Primer	Mutation	Sequence (5´-3´)
CrmD37	K44A	GGTACAGACTATAACAGTAATAATCTATGTTGT <u>GC</u> ACAATGCAATCCTGGAAT
CrmD38	K44A	ATTCCAGGATTGCATTGT <u>GC</u> ACAACATAGATTATTACTGTTATAGTCTGTACC
CrmD39	P68A	TACAAAATGTGACAAGTGC <u>G</u> CAGATGACACCTTTACATC
CrmD40	P68A	GATGTAAAGGTGTCATCTG <u>C</u> GCACTTGTCACATTTTGTA
CrmD41	N77A	AGATGACACCTTTACATCCATTCCT <u>GC</u> TCATAGTCCCGCGTG
CrmD42	N77A	CACGCGGGACTATGA <u>GC</u> AGGAATGGATGTAAAGGTGTCATCT
CrmD43	N77F	AGATGACACCTTTACATCCATTCCT <u>TT</u> TCATAGTCCCGCGTG
CrmD44	N77F	CACGCGGGACTATGA <u>AA</u> AGGAATGGATGTAAAGGTGTCATCT
CrmD45	Q93A/E95A	AAGTTGTCGAGGCAAATGTAGCAGTAAT <u>GC</u> AGTAG <u>C</u> GACTAAATCGTGTAGTAAC
CrmD46	Q93A/E95A	GTTACTACACGATTTAGTC <u>G</u> CTACT <u>GC</u> ATTACTGCTACATTTGCCTCGACAACTT
CrmD47	Q103A/D104A/R105A	${\tt GTAATCAAGTAGAGACTAAATCGTGTAGTAACACA} \underline{{\tt GC}} {\tt GG} \underline{{\tt C}} \underline{{\tt GC}} {\tt AGTATGTGTCTGTGCATC}$
CrmD48	Q103A/D104A/R105A	${\tt GATGCACAGACACATACT} \underline{{\tt GC}} \underline{{\tt GG}} \underline{{\tt CC}} \underline{{\tt GT}} \underline$
CrmD49	Y141A/S142A/S143A	${\tt GTGGTTCTGGTTACGGTGTATATGGC} \underline{{\tt GC}} {\tt CA} \underline{{\tt G}} {\tt CTAAAGGAGATGTAATATGTAAAAAGT}$
CrmD50	Y141A/S142A/S143A	ACTTTTTACATATTACATCTCCTTTAGCTGCGGCGCCATATACACCGTAACCAGAACCAC
CrmD51	D35A/Y36A	CCCATTAATGGGAAATGTAACGGTACAG <u>C</u> C <u>GC</u> TAACAGTAATAATCTATGTTGTAAACAA
CrmD52	D35A/Y36A	${\tt TTGTTTACAACATAGATTATTACTGTTA} \underline{{\tt GC}} {\tt GG} {\tt CTGTACCGTTACATTTCCCATTAATGGG}$
CrmD53	F72A/T73A/S74A	GACAAGTGCCCAGATGACACC <u>GC</u> T <u>G</u> CA <u>G</u> CCATTCCTAATCATAGTCCCG
CrmD54	F72A/T73A/S74A	CGGGACTATGATTAGGAATGG <u>C</u> TG <u>C</u> A <u>GC</u> GGTGTCATCTGGGCACTTGTC
CrmD55	H78A/S79A	CCCAGATGACACCTTTACATCCATTCCTAAT <u>GC</u> TCCCGCGTGTCTAAGT
CrmD56	H78A/S79A	ACTTAGACACGCGGGA <u>GC</u> A <u>GC</u> ATTAGGAATGGATGTAAAGGTGTCATCTGGG
CrmD57	R86A/K86A	TAGTCCCGCGTGTCTAAGTTGT <u>GC</u> AGGC <u>GC</u> ATGTAGCAGTAATCAAGTAGAG
CrmD58	R86A/K86A	CTCTACTTGATTACTGCTACAT <u>GC</u> GCCT <u>GC</u> ACAACTTAGACACGCGGGACTA
CrmD59	S90A/S91A	GTGTCTAAGTTGTCGAGGCAAATGT <u>GC</u> C <u>GC</u> TAATCAAGTAGAGACTAAATCGTGT
CrmD60	S90A/S91A	ACACGATTTAGTCTCTACTTGATTA <u>GC</u> G <u>GC</u> ACATTTGCCTCGACAACTTAGACAC
CrmD61	E116A/F117A/E118A	TGCATCCGGATACTACTGCG <u>CAGC</u> TG <u>C</u> AGGATCAAACGGTTGCAGGC
CrmD62	E116A/F117A/E118A	GCCTGCAACCGTTTGATCCT <u>G</u> CA <u>GC</u> T <u>G</u> CGCAGTAGTATCCGGATGCA
CrmD63	D146A/I148A	GTATATGGCTACTCATCTAAAGGAG <u>C</u> TGTA <u>GC</u> ATGTAAAAAGTGTCCGGGTAATATA
CrmD64	D146A/I148A	TATATTACCCGGACACTTTTTACAT <u>GC</u> TACA <u>G</u> CTCCTTTAGATGAGTAGCCATATAC

Supplementary table 1. Sequences of the primers used for the generation of CrmD mutants. The substituted bases are underlined.



Figure S1. CrmD D35A/Y36A mutant displays the same oligomeric state than wild type CrmD. The oligomeric state of wild type CrmD (WT) and the CRD1 mutant D35A/Y36A was compared by size exclusion chramathography (A) and electrophoresis in native gel (B). In A, 100 µg of WT (orange) and D35A/Y36A (blue) CrmD mutant in 1 ml of PBS were eluted in a Superdex 200 gel filtration column. In B, 500 ng of WT and D35A/Y36A were analyzed in a native gel using the PhastGel System. Proteins were loaded in a 12.5% homogeneous gel with native buffer strips. Subsequently, the gel was stained with coomasie blue.