

Supplementary Materials for

**Antioxidant Activity and Antibacterial Evaluation of New Thiazolin-4-one Derivatives as Potential Tryptophanyl-tRNA Synthetase Inhibitors**

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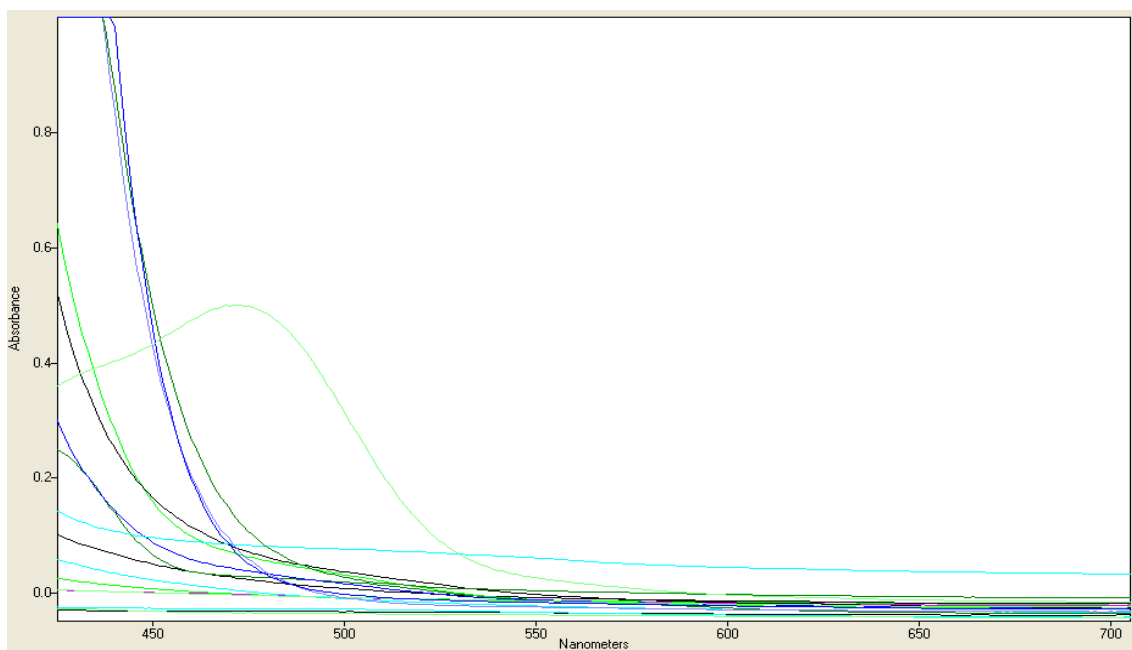


Figure S1. Overlapping of the absorption spectra of the compounds in the region 430-700 nm. None of the tested compounds have absorption peaks near the wavelengths at which the antioxidant and antiradical assays were performed (517 nm, 593 nm, 695 nm and 700 nm).



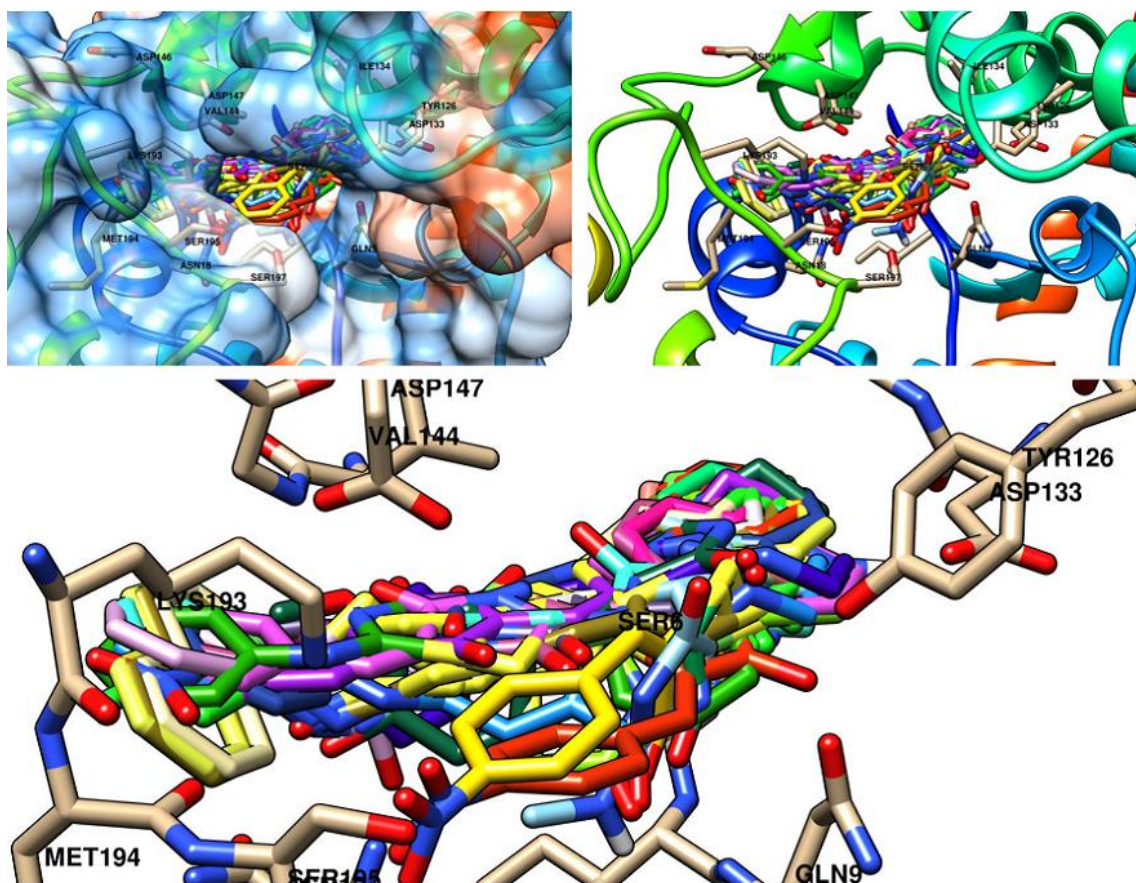


Figure S3. Docking poses of the screened compounds in the active site of *S. aureus* TrpRS: top left image – the target is depicted as thin sticks with secondary structure drawn as ribbon, with simulation of the molecular surface for a better understanding of tridimensional positioning in the active site of enzyme, meanwhile ligands are figured as sticks; top right image - ribbons and bars representation; down image - bars representation

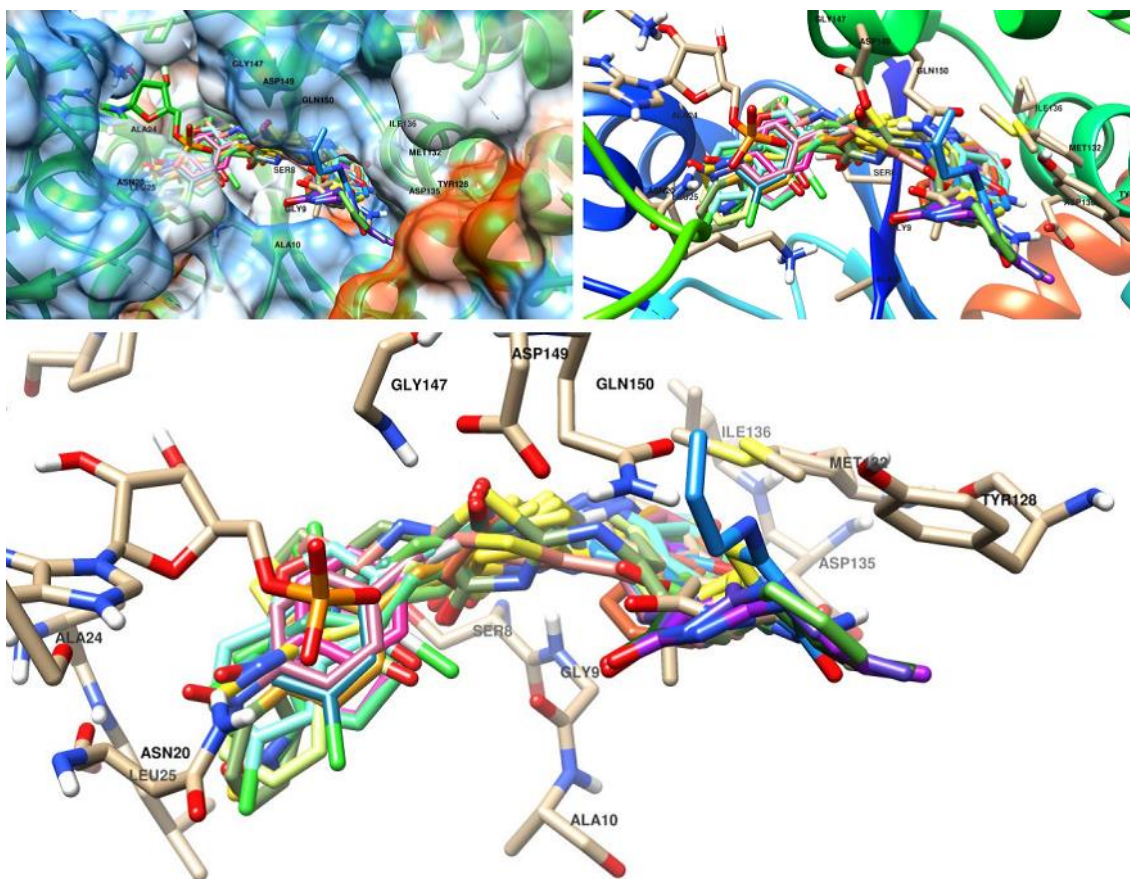


Figure S4. Docking poses of the screened compounds in the active site of *E. coli* TrpRS: top left image – the target is depicted as thin sticks with secondary structure drawn as ribbon, with simulation of the molecular surface for a better understanding of tridimensional positioning in the active site of enzyme, meanwhile ligands are figured as sticks; top right image - ribbons and bars representation; down image - bars representation

The sequence alignment between the primary sequence of P67592\_1I6K (*S. aureus*) and that of PDB 5V01 (*E. coli*) revealed a certain degree of similitude between these two structures, but also some significant differences (Figures S5, S6 and Table T1). As results of the alignment between the two sequences, EMBOSS Stretcher returned the following scores: identity=49.2% (164/333), similarity=66.1% (220/333) and gaps=5.1% (17/333). A degree of conservation between the two structures can be found, but some mutations can be ascertained, even in the catalytic site – the 8<sup>th</sup> amino acid is isoleucine in P67592\_1I6K (*S. aureus*) and alanine in 5V01 (*E. coli*). We can



conclude that, strictly regarding the primary structure, there are significant differences between the two enzymes.

P67592_1I6K	1	METL <b>FSGIQ</b> PSGIPT <b>IGNY</b> IGALKQFVDVQNDYDCYFCIVDQ <b>HAITMPQD</b>	50
		...:      .       .           :     :     :     .   :               : .	
5V0I	1	KPIV <b>FSGAQ</b> PS--LT <b>IGNY</b> MGALRQWVNMQDDY <b>HC</b> IYCIVDQ <b>HAITVRQD</b>	48
P67592_1I6K	51	RLKLRKQTRQLAAIYLASGIDPDKATLFI <b>Q</b> SEVPAHVQAGWMLTTIASVG	100
		. .         .   . . . .   :       .         :   :   :     .   .   .   .   . . . .	
5V0I	49	AQKLRKATLDTLALYLACGIDPEKSTIFV <b>Q</b> SHVPEHAQLGWALNCYTYFG	98
P67592_1I6K	101	ELE <del>R</del> MTQYKDKAQA <b>Q</b> AVEGIPAGLLTY <b>PPLMAAD</b> IVLYNTNIV <b>PVGDDQ</b> KQ	150
		.       . . . .   .   .         . .     .           :     .     :         :	
5V0I	99	ELSRMT-----AENINAGLFDY <b>PV</b> L <b>MAAD</b> ILLYQTNLVP <b>VGEDQ</b> KQ	139
P67592_1I6K	151	<b>H</b> IELTRNLVDRFNSRYNDVLVKPEIRMPK <b>VGGRVMSLQDPTRKMSK</b> SDDN	200
		:     :   : . .       : .   : . . .     . . :     .   .           . :     :	
5V0I	140	<b>H</b> LELSRDIAQRFNALYGDIFKVPEPFIPKSGAR <b>VMSLLEPTKMSK</b> SDDN	189
P67592_1I6K	201	AKNFISLLDEPNVA <b>AK</b> IKSAVTDSDG-- <b>I</b> IKFDRDNKPGITNLIS <b>IYAG</b>	248
		. .   .   .     : :   . . . .         .           . : : :   .     .   :     :   .   . . .	
5V0I	190	RNNVIGLLEDPKSVVKKIKRAVTDSD <b>E</b> PPVVRVYDVQNKAGVSNLLDILSA	239
P67592_1I6K	249	LTDMP <b>IKDIEAKYEGEGY</b> GKFKGDLAEIVKAFLVEF <b>QEKY</b> ESFYNSDK-L	297
		:   . . .   . : :   . :     : .     .     : :   : .   . .   .   .     :   .   .   . .	
5V0I	240	VTGQ <b>S</b> IPELEK <b>Q</b> FEGKMYGHLKGEVADAVSGMLTELQERYHRFRNDE <b>AF</b> L	289
P67592_1I6K	298	DDILDQGRDKAHKVSFKTVKKMEKAMGL---GR	327
		. . : . .   . :     . . . . :   :   . : :   :   . . .	
5V0I	290	QQVMKDGAEKASAHASRTLKAVYEAIGFVAKPX	322

Figure S5. Sequence alignment between the primary sequences of P67592\_1I6K (*S. aureus*) and PDB 5V0I (*E. coli*). Conserved residues are depicted with a vertical line between the two sequences. Following the BLASTP analysis using the two sequences, the amino acids involved in catalyzing the reaction are depicted in red.

The enzyme 1I6K\_P67592 of *S. aureus* has a smaller catalytic site compared to PDB 5V0I from *E. coli*, both in terms of volume (966.98 Å<sup>3</sup> vs. 1541.63 Å<sup>3</sup>), inner surface (1043 Å<sup>2</sup> vs. 1810.80 Å<sup>2</sup>) and depth (27.73 Å vs. 30.29 Å). It has a “Y”-shape, with the distal region heading to the outside of the enzyme, while the *E. coli* enzyme has multiple pockets, with ramifications from its center of catalytic site. The existence of multiple branches can lead to the existence of other sites with unknown functions,

besides the catalytic site. The effect of binding ligands in these pockets may be difficult to predict regarding the possibility and the manner in which these bindings might affect the enzyme activity modulation.

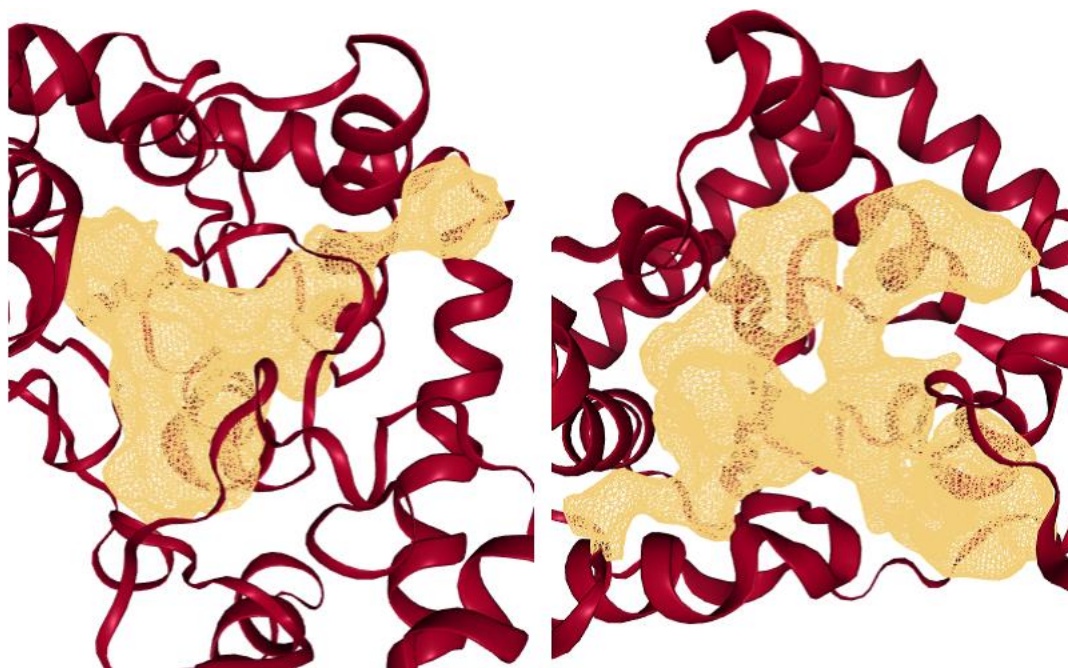


Figure S6. The internal accessible volume in the catalytic site of tryptophanyl-tRNA synthetases used in the molecular docking study. Binding pockets of 1I6K\_P67592 from *S. aureus* (left) and of PDB 5V0I from *E. coli* (right) are depicted in yellow mesh.

Table T1. Comparative description of the two tryptophanyl-tRNA synthetases

Parameter	Enzyme binding pocket	
	1I6K_P67592 ( <i>S. aureus</i> )	PDB 5V0I ( <i>E. coli</i> )
Volume (Å <sup>3</sup> )	966.98	1541.63
Surface (Å <sup>2</sup> )	1043.82	1810.80
Depth (Å)	27.73	30.29
Hydrogen bond donors	31	39
Hydrogen bond acceptors	65	85
Hydrophobic interactions	28	76
Hydrophobicity ratio	0.23	0.38
Non-polar amino acid ratio	0.37	0.50
Polar amino acid ratio	0.43	0.37
Positive amino acid ratio	0.14	0.10
Negative amino acid ratio	0.06	0.03