Supporting Information:

Sub lethal levels of platinum nanoparticle cures plasmid and in combination with carbapenem, curtails carbapenem resistant *Escherichia coli*

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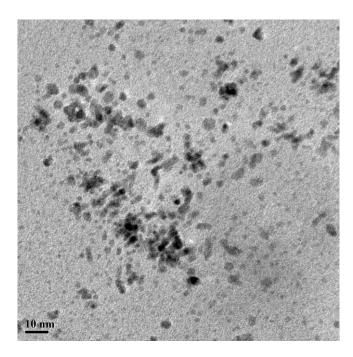
Antibiotic/Compound	Minimum Inhibitory Concentration (µg/ml)
Ceftriaxone	>128
Ciprofloxacin	256
Colistin	32
Erythromycin	256
Gentamicin	256
Kanamycin	>256
Levofloxacin	32
Meropenem	>32
Norfloxacin	>128
Streptomycin	16
Tetracycline	128
Tigecycline	>128
Tobramycin	128

Supplementary Table 1: Antimicrobial profiling of XDR U3790

Supplementary Table 2: Output from the plasmid finder tool used for identification of plasmid from the contig Sequences

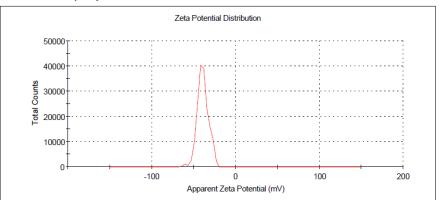
Plasmid	Identity	Query / Template length	Contig	Position in contig	Note	Accession number
IncFIA	99.74	388 / 388	MyFirstAss embly_rep_ c200	3592397 9		<u>AP001918</u>
IncFIB(AP00 1918)	98.39	682 / 682	MyFirstAss embly_rep_ c201	$embly_rep_ = \begin{bmatrix} 5248592\\ 9 \end{bmatrix}$		<u>AP001918</u>
IncFII(pAM A1167- NDM-5)	99.62	261 / 261	MyFirstAss embly_rep_ c213	7730799 0	pAMA1 167- NDM-5	<u>CP024805.</u> <u>1</u>
Incl1	100	142 / 142	MyFirstAss embly_rep_ c206	1795418 095	Alpha	<u>AP005147</u>
p0111	98.53	885 / 885	MyFirstAss embly_rep_ c199	3051231 396		<u>AP010962</u>

Supplementary Figure 1: TEM image and zeta potential of pectin capped PtNPs

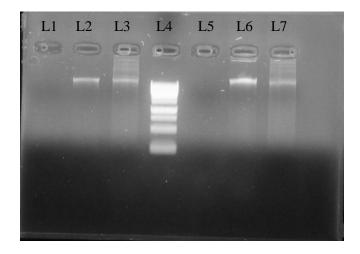


Results

			Mean (mV)	Area (%)	St Dev (mV)
Zeta Potential (mV):	-38.9	Peak 1:	-38.7	98.7	6.42
Zeta Deviation (mV):	6.74	Peak 2:	-59.8	1.3	2.50
Conductivity (mS/cm):	0.241	Peak 3:	0.00	0.0	0.00
Result quality :	Good				



Supplementary Figure 2: (The full length gel image of Figure 1B) Gel picture showing absence of plasmid in cured SCV strain (L1, L5) and its presence in uncured strain (L2, L6) and in wild type (L3, L7), 1 KB ladder Molecular weight marker (L4).



Supplementary Figure 3: **ABST test showing plasmid curing restores sensitivity to meropenem in SCV.** Antibiotic susceptibility test was performed for the wild type U3790, large colony variant (LCV) and small colony variant (SCV) by exposing the strain to PtNPs and the zone of inhibition was measured. SCV exhibited a larger zone of inhibition (16mm) relative to its wild type and LCV (10mm).

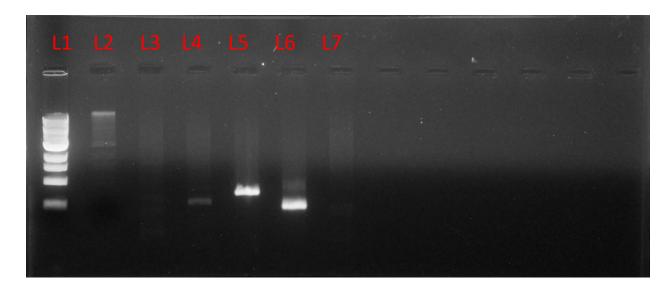


Wild type

LCV

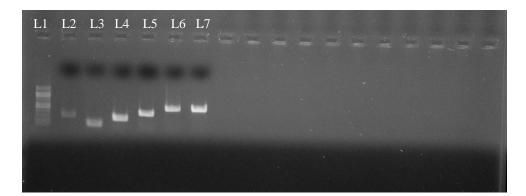
SCV

Supplementary Figure 4: **PCR amplification of ESBL and mcr-1 genes.** Plasmid harbored carbapenem resistant genes. *bla*_{OXA23}, *bla*_{OXA 48} and *bla*_{NDM} genes were amplified by PCR using plasmid DNA as template.

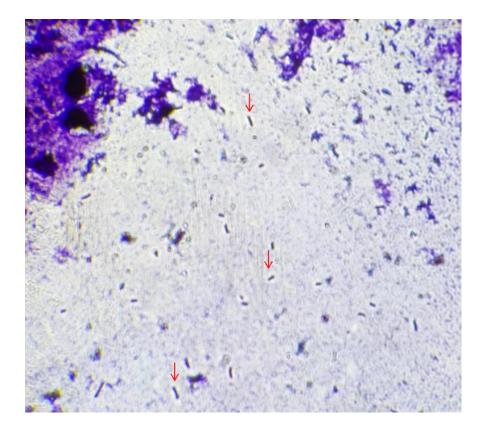


- L1: Molecular weight marker
- L2: Plasmid DNA template
- L3: bla_{KPC}
- L4: *bla*_{OXA23}
- L5: *bla*_{OXA48}
- L6: *bla*_{NDM}
- L7: bla_{VIM}

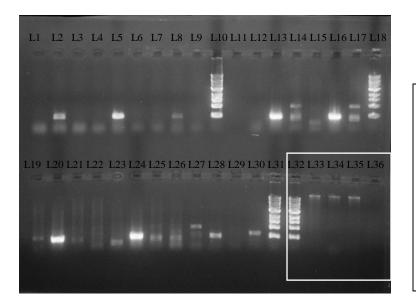
Supplementary Figure 5: Detection of Incompatibility groups in plasmid. PCR amplification of plasmid from U3790 as template showed the presence of IncI1, IncFII, IncFIA, IncFIB and pO111.

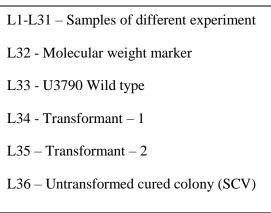


Lane Description: L1: Molecular weight marker L2: Other sample L3: IncI1 L4: IncFII L5: IncFIA L6: IncFIB L7: pO111 **Supplementary Figure 6**: **Capsulated strain**: Capsular staining indicates that U3790 possesses capsule as evidenced by halo around bacilli, which might have prevented the action of SDS/AO on plasmid.



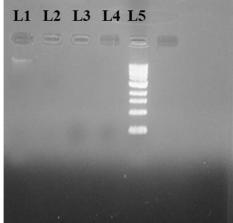
Supplementary Figure 7: (The full length gel image of Figure 2b) Plasmid extracted from transformants on LB + antibiotic plate showed reappearance of plasmid band.





Supplementary Figure 8: PtNPs cures plasmid from *E.coli* strain. 20 μ M of PtNPs was effective in eliminating plasmid from *E.coli* DH5 α strain harboring yeast expression vector p2GB42/Kan

Plasmid Gel Image



L1- Plasmid from E.coli 17260

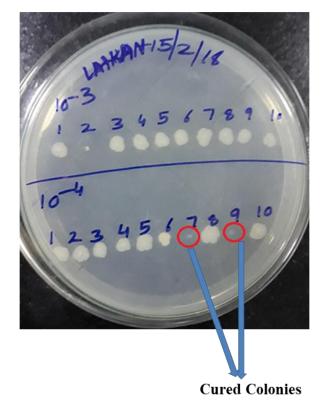
L2- Plasmid from *E.coli* 17260 treated with Nanoparticle(0.02mM)

L3- Plasmid from *E.col*i 17260 treated with SDS

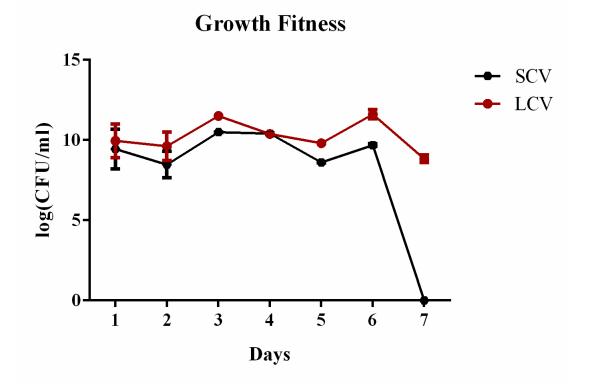
L4- Plasmid from *E.coli* 17260 treated with AO

L5- Molecular Weight Marker

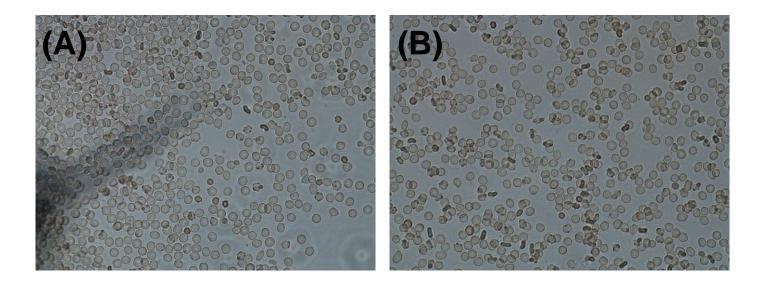
Spot Plating on Kanamycin Plates



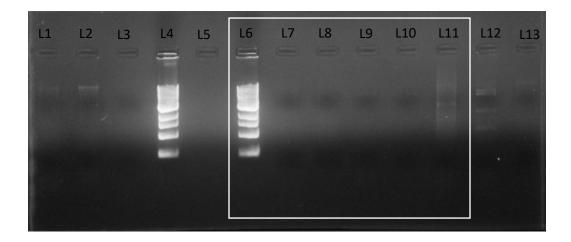
Supplementary Figure 9: Growth fitness assay revealed that SCV displayed lower fitness relative to LCV.



Supplementary Figure 10: Hemocompatibility assay. (A) Untreated human RBC and (B) human RBC treated with 200 μ M PtNPs. RBC were incubated with 200 μ M PtNPs for 30min and evaluated under microscope for morphological changes. No damage to the cells were observed, which shows intrinsic hemocompatibility.



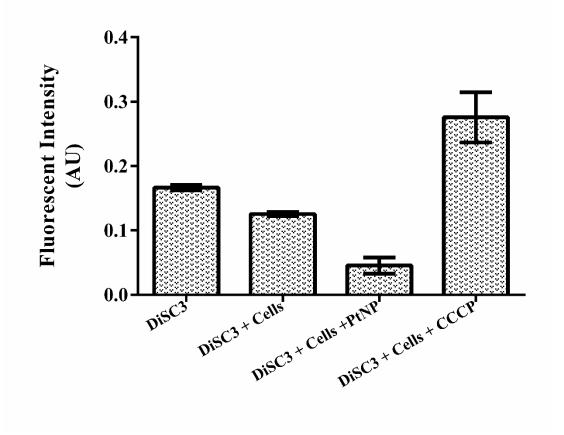
Supplementary Figure 11: **PtNP along with antibiotic cures plasmid** *in vivo*. SCVs isolated from infected muscle tissue of fish lacks plasmid Molecular weight marker (L6), SCV from zebrafish muscle tissue treated with PtNP alone (L7 & L8) and PtNP + Meropenem (L9&L10) were devoid of the plasmid relative to bacteria treated with meropenem (L11)



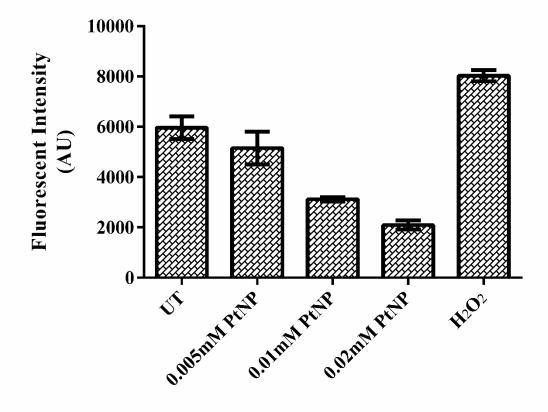
KEY:

- L1 to L5 & L12, L13 Samples of different experiment
- L6 Molecular weight Marker
- L7 & 8 SCV from zebrafish muscle treated with PtNP alone
- L9 & 10 SCV from zebrafish muscle treated with Meropenem +PtNPs
- L11 SCV from zebrafish muscle treated with meropenem

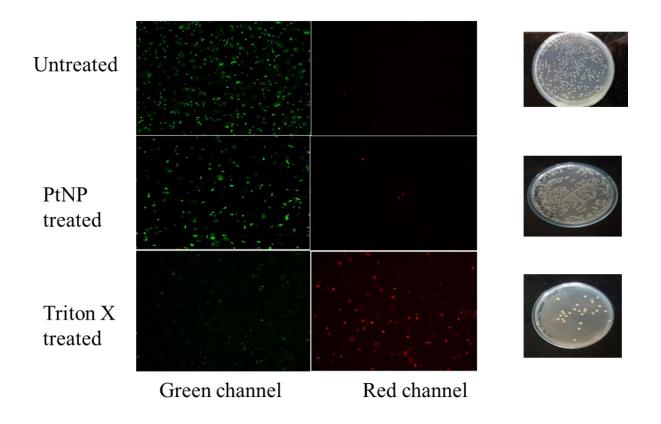
Supplementary Figure 12: PtNPs does not perturb Membrane potential in carbapenem resistant *E.coli*. Cells were incubated with PtNPs and Disc3 for 1hr and the fluorescence were measured at Ex605nm and Em665nm. The experiment was performed in triplicates and the error bar represents the mean standard error from three independent experiments.



Supplementary Figure 13: PtNPs at sub MIC levels does not induce intracellular ROS in *E.coli*. Cells were treated with PtNPs and H_2O_2 , as positive control. The fluorescence intensity is measured with excitation and emission at 495nm and 529nm. ROS generation was determined by the conversion of DCFDA to DCF. The experiment was performed in triplicates and the error bar represents the mean standard error from three independent experiments.



Supplementary Figure 14: Platinum NP (20 μ M) treatment for 24h does not affect cell viability of *E.coli*. Proportion of propidium iodide stained cells were negligible in PtNP treatment relative to Triton X 100 treatment and plating reveals reduction in cell density for Triton X 100 treatment.



Supplementary Figure 15: (The full length gel image of Figure 6) PtNPs induce DNA cleavage in presence and absence of gyrase.

