# **SI Appendix**

# Singlet Oxygen Mediated Iron-based Fenton-like Catalysis under Nanoconfinement

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# **Part S1 Supplementary Discussion**

### Effect of pH on <sup>1</sup>O<sub>2</sub> generation and MB removal

In this section, we would like to discuss the effect of pH (from 2.0 - 9.0) on the MB removal in the Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system from three perspectives, *i.e.*, the reaction between  ${}^{1}O_{2}$  and MB, the generation of  ${}^{1}O_{2}$ , the adsorption of MB.

*Reaction between*  ${}^{1}O_{2}$  *and MB*. It is accepted that pH would affect a chemical reaction through the protonation/deprotonation of the reactants (S1). In the case of reaction between  ${}^{1}O_{2}$  and MB, as the status of MB is constant in the pH range of 2.0 - 9.0 (S2), we exclude the pH factor.

 ${}^{1}O_{2}$  generation. Based on the Haber-Weiss cycle,  ${}^{1}O_{2}$  should be generated from  $HO_2 \cdot O_2 \cdot (S3)$ . We are convinced by two facts, one is MB could not be degraded by HO<sub>2</sub>•/O<sub>2</sub>•-, which is generated in xanthine-xanthine oxidase system (SI Appendix, Fig. S12), the other is the addition of a typical  $HO_2 \bullet / O_2 \bullet^-$  quenching molecule, superoxide dismutase (SOD), effectively suppressed MB degradation in our Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system (SI Appendix, Fig. S13). These two facts provide solid evidence for the  ${}^{1}O_{2}$  generation through HO<sub>2</sub>•/O<sub>2</sub>• as the intermediate. We then investigated the generation of HO<sub>2</sub>•/O<sub>2</sub>• and <sup>1</sup>O<sub>2</sub> under different pH conditions. The normalised double integration of the ESR spectra was used to semi-quantify both  $HO_2 \bullet /O_2 \bullet^-$  and  $^1O_2$  (S4-6). SI Appendix, Fig. S14A shows that the amount of  $HO_2 \bullet /O_2 \bullet^$ increases dramatically from pH 2.0 to 3.0, and then remains almost stable from pH 3.0 to 9.0. SI Appendix, Fig. S14B shows that the amount of  ${}^{1}O_{2}$  increases steadily from pH 2.0 to 6.0, and then remains stable from pH 6.0 to 9.0. Based on the pKa value of  $HO_2 \bullet / O_2 \bullet$  (4.8), we could calculate the amount of  $O_2 \bullet$  at various pH values, and plotted them versus the amount of  ${}^{1}O_{2}$  in SI Appendix, Fig. S14C, showing a linear relationship. Meanwhile, the amount of  ${}^{1}O_{2}$  is linearly proportional to the MB degradation constant  $k_{app}$ , as shown in SI Appendix, Fig. S14D. Consequently, we propose that pH affects the <sup>1</sup>O<sub>2</sub> generation for MB degradation through affecting the protonation/ deprotonation status of HO<sub>2</sub>•/O<sub>2</sub>•-.

Adsorption of MB. SI Appendix, Fig. S15A shows the improved MB adsorption as the increase of pH value for Fe<sub>2</sub>O<sub>3</sub>@FCNT-H, with the maximum adsorption capacity at equilibrium ( $q_{e,cal}$ ) positively correlated with MB degradation  $k_{app}$  (SI Appendix, Fig. S15B). Since we have shown an adsorption-dependent removal feature of Fe<sub>2</sub>O<sub>3</sub>@FCNT-H in Fig. 6A in the main text, we reckon that pH could affect the MB

adsorption, and thus affect the MB removal.

Thus, for the  $Fe_2O_3@FCNT-H/H_2O_2$  system, pH plays a crucial role in the MB removal through affecting the  ${}^1O_2$  generation as well as the MB adsorption.

### Part S2 Supplementary Experimental Section

#### Text S1 The intermediate products detection by UHPLC/MS

An UHPLC/MS with an electron spray ionization (ESI) source was employed to identify the intermediate products.

(i) The intermediate products from the oxidation of DPA

$$(1)$$

10 µM DPA was oxidized in the Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system. The reaction was conducted in a 10-mL Teflon-line screw-cap glass vial under magnetic stirring at  $293.2 \pm 0.3$  K. 50 mM ACN was added to ensure the dissolution of DPA. The reaction aliquot was withdrawn at the end of the reaction (60 min) and filtered through a PTFE membrane (0.22 µm) to remove the solid catalysts for analysis. UHPLC/MS with an ESI source in the positive ionization mode was used for analysis. An Agilent Eclipse plus C18 column (4.6  $\times$  100 mm, 3.5  $\mu$ m particle size) was used for UHPLC separation. The flow rate was set at 0.2 mL·min<sup>-1</sup>. To eliminate the possible contamination of mass spectrometer by the oxidants and other impurities in the samples, the UHPLC effluent in the first 2 minutes was diverted to the waste through a switching valve between the UHPLC column outlet and the mass detector inlet. The gradient mobile phase consisted of ACN (A) and Milli-Q water (B), which increased linearly from 65/35 to 95/5 (v/v, A/B) in first 25 min and held for 10 min, and then returned to 65/35 in 0.5 min, and kept for 5 min for re-equilibration. The MS parameters were optimized and set as follows: scan type, full mass; resolution, 35000; sheath gas flow rate, 20 arbitrary units; aux gas flow rate, 10 arbitrary units; spray voltage, 3000 V; capillary temperature, 593.2 K.

(ii) The intermediate products from the oxidation of FFA

$$(2)$$

A relatively high concentration of FFA (1 or 5 mM) was oxidized in the

Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system. The reaction was conducted in a 10-mL Teflon-line screw-cap glass vial under magnetic stirring at 293.2  $\pm$  0.3 K. The reaction aliquots were withdrawn after 60 min and only 2.0% and 3.4% of FFA were degraded, respectively. The samples were filtered through a hydrophilic polyether sulfone (PES) membrane (0.22 µm) to remove the solid catalysts for subsequent analysis. The UHPLC/MS with an ESI source in the negative ionization mode was employed for analysis. An Agilent Eclipse plus C18 column (4.6  $\times$  100 mm, 3.5  $\mu$ m particle size) was used for UHPLC separation. The flow rate was set at 0.2 mL·min<sup>-1</sup>. To eliminate the possible contamination of mass spectrometer by the oxidants and other impurities in the samples, the UHPLC effluent in the first 2 minutes was diverted to the waste through a switching valve between the UHPLC column outlet and the mass detector inlet. The gradient mobile phase consisted of MeOH (A) and Milli-Q water (B), which increased linearly from 5/95 to 20/80 (v/v, A/B) in first 20 min and hold for 15 min, and then returned to 5/95 in 0.5 min, and kept for 5 min for re-equilibration. The MS parameters were optimized and set as follows: scan type, full mass; resolution, 35000; sheath gas flow rate, 35 arbitrary units; aux gas flow rate, 10 arbitrary units; spray voltage, 2500 V; capillary temperature, 573.15 K.

#### Text S2 Calculation of the concentration of products

The standard sample of the products (*i.e.*, C<sub>5</sub>H<sub>6</sub>O<sub>4</sub>, C<sub>4</sub>H<sub>4</sub>O<sub>3</sub>, and C<sub>5</sub>H<sub>6</sub>O<sub>3</sub>) from the oxidation of FFA by <sup>1</sup>O<sub>2</sub> cannot be readily available. Thus, the calculation of their concentrations from the UHPLC peak area was conducted on the basis of the molar adsorption coefficients using FFA as a reference at 219 nm, as reported by Richard et al (S7). The molar adsorption coefficient of C<sub>5</sub>H<sub>6</sub>O<sub>4</sub>, C<sub>4</sub>H<sub>4</sub>O<sub>3</sub>, C<sub>5</sub>H<sub>6</sub>O<sub>3</sub>, and FFA was 1320 ± 40, 2000, 7500 ± 200, and 8100 ± 60 M<sup>-1</sup>·cm<sup>-1</sup>, respectively (S8). Thus, the concentration of C<sub>5</sub>H<sub>6</sub>O<sub>4</sub>, C<sub>4</sub>H<sub>4</sub>O<sub>3</sub>, and C<sub>5</sub>H<sub>6</sub>O<sub>3</sub> was calculated on the basis of the Lambert-Beer law:

$$A = \varepsilon bc \tag{3}$$

Where A is the absorbance (here is the peak area),  $\varepsilon$  represents the molar adsorption coefficient, b is the optical depth through the sample, c is the concentration of the sample.

$$\frac{A_{product}}{A_{FFA}} = \frac{\varepsilon_{product}}{\varepsilon_{FFA}} \times \frac{c_{product}}{c_{FFA}}$$
(4)

$$c_{product} = \frac{A_{product}}{A_{FFA}} \times \frac{\varepsilon_{FFA}}{\varepsilon_{product}} \times c_{FFA}$$
(5)

We obtained  $c_{\text{FFA}}$  /  $A_{\text{FFA}}$  of 1/24330 from the inverse slope of the standard calibration of FFA. Thus, the above equation can be further simplified:

$$c_{product} = \frac{\varepsilon_{FFA} \times A_{product}}{24330 \times \varepsilon_{product}}$$
(6)

According to eq 6, the concentration of the products can be calculated.

#### **Text S3 Chemicals and materials**

Multi-walled carbon nanotubes (MWCNTs, I.D. 5-10 nm, O.D. 10-20 nm, length 10-30 µm) were purchased from Chengdu Organic Chemicals Co., Ltd. (China) with purity of  $\geq$  98%. Mesoporous silica (SBA-15, I.D. 6-11 nm), and graphene oxide (GO) were obtained from Nanjing XFNANO Materials Tech Co., Ltd. (China). Aniline, dimethyl sulfoxide (DMSO), superoxide dismutase (SOD), furfuryl alcohol (FFA), tertiary butanol (TBA), and 1,10-phenanthroline of analytical grade or better were obtained from Shanghai Aladdin Biochemical Technology Co., Ltd. (China). Fe(NO<sub>3</sub>)<sub>3</sub>.9H<sub>2</sub>O, HNO<sub>3</sub>, HCl, H<sub>2</sub>O<sub>2</sub> (in water), acetone, methylene blue (MB), methyl violet (MV), crystal violet (CV), chrysoidine, malachite green oxalate (MGO), methyl orange (MO), and chromotrope 2R (C2R) were received from Sinopharm Chemical Reagent Co., Ltd. (China) and used without further purification. Their basic properties S3. are shown in SI Appendix, Table 4-Chlorophenol (4-CP), 2,2,6,6-tetramethyl-4-piperidinol (TEMP), 5,5-dimethyl-1-pyrolin-N-oxide (DMPO), and 9,10-diphenylanthracene (DPA) of ACS reagents were purchased from Sigma-Aldrich (U.S.A). Methanol (MeOH) and acetonitrile (ACN) of HPLC grade were purchased from Merck (Germany). All of the stock solutions were prepared in ultrapure water (18.25 MΩ.cm) except insoluble DPA. 1 mM DPA stock solution was prepared using ACN as the solvent.

### Text S4 The transfer of XRD patterns

The crystalline structures of the catalysts were determined by X-ray diffraction analysis (XRD, D-MAX Rapid-II, Rigaku, Japan) using Mo K $\alpha$  radiation ( $\lambda = 0.7093$  Å). For better analysis and comparison with the data from XRD in Cu K $\alpha$  radiation, the obtained XRD spectra were transferred according to the Bragg equation as follows :

$$2d\sin\theta = n\lambda \tag{7}$$

Where *d* is the interplanar crystal spacing, Å;  $\theta$  is the angle between X ray and the crystal surface; n represents diffraction series;  $\lambda$  is the wavelength of X ray. For a

given sample, d-spacing will not change regardless of the X ray species and thus the transformation of  $\theta$  in different model can be achieved as follows:

$$\frac{\sin \theta_{Cu}}{\sin \theta_{Mo}} = \frac{\lambda_{Cu}}{\lambda_{Mo}}$$
(8)

$$\theta_{Mo} = \arcsin(\frac{\lambda_{Cu}}{\lambda_{Mo}} \times \sin \theta_{Mo}) \tag{9}$$

#### Text S5 Evaluating the stability and reusability of Fe<sub>2</sub>O<sub>3</sub>@FCNT-H

For the consecutive MB addition runs without regeneration, the degradation experiment was conducted in a 500-mL conical flask. The reaction was initiated by simultaneous addition of  $4.0 \times 10^{-2}$  g·L<sup>-1</sup> catalyst suspension and 50 mM H<sub>2</sub>O<sub>2</sub> into the solution containing 10 µM MB. The reaction aliquots of 4 mL were periodically sampled by pipette and filtered through a membrane (0.22 µm) to remove the solid catalysts for MB analysis. After 60-min reaction, MB was completely degraded, and  $10 \ \mu M$  MB was added again. For the regeneration experiment, to avoid the loss of catalysts in the cyclic runs, a large number of catalysts were prepared in an amplifying reactor. Specially, 0.14 g Fe<sub>2</sub>O<sub>3</sub>@FCNT-H was divided equally into two 1.75 L solutions containing 10 µM MB. The reaction was initiated by adding 50 mM H<sub>2</sub>O<sub>2</sub>. After 60-min reaction, the catalysts were collected by suction filtration, followed by rinsing with ethanol (EtOH) and ultrapure water in sequence for five times, the catalysts were lyophilized. The obtained solids were denoted as Fe<sub>2</sub>O<sub>3</sub>@FCNT-H-2nd, and we got Fe<sub>2</sub>O<sub>3</sub>@FCNT-H-3rd following the same procedure mentioned above. For each run, 10 mg Fe<sub>2</sub>O<sub>3</sub>@FCNT-H-x (x = 2nd, 3rd) was weighed by micro-balance (XP56, Mettler Toledo, Switzerland) and used for the reaction conducted in a 250 mL conical flask, where the MB solution was sampled and analyzed.

#### Text S6 The operation methods for reactive intermediates analysis by ESR

*Qualitative analysis.* The reactions were carried out in 1 mL centrifuge tubes.  $1.5 \times 10^{-2}$  g·L<sup>-1</sup> catalyst (*i.e.*, Fe<sub>2</sub>O<sub>3</sub>, Fe<sub>2</sub>O<sub>3</sub>/FCNT-L, or Fe<sub>2</sub>O<sub>3</sub>@FCNT-H), 50 mM H<sub>2</sub>O<sub>2</sub> were added orderly to 160 µL solution containing 50 mM DMPO/ TEMP to made up the volume to 200 µL. After ca. 5 min, the solutions were sucked to capillary tubes and determined on an ESR A300 spectrometer.

*Semi-quantitative analysis.* We conducted normalised double integration of the obtained ESR spectra to semi-quantify the concentration of the generated  $HO_2 \cdot /O_2^{-1}$  and  $^1O_2$  on WINEPR Processing system (S4-6). The integral intervals were set as

3455 - 3508 G for the DMPO-OOH signal and 3457 - 3505 G for the TMPN signal. As for HO<sub>2</sub>•/O<sub>2</sub>•, MeOH and water (20:1, v:v) was mixed and adjusted the solution pH to 2.0 - 9.0 by 0.01 M HCl and 0.01 M NaOH in advance. In the detection process, 50 mM DMPO,  $1.5 \times 10^{-2}$  g·L<sup>-1</sup> Fe<sub>2</sub>O<sub>3</sub>@FCNT-H, and 50 mM H<sub>2</sub>O<sub>2</sub> were added orderly to 300 µL solution with different pH. The DMPO-OOH signal was detected after 60 min of reaction. As for <sup>1</sup>O<sub>2</sub> production, the pH of 100 mM TEMP solution is 10.1 and thus the solution pH is adjusted to ca. 4.8 in advance. In the detection process, 10 mM TEMP,  $1.5 \times 10^{-2}$  g·L<sup>-1</sup> Fe<sub>2</sub>O<sub>3</sub>@FCNT-H, and 50 mM H<sub>2</sub>O<sub>2</sub> were added orderly to 300 µL solution with different pH. The TMPN signal was detected after 60 min. **Part S3 Supplementary Figures** 



**Fig. S1.** TEM images of (*A*) FCNT-L and (*B*) FCNT-H. The insets are inner diameter distribution of both CNTs.



**Fig. S2.** STEM-HADDF images of (*A*) Fe<sub>2</sub>O<sub>3</sub>/FCNT-L and (*B*) Fe<sub>2</sub>O<sub>3</sub>@FCNT-H.



**Fig. S3.** <sup>57</sup>Fe Mössbauer spectra of both  $Fe_2O_3/FCNT-L$  and  $Fe_2O_3@FCNT-H$  at room temperature. Both samples show the central doublets in the spectra which may due to the ultrafine nanoparticles as verified by TEM images. The spectra confirm the sole presence of  $Fe^{3+}$  species by the uniform isomer shift (IS) of ca.0.36 mm·s<sup>-1</sup> relative to alpha-iron (S9-12). Combined with the quadrupole splitting (QS) of the fitted doublets in the range of 0.69 mm·s<sup>-1</sup> - 1.23 mm·s<sup>-1</sup>, we can confirm the existence of  $FeO_6$ -type octahedral coordination for both samples (S9-11).



Fig. S4. Raman spectra of the catalysts.



**Fig. S5**. The molecular mass spectra of the detected (*A*)  $C_5H_6O_4$ , (*B*)  $C_4H_4O_3$ , (*C*)  $C_5H_6O_3$ , and (*D*) FFA in negative ionization mode by UHPLC/MS.



**Fig. S6.** The kinetics of MB degradation in both  $Fe_2O_3@FCNT-H/H_2O_2$  system and  $Fe_2O_3/FCNT-L/H_2O_2$  system. Reaction conditions: pH = 5.0, T = 293.2 K,  $[Fe_2O_3@FCNT-H] = [Fe_2O_3/FCNT-L] = 1.5 \times 10^{-2} \text{ g}\cdot\text{L}^{-1}$ ,  $[MB] = 10 \ \mu\text{M}$ ,  $[H_2O_2] = 50 \ \text{mM}$ .



Fig. S7. TOC removal during MB degradation in both Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system and Fe<sub>2</sub>O<sub>3</sub>/FCNT-L/H<sub>2</sub>O<sub>2</sub> system. Reaction conditions: pH = 5.0, T = 293.2 K, [Fe<sub>2</sub>O<sub>3</sub>@FCNT-H] = [Fe<sub>2</sub>O<sub>3</sub>/FCNT-L] =  $1.5 \times 10^{-2} \text{ g} \cdot \text{L}^{-1}$ , [MB] =  $10 \mu$ M, [H<sub>2</sub>O<sub>2</sub>] = 50 mM.



**Fig. S8.** The degradation of MB in the different systems. Reaction conditions: pH = 5.0, T = 293.2 K, [catalysts] =  $1.5 \times 10^{-2}$  g·L<sup>-1</sup>, [MB] =  $10 \mu$ M, [H<sub>2</sub>O<sub>2</sub>] = 50 mM. The Fe loading is 2 wt. % for all the catalysts.



Fig. S9. MB removal under different conditions. The effect of (A-B) catalyst dosage, (C-D) H<sub>2</sub>O<sub>2</sub>

concentration, (E-F) reaction temperature, and (G-H) initial MB concentration on MB degradation and corresponding degradation kinetics in Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system. The insets show the relationship between (B) catalyst dosage, (D) H<sub>2</sub>O<sub>2</sub> concentration, (F) reaction temperature and  $k_{app}$  of MB degradation. The inset in (H) shows the relationship between the concentration of removed MB and initial MB concentration. Experimental conditions: pH = 5.0, T = 278.2 - 313.2K,  $[Fe_2O_3@FCNT-H] = 0.5 - 4.0 \times 10^{-2} \text{ g.L}^{-1}$ ,  $[MB] = 5 - 60 \mu\text{M}$ ,  $[H_2O_2] = 0 - 500 \text{ mM}$ . These results show that the increase of catalyst dosage (A) and  $H_2O_2$  concentration (B) could remarkably increase  ${}^{1}O_{2}$  production and subsequently enhance MB degradation. The linear correlations between the catalyst dosage/ H<sub>2</sub>O<sub>2</sub> concentration and  $k_{app}$  values (B and D, r > 0.99) suggest the direct generation of <sup>1</sup>O<sub>2</sub> from H<sub>2</sub>O<sub>2</sub> activation by Fe<sub>2</sub>O<sub>3</sub>@FCNT-H. Results in (E-F) show that the increase of reaction temperature also effectively accelerate the MB degradation and the apparent activation energy could be calculated based on Arrhenius equation to be 50.7 kJ·mol<sup>-1</sup>. As the increase of initial MB concentration, the  $k_{app}$  value decreases, however, the total removed quantity increases. Results in the inset of (H) indicate the amount of 1O2 generated by the Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system under the present condition could degrade ca. 20 µM MB within 60 min.



**Fig. S10.** The MB degradation kinetics under different solution pH in Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system. Reaction conditions: pH = 3.0 - 9.0, T = 293.2 K, [catalysts] =  $1.5 \times 10^{-2}$  g·L<sup>-1</sup>, [MB] = 10  $\mu$ M, [H<sub>2</sub>O<sub>2</sub>] = 50 mM.



**Fig. S11.** The effect of solution pH on (*A*) MB removal by Fe<sub>2</sub>O<sub>3</sub>/FCNT-L with/without H<sub>2</sub>O<sub>2</sub> and (*B*) degradation kinetics. Reaction conditions: pH = 3.0 - 9.0, T = 293.2 K, [Fe<sub>2</sub>O<sub>3</sub>/FCNT-L] = 1.5  $\times 10^{-2}$  g·L<sup>-1</sup>, [MB] = 10 µM, [H<sub>2</sub>O<sub>2</sub>] = 50 mM.



**Fig. S12.** MB removal in xanthine-xanthine oxidase system. Reaction conditions: pH = 7.8, T = 293.2 K, [phosphate buffer] = 10 mM, [MB] = 10  $\mu$ M, [xanthine] = 1 mM, [xanthine oxidase] = 125 mU·mL<sup>-1</sup>.



**Fig. S13.** The inhibition efficiency of the catalytic activity of Fe<sub>2</sub>O<sub>3</sub>@FCNT-H using different quenching molecules. Reaction conditions: pH = 5.0, T = 293.2 K, [Fe<sub>2</sub>O<sub>3</sub>@FCNT-H] =  $1.5 \times 10^{-2}$  g·L<sup>-1</sup>, [MB] = 10 µM, [H<sub>2</sub>O<sub>2</sub>] = 50 mM, [TBA] = [FFA] = [DMSO] = 50 mM, [SOD] = 328 U·mL<sup>-1</sup>.



**Fig. S14.** The effect of pH on HO<sub>2</sub>•/O<sub>2</sub>• and <sup>1</sup>O<sub>2</sub> generation. The effect of pH on (*A*) electron spins of HO<sub>2</sub>•/O<sub>2</sub>• trapped by DMPO and (*B*) electron spins of <sup>1</sup>O<sub>2</sub> trapped by TEMP in Fe<sub>2</sub>O<sub>3</sub>@FCNT-H/H<sub>2</sub>O<sub>2</sub> system. (*C*) The relationship between the calculated electron spins of O<sub>2</sub>• and electron spins of <sup>1</sup>O<sub>2</sub>. (*D*) The relationship between electron spins of <sup>1</sup>O<sub>2</sub> and  $k_{app}$  value. Reaction conditions: pH = 2.0 - 9.0, T = 293.2 K, [Fe<sub>2</sub>O<sub>3</sub>@FCNT-H] = 1.5 × 10<sup>-2</sup> g·L<sup>-1</sup>, [H<sub>2</sub>O<sub>2</sub>] = 50 mM, [DMPO] = 50 mM for (*A*) and [TEMP] = 50 mM for (*B*).



**Fig. S15.** (*A*) The adsorption kinetics of MB by  $Fe_2O_3@FCNT$ -H and (*B*) the relationship between  $q_{e,cal}$  and  $k_{app}$  under different solution pH. Reaction conditions: pH = 2.0 - 9.0, T = 293.2 K,  $[Fe_2O_3@FCNT$ -H] =  $1.5 \times 10^{-2} \text{ g}\cdot\text{L}^{-1}$ ,  $[MB] = 10 \ \mu\text{M}$ .



**Fig. S16.** The degradation (*A*) and the adsorption kinetics (*B*) of target molecules by  $Fe_2O_3@FCNT-H$  with and without  $H_2O_2$ . Reaction conditions: pH = 5.0, T = 293.2 K,  $[Fe_2O_3@FCNT-H] = 1.5 \times 10^{-2} \text{ g}\cdot\text{L}^{-1}$ , [target molecules] = 10  $\mu$ M,  $[H_2O_2] = 0/50$  mM.



Fig. S17. The adsorption kinetics of target molecules onto Fe<sub>2</sub>O<sub>3</sub>@FCNT-H. Reaction conditions: pH = 5.0, T = 293.2 K, [Fe<sub>2</sub>O<sub>3</sub>@FCNT-H] =  $1.5 \times 10^{-2}$  g·L<sup>-1</sup>, [Al<sup>3+</sup>] = 0.1 - 1 mM, [target molecules] =  $10 \mu$ M.



**Fig. S18.** The change of UV-vis absorbance of MB in the presence of MO, aniline, and/or C2R as a function of reaction time. Reaction conditions: pH = 5.0, T = 293.2 K,  $[Fe_2O_3@FCNT-H] = 1.5 \times 10^{-2} \text{ g}\cdot\text{L}^{-1}$ ,  $[target molecules] = 10 \,\mu\text{M}$ ,  $[H_2O_2] = 50 \,\text{mM}$ .



Fig. S19. Effect of light irradiation and O<sub>2</sub> on MB removal. Reaction conditions: pH = 5.0, T = 293.2 K,  $[Fe_2O_3@FCNT-H] = 1.5 \times 10^{-2} \text{ g}\cdot\text{L}^{-1}$ ,  $[MB] = 10 \ \mu\text{M}$ ,  $[H_2O_2] = 50 \ \text{mM}$ .



Fig. S20. Fe K-edge XANES of Fe<sub>2</sub>O<sub>3</sub>, Fe foil, Fe<sub>2</sub>O<sub>3</sub>@FCNT-H, and Fe<sub>2</sub>O<sub>3</sub>@FCNT-H in the presence of  $H_2O_2$ .

# Part S4 Supplementary Tables

Classification	Compound*	Solvent	рН	$k^{1}O_{2}$ (M <sup>-1</sup> ·s <sup>-1</sup> ) <sup>§</sup>	Ref.
	MB	H <sub>2</sub> O	5.9	3.0×10 <sup>8</sup>	(S13)
D	eosin	ACN	-	5.0×10 <sup>7</sup>	(S14)
Dyes	rose bengal	ACN	-	$7.2 \times 10^{7}$	(S14)
	leucomalachite green	toluene	-	$2.0 \times 10^{8}$	(S15)
	phenol <sup>†</sup>	$H_2O$	$\leq 8.0$	$2.6\pm4.0\times10^{6}$	(S1)
	4-chlorophenol <sup>†</sup>	$H_2O$	≤7.4	$6.0\pm3.6\times10^{6}$	(S1)
	2-nitrophenol <sup>†</sup>	$H_2O$	≤5.2	$1.3\pm1.1\times10^{6}$	(S1)
	4-methylphenol <sup>†</sup>	$H_2O$	≤8.3	$9.6\pm2.8\times10^{6}$	(S1)
	1,3-benzenediol	EtOH	-	$7.9 \times 10^{5}$	(S16)
<b>A</b>	1,4-benzenediol	pyridine	-	$2.5-4.0 \times 10^{7}$	(S16)
Aromatic	2,6-di-t-butylphenol	MeOH	-	$1.0 \times 10^{6}$	(S17)
compounds	2,4,6-triphenylphenol	MeOH	-	$2.5 \times 10^{8}$	(S17)
	1,2,4-trimethoxybenzene	MeOH	-	$1.8 \times 10^{7}$	(S17)
	2,4,6-trichlorophenol <sup>†</sup>	$H_2O$	≤7.4	$1.7\pm0.7\times10^{7}$	(S1)
	aniline	MeOH	-	$2.0 \times 10^{9}$	(S18)
	naphthalene	1-BuOH <sup>‡</sup>	-	$5.2 \times 10^8$	(S15)
	9,10-diphenylanthracene	benzene	-	$1.2 \times 10^{6}$	(S19)
	histidine	$H_2O$	8.5	$6.6 \times 10^7$	(S20)
A mine eside	methionine	$H_2O$	8.5	$2.2\pm0.7\times10^{7}$	(S20)
Amme actus	tyrosine	$H_2O$	8.5	$0.9 \times 10^{7}$	(S20)
	tryptophan	$H_2O$	8.5	$1.8 \times 10^{8}$	(S20)
	furfuryl alcohol	$H_2O$	5.0-12.0	$1.2 \times 10^{8}$	(S21)
Unternovalia	2,5-dimethylfuran	$H_2O$	7.0	$8.2 \times 10^{8}$	(S15)
aomnounda	cyanohemoglobin	$H_2O$	7.0	$1.2 \times 10^{9}$	(S15)
compounds	porphyrin	CHCl <sub>3</sub>	-	$4.2 \times 10^{8}$	(S15)
	cytochrome C	$H_2O$	7.4	$1.4 \times 10^{8}$	(S15)
	β-carotene	MeOH	-	$9.3 \times 10^{8}$	(S15)
	crocetin	$H_2O$	7.8	$5.7 \times 10^9$	(S15)
Olefins	oleic acid	ACN	-	$1.6 \times 10^{5}$	(S15)
	1,4-dioxene	Ace <sup>‡</sup>	-	2.2-3.6×10 <sup>5</sup>	(S15)
	polyisoprene	CHCl <sub>3</sub>	-	$1.2 \times 10^{5}$	(S15)
	azide ion	$H_2O$	7.0	$4.5-6.4 \times 10^8$	(S15)
Inorganic	iodide ion	$H_2O$	7.0	$8.7 \times 10^{5}$	(S15)
compounds	nitrate ion	$H_2O$	8.3	$3.1 \times 10^{6}$	(S15)
	O2*-	DMSO	-	$1.6 \times 10^9$	(S15)

Table S1. <sup>1</sup>O<sub>2</sub> - mediated oxidation of various organic and inorganic compounds

\* Various compounds have been reported to be oxidized by  ${}^{1}O_{2}$  generated from different systems. For examples,  $MoO_{4}{}^{2}$  - exchanged layered double hydroxides could effectively catalyze  $H_{2}O_{2}$  decomposition to produce  ${}^{1}O_{2}$  to oxidize chemicals including 2,3-dimethyl-2-butene, olefins, and allylic alcohols, through either the ene hydroperoxidation or the [2+4] cycloaddition (S23). Pharmaceutical compounds such as cimetidine, propranolol, ranitidine, sulfamethoxazole could be effectively degraded by  ${}^{1}O_{2}$  generated from either alkaline periodate (S24) or peroxymonosulfate activation by benzoquinone (S25). Dissolved organic matter could also be partially degraded by  ${}^{1}O_{2}$ , resulting in slower bacterial growth (S26). Apoptosis of the cancer cells was reported in the *in vitro* photodynamic therapy due to  ${}^{1}O_{2}$  generation from ultrathin black phosphorus nanosheets under light irradiation (S27).

<sup>†</sup> Phenolate anions dominate at higher pH values above  $pK_a$ . The rate constants for the reactions between phenolate anions and  ${}^1O_2$  are always one order of magnitude higher than those involving the undissociated phenols (S1).

<sup>‡</sup> 1-BuOH is the abbreviation of 1-butanol, and Ace is the abbreviation of acetone.

<sup>§</sup>The values of rate constant are the sum of that of the chemical oxidation  $(A + {}^{1}O_{2} \rightarrow Products)$ and the physical quenching of  ${}^{1}O_{2}$  by the target molecules  $(A + {}^{1}O_{2} \rightarrow A + {}^{3}O_{2})$ .

NO.	Reactions	$k (\mathbf{M}^{-1} \cdot \mathbf{s}^{-1})$	Ref.	
<b>S</b> 1	$Fe(III) + H_2O_2 \rightarrow Fe(II) + HO_2 \bullet + H^+$	1.0-10×10 <sup>-3</sup>	(S28)	
S2	$Fe(III) + HO_2 \bullet \rightarrow Fe(II) + H^+ + O_2$	$<2.0\times10^{3}$	(S28)	
<b>S</b> 3	$Fe(II) + HO_2 \bullet \rightarrow Fe(III) + HO_2^-$	$1.2 \times 10^{6}$	(S29)	
<b>S</b> 4	$Fe(III) + O_2 \xrightarrow{\cdot} \rightarrow Fe(II) + O_2$	$1.5 \times 10^{8}$	(S29)	
<b>S</b> 5	$Fe(II) + O_2^{-} + 2H^+ \rightarrow Fe(III) + H_2O_2$	$1.0 \times 10^{7}$	(S28)	
<b>S</b> 6		$k_{+} 1.0 \times 10^{10}$	(S28)	
	$H^{+} + O_2^{-} \leftrightarrow HO_2^{-}$	$k_{-} 1.58 \times 10^{5}$		
<b>S</b> 7	$HO_2 \bullet + H_2O_2 \longrightarrow HO \bullet + H_2O + O_2$	3	(S30)	
<b>S</b> 8	$HO_2 \bullet + HO_2 \bullet \longrightarrow H_2O_2 + O_2$	$8.3 \times 10^{5}$	(S29)	
<b>S</b> 9	$O_2^{\bullet} + H_2O_2 \rightarrow HO^{\bullet} + OH^- + O_2$	0.13	(S31)	
S10	$O_2^{\bullet} + HO_2^{\bullet} \rightarrow HO_2^{-} + O_2$	$9.7 \times 10^{7}$	(S29)	
<b>S</b> 11	$\mathrm{HO}\bullet + \mathrm{H}_2\mathrm{O}_2 \to \mathrm{HO}_2\bullet + \mathrm{H}_2\mathrm{O}$	3.3×10 <sup>7</sup>	(S28)	
S12	$\mathrm{HO}\bullet + \mathrm{HO}_{2}\bullet \longrightarrow \mathrm{H}_{2}\mathrm{O} + \mathrm{O}_{2}$	7.1×10 <sup>9</sup>	(S28)	
S13	$HO \bullet + O_2 \bullet \to OH^- + O_2$	$1.01 \times 10^{10}$	(S28)	
S14	$\mathrm{HO}\bullet + \mathrm{HO}\bullet \to \mathrm{H}_2\mathrm{O}_2$	$5.2 \times 10^9$	(S28)	

Table S2. Reactions involving HO<sub>2</sub>•/O<sub>2</sub>• in the Haber-Weiss cycle

Compound	CAS No.	Structure*	Formula	Molar mass (g·mol <sup>-1</sup> )	Collision dimension <sup>†</sup> (Å ×Å ×Å)	Maximum absorption wavelength (nm)
methylene blue	7220-79-3	jijiji	$C_{16}H_{18}ClN_3S$	319.9	14.3×7.6×3.8	664
methyl violet	8004-87-3		C <sub>24</sub> H <sub>28</sub> ClN <sub>3</sub>	393.96	14.5×16.4×6.1	579
crystal violet	548-62-9		C <sub>25</sub> H <sub>30</sub> ClN <sub>3</sub>	407.99	15.0×16.6×6.2	583
malachite green oxalate	569-64-2	the states	C <sub>23</sub> H <sub>25</sub> ClN <sub>2</sub>	364.92	14.1×14.1×6.0	616
chrysoidine	532-82-1	××××	C <sub>12</sub> H <sub>12</sub> N <sub>4</sub> HCl	248.71	14.7×8.1×3.4	453
chromotrope 2R	4197-07-3	A.A.A.A.A.A.A.A.A.A.A.A.A.A.A.A.A.A.A.	$C_{16}H_{10}N_2Na_2O_8S_2$	468.37	17.9×10.0×5.8	507
methyl orange	547-58-0	Fit-tty	$C_{14}H_{14}N_3NaO_3S$	327.33	19.4×5.4×7.4	464
aniline	62-53-3		C <sub>6</sub> H <sub>7</sub> N	93.13	8.4×7.0×3.4	263

\*The geometries of the selected contaminants were optimized in the gas phase using the Gaussian 16 software package (Gaussian, Inc.) using hybrid density functional theory (DFT), B3LYP and the 6-311G basis set. <sup>†</sup> The collision dimension estimation was carried out by Chemcraft based on the optimized geometries.

Compound	Mobile phase	Flow rate	Wavelength	Column
	(MeOH/H <sub>2</sub> O, v/v)	(mL·min <sup>-1</sup> )	( <b>nm</b> )	
FFA	5/95	1.0	219	C18, 4.6 ×100 mm,
				3.5 µM particle size
aniline	60/40	0.2	263	C18, 2.1 × 50 mm,
				1.9 µM particle size
4-CP	50/50	0.2	280	C18, 2.1 × 50 mm,
				1.9 µM particle size

Table S4. The detection methods of organic molecules by UHPLC

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