL) and 1 M aq. K₂CO₃ (30 mL). The organic layer was separated, dried over anhydrous MgSO₄, and evaporated under reduced pressure. The obtained oily materials were purified by column chromatography $(SiO_2, hexanes:CH_2Cl_2/2:1)$. The butanoate $(27c)$,⁷ benzoate $(27g)$,⁸ 4bromobenzoate (27h),⁹ and 4-chlorobenzoate (27i) derivatives have previously been reported. The physical appearance and spectral data of all esters are listed below:

4-Nitrobenzyl butanoate (27c). White solid (162 mg, 73%). ¹H NMR (CDCl₃, Fig. S25) δ 8.14 (d, *J* = 8.8 Hz, 2H), 7.46 (d, *J* = 8.4 Hz, 2H), 5.16 (s, 2H), 2.33 (t, *J* = 7.6 Hz, 2H), 1.63 (sx, *J* = 7.6 Hz, 2H), 0.90 (t, J = 7.6 Hz, 3H); ¹³C NMR (CDCl₃, Fig. S26) δ 173.0, 147.5, 143.5, 128.2, 123.7, 64.4, 35.9, 18.3, 13.6.

4-Nitrobenzyl cyclobutanecarboxylate (27d). Colorless oil (208 mg, 88%). ¹H NMR (CDCl₃, Fig. S27) d 8.20 (d, *J* = 8.4 Hz, 2H), 7.48 (d, *J* = 8.4 Hz, 2H), 5.18 (s, 2H), 3.21 (p, *J* = 8.4 Hz, 1H), 2.32-2.18 (m, 4H), 2.02-1.88 (m, 2H); 13C NMR (CDCl3, Fig. S28) d 174.9, 147.6, 143.5, 128.2, 123.8, 64.5, 37.9, 25.2, 18.4.

4-Nitrobenzyl cyclopentanecarboxylate (27e). Colorless oil (155 mg, 62%). ¹H NMR (CDCl₃, Fig. S29) d 8.13 (d, *J* = 8.4 Hz, 2H), 7.45 (d, *J* = 8.4 Hz, 2H), 5.14 (s, 2H), 2.78 (p, *J* = 8.4 Hz, 1H), 1.85-1.52 (m, 8H); ¹³C NMR (CDCl₃, Fig. S30) δ 176.2, 147.5, 143.7, 128.1, 123.6, 64.5, 44.6, 43.6, 29.9, 29.4, 25.7.

4-Nitrobenzyl cyclohexanecarboxylate (27f). Colorless oil (105 mg, 40%). ¹H NMR (CDCl₃, Fig. S31) δ 8.17 (d, *J* = 8.0 Hz, 2H), 7.46 (d, *J* = 8.0 Hz, 2H), 5.16 (s, 2H), 2.34 (tt, *J₁* = 11.2 Hz, *J₂* = 3.2 Hz, 1H), 1.90 (d, *J* = 12.0 Hz, 2H), 1.72 (m, 2H), 1.61 (m, 1H), 1.43 (q, *J* = 12.0 Hz, 2H), 1.30- 1.16 (m, 3H); ¹³C NMR (CDCl₃, Fig. S32) δ 175.5, 147.5, 143.7, 128.1, 123.7, 64.4, 43.0, 28.9, 25.6, 25.3.

4-Nitrobenzyl benzoate (27g). White solid (222 mg, 86%). ¹H NMR (CDCl₃, Fig. S33) δ 8.24 (d, *J* = 8.8 Hz, 2H), 8.07 (d, *J* = 8.4 Hz, 2H), 7.59 (m, 3H), 7.46 (t, *J* = 8.0 Hz, 2H), 5.45 (s, 2H); 13C NMR (CDCl₃, Fig. S34) δ 166.1, 147.7, 143.3, 133.5, 129.7, 129.4, 128.6, 128.3, 123.9, 65.2.

4-*Nitrobenzyl 4-bromobenzoate (27h)*. White solid (226 mg, 67%). ¹H NMR (CDCl₃, Fig. S35) δ 8.24 (d, *J* = 8.4 Hz, 2H), 7.92 (d, *J* = 8.4 Hz, 2H), 7.60 (d, *J* = 8.4 Hz, 2H), 7.58 (d, *J* = 8.8 Hz, 2H), 5.43 (s, 2H); 13C NMR (CDCl3, Fig. S36) d 165.4, 147.8, 143.0, 131.9 (2 carbons), 131.2 (2 carbons), 128.7, 128.4 (2 carbons), 128.3, 123.9 (2 carbons), 65.4.

4-*Nitrobenzyl* 4-*chlorobenzoate (27i)*. White solid (276 mg, 74%). ¹H NMR (CDCl₃, Fig. S37) δ 8.23 (d, *J* = 8.8 Hz, 2H), 8.01 (d, *J* = 8.8 Hz, 2H), 7.58 (d, *J* = 8.8 Hz, 2H), 7.42 (d, *J* = 8.8 Hz, 2H), 5.43 (s, 2H); 13C NMR (CDCl3, Fig. S38) d 165.2, 147.8, 143.0, 140.0, 131.1 (2 carbons), 128.9 (2 carbons), 128.4 (2 carbons), 127.9, 123.9 (2 carbons), 65.4.

4-Nitrobenzyl 4-chloro-2-fluorobenzoate (27k). Brown solid (197 mg, 64%). ¹H NMR (CDCl₃, Fig. S39) d 8.24 (d, *J* = 8.4 Hz, 2H), 7.92 (m, 1H), 7.60 (d, *J* = 8.0 Hz, 2H), 7.22 (dd, *J1* = 8.4 Hz, J_2 = 2.0 Hz, 1H), 7.20 (dd, J_1 = 11.2 Hz, J_2 = 2.0 Hz, 1H), 5.43 (s, 2H); ¹³C NMR (CDCl₃, Fig. S40) δ 163.23-163.16 (${}^{3}J_{C\text{-}F}$ = 7.6 Hz), 163.20-160.61 (${}^{1}J_{C\text{-}F}$ = 258.8 Hz), 147.8, 142.7, 140.68-140.57 (${}^{3}J_{C\text{-}F}$ = 10.6 Hz), 133.1, 128.3 (2 carbons), 124.83-124.80 (${}^{4}J_{C\text{-}F}$ = 3.8 Hz), 123.9 (2 carbons), 118.04-117.78 (${}^{2}J_{C-F}$ = 25.8 Hz), 116.61-116.51 (${}^{3}J_{C-F}$ = 9.9 Hz), 65.6.

4-*Nitrobenzyl 2-cyclohexylacetate (271)*. Colorless oil (98 mg, 35%). ¹H NMR (CDCl₃, Fig. S41) d 8.14 (d, *J* = 7.6 Hz, 2H), 7.46 (d, *J* = 7.6 Hz, 2H), 5.15 (s, 2H), 2.23 (d, *J* = 6.8 Hz, 2H), 1.74 $(sp, J = 4.0 \text{ Hz}, 1H)$, 1.60 (m, 5H), 1.21-1.06 (m, 3H), 0.95 (m, 2H); ¹³C NMR (CDCl₃, Fig. S42) d 172.5, 147.5, 143.5, 128.2, 123.7, 64.4, 41.8, 34.8, 32.9, 26.0, 25.9.

4-Nitrobenzyl 1-adamantanecarboxylate (27m). Colorless oil (112 mg, 37%). ¹H NMR (CDCl₃, Fig. S43) d 8.19 (d, *J* = 8.4 Hz, 2H), 7.47 (d, *J* = 8.4 Hz, 2H), 5.21 (s, 2H), 2.66 (t, *J* = 5.6 Hz, 1H), 2.29 (m, 2H), 2.06 (d, *J* = 2.1 Hz, 2H), 1.81 (m, 5H), 1.63-1.54 (m, 5H); 13C NMR (CDCl3, Fig. S44) d 177.0, 147.5, 143.9, 127.9, 123.8, 64.5, 53.7, 46.8, 46.4, 44.2, 43.6, 43.5, 37.4, 37.3, 34.6.

4-*Nitrobenzyl lithocholate (27n)*. White solid (215 mg, 41%). ¹H NMR (CDCl₃, Fig. S45) δ 8.20 (d, *J* = 8.8 Hz, 2H), 7.49 (d, *J* = 8.8 Hz, 2H), 5.17 (s, 2H), 3.60 (p, *J* = 4.8 Hz, 1H), 2.43 (m, 1H), 2.28 (m, 1H), 1.92 (d, *J* = 11.6 Hz, 1H), 1.77-1.63 (m, 5H), 1.49 (m, 2H), 1.46 (m, 2H), 1.23 (m, 8H), 1.09 (m, 3H), 1.05-0.93 (m, 5H), 0.89 (s, 3H), 0.88 (s, 3H), 0.59 (s, 3H); 13C NMR (CDCl3, Fig. S46) d 173.8, 147.6, 143.4, 128.4, 123.8, 71.8, 64.6, 56.4, 55.8, 42.7, 42.0, 40.4, 40.1, 36.4, 35.8, 35.3, 35.3, 34.5, 31.1, 30.9, 30.5, 28.2, 27.1, 26.4, 24.2, 23.3, 20.8, 18.2, 12.0.

General procedure for the synthesis of 4-nitrobenzyl thioethers (28g-i,k,p). The thiol derivative (1.1 mmol) and K_2CO_3 (150 mg, 1.1 mmol) were added to EtOH (15 mL). The reaction mixture was stirred for 20 min, and then 4-nitrobenzylbromide (216 mg, 1 mmol) was added portion-wise. The reaction mixture was continuously stirred at reflux temperature for 2 h, and then cooled down to room temperature. The organic materials were collected by filtration and concentrated under reduced pressure. The obtained brownish oily materials were purified by column chromatography (SiO₂, hexanes: $CH_2Cl_2/1:1$). 4-nitrobenzylbenzyl thioether $(28g)^{10}$ has previously been reported. The physical appearance and spectral data for all thioethers are listed below:

Benzyl(4-nitrobenzyl)sulfane (28g). Light yellow solid (153 mg, 59%). ¹H NMR (CDCl₃, Fig. S47) d 8.14 (d, *J* = 8.8 Hz, 2H), 7.40 (d, *J* = 8.8 Hz, 2H), 7.30 (d, *J* = 6.4 Hz, 2H), 7.26 (m, 3H), 3.64 (s, 2H), 3.60 (s, 2H); 13C NMR (CDCl3, Fig. S48) d 146.9, 146.1, 137.3, 129.8, 129.0, 128.6,127.3, 123.7, 35.8, 34.9.

4-Bromobenzyl(4-nitrobenzyl)sulfane (28h). White solid (246 mg, 73%). ¹H NMR (CDCl₃, Fig. S49) d 8.15 (d, *J* = 8.4 Hz, 2H), 7.42 (d, *J* = 8.4 Hz, 2H), 7.40 (d, *J* = 8.8 Hz, 2H), 7.12 (d, *J* = 8.8

Hz, 2H), 3.62 (s, 2H), 3.53 (s, 2H); ¹³C NMR (CDCl₃, Fig. S50) δ 147.0, 145.7, 136.4, 131.7 (2 carbons), 130.6 (2 carbons), 129.7 (2 carbons), 123.8 (2 carbons), 121.2, 35.2, 34.9.

4-Chlorobenzyl(4-nitrobenzyl)sulfane (28i). White solid (210 mg, 71%). ¹H NMR (CDCl₃, Fig. S51) d 8.15 (d, *J* = 8.4 Hz, 2H), 7.39 (d, *J* = 8.0 Hz, 2H), 7.27 (d, *J* = 8.4 Hz, 2H), 7.18 (d, *J* = 8.0 Hz, 2H), 3.62 (s, 2H), 3.55 (s, 2H); ¹³C NMR (CDCl₃, Fig. S52) δ 147.0, 145.7, 135.8, 133.1, 130.2 (2 carbons), 129.7 (2 carbons), 128.8 (2 carbons), 123.7 (2 carbons), 35.1, 34.9.

4-Chloro-2-fluorobenzyl(4-nitrobenzyl)sulfane (28k). Note: For the preparation of compound **3d**, 4-nitrobenzyl bromide (108 mg, 0.50 mmol), the thiol derivative (97 mg, 0.55 mmol), and K_2CO_3 (76 mg, 0.55 mmol) were used. Yellow solid (112 mg, 72%). ¹H NMR (CDCl₃, Fig. S53) δ 8.15 (d, *J* = 8.4 Hz, 2H), 7.44 (d, *J* = 8.8 Hz, 2H), 7.21 (m, 1H), 7.07 (m, 2H), 3.70 (s, 2H), 3.60 (s, 2H); 13C NMR (CDCl3, Fig. S54) d 161.72-159.24 (1 *JC-F* = 248.9 Hz), 147.1, 145.5, 133.97-133.87 $(^3J_{C-F} = 9.8$ Hz), 131.57-131.52 ($^3J_{C-F} = 4.6$ Hz), 129.7 (2 carbons), 124.72-124.68 ($^4J_{C-F} = 3.8$ Hz), 123.8 (2 carbons), 123.72-123.56 (${}^{3}J_{C-F}$ = 15.2 Hz), 116.47-116.22 (${}^{2}J_{C-F}$ = 22.1 Hz), 35.5, 28.31-28.28 $(3J_{C-F} = 3.0 \text{ Hz})$.

Cyclohexyl(4-nitrobenzyl)sulfane (28p). Faint yellow oil (65 mg, 26%). ¹H NMR (CDCl₃, Fig. S55) d 8.13 (d, *J* = 8.8 Hz, 2H), 7.46 (d, *J* = 8.8 Hz, 2H), 3.78 (s, 2H), 2.51 (p, *J* = 6.8 Hz, 1H), 1.88 (m, 2H), 1.71 (m, 2H), 1.56 (m, 1H), 1.26 (m, 5H); 13C NMR (CDCl3, Fig. S56) d 147.0, 146.8, 129.5, 123.7, 43.2, 34.0, 33.3, 25.9, 25.7.

BIOLOGICAL STUDIES

Optimization of the conditions for CNR activity. The CNR enzyme was purified as previously reported.11 The nitroreduction activity of CNR was observed using the absorbance of NADPH (340 nm, $\epsilon = 6,220$ M⁻¹cm⁻¹), which is used to reduce the flavin mononucleotide (FMN) of the enzyme after oxidation due to the ongoing reaction. Reactions (200 µL total volume) consisting of CAM (150 μ M), Tris-HCl (50 mM, pH 8.0 adjusted at room temperature), and NADPH (0.75 mg/mL, 1.0 mM), were initiated by addition of CNR (1 µM). Reactions were monitored taking readings every 30 s for 60 min. The initial rates were determined using the first 2-5 min of the reaction.

pH: The conditions above were used with various pHs to determine the optimum pH for CNR activity. Buffers and pHs used were as follows: 100 mM citric acid buffer (pH 3, 3.5, 4, 4.5, 5, 5.5, 6), 100 mM sodium phosphate (6.0, 6.5, 7, 7.5, 8.0), and 50 mM Tris-HCl (pH 6.8, 7.5, 8.0, 8.4, 9.0). Each reaction contained one of the buffers listed above, NADPH (750 μ M), CAM (500 μ M), and was initiated with the addition of CNR (0.4 μ M). Reactions were incubated at 25 °C and monitored as above in this section. These data are presented in Fig. S57.

Temperature: The temperature for optimum enzyme activity was tested by monitoring reactions at 20, 25, 30, 37, 42, 57, and 60 °C. Reactions contained Tris pH 8.0 (50 mM), NADPH (750 µM), CAM (500 μ M), and were initiated by the addition of CNR (0.4 μ M). Reactions were monitored as above in this section. These data are presented in Fig. S58.

Determination of kinetic parameters of CAM. The kinetic parameters of CAM were determined using reactions containing Tris-HCl (50 mM, pH 8.0 adjusted at room temperature), NADPH (750 μ M), CAM (0, 1, 5, 10, 50, 100, 250, 500, or 2500 μ M), and CNR (0.4 μ M). Reactions were incubated at 37 or 50 °C and monitored for 30 min taking measurements every 30 s. The first 2-5 min of reaction were used to determine the initial rates. The resulting data were fit with a Michaelis-Menten plot and kinetic parameters were determined using SigmaPlot 13. These data are presented in Fig. 1.

Monitoring of nitroreduction by the CNR enzyme by UV-Vis assays. Reactions (200 µL total volume) consisting of nitro-containing compounds (synthesized, Scheme 2; commercially available, Fig. S59) (150 µM), Tris-HCl (50 mM, pH 8.0 adjusted at room temperature), and NADPH (0.75 mg/mL, 1.0 mM) were initiated by addition of CNR (1 μ M). Reactions were monitored taking readings every 30 s for 60 min. The initial rates were determined using the first 5 min of the reaction and were normalized to the rate for chloramphenicol (CAM), a known substrate of the CNR enzyme.

Large-scale reduction reactions for LCMS. Reactions contained Tris-HCl (50 mM, pH 8.0) adjusted at room temperature), NAPDH (100 μ M), glucose (40 mM), glucose dehydrogenase (4 μ M, used with glucose to regenerate NADPH),¹² CNR (5 μ M), and the compound of interest (1 mM). Reactions were incubated at 37 °C for 24 h. After incubation, reactions were cooled and extracted with 3×10 mL of EtOAc. The organic fractions were pooled and dried over MgSO₄. The remaining solvent was removed *in vacuo*. The residue was redisolved in 1 mL of MeOH and analyzed by LCMS. These data are presented in Fig. S60.

Large-scale reduction reactions for RP-HPLC time course. Reactions were identical to those used in preparation for LCMS above. After 10 min, 30 min, 60 min, 3 h, 6 h, and 24 h, a 0.5-mL aliquot of the reaction was extracted and dried as above. The resulting residues were dissolved in 100 µL of MeOH. A 10-µL portion was injected onto the RP-HPLC and run with general method 1. Results of the RP-HPLC time course are shown in Fig. S61.

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Fig. S7: 1H NMR spectrum for compound **26e** in CDCl3 (400 MHz).

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Fig. S23: 1H NMR spectrum for compound **26q** in CDCl3 (400 MHz).

Fig. S27: 1H NMR spectrum for compound **27d** in CDCl3 (400 MHz).

Fig. S30: 13C NMR spectrum for compound **27e** in CDCl3 (100 MHz).

Fig. S32: 13C NMR spectrum for compound **27f** in CDCl3 (100 MHz).

Fig. S35: 1H NMR spectrum for compound **27h** in CDCl3 (400 MHz).

Fig. S43: 1H NMR spectrum for compound **27m** in CDCl3 (400 MHz).

S27

Fig. S57: Rate of reaction of CNR with CAM at different pHs in increments of 0.5 (pH 3-6 using citrate buffer; pH 6-8 using sodium phosphate buffer; pH 7-9 using Tris-HCl buffer).

Fig. S58: Rate of reaction of CNR with CAM at different temperatures.

Fig. S59: Structures of synthesized nitro-containing molecules tested with CNR. *Note*: The synthetic schemes along with yields are presented in Scheme 2.

Fig. S60: MS traces of CNR-catalyzed large-scale reduction reactions of **A.** compound **26h** showing the masses of the reduced products 291.0 (M+, amine) and 308.0 (hydroxylamine, [M+H]+), **B.** compound **27h** with the deprotection mass of 199.0 ([M-H]-), **C.** compound **27i** having the deprotected mass of 173.0 (([M-H]-), **D.** compound **27k** with a mass of 173.0 (([M-H]-) indicating deprotection, and **E.** compound **28h** showing the reduction to the amine with a mass of 308.0 ($\overline{[M+H]}^+$).

Fig. S61: RP-HPLC traces of a reaction of compound **27h** with CNR and NADPH. Samples were taken at 10 min, 30 min, 60 min, 3 h, 6 h, and 24 h.

