

Supplementary Online Content

Bichsel CA, Goss J, Alomari M, et al. Association of somatic *GNAQ* mutation with capillary malformations in a case of choroidal hemangioma. *JAMA Ophthalmol*. Published online October 25, 2018. doi:10.1001/jamaophthalmol.2018.5141

eMethods

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eFigure 2. Macroscopic View of the Patient's Eye and the Retinoblastoma-Affected Eye

eFigure 3. The Patient's Ocular Tissue is Negative for *GNAQ* Q209L, Q209P and Q209H

eFigure 4. Tissue Sections Stained for the Infantile Hemangioma Marker GLUT1

This supplementary material has been provided by the authors to give readers additional information about their work.

Supplemental methods

Choroidal tissue isolation, genomic DNA extraction and droplet digital PCR

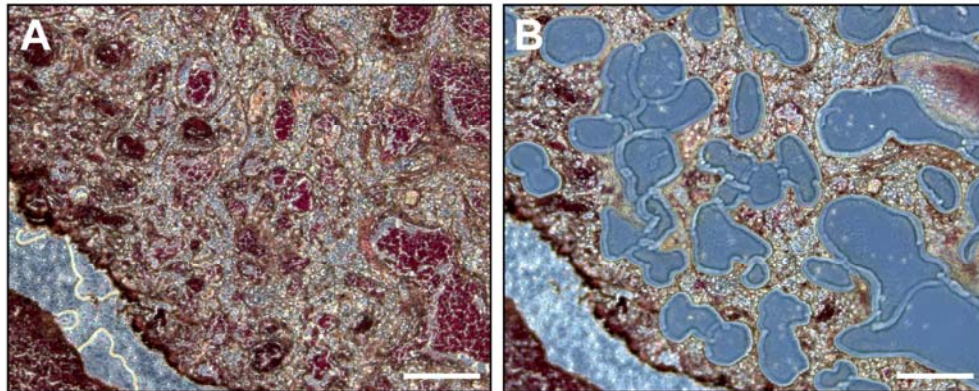
Immunofluorescent Staining of FFPE sections

Supplemental list of ddPCR primers and probes

Primer sequences

Probe sequences

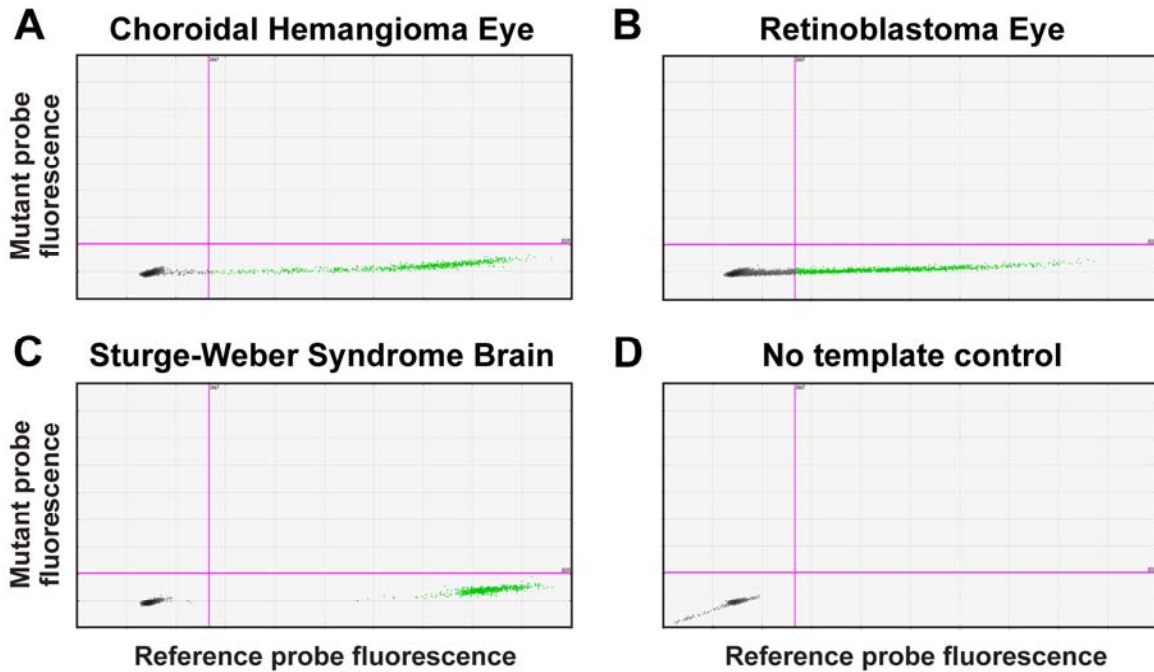
Supplemental Figures



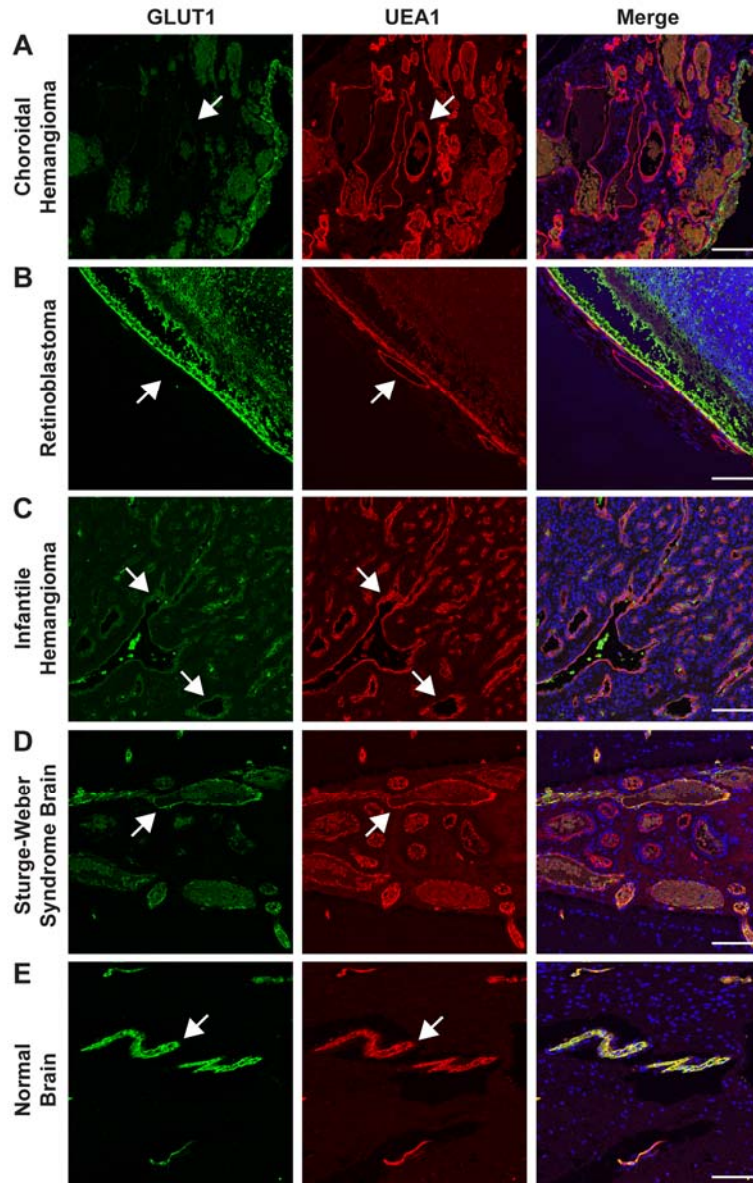
eFigure 1. The patient's choroidal tissue section before and after laser-capture microdissection. H&E stained choroidal tissue section of the patient showing a plethora of abnormal blood vessels in the choroid (A). Same section after blood vessel isolation by laser-capture microdissection (B). Scale bars: 200 μ m.



eFigure 2. Macroscopic view of the patient's eye and the retinoblastoma-affected eye. H&E stained ocular tissue section of the patient's eye (bottom) and a retinoblastoma-affected eye (top). Scale bar: 10 mm.



eFigure 3. The patient's ocular tissue is negative for *GNAQ* Q209L, Q209P and Q209H. DDPCR with pooled probes detecting the *GNAQ* Q209L, Q209P and Q209H mutations in samples from choroidal hemangioma eye (A), retinoblastoma eye (B) and Sturge-Weber brain tissue (C). Water was used as no template control (D). All samples were negative for the three *GNAQ* Q209 mutations.



eFigure 4. Tissue sections stained for the infantile hemangioma marker GLUT1.

FFPE sections stained for GLUT1 (green), the endothelial marker Ulex Europaeus Agglutinin I (red), the nuclear stain Hoechst (blue) in the SWS patient's choroidal vessels (A), retinoblastoma choroidal vessels (B), infantile hemangioma (C), SWS-affected leptomeninges (D), normal brain leptomeninges (E). Scale bars: 100 μ m.

Supplemental methods

Choroidal tissue isolation, genomic DNA extraction and droplet digital PCR

Formalin fixed paraffin-embedded (FFPE) tissue from this patient's enucleation was obtained and 15 µm sections were cut and placed onto PEN-membrane slides (Leica Biosystems). Slides were stained with hematoxylin and eosin, and laser-capture microdissection (Leica) was used to isolate choroidal vessels (eFigure 1). Similarly, choroidal vessels were obtained from FFPE tissue from an eye with a normal choroid that was enucleated because of retinoblastoma (eFigure 2). Genomic DNA isolated from a SWS brain specimen and normal lung tissue from different individuals were used as controls.

Choroidal vessels laser-captured from two to five tissue slides were pooled. For whole eye tissue, the entire section was scraped from a slide. Genomic DNA was extracted with the Agencourt FormaPure DNA kit (Beckman-Coulter). 30-90 ng of genomic DNA was then used for droplet digital PCR (ddPCR). Briefly, the DNA was mixed with ddPCR SuperMix (BioRad), primers for a 74bp amplicon in GNAQ Exon4, FAM-labeled quenched mutant probes and HEX-labeled quenched wild-type probes (supplemental list), divided into droplets (QX200 AutoDG) and amplified for 40 polymerase chain reaction cycles using 60°C as annealing temperature. The droplet fluorescence was read in a QX200 droplet reader (BioRad) and analyzed in QuantaSoft (BioRad).

Immunofluorescent Staining of FFPE sections

The FFPE sections were de-paraffinized, and antigen retrieval was performed in citrate buffer at pH 6.0 for 20 minutes at 95°C. The sections were incubated with primary antibodies (40 µg/ml (1:50) anti-GLUT1 (ab40084 Abcam), 40 µg/ml (1:5000) mouse-IgG (I-2000 Vector) for control, and 20 µg/ml (1:100) rhodamine-labeled Ulex Europaeus Agglutinin I, (RL-1062 Vector)) for one hour at room temperature in Tris-NaCl buffer with 0.5% blocking reagent (FP1012 Perkin Elmer), washed in Tris-NaCl buffer with 0.05% Tween-20 (BioRad), and incubated with the secondary antibody (1:200), Alexa647 goat anti-mouse IgG (A21236 Thermo Fisher) and Hoechst (1:1000) (33342 Thermo Fisher) for one hour at room temperature, washed and mounted (P36965 Thermo Fisher). Images were taken with a Zeiss LSM 880 confocal microscope.

Supplemental list of ddPCR primers and probes

Primer sequences

GNAQ R183 fwd: 5'-CCTGCCTACGCAACAAGAT-3'
rev: 5'-GTAAGTCAAAGGGGTATTCGAT-3'
GNAQ Q209 fwd: 5'-GTTAACCTTGCAGAATGGTTCG-3'
rev: 5'-GACATTTTCAAAGCAGTGTATCC-3'

Probe sequences

GNAQ R183 reference:
5'-/5HEX/TGCTTAGAG/ZEN/TTCGAGTCCCCACC/3IABkFQ/-3'
GNAQ R183Q mutant:
5'-/56-FAM/TGCTTAGAG/ZEN/TTCAGTCCCCACC/3IABkFQ/-3'
GNAQ Q209 reference:
5'-/5HEX/CTTCTCTCT/ZEN/GACCTTTGGCCCCCTA/3IABkFQ/-3'
GNAQ Q209L mutant:
5'-/56-FAM/TCTCTCT/ZEN/GACCTTAGGCCCCCTAC/3IABkFQ/-3'
GNAQ Q209P mutant:
5'-/56-FAM/TCTCTCT/ZEN/GACCTTGGGCCCCCTAC/3IABkFQ/-3'
GNAQ Q209H mutant:
5'-/56-FAM/CTCTCTGAC/ZEN/CTGTGGCCCCCT/3IABkFQ/-3'