## **Supplementary Note 1**

We first sought to apply Monocle 3 to a single major cell type, cluster 25, whose 26,559 cells we annotate as limb bud mesenchyme on the basis of *Hoxd13, Fgf10* and *Lmx1b* expression (**Supplementary Table 4**). Visualizing the trajectory of cells of this cluster illustrates the dramatic expansion of limb mesenchymal cells over developmental time, with the main outgrowth between E10.5 and E12.5 (**Extended Data Fig. 7a**). Gene expression is highly dynamic during this expansion, with the levels of 4,763 protein-coding genes changing (FDR of 1%; **Supplementary Table 7**). The early stages of limb mesenchyme development are characterized by expression of some expected genes such as *Tbx151* , and *Gpc32* and the later stages by *Msx13* , *Epha44* and *Dach15* (**Extended Data Fig. 7b**), but the vast majority of dynamically expressed genes are novel. Transcription factors significantly upregulated during limb mesenchyme development included those with roles in chondrocyte differentiation (*e.g. Sox96* and *Yap17* ), muscle differentiation (*e.g. Tead48* ), and wound healing and limb regeneration (*e.g. Smarcd19* ) (**Extended Data Fig. 7c**).

Interestingly, forelimb and hindlimb cells were not obviously separated by unsupervised clustering (**Extended Data Fig. 7d**) or trajectory analysis (**Extended Data Fig. 7e**), but could be distinguished by the mutually exclusive expression of *Tbx5* in forelimb (2,085 cells, 7.9% of all limb mesenchyme cells) and *Pitx1* in hindlimb (1,885 cells, 7.1% of all limb mesenchyme cells) with only 22 cells expressing both markers (0.08% of all limb mesenchyme cells vs.  $\sim 0.6\%$ expected if they were independent; **Extended Data Fig. 7f**)<sup>10</sup>. 285 genes were differentially expressed between cells assigned to the forelimb and hindlimb in this way (**Extended Data Fig. 7g**, **Supplementary Table 8**). Known marker genes such as *Tbx4* and the genes of the Hoxc cluster (*Hoxc4-10*) <sup>11</sup> were upregulated in hindlimb cells as expected, but we also identified genes not previously shown to be differentially expressed. For example, we observed *Epha3* and *Hs3st3b1*  to be 5-fold enriched in forelimb, and *Pcdh17* and *Igf1* to be 3-fold enriched in hindlimb.

Although developmental time is a major axis of variation in the limb mesenchyme trajectory (**Extended Data Fig. 7a**), there is clearly additional structure. At least some of this appears to correspond to the two main spatial axes of limb development: the proximal-distal axis (the primary direction of outgrowth) and the anterior-posterior axis (corresponding to the five digits)<sup>10</sup>. With Monocle 3, we applied Moran's I test<sup>12</sup> to detect genes exhibiting autocorrelation across the limb mesenchyme trajectory (*i.e.* genes expressed in similar regions of the principal graph). We found, for example, that cells expressing *Sox6* and *Sox9* (proximal markers)13,14*, Hoxd13* and *Tfap2b*  (distal markers)15, *Pax9* and *Alx4* (anterior markers), and *Shh* and *Hand2* (posterior markers), were differentially distributed across the trajectory (**Extended Data Fig. 7h**, **Extended Data Fig. 7i**). Whole-mount *in situ* hybridization of *Hoxd13* (a known distal marker) and *Cpa2* (a novel marker whose distribution in the Monocle 3 trajectory was similar to that of known distal markers), confirmed that both genes are expressed in the distal limb mesenchyme between E10.5 and E13.5 (**Extended Data Fig. 7j-l**). Altogether, we identified 1,783 genes exhibiting variable expression across the limb mesenchymal trajectory (FDR of 1%; Moran's  $I > 0.01$ ). These genes clustered into eight patterns of expression, several of which matched the distributions of known markers for the proximal-distal and anterior-posterior axes (**Extended Data Fig. 7m**, **Supplementary Table** 

**9**). These analyses illustrate how this single cell atlas of mouse organogenesis can be used to characterize the spatiotemporal dynamics of gene expression in specific systems.

## **References:**

- 1. Singh, M. K. *et al.* The T-box transcription factor Tbx15 is required for skeletal development. *Mech. Dev.* **122,** 131–144 (2005).
- 2. Paine-Saunders, S., Viviano, B. L., Zupicich, J., Skarnes, W. C. & Saunders, S. glypican-3 controls cellular responses to Bmp4 in limb patterning and skeletal development. *Dev. Biol.* **225,** 179–187 (2000).
- 3. Hara, K. & Ide, H. Msx1 expressing mesoderm is important for the apical ectodermal ridge (AER)-signal transfer in chick limb development. *Dev. Growth Differ.* **39,** 705–714 (1997).
- 4. Lupiáñez, D. G. *et al.* Disruptions of Topological Chromatin Domains Cause Pathogenic Rewiring of Gene-Enhancer Interactions. *Cell* **161,** 1012–1025 (2015).
- 5. Davis, R. J. *et al.* Dach1 mutant mice bear no gross abnormalities in eye, limb, and brain development and exhibit postnatal lethality. *Mol. Cell. Biol.* **21,** 1484–1490 (2001).
- 6. Akiyama, H., Chaboissier, M.-C., Martin, J. F., Schedl, A. & de Crombrugghe, B. The transcription factor Sox9 has essential roles in successive steps of the chondrocyte differentiation pathway and is required for expression of Sox5 and Sox6. *Genes Dev.* **16,** 2813–2828 (2002).
- 7. Deng, Y. *et al.* Yap1 Regulates Multiple Steps of Chondrocyte Differentiation during Skeletal Development and Bone Repair. *Cell Rep.* **14,** 2224–2237 (2016).
- 8. Joshi, S. *et al.* TEAD transcription factors are required for normal primary myoblast differentiation in vitro and muscle regeneration in vivo. *PLoS Genet.* **13,** e1006600 (2017).
- 9. Knapp, D. *et al.* Comparative transcriptional profiling of the axolotl limb identifies a tripartite regeneration-specific gene program. *PLoS One* **8,** e61352 (2013).
- 10. Zeller, R., López-Ríos, J. & Zuniga, A. Vertebrate limb bud development: moving towards integrative analysis of organogenesis. *Nat. Rev. Genet.* **10,** 845–858 (2009).
- 11. Nishimoto, S., Minguillon, C., Wood, S. & Logan, M. P. O. A combination of activation and repression by a colinear Hox code controls forelimb-restricted expression of Tbx5 and reveals Hox protein specificity. *PLoS Genet.* **10,** e1004245 (2014).
- 12. Moran, P. A. P. Notes on continuous stochastic phenomena. *Biometrika* **37,** 17–23 (1950).
- 13. Vargesson, N., Luria, V., Messina, I., Erskine, L. & Laufer, E. Expression patterns of Slit and Robo family members during vertebrate limb development. *Mech. Dev.* **106,** 175–180 (2001).
- 14. Chimal-Monroy, J. *et al.* Analysis of the molecular cascade responsible for mesodermal limb chondrogenesis: Sox genes and BMP signaling. *Dev. Biol.* **257,** 292–301 (2003).
- 15. Petit, F., Sears, K. E. & Ahituv, N. Limb development: a paradigm of gene regulation. *Nat. Rev. Genet.* **18,** 245–258 (2017).