Supporting Information

Identification of Factors Complicating Bioluminescence Imaging

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Figure S1. (A) Representative bioluminescence images of BALB/c mice (from left to right: blank, i.v. injection of DTZ, i.p. injection of DTZ, and i.v. or i.p. injection of DTZ after i.v. injection of Antares2-expressing HEK 293T cells). These are identical to the images in Figure 3B but are presented in grayscale. (B) Quantitative analysis of signals integrated over areas. The y axis is in a logarithmic scale. Data are presented as mean and s.d. of three biological replicates.



Figure S2. (A-C) *Ex vivo* bioluminescence of organs harvested from mice at 1, 5, 24 hours after i.v. injection of HEK293T cells expressing (A) teLuc, (B) Antares2, or (C) Akaluc (n = 1 for each group). Signals were integrated over the area of each organ. (D) Bioluminescence images of extracted organs from mice with i.v. injected teLuc-labeled HEK 293T cells (top row) or untransfected HEK 293T cells (bottom row). H: heart, Lg: lung, Lv: liver, P: spleen, S: stomach, In: intestine, K: kidney, T: testis, and B: bladder.



Figure S3. (A) Bioluminescence kinetics of purified 1 nM teLuc or Anatres2 in either PBS or mouse serum post injection of 25 μ M DTZ. (B) Bioluminescence kinetic of purified 1 μ M Akaluc in PBS or mouse serum post injection of 100 μ M AkaLumine with or without 5 mM ATP. (CD) Bioluminescence intensities (C) and kinetics (D) of purified 1 μ M FLuc in either PBS or mouse serum post injection of 100 μ M D-luciferin with or without 5 mM ATP.



Figure S4. (A) Fluorescence imaging of live mammalian cells after incubation with indicated luciferins for 10 min. 100 μ M AkaLumine-HCl, 200 μ M D-luciferin, or 50 μ M DTZ was added to cell lines, including HeLa, HEK 293T, and SH-SY5Y. These concentrations were chosen since each compound has different cell permeability and previous publications used these concentrations for corresponding assays.¹⁻⁴ Images were captured with 405-nm or 488-nm laser excitation, and emission was collected from 510 nm to 600 nm (scale bars: 20 μ m). (**BC**) Emission spectra of AkaLumine-treated HEK 293T cells with 405 nm (B) or 488 nm (C) laser excitation. (**D**) Emission spectrum of D-luciferin-treated HEK 293T cell with 405-nm laser excitation. (**E**) Fluorescence emission of 1 mM D-luciferin (gray) or AkaLumine (black) in PBS with 400 nm excitation. (**F**) Fluorescence emission of 1 mM AkaLumine (black) in PBS with 580 nm excitation.



Figure S5. (A) Morphological changes of HeLa cells in response to addition of 100 μ M AkaLumine at 0 min. Cells were illuminated under a Leica SPE confocal scanning microscope with a 10-mW 488-nm laser (10% output). The scanning line speed was 600 Hz for a 275 μ m*275 μ m area under a 63x objective. The acquisition interval was 5 s. The blebbing of cell membrane and the formation of extracellular vesicles were observed at 15 and 30 min. (B) HeLa cells in the absence of AkaLumine but otherwise treated the same as cells in panel A. (C) HeLa cells treated with 100 μ M AkaLumine but in the absence of 488-nm excitation. The data suggest that AkaLumine may act as a photosensitizer in stained cells.



Figure S6. (A) PercevalHR fluorescence in HEK 293T upon treatment with D-luciferin or DTZ in the absence of FLuc or teLuc, showing no change in ATP occupancy. (B) pHRFP fluorescence in HEK 293T upon treatment with D-luciferin or DTZ in the presence of the corresponding luciferase, showing that the corresponding bioluminescence reactions do not affect intracellular pH.



Figure S7. Fluorescence imaging (excitation: 605/25 nm bandpass; emission: 660/25 nm bandpass) of organs harvested from (**A**) an untreated BALB/c mouse, or (**B**) a BALB/c mouse i.v. injected with 1.5 µmol AkaLumine-HCl. Near-infrared (NIR) background fluorescence, mostly in the gastrointestinal tract, was observed for mice, because chlorophyll from plant-based diet (e.g., alfalfa) is fluorescence in this spectral range.⁵ In addition to the background, the AkaLumine-injected mouse showed high NIR fluorescence in the liver. H: heart, Lg: lung, Lv: liver, P: spleen, S: stomach, In: intestine, K: kidney, and B: bladder.

References:

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