

## **Supplementary information**

### **Long noncoding RNAs in vascular smooth muscle cells regulate vascular calcification**

Geon Jeong<sup>1,2,3</sup>, Duk-Hwa Kwon<sup>1,4</sup>, Sera Shin<sup>1,4</sup>, Nakwon Choe<sup>1,4</sup>, Juhee Ryu<sup>1,2,3,4</sup>, Yeong-Hwan Lim<sup>1,2,3</sup>, Jaetaek Kim<sup>1,5</sup>, Woo Jin Park<sup>1,6</sup>, Hyun Kook<sup>1,3,4,\*</sup>, Young-Kook Kim<sup>1,2,3,\*</sup>

**Supplementary Table S1.** A list of lncRNAs identified in this study, along with their expression change after induction of vascular calcification in VSMCs. Those lncRNAs with an average FPKM value greater than 10 and not 0 in any sample are shown. The lncRNAs presented in Figure 2A are highlighted.

**Supplementary Table S2.** Full-length sequences and the exon-intron boundary of lncRNAs. The exon-exon junction is indicated with asterisk (\*).

**(A) Linc00116**

TGGACTGGAGGAAGCGGAGGCTGCAGGACAAGCTGGCGACGACTCAGAAAAAGCTGGA  
CCTGGCCTGAGCACGTTGGTTCTCTCCAAGCCTCGTCAGCCGTATCCGGGGAGCTGC  
GCGACTACACGCCCTGAGTCCCAGCTCGTCTTGACGGCTTGCAGGAACGTGGCTGCT  
TCCAGACCTCAGAAAGAAAATAGTTTGTCTCGCTAACAGCCTGTGCTCAG\*CTTGTG  
AAGATGGATATAAAGCACGGAAAGAAAACATTCTTGACTTATGAATCTATTAA  
AATAAAAAATTAAACATC

**(B) Lrrc75a-as1**

CTCTCGGCCATCATGGGATCATGTTCCCTCATCTTCCTGTAGCTGGTACTTCGCCATGGCC  
TGCACTCGGCGGCCCGCAGCCATGCTGAGCCCCGGGCCGGTCTCCAGCACCAGCCCTGGG  
CGCTGCGCAGATCTAGGGGAGCCTG\*ACAGGACGTTGACATCGTGGAAATAGGAGCACTAT  
CCCACCATGAG\*GTTGGGAAGTTAGGGTGGAAAGATTGCCAGCAGCCATGGCCTCCATGA  
G\*GGTGTCCCTTCAAGTTGAGAACGCACTGGGGACTGCAGATGGTGTGACAGCAGC  
TGAG\*GCTACATTCCCAGGAGTCTCCAGCCATCACCCAAAAGCAAGGTGTGGTCAGCC  
AACTTTGTTCTGGACAGGCACGGTGGTGGCCTATCTCCCACAGGATCTCTGTACCAT  
TTAATAAAAGTTTGTAAAGACTGAT

**(C) Snhg1**

TCATTTCTTGTCTGG\*GTTAAGGTACTCTTACATGCGAAAGAATGGGAGCACGC  
TCTCATTCAAA\*ATATGGCTAATGTCAAAGCCGTTGCACCTGGAGCCAGGCCTGTTCA  
ATTTTACAAATAGCAAGTTTCTTACAATCAG\*GGTACGGTGGCAACGTGACTTGAAAC  
CCCCAGAACATACCCGGGATGAATGCAG\*GTTTCTGGTGGTGGATTATCTTAAAG\*GA  
GGAGGTTGTTCTGCACGCCATCCATCTCCAGCGTTGAAG\*GTTCTGTTAGCAGTGCTGT  
TACTTGGTGTCTCAGAG\*ATATGGTACTGGCCGAGCGCAAAGGAGCCATGCCAG\*GAAA  
ACTAAGAGCACATCTTCATAAAAGTCCAGGCTAAAGTAAG\*GTCTGAGGAGAGGGATGAG  
GTGTGTGTTAGAG\*GAACTGAAATCAAAGATCCGATTACCTAGGCCAGTGGCACGGCAA

ACTTCAGAACCCATGGAATACAGCCTGTTGATTCTTGTGGATTTAGCATCTGAGCAG  
TATCAGTGGCACTCCAAGTGAAATGGTGATCTCTTCAGTACCGCTGGAGCAGATTAATA  
TTTATTCACTGCCAGAGGCTATATTCTGCCCCCCCATAACCCCTCCTCTCCTTA  
GACTACTTAAGATTGAAATATTGCAACTCCAAATTATCTCCTGCTTACCACTTTTT  
TTCGGAGCTGGGACCAAACCCAGGGCCTGCGCTCCTAGGTAAGCGCTTACCACTGA  
GCTAAATCCCCAGCCCCGCTTACACATTGTTAATCTGATCCTCCGCAGTTATTGTGCT  
GTTCTTTGTTCTGGCAATAATGCTTTATGCTA

**(D) Snhg16**

CTTCCGAGGTCTCTGCCCGTGCACTAAAGTTATGGCCGGTCTGTCCGTGGTTCACTC  
TCTGAAAGGCCTAGGGTTGGGTGATG\*GCACGCCATTGCACATGGGACGCAAGCCGCA  
GGTCCATTTCAGAAGTAATGGCTGGATCCCTGGAAGCAGATGGACTCGGTGTGGTTAGA  
GGCAGAGGTAGATGATGCTGCGTATTGAGCCCTCCTCAGCCCACTTG\*ATTGAGGTGT  
TGCGCGGTGAGGAAGGGCCTCGTGACCAGTCTGCATG\*GGGGAGGCTGAACCATAAGTT  
CTGGGCCTCAAGGGTGGCTGCTGCAGCCACGGGGCAGCCCTCCTGCGCCTGGACTTCT  
GCCTGCGAGCAGAGGAGCTCCACACACAGCTCGTCAGAGCATCCGTGCTCTGCGCTAA  
GTGGCAG\*ATTCCAGGACGGCTCCAGGGGCATCATCGAAAGGCGTGGCACATCAGGGC  
CTTCAGGATGGCTGCGTCCTG\*GTTATTGCCCTGTGGTGCAGAGAGAACACACAGCCT  
GCTGCTGCTCCTGCCTTAATTCCAAAGAGCCATCAAAAGACCTGGACTTGTCTGC  
AAGAGGAGCTAGCAAATGGAGTGACTGTGGTCACCCCTGGAAGCCTTGACAGCTGGG  
AGGCCCTGCTGAGCTTAGAAAACCATCTGATAGATTCCCTGAGTAAGGATTGGCTTGT  
ATTGAAAGCACAAATAACTTGATAAAATTAAA

**Supplementary Table S3.** The sequences of PCR primers to measure lncRNA and mRNA expression.

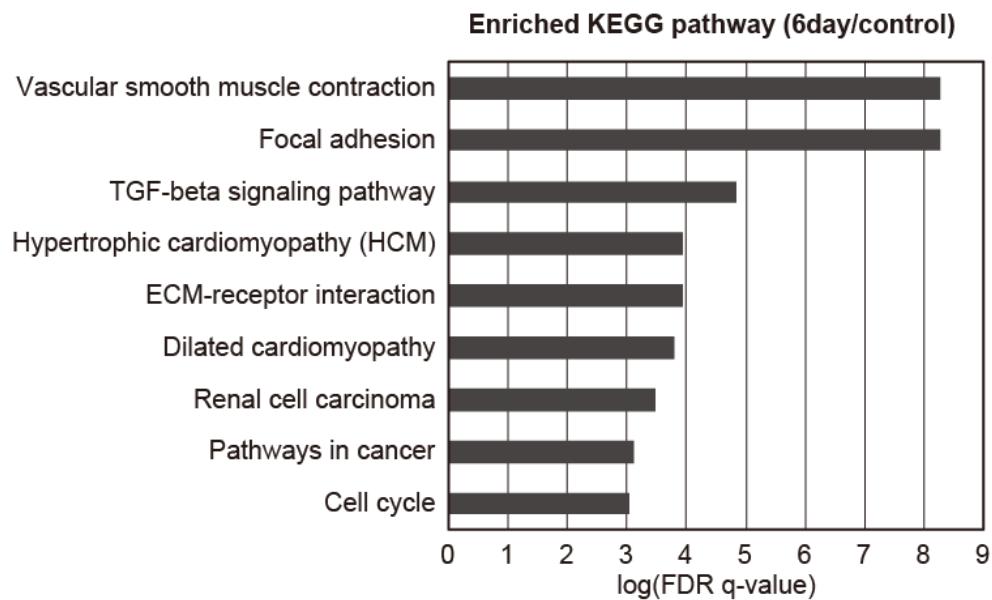
Linc00116 forward primer	5'-TTGCTTCCAGACACCTCAGAAAG-3'
Linc00116 reverse primer	5'-TCCC GTGCTTATATCCATCTC-3'
Lrrc75a-as1 forward primer	5'-TTTCGCCATGGCCTGCACTC-3'
Lrrc75a-as1 reverse primer	5'-GCCATGGCTGCTGGCAATCT-3'
Snhg1 forward primer	5'-CAAAGATCCGATTACCTAGGCC-3'
Snhg1 reverse primer	5'-CACAAAGGAATCAACAGGCTG-3'
Snhg16 forward primer	5'-TCGAAAGGCGTGGCACATCA-3'
Snhg16 reverse primer	5'-CTCTGGCACACAGGGCAAT-3'
Malat1 forward primer	5'-AGTTCTGTTGGCACACTGGGT-3'
Malat1 reverse primer	5'-CCAAAGCTGCACTGTGCTGTA-3'
Gas5 forward primer	5'-ACATGTGGTCCCTTAAGTG-3'
Gas5 reverse primer	5'-GGCAAATCTCTGTTCAAGCATC-3'
Runx2 forward primer	5'-CGATGGGACCGTGGTTACCG-3'
Runx2 reverse primer	5'-GTCCGCTCCGGCCTACAAAT-3'
Msx2 forward primer	5'-GGCCTCTCGTCAAGCCCTTC-3'
Msx2 reverse primer	5'-AGTATCTGCCGGGCTCCTGT-3'
Bmp2 forward primer	5'-CGACGTCCCTCAGCGAGTTG-3'
Bmp2 reverse primer	5'-GGGTGGGTCTCTGCTTCAGG-3'
Pre-Gapdh forward primer	5'-CCATGGTGCAGCGATGCTTT-3'
Pre-Gapdh reverse primer	5'-ACGGCCAATCTGAGGCAAG-3'
Gapdh forward primer	5'-ATGACTCTACCCACGGCAAG-3'
Gapdh reverse primer	5'-CTGGAAGATGGTGATGGTT-3'
Actb forward primer	5'-CACTTCTACAATGAGCTGCG-3'
Actb reverse primer	5'-CTGGATGGCTACGTACATGG-3'

**Supplementary Table S4.** The sequences of gene-specific primers (GSPs) used in RACE experiments.

Linc00116 5' GSP	5'-GAGCACAGGCTGTTAGCGAAGAC-3'
Linc00116 Nested 5' GSP	5'-AAGAGCAAGCCACGTTCCCTGCAAG-3'
Lrrc75a-as1 5' GSP	5'-CTATTCCACGATGTCAACGTCCTG -3'
Lrrc75a-as1 Nested 5' GSP	5'-AGGCTCCCCTAGATCTGCGCAG-3'
Snhg1 5' GSP	5'-TTCAAGTCACGTTGCCACCGTACC-3'
Snhg1 Nested 5' GSP	5'-GTGCAAACGGCTTGACATTAGCC-3'
Snhg16 5' GSP	5'-CCAGGGATCCAGCCATTACTTCTG-3'
Snhg16 Nested 5' GSP	5'-GCTTGCCTCCCATGTGCAATGG-3'
Linc00116 3' GSP	5'-GGCTTGCTTCCAGACCTCAGAAAG-3'
Linc00116 Nested 3' GSP	5'-TCTTCGCTAACAGCCTGTGCTCAG-3'
Lrrc75a-as1 3' GSP	5'-ATTCCCAGGAGTCTCCAGGCCATC-3'
Lrrc75a-as1 Nested 3' GSP	5'-AAAGCAAGGTGTGGTCAGCCAAC-3'
Snhg1 3' GSP	5'-TCTGAGGAGAGGGATGAGGTGTG-3'
Snhg1 Nested 3' GSP	5'-CAAAGATCCGATTACCTAGGCCAG-3'
Snhg16 3' GSP	5'-TGGAGTGACTGTGGTCACCCTGG-3'
Snhg16 Nested 3' GSP	5'-ACCCTGGAAGCCTTGACAGCTG-3'

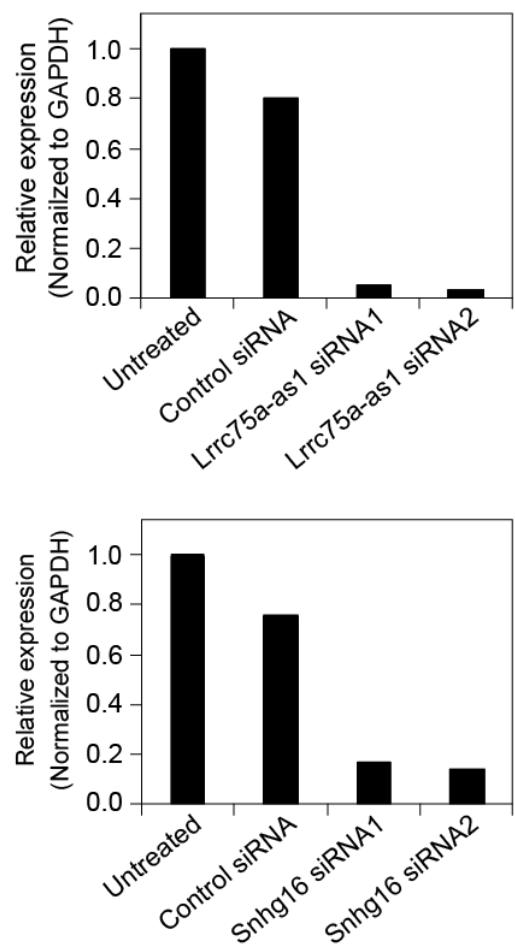
**Supplementary Table S5.** The sequences of PCR primers for cloning of lncRNAs.

Linc00116 forward primer	5'-TGGACTGGAGGAAGCGGGAGGCTGC-3'
Linc00116 reverse primer	5'-GATGTTAAAATTTTATTAA-3'
Lrrc75a-as1 forward primer	5'-CTCTGCGGCCATCATGGGATCAT-3'
Lrrc75a-as1 reverse primer	5'-ATCAGTCTTATAAACAAAACTTT-3'
Snhg1 forward primer	5'-TCATTTCTTGTCTGGTTT-3'
Snhg1 reverse primer	5'-TAGCATAAAAGCATTATTGCC-3'
Snhg16 forward primer	5'-CTTCCGAGGTCTCTGCCGC-3'
Snhg16 reverse primer	5'-TTAATTTATCAAGTTATTG-3'



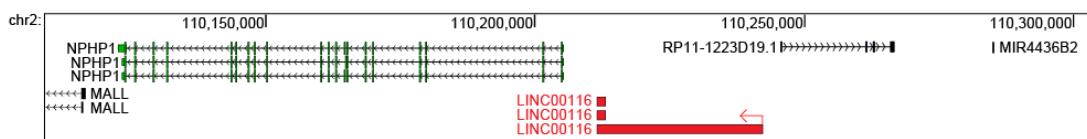
**Supplementary Figure S1.** Pathway analysis of RNA-seq data. The decreased mRNAs at the day 6 sample compared to the control sample with fold change (6 day/control) below 0.5 were selected. Pathway analysis was performed using the Kyoto Encyclopedia of Genes and Genomes (KEGG) gene set analysis (reference) at MSigDB database<sup>22,23</sup>.

Measurement of lncRNA after siRNA treatment

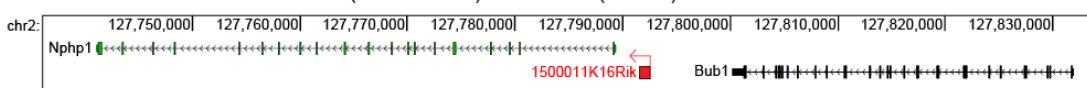


**Supplementary Figure S2.** Measurement of lncRNA level after the treatment with siRNA for each gene.

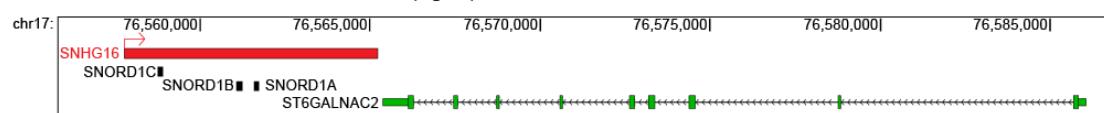
**A Gene locus of LINC00116 on human (hg38)**



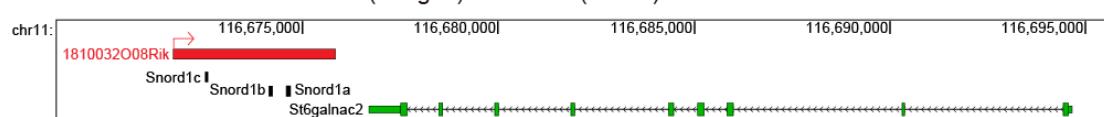
**B Gene locus of 1500011K16Rik(LINC00116) on mouse (mm10)**



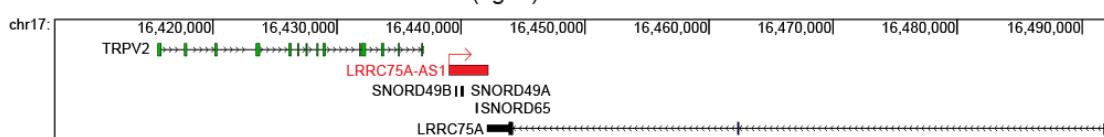
**C Gene locus of SNHG16 on human (hg38)**



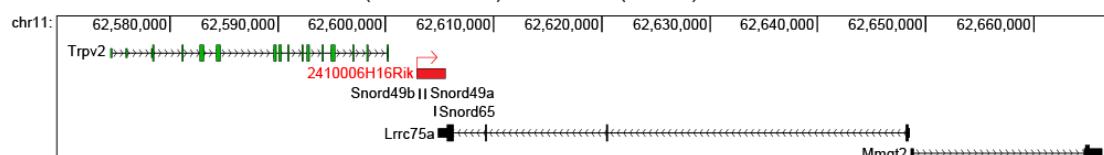
**D Gene locus of 1810032O08Rik(Snhg16) on mouse (mm10)**



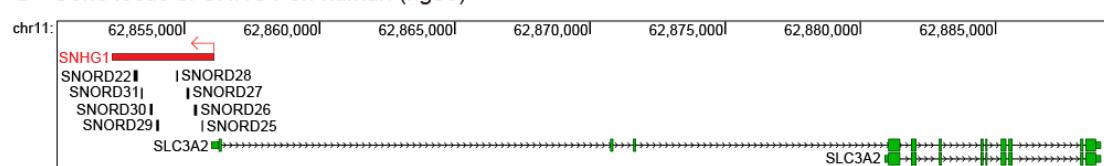
**E Gene locus of LRRC75A-AS1 on human (hg38)**



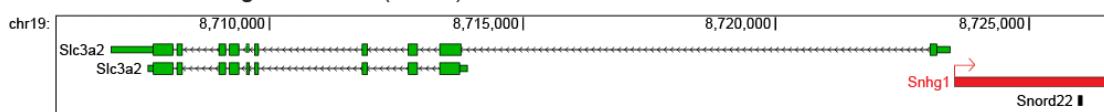
**F Gene locus of 2410006H16Rik(Lrrc75a-as1) on mouse (mm10)**



**G Gene locus of SNHG1 on human (hg38)**

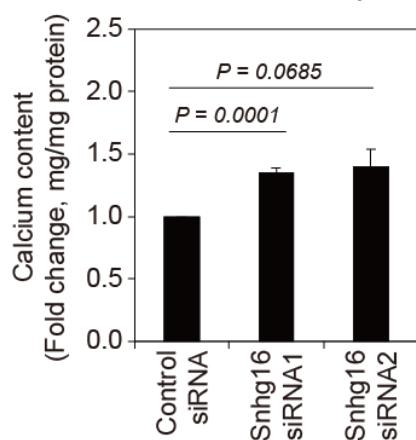


**H Gene locus of Snhg1 on mouse (mm10)**

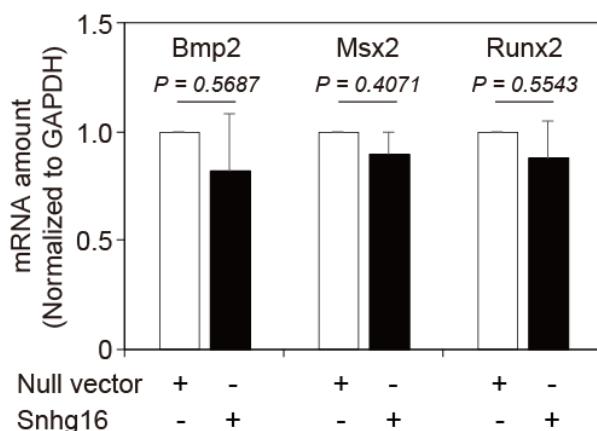


**Supplementary Figure S3.** The human and mouse genomic regions near lncRNAs analyzed in this study.

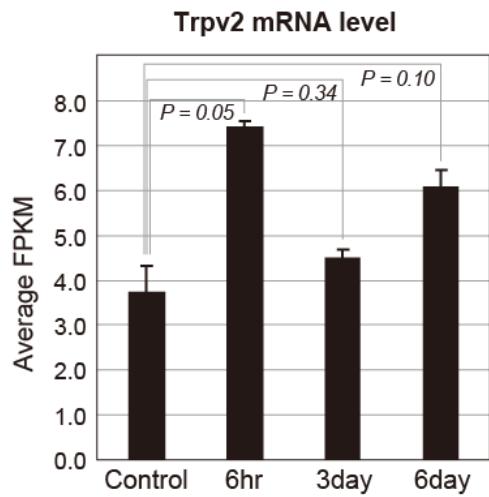
**A** Measurement of calcium deposition after Snhg16 knockdown



**B** Change of osteoblast related factors after Snhg16 overexpression



**Supplementary Figure S4.** (a) Measurement of calcium deposition after knockdown of Snhg16. Six independent experiments were performed, and the P value was calculated by a two-sided paired *t*-test.  
(b) Expression change of the osteoblast-related factors after overexpression of Snhg16. Error bars indicate standard error between three independent experiments.



**Supplementary Figure S5.** The expression level of Trpv2, the neighboring gene of Lrrc75a-as1. FPKM values from RNA-seq data were averaged. Error bars indicate the deviation from two samples.