

**Supplemental Table 1. Statistics of X-ray diffraction and structure refinement <sup>a</sup>**

	Native	Pt SAD	S SAD
<b>Data collection statistics</b>			
Space group	P4 <sub>3</sub> 2 <sub>1</sub> 2	P4 <sub>3</sub> 2 <sub>1</sub> 2	P4 <sub>3</sub> 2 <sub>1</sub> 2
$\alpha, \beta, \gamma, ^\circ$	90, 90, 90	90, 90, 90	90, 90, 90
Unit cell (a, b, c), Å	174.7, 174.7, 104.6	173.8, 173.8, 104.6	174.5, 174.5, 104.7
Resolution range (Å)	50.0-2.50 (2.56-2.50) <sup>b</sup>	50.0-3.50 (3.59-3.50)	50.0-3.30 (3.39-3.30)
Completeness (%)	98.7 (87.2)	99.9 (100.0)	99.9 (100.0)
Number unique reflections	55,726 (3,570)	38,576 (2,865)	46,450 (3,464)
Redundancy	14.1 (10.9)	15.6 (15.8)	13.4 (13.4)
R <sub>merge</sub> (%) <sup>c</sup>	15.5 (519)	15.9 (135)	10.9 (57.7)
I/ $\sigma$ (I)	13.3 (0.41)	16.3 (2.99)	21.6 (5.35)
CC <sub>1/2</sub> (%) <sup>d</sup>	99.9 (11.4)	99.9 (86.6)	99.9 (95.1)
Wavelength (Å)	0.97931	1.06975	1.90745
<b>Refinement statistics</b>			
R <sub>work</sub> (%) <sup>e</sup>	19.9 (37.1)		
R <sub>free</sub> (%)	24.2 (38.5)		
Bond RMSD (Å)	0.005		
Angle RMSD (°)	0.71		
Ramachandran plot <sup>f</sup> (Favored/allowed/outlier)	95.6/4.2/0.2		
Number of atoms			
Protein	7273		
Ligand	87		
Water	100		
B factor			
Protein	101.3		
Ligand	140.2		
Water	82.9		
Molprobit percentile (Clash/Geometry)	99/99		
PDB	6N29		

a. Anomalous data was collected at 1.06975 Å for platinum SAD and at 1.90745 for sulfur SAD. Only the 1.06975 Å data was strong enough to be useful for phasing or refinement. However, the 1.90745 data was useful for confirming locations of sulfurs in disulfide bonds and the identity and location of the Ca ion as shown in Supplemental Fig. 3.

b. The numbers in parentheses refer to the highest resolution shell.

c.  $R_{\text{merge}} = \sum_h \sum_i |I_i(h) - \langle I(h) \rangle| / \sum_h \sum_i I_i(h)$ , where  $I_i(h)$  and  $\langle I(h) \rangle$  are the  $i^{\text{th}}$  and mean measurement of the intensity of reflection  $h$ .

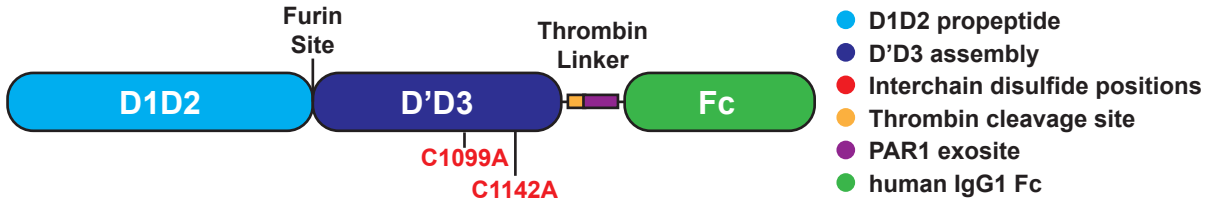
d. Pearson's correlation coefficient between average intensities of random half-datasets for each unique reflection<sup>1</sup>.

e.  $R_{\text{factor}} = \sum_h ||F_{\text{obs}}(h)| - |F_{\text{calc}}(h)|| / \sum_h |F_{\text{obs}}(h)|$ , where  $F_{\text{obs}}(h)$  and  $F_{\text{calc}}(h)$  are the observed and calculated structure factors, respectively. No  $I/\sigma(I)$  cutoff was applied.

f. Calculated with MolProbity<sup>2</sup>.

## References.

1. Karplus PA, Diederichs K. Linking crystallographic model and data quality. *Science*. 2012;336(6084):1030-1033.
2. Davis IW, Leaver-Fay A, Chen VB, et al. MolProbity: all-atom contacts and structure validation for proteins and nucleic acids. *Nucleic Acids Res*. 2007;35:W375-383.

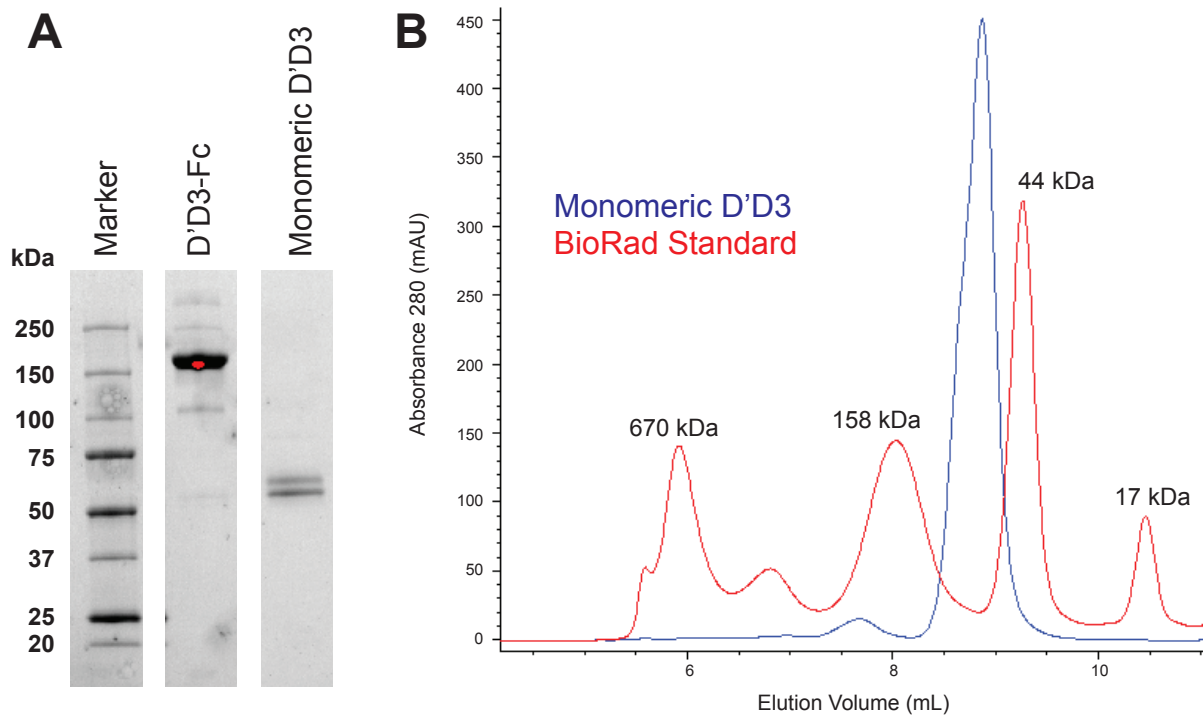


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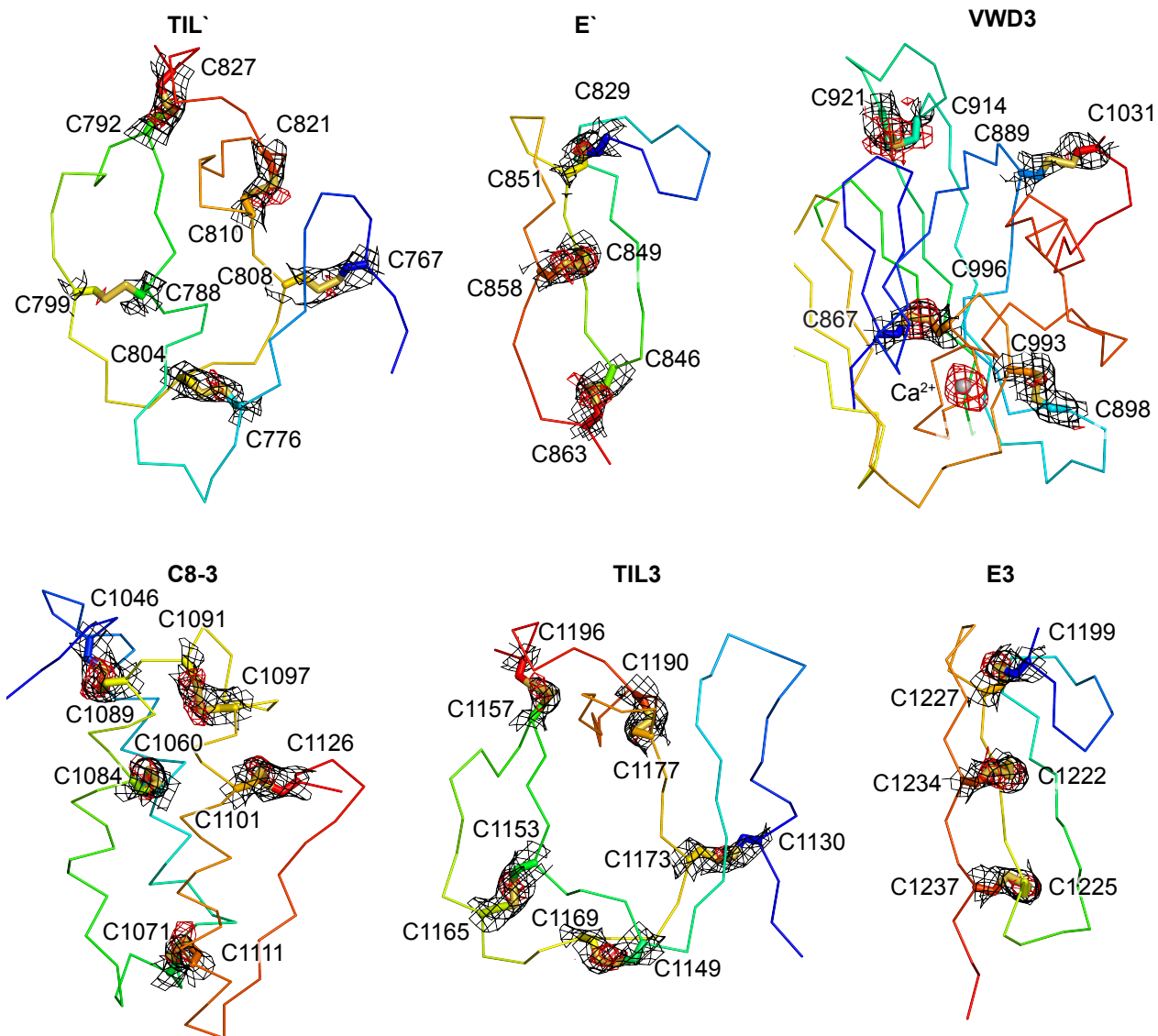
1  MIPARFAGVL  LALALILPGT  LCAEGTRGRS  STARCSLFGS  DFVNTFDGSM  YSFAGYCSYL
61  LAGGCQKRSF  SIIGDFQNGK  RVSLSVYLGE  FFDIHLFVNG  TVTQGDQQRVS  MPYASKGLYL
121  ETEAGYYKLS  GEAYGFVARI  DGSGNFQVLL  SDRYFNKTCG  LCGNFNIFAE  DDFMTQEGTL
181  TSDPYDFANS  WALSSGEQWC  ERASPPSSSC  NISSGEMQKG  LWEQCQLLKS  TSVFARCHPL
241  VDPEPFVALC  EKTLCECAGG  LECACPALLE  YARTCAQEGM  VLYGWTDHSA  CSPVCPAGME
301  YRQCVSPCAR  TCQSLHINEM  CQERCVDGCS  CPEGQLLDEG  LCVESTTEPC  VHS GKRYPPG
361  TSLSRDCNTC  ICRNSQWICS  NEECPGECLV  TGQSHFKSFD  NRYFTTFSGIC  QYLLARDCQD
421  HSFSIVIETV  QCADDRDAVC  TRSVTVRLPG  LHNSLVKCLKH  GAGVAMDGQD  IQLPLLKGD
481  RIQHTVTASV  RLSYGEDLQM  DWDGRGRLLV  KLSPVYAGKT  CGLCGNYNGN  QGDDFLTPSG
541  LAEPRVEDFG  NAWKLGDCQ  DLQKQHS DPC  ALNPRMTRFS  EEACAVLTSP  TFEACHRAVS
601  PLPYLRNCRY  DVCSCSDGRE  CLCGALASYA  AACAGRGVRV  AWREPGRCEL  NCPKGQVYLQ
661  CGTPCNLTCT  SLSYPDEECN  EACLEGCFCP  PGLYMDERGD  CVPKAQCPCY  YDGEIFQPED
721  IFSDHHTMCY  CEDGFMHCTM  SGVPGSLLPD  AVLSSPLSHR  SKRSLSCRPP  MVKLVCPADN
781  LRAEGLECTK  TCQNYDLECM  SMGCVSGCLC  PPGMVRHENR  CVALERCPCF  HQGKEYAPGE
841  TVKIGCNTCV  CRDRKWNCTD  HVC DATCSTI  GMAHYLTFDG  LKYLFPGECQ  YVLVQDYCGS
901  NPGTFRILVG  NKGCSHPSVK  CKKRV TILVE  GGEIELFDGE  VNVKRPMKDE  THFEVVE SGR
961  YIILLGKAL  SVVWDRHLSI  SVVLKQTYQE  KVCGLCGNFD  GIQNNDLTSS  NLQVEEDPVD
1021  FGNSWKVSSQ  CADTRKVPLD  SSPATCHNNI  MKQTMVDSSC  RILTSDFVQD  CNKLVDP EPY
1081  LDVCIYDTCS  CESIGDCAAF  CDTIAAYAHV  CAQH GKVV TW  RTATLCPQSC  EERNLRENGY
1141  EAEWRYNSCA  PACQVTCQHP  EPLACPVQCV  EGCHAHCPPG  KILDELLQTC  VDPEDCPVCE
1201  VAGRRFASGK  KVTLNPSDPE  HCQICHCDVV  NLTCEACQEP  GGLVPRS FLL  RNPNDKYEPF
1261  WEDEESDKTH  TCPPCPAPEL  LGGPSVFLFP  PKPKDTLMIS  RTPEVTCVVV  DVSHEDPEVK
1321  FNWYVDGVEV  HNAKTKPREE  QYNSTYRVVS  VLTVLHQDWL  NGKEYKCKVS  NKALPAPIEK
1381  TISKAKGQPR  EPQVYTLPPS  RDELTKNQVS  LTCLVKGFYP  SDIAVEWESN  GPENNYKTT
1441  PPVLDS DGSF  FLYSKLTVDK  SRWQQGNVFS  CSVMHEALHN  HYTQKSL SLS  PGK

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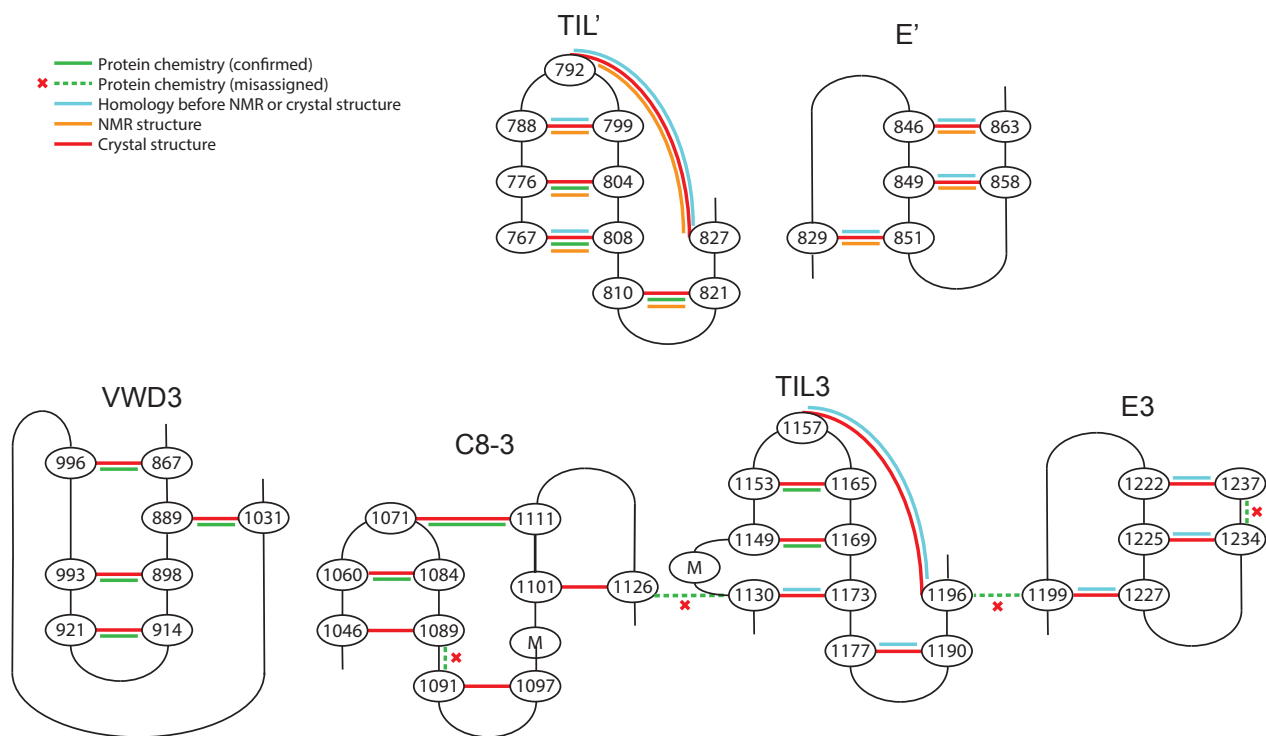
**Supplemental Figure 1.** Cartoon representation of the D1D2D'3-Fc expression construct with its sequence below and colored according to the regions in the cartoon. The plasmid containing the construct, pSYN-VWF-039, has been deposited at AddGene (Watertown, MA) with ID 120292.



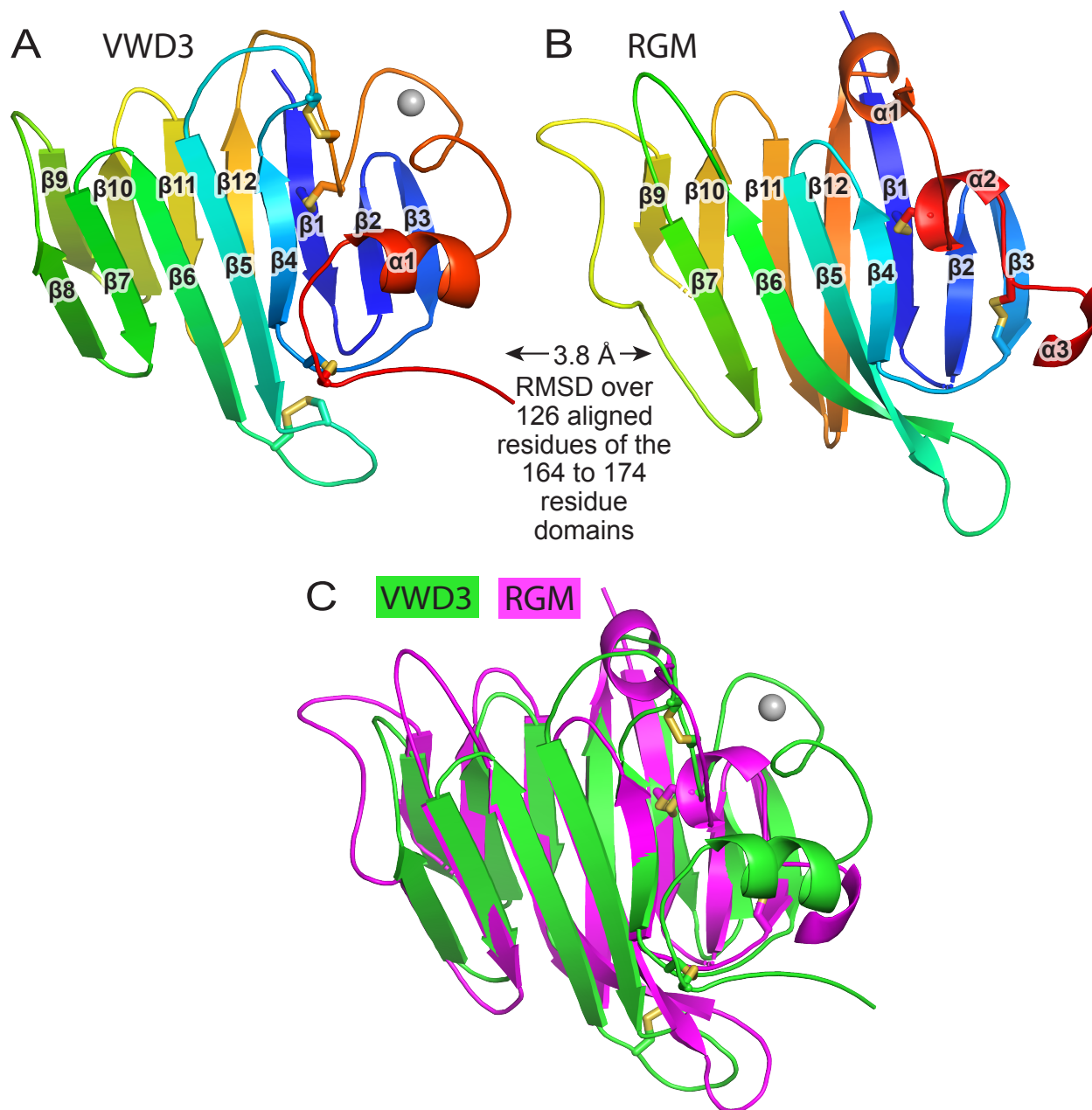
**Supplemental Figure 2.** A. Non-reducing SDS-PAGE showing purified D'D3-Fc prior to thrombin and Endo Hf cleavage and the final, thrombin and endo Hf-cleaved D'D3 monomer. B. The same D'D3 monomer was run on analytical size exclusion chromatography to confirm its purity and monomeric state. BioRad standard is shown for comparison.



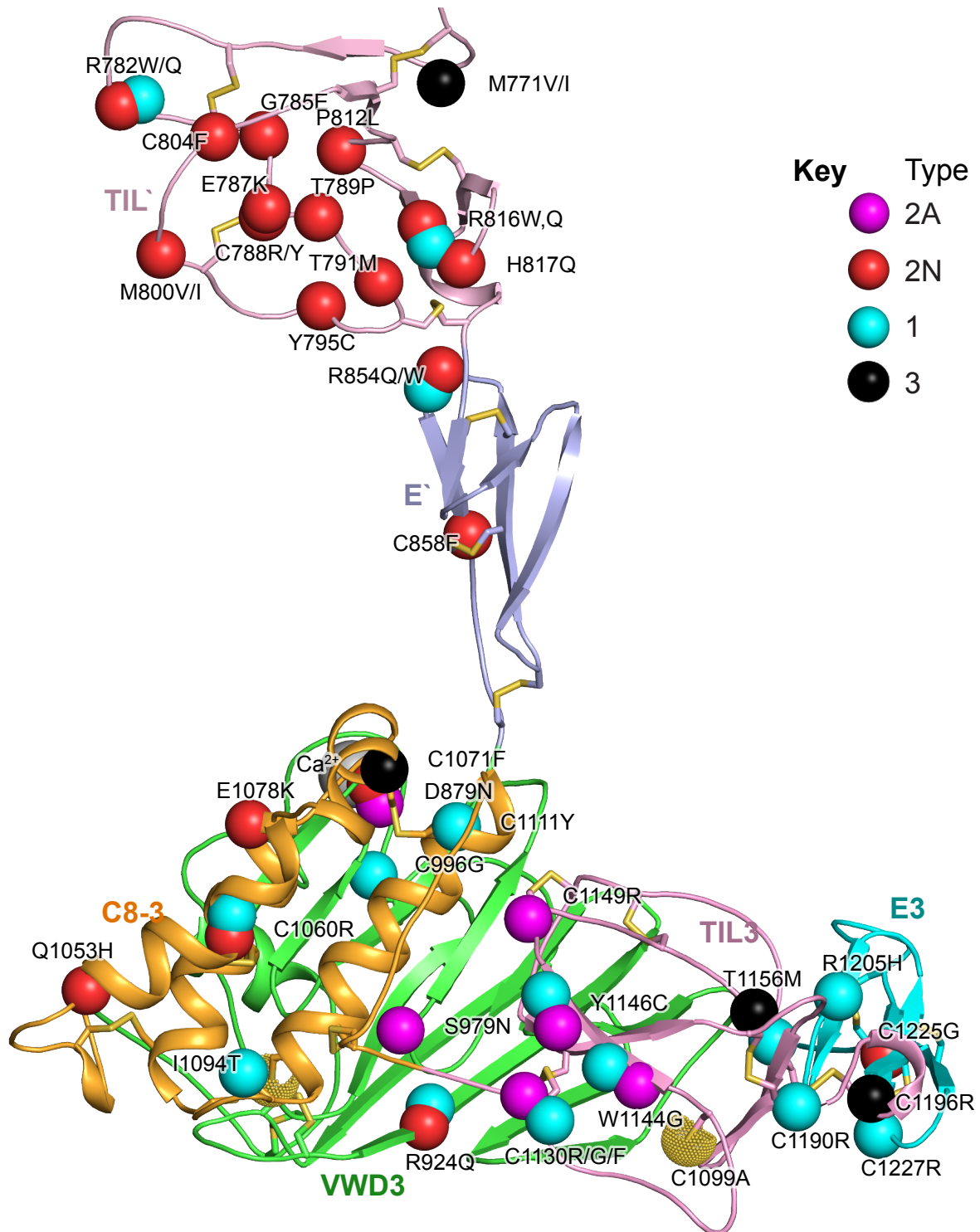
**Supplemental Figure 3.** Electron density around cysteine sulfurs and Ca<sup>2+</sup> in D'D3. Anomalous difference maps from 1.9 Å-wavelength diffraction contoured at 3  $\sigma$  (red mesh) and carved at 2 Å around cysteine residues and Ca<sup>2+</sup> indicates the position of sulfur and calcium atoms. Composite omit simulated annealing maps contoured at 1  $\sigma$  (black mesh) are similarly carved within 2 Å of cysteine residues. The densities confirm evidence for the disulfide bonds defined in the structure. The identity of the metal ion as Ca<sup>2+</sup> is confirmed by the anomalous diffraction at 1.9 Å-wavelength, together with excellent agreement of the 2Fo-Fc maps from diffraction at 0.979 Å-wavelength with the density expected for Ca<sup>2+</sup>, the metal-oxygen distances of 2.3 and 2.4 Å, the propensity of Ca<sup>2+</sup> to coordinate carbonyl oxygens, and the octahedral coordination geometry.



**Supplemental Figure 4.** Graphical representation of the disulfide bonds defined by different methods in the domains of D'D3. Green lines denote disulfide bonds that were determined by protein chemistry (Marti, T., Rosselet, S. J., Titani, K., and Walsh, K. A. (1987) *Identification of disulfide-bridged substructures within human von Willebrand factor. Biochemistry* 26, 8099-8109) and subsequently confirmed (solid lines) or found to be incorrect (dashed lines with red asterisks). Cyan lines show all disulfide bonds predicted by homology and loss or absence of pairs of cysteines (Zhou, Y. F., Eng, E., Zhu, J., Lu, C., Walz, T., and Springer, T. A. (2012) *Sequence and structure relationships within von Willebrand factor. Blood* 120, 449-458). Orange lines show disulfides found in the NMR structure of D' (Shiltagh, N., Kirkpatrick, J., Cabrita, L. D., McKinnon, T. A., Thalassinou, K., Tuddenham, E. G., and Hansen, D. F. (2014) *Solution structure of the major factor VIII binding region on von Willebrand factor. Blood* 123, 4143-4151). Red lines show disulfide bonds found here in the crystal structure.



**Supplemental Figure 5.** Structural homology of the VWD domains in VWF D3 and repulsive guidance molecules (RGM). (A and B) Superimposed domains are shown in rainbow in identical orientations or (C) in different colors. Representations are as in Fig. 1. RGM domain is from PDB ID code 4BQ6. Superposition was with the Deep Align server (Wang, S., Ma, J., Peng, J., and Xu, J. (2013) *Protein structure alignment beyond spatial proximity. Sci. Rep.* 3, 1448).



**Supplemental Figure 6.** Locations of VW disease mutations within D'D3 are shown as spheres at mutated residues. Each type of mutation is color coded in the key. When more than two disease types are reported for a residue, a sphere is shown for each disease type. To facilitate display, mutated residues are displayed with spheres on amino acid residue backbone atoms Ca (type 2N), C (type 1), and N (type 2A and 3, which currently have no overlapping residues). Mutations were taken on Aug. 19, 2018 from the International Society for Hemostasis and Thrombosis Database. Hampshire, D. J., and Goodeve, A. C. (2011) *The International Society on Thrombosis and Haematosi von Willebrand disease database: an update. Semin. Thromb. Hemost.* 37, 470-479.



**Supplemental Video 1.** D'D3 structure. D'D3 is rotated. Figures prepared with PyMol show ribbon cartoon and disulfides and N-glycans with white carbons in stick, red oxygens, blue nitrogens, and yellow sulfurs.  $\text{Ca}^{2+}$  is shown as a silver sphere. A1099 and A1142 are modeled as cysteines using the one allowed rotameric position of the S atom, as discussed in Results. Dot surfaces around the S atom emphasize that there is room for it in the structure and that it is a model.

**Supplemental Video 2.** Structure around the buried cysteine residues required for D3 dimerization. A1099 and A1142 are shown as cysteines as described in the legend for Video 1, and sidechains of residues that bury the Cys sidechains are shown as stick. The movie zooms in to view the position of each mutated Cys residue, first residue 1099 in C8-3 and then residue 1142 in TIL3.