

Supporting Information

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Structure-Activity Relationship of NF023 Derivatives Binding to XIAP-BIR1

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Author Contributions

L.S. Conceptualization:Equal; Data curation:Lead; Methodology:Equal

Experimental Section

Infrared spectra (IR) were measured on a Fourier transform infrared spectrometer (FT-IR). Absorption intensities are recorded by the following abbreviations: s, strong; m, medium; and w, weak. Proton NMR spectra were obtained on a Varian Mercury-400 (400 MHz) or a Bruker AV-400 (400 MHz) spectrometer by use of dimethylsulfoxide-*d*₆ as the solvent. Carbon-13 NMR spectra were obtained on a Varian Mercury-400 (100 MHz) or a Bruker AV-400 (100 MHz) spectrometer. The residual solvent peaks, δ_H 2.50 ppm and δ_C 39.5 ppm for DMSO-*d*₆, were used as references. Multiplicities are recorded by the following abbreviations: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; J, coupling constant (hertz). High-resolution mass spectra were obtained by means of a VARIAN-901 mass spectrometer.

Tetrasodium 4,4'-Carbonylbis(imino)bis-1,5-naphthalenedisulphonate (3).

mp (recrystallized from H₂O/EtOH) 193.6–194.2 °C; ¹H NMR (DMSO-*d*₆, 400 MHz) δ 8.80 (d, *J* = 8.0 Hz, 2 H, H-2), 8.05 (d, *J* = 8.0 Hz, H-3), 7.65 (d, *J* = 8.4 Hz, 2 H, ArH), 7.22 (t, *J* = 8.0 Hz, 2 H, H-7), 7.07 (s, 2 H, NH), 6.45 (d, *J* = 8.4 Hz, 2 H, ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 154.79 (C=O), 147.18, 143.20, 132.48, 130.52, 129.89, 126.44, 124.69, 122.92, 118.40, 107.36; IR (KBr) 3087 (w), 2989 (w), 1580 (s, C=O), 1423 (s), 1189 (m), 1042 (s), 835 (s) cm⁻¹; UV (Water): λ_{max} 253 (ϵ 11,804), 216 (ϵ 11,720), 342 (ϵ 5,171); HRMS (ESI) *m/z* [M + H]⁺ calcd for C₂₁H₁₂N₂Na₄O₁₃S₄ + H 720.8891, found 720.8896.

Tetrasodium 4,4'-(Carbonylbis(imino-3,1-phenylenecarbonylimino))bis-1,5-naphthalenedisulphonate (5a).

mp (recrystallized from H₂O/EtOH) 220.2–220.6 °C; ¹H NMR (DMSO-*d*₆, 400 MHz) δ 12.54 (s, 2 H, NH), 10.55 (s, 2 H, NH), 9.10 (d, *J* = 8.8 Hz, 2 H, ArH), 8.29 (d, *J* = 8.8 Hz, 2 H, ArH), 8.07–8.01 (m, 6 H, ArH), 7.83–7.79 (m, 4 H, ArH), 7.45 (t, *J* = 8.8 Hz, 2 H, ArH), 7.35 (t, *J* = 8.0 Hz, 2 H, ArH); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 165.70 (C=O), 153.17 (C=O), 141.71, 140.98, 140.28, 136.21, 134.53, 131.57, 130.70, 128.21, 127.04, 124.57, 123.48, 123.45, 122.39, 120.88, 120.68, 118.11; IR (KBr) 3355 (br, NH), 2919 (m), 1652 (m, C=O), 1575 (s), 1545 (s), 1524 (s), 1193 (s), 1044 (s), cm⁻¹; UV (Water): λ_{max} 234 (ϵ 11,641), 311 (ϵ 6,872); HRMS (ESI) *m/z* [M + Na]⁺ calcd for C₃₅H₂₂N₄Na₄O₁₅S₄ + Na 980.9447, found 980.9446.

Tetrasodium 4,4'-(Carbonylbis(imino-3,1-(4-methylphenylene)carbonylimino)bis-1,5-naphthalenedisulphonate (5b).

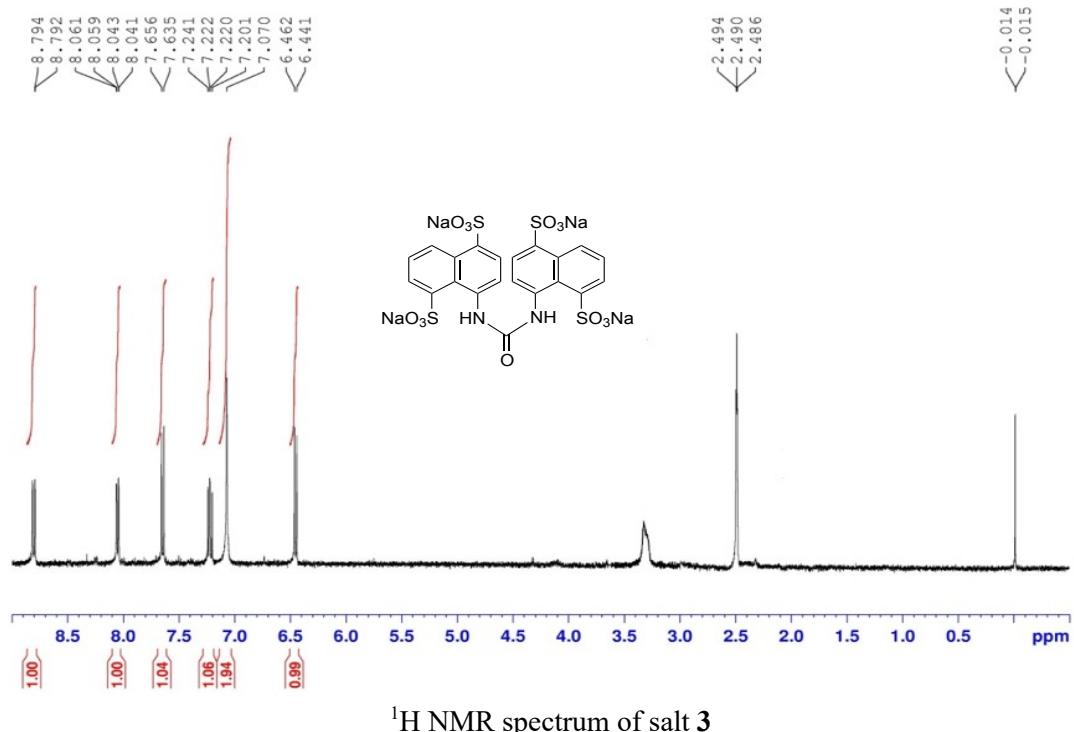
mp (recrystallized from H₂O/EtOH) 224.4–224.8 °C; ¹H NMR (DMSO-*d*₆, 400 MHz) δ 12.50 (s, 2 H, NH), 9.09 (s, 2 H, NH), 9.06 (d, *J* = 7.6 Hz, 2 H, ArH), 8.35 (s, 2 H, H-2'), 8.28 (d, *J* = 8.0 Hz, 2 H, ArH), 8.02–8.00 (m, 4 H, ArH), 7.84 (d, *J* = 7.6 Hz, 2 H, ArH), 7.45 (t, *J* = 8.0 Hz, 2 H, H-7), 7.26 (d, *J* = 8.0 Hz, 2 H, ArH), 2.39 (s, 6 H, 2 × CH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 165.61 (C=O), 153.32 (C=O), 141.77, 140.82, 137.51, 134.66, 133.74, 132.18, 131.58, 130.69, 129.68, 127.01, 126.90, 124.61, 123.47, 123.42, 122.37, 122.29, 18.48 (CH₃); IR (KBr) 3354 (br, NH), 3048 (w), 2914 (s), 2848 (m), 1635 (s, C=O), 1575 (m), 1424 (m), 1225 (w) cm⁻¹; UV (Water): λ_{max} 236 (ε 15,050), 310 (ε 10,288); HRMS (ESI) *m/z* [M + Na]⁺ calcd for C₃₇H₂₆N₄Na₄O₁₅S₄ + Na 1008.9760, found 1008.9765.

Disodium 4-(3-[3-(Ethoxylthioxoimino)benzamido]-4-tolylamido)naphthalene-1,5-disulphonate (9).

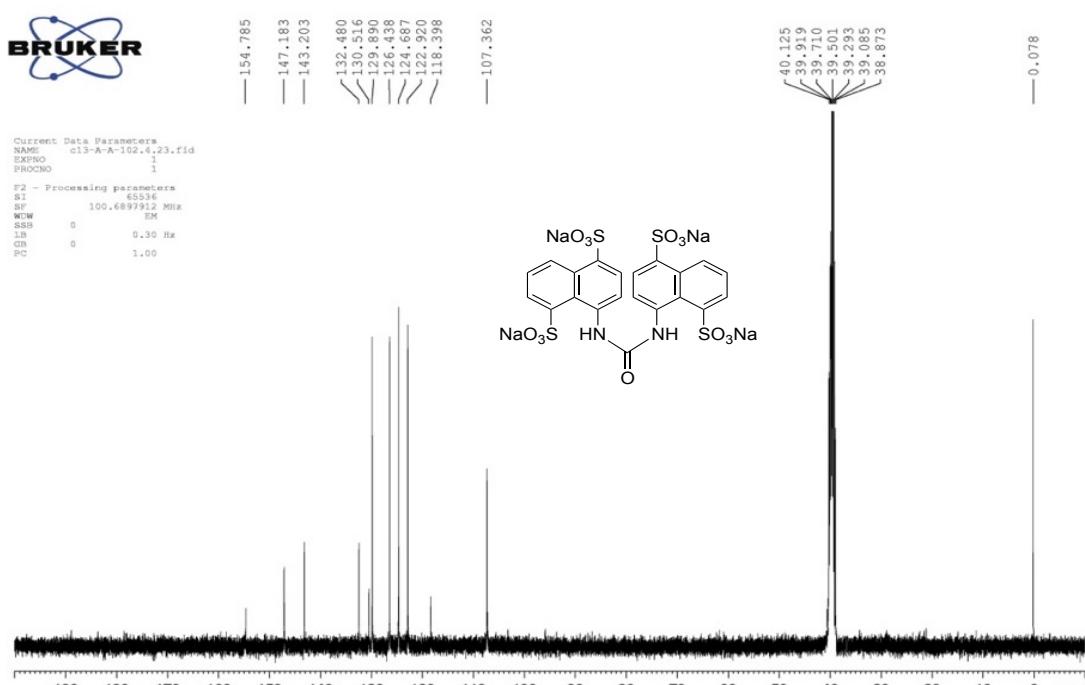
mp (recrystallized from H₂O/EtOH) 205.4–205.6 °C; ¹H NMR (DMSO-*d*₆, 400 MHz) δ 8.89 (d, 1 H, *J* = 8.8 Hz, ArH), 8.31 (d, *J* = 8.8 Hz, 1 H, ArH), 7.63 (d, *J* = 7.6 Hz, 1 H, ArH), 7.44 (s, 1 H, ArH), 7.39 (s, 1 H, ArH), 7.30–7.26 (m, 2 H, ArH), 6.88 (t, *J* = 7.6 Hz, 1 H, ArH), 6.67 (d, *J* = 7.6 Hz, 1 H, ArH), 6.60–6.43 (m, 3 H, ArH), 4.11 (q, *J* = 6.8 Hz, 2 H, OCH₂), 2.05 (s, 3 H, CH₃), 1.03 (t, *J* = 6.8 Hz, 3 H, CH₃); ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 183.55 (C=S), 165.65 (C=O), 165.15 (C=O), 147.03, 144.58, 139.99, 136.26, 135.29, 133.62, 131.96, 130.28, 128.98, 127.89, 126.99, 125.64, 125.39, 123.26, 122.78, 122.71, 122.71, 120.10, 118.31, 116.72, 114.62, 113.36, 66.03, 18.53, 14.51; IR (KBr) 3346 (br, NH), 2927 (s), 1646 (s, C=O), 1574 (w), 1544 (m), 1422 (m), 1334 (m), 1227 (m) cm⁻¹; HRMS (ESI) *m/z* [M + H]⁺ calcd for C₂₈H₂₃N₃Na₂O₉S₃ + H 688.0470, found 688.0478.

¹H-NMR, ¹³C-NMR, and IR Spectra as well as HRMS Data of Sodium Organosulfonates 3, 5a, 5b, and 9

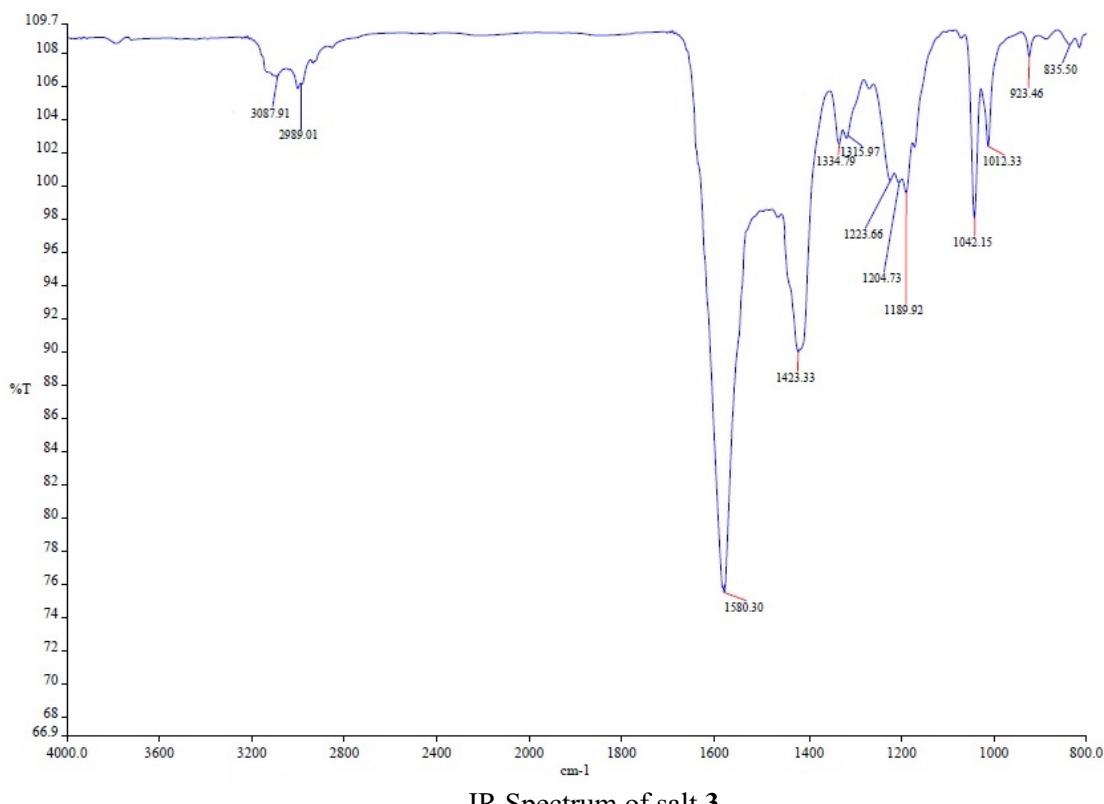
Compound 3



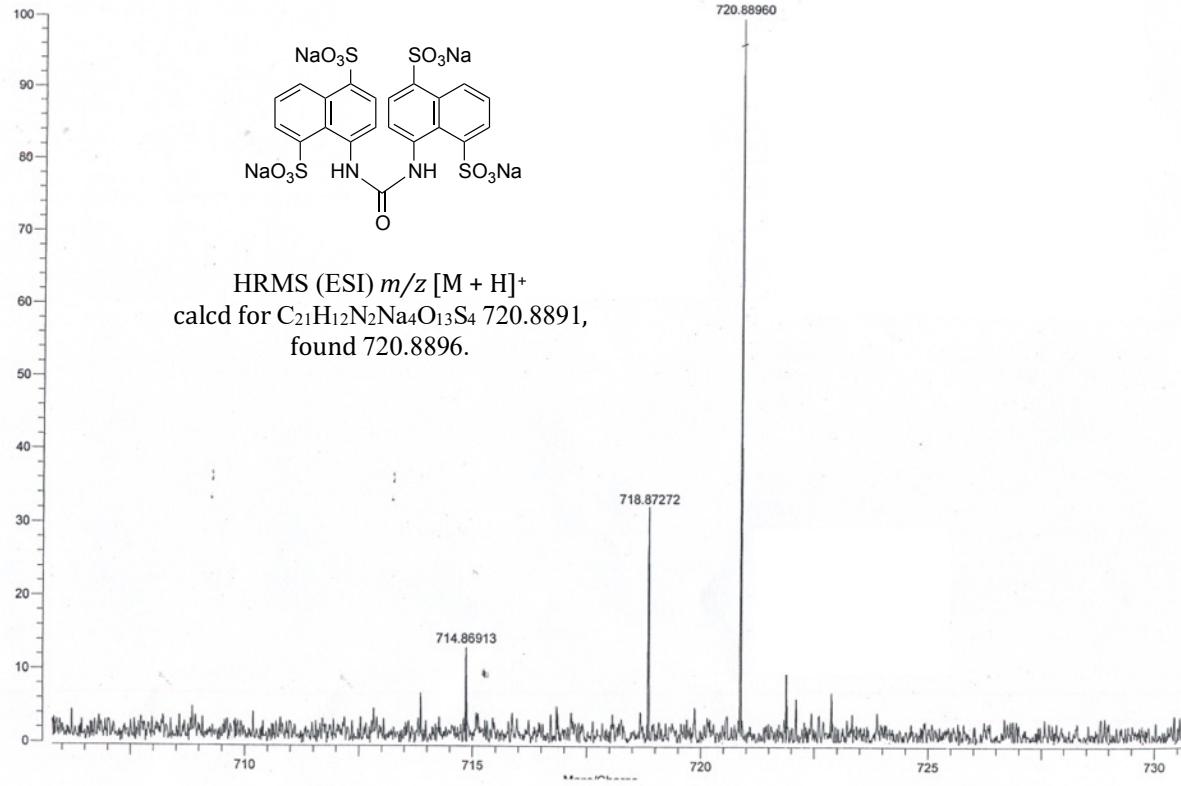
¹H NMR spectrum of salt 3



¹³C NMR spectrum of salt 3

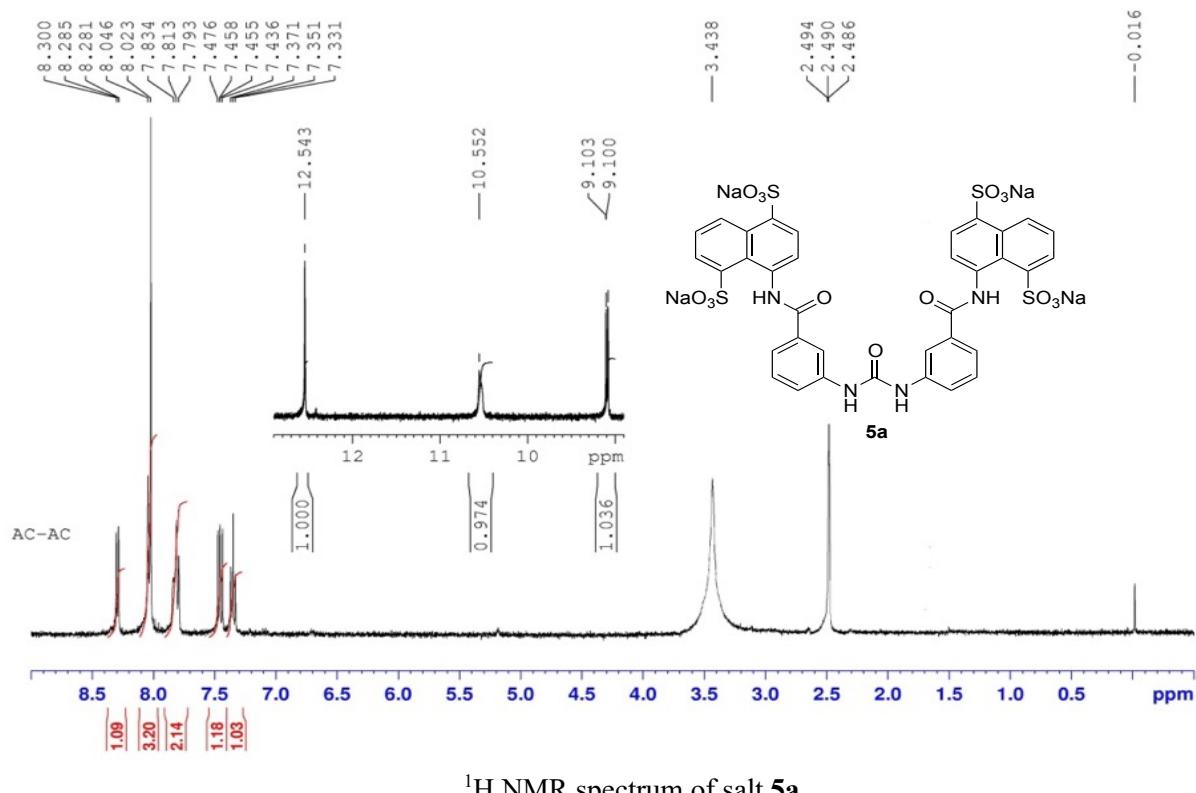


IR Spectrum of salt 3

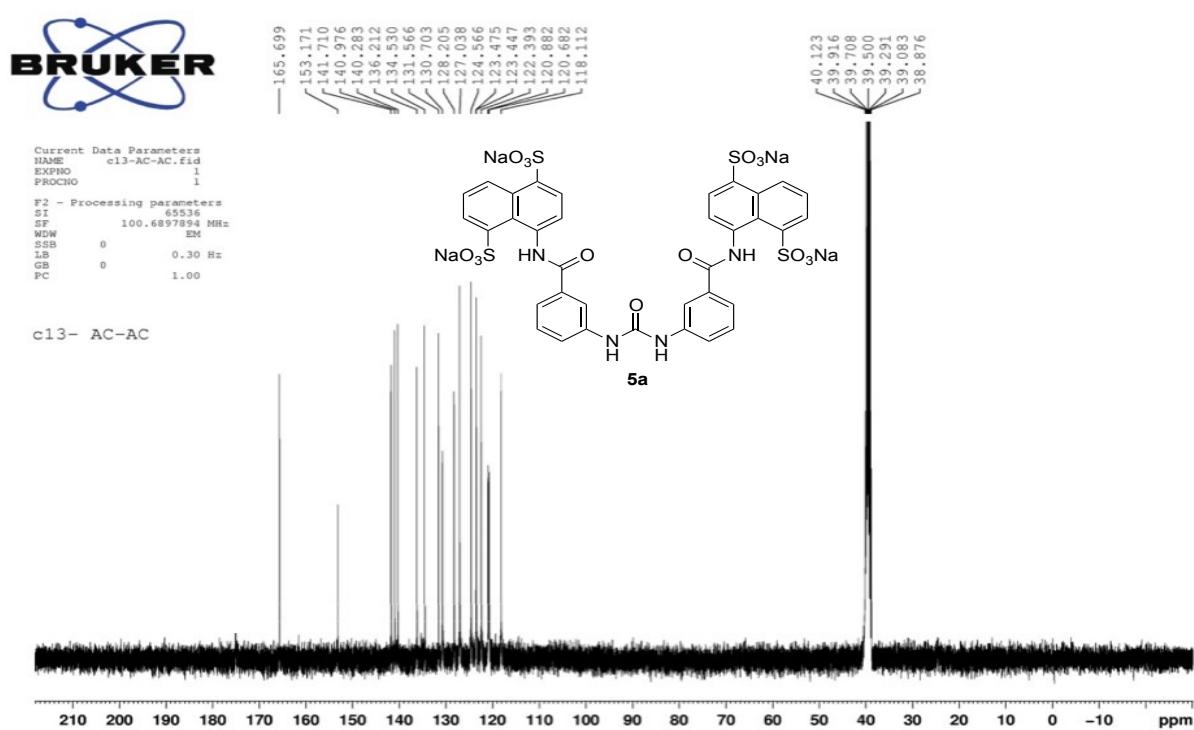


HRMS of salt 3

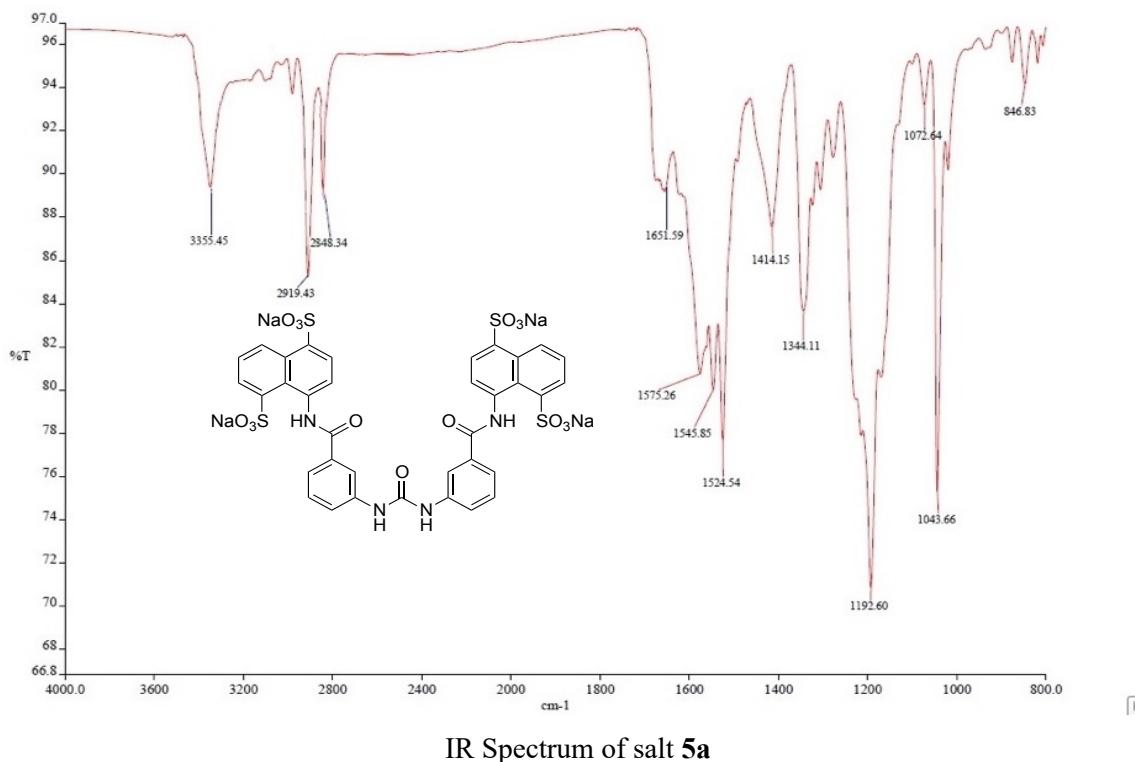
Compound 5a



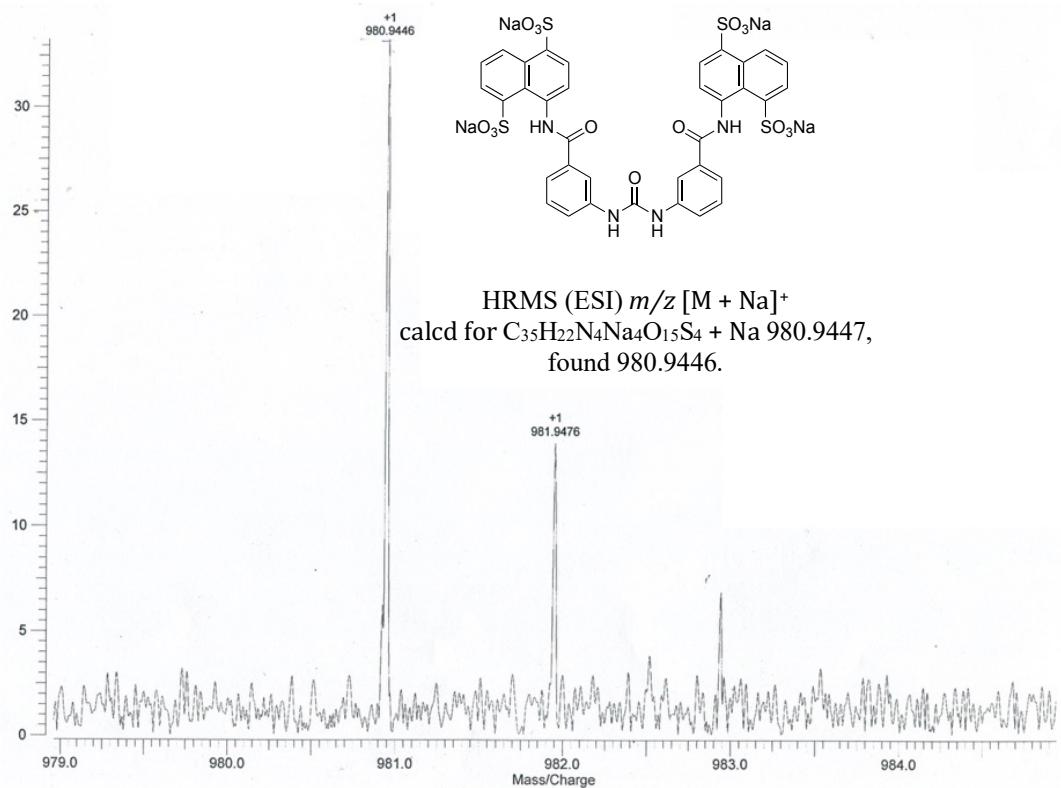
¹H NMR spectrum of salt 5a



¹³C NMR spectrum of salt 5a

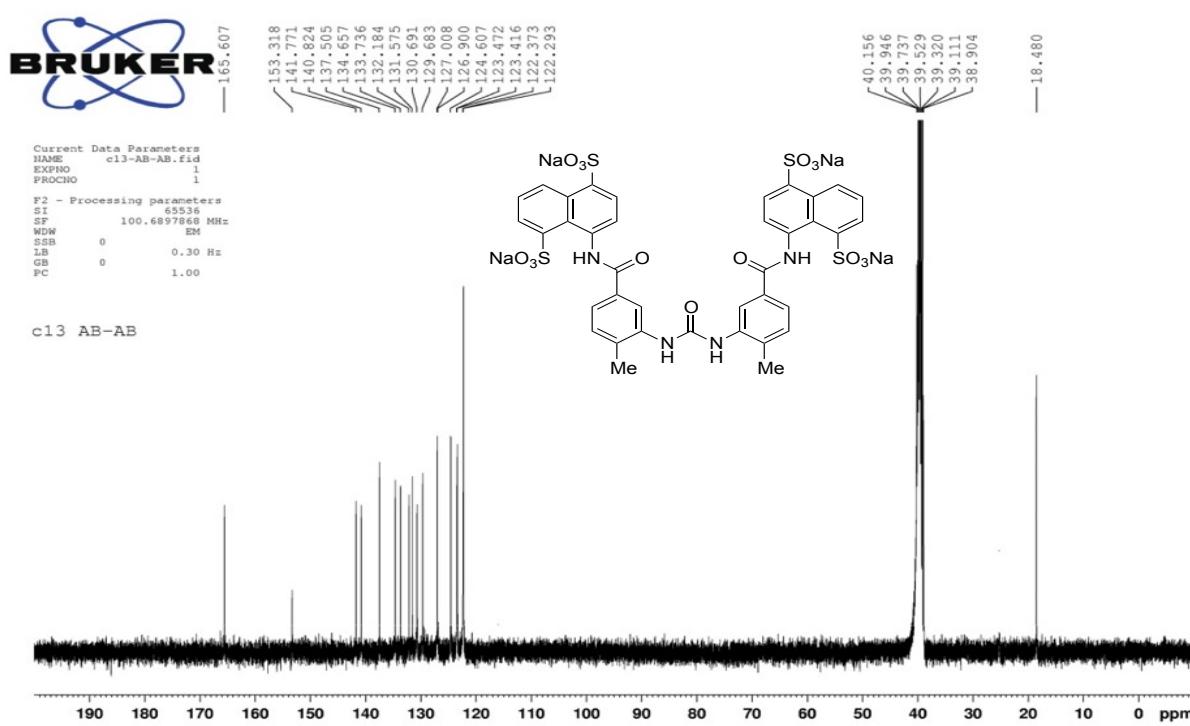
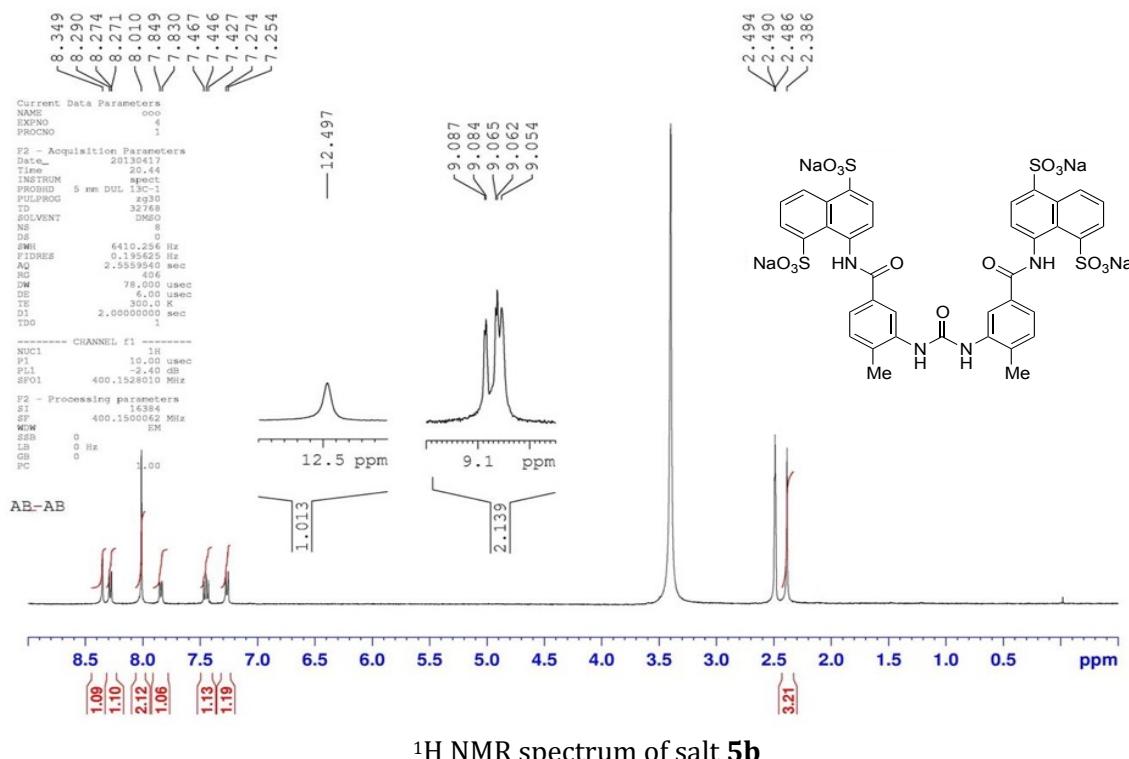


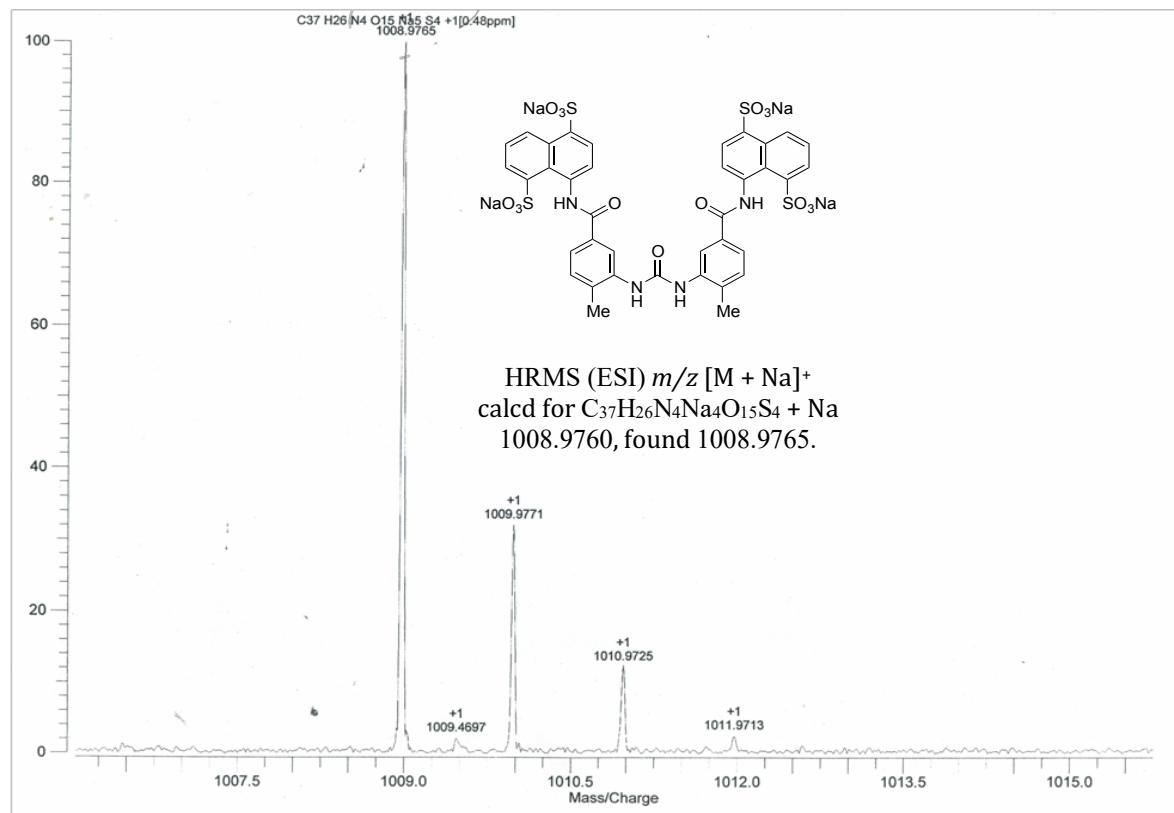
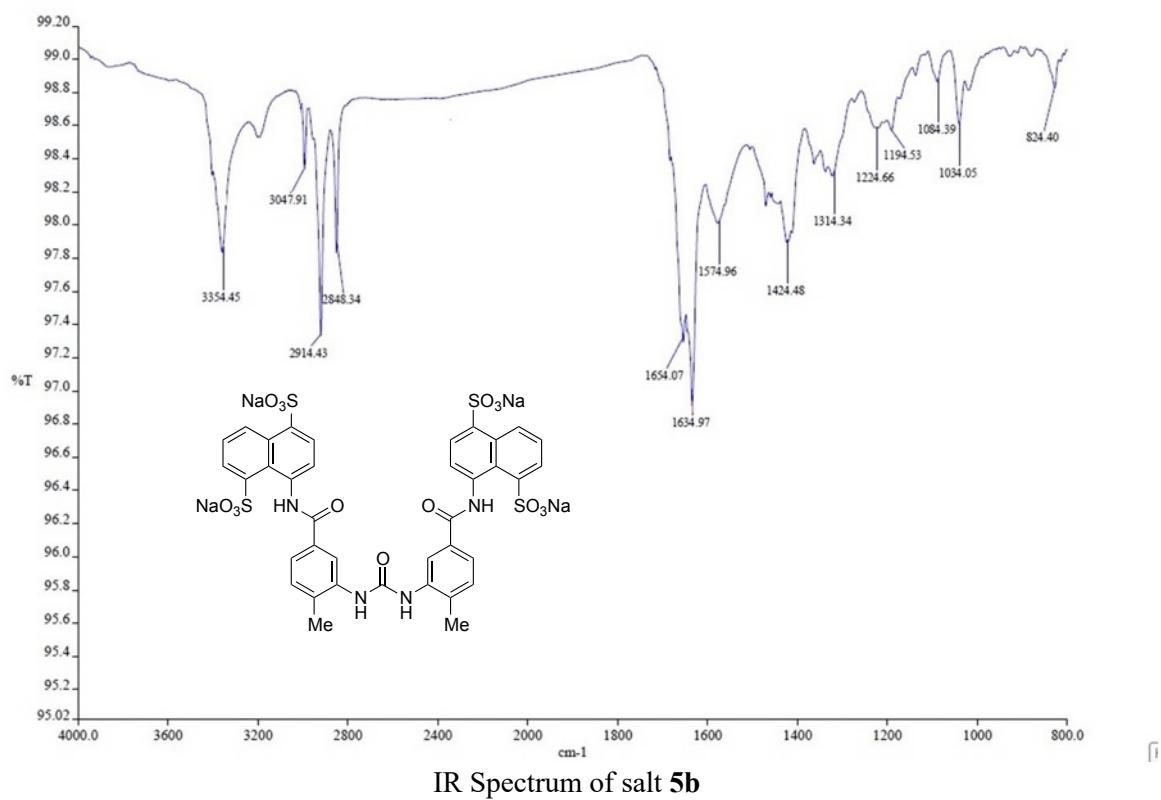
IR Spectrum of salt **5a**



HRMS of salt **5a**

Compound 5b

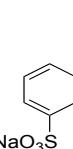




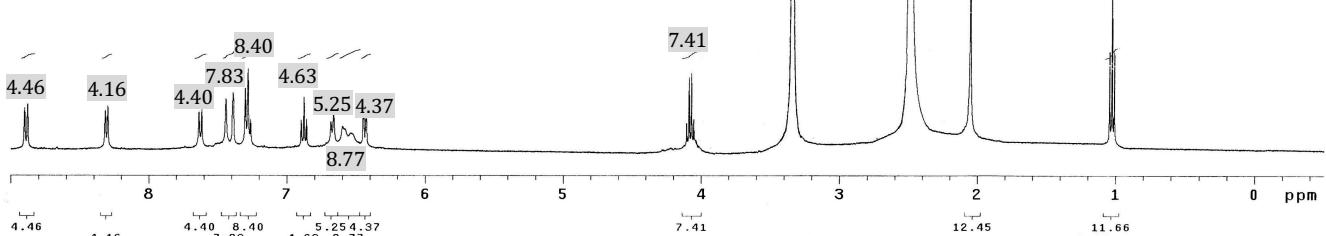
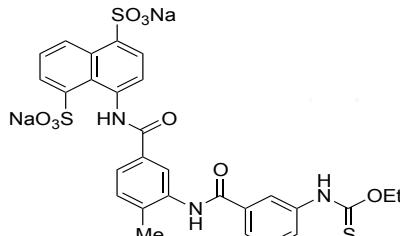
HRMS of salt 5b

Compound 9

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solvent	DMSO	dn	H1
file	exp	dprw	30
ACQUISITION		dof	0
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tn	H1	dmn	
at	2.700	mf	200
np	32768	dseq	
sw	6000.6	dres	1.0
fb	3400	homo	n
bs	4	PROCESSING	
tpwrr	55	1b	0.10
pw		wkfile	
d1	1.000	proc	ft
tof	0	fn	65536
nt	1000	math	f
ct	148		
alock		werr	
gain	not used	wexp	
FLAGS		wbs	
ii	n	wnt	
in	n		
dp	y		
hs	nn		
DISPLAY			
sp	-200.1		
wp	3799.8		
vs	973		
sc	0		
wc	250		
hzmm	15.20		
ls	732.80		
rfl	1565.6		
rfp	995.9		
th	4		
ins	100.00		
nm	ph		



O=C1C=C(C=C1N)C(=O)c2ccc(O[N+](=O)[O-])cc2



*The integral values are presented in the gray boxes.

¹H NMR spectrum of salt 9

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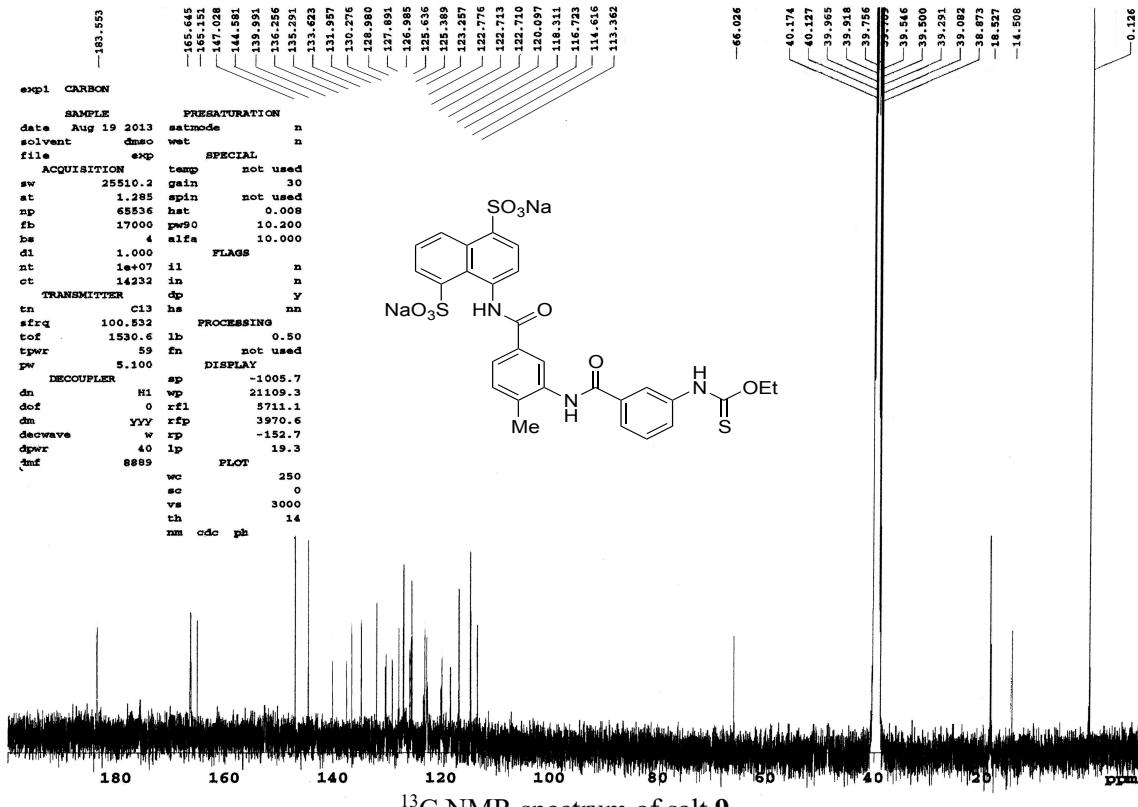
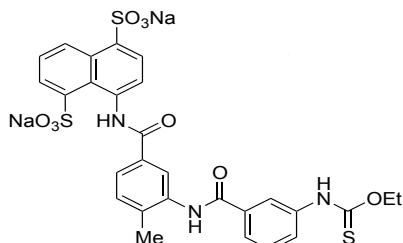
exp1 CARBON                                —183.553

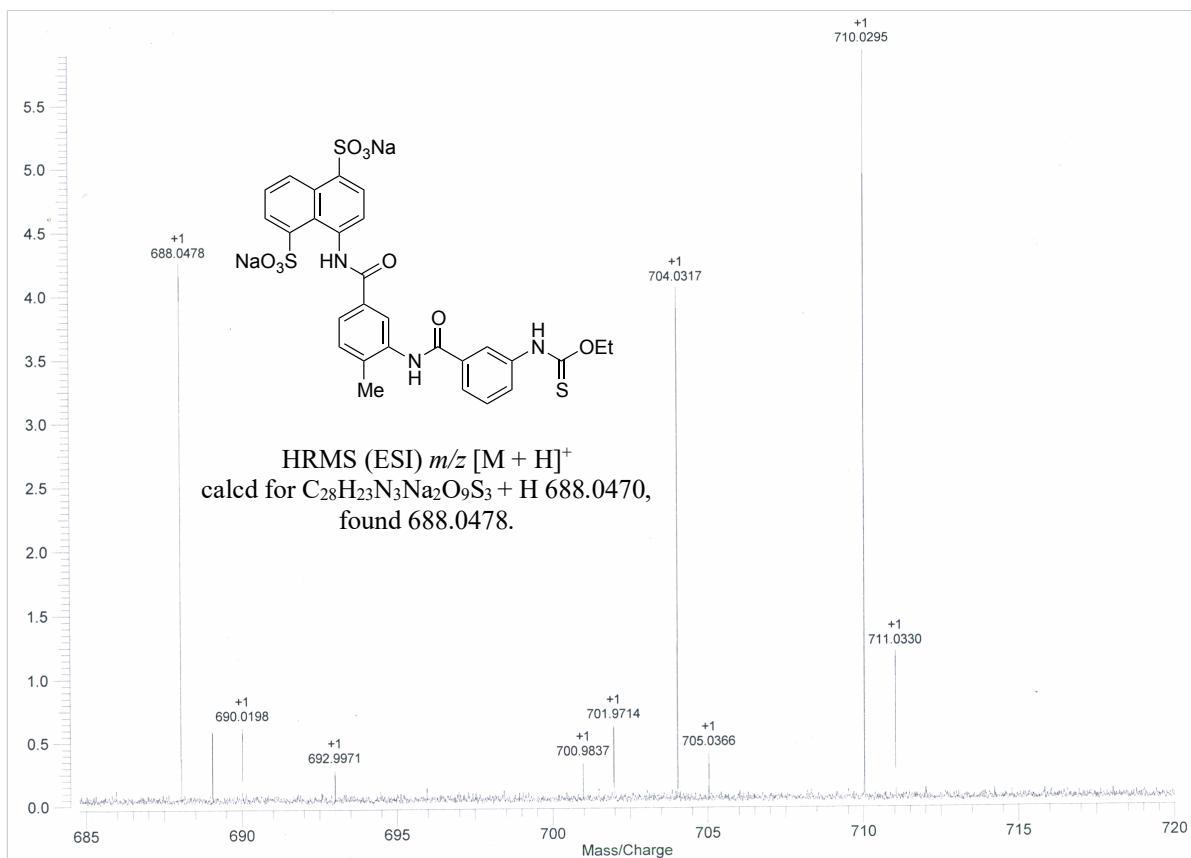
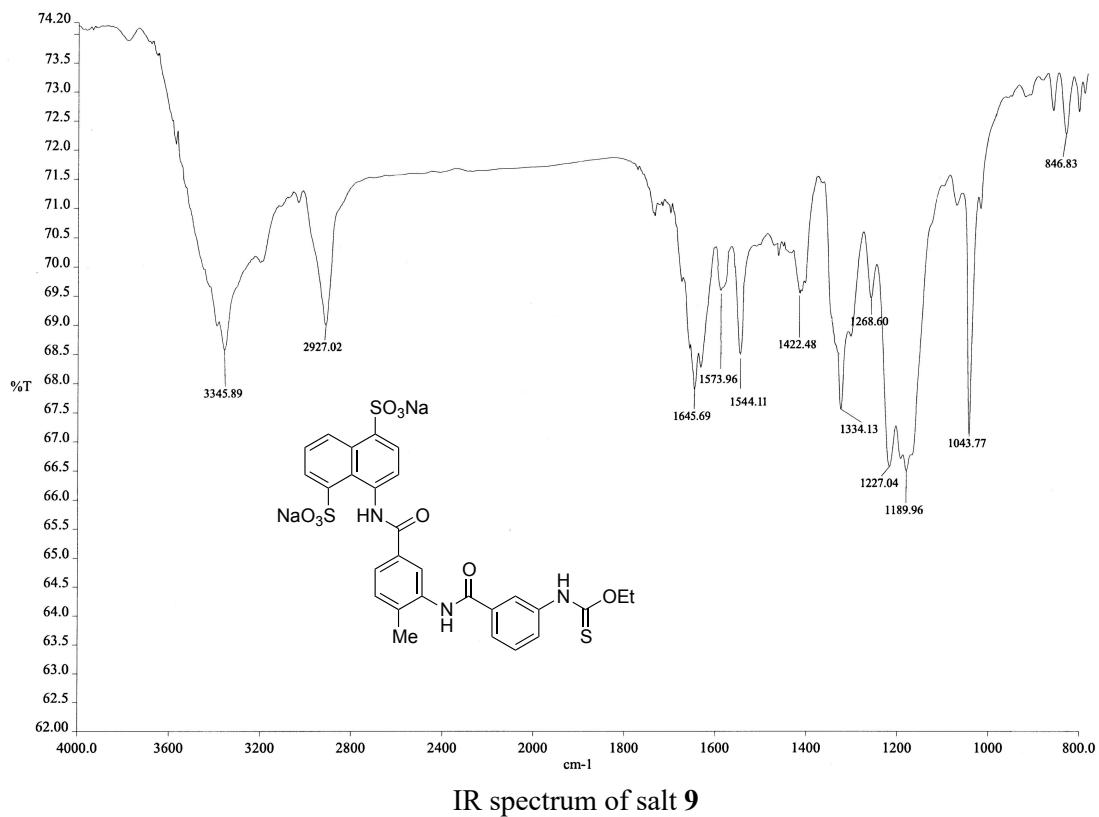
SAMPLE                                     PRESATURATION
date Aug 19 2013   satmode    n
solvent dmso   wet
file      exp   SPECIAL

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st      1.285   spin      not used
np      65536   hat       0.008
fb      17000   pw90     10.200
bs      4        alfa     10.000
d1      1.000   FLAGs
nt      1e+07   il       n
ct      14232   in       n
          nn
TRANSMITTER          dp      y
tn      C13   hs
sfreq  100.532   PROCESSING
tot      1530.6   lb      0.50
tpwr     59   fn   not used
pw      5.100   DISPLAY

DECOUPLER           ap      -1005.7
dn      H1   wp     21109.1
dof      0   rrf1    5711.1
dm      YYY   rrp     3970.6
decwave   rp     -152.7
dpwr     40   lp      19.3
jmf      8889   PLOT
          wc      250
          sc      0
          vs     3000
          th      14

```





HRMS of salt 9

Table S1. XIAP-BIR1 thermal stability and affinity for NF023 like molecules moieties.

Sample	Compounds Chemical Structure	Melting Temperature (°C)	ΔT_m (°C) ^a
XIAP-BIR1	/	63.5 ± 0.9	/
XIAP-BIR1/NF023		56.7 ± 1.0	-6.8
XIAP-BIR1/Suramin		50.0 ± 0.5	-13.5
XIAP-BIR1/3		63.0 ± 0.3	-0.5
XIAP-BIR1/5a		56.2 ± 0.3	-7.3
XIAP-BIR1/5b		56.2 ± 0.3	-7.3
XIAP-BIR1/6		60.5 ± 0.1	-3.0
XIAP-BIR1/7		60.5 ± 0.2	-3.0
XIAP-BIR1/9		62.5 ± 1.5	-1.0
XIAP-BIR1/10		61.2 ± 0.6	-2.3
XIAP-BIR1/NAF2		61.0 ± 0.3	-2.5

^a ΔT_m is calculated as the difference from the apoprotein melting temperature.

Thermal shift assays to monitor unfolding of 80 μ M XIAP-BIR1 (~1 mg/ml) upon incubation with 1.0 mM of each compound were conducted in a MiniOpticon Real-Time PCR Detection System (Bio-Rad). The fluorescent dye Sypro Orange was used to monitor protein unfolding. The sample plates were heated from 15 to 95 °C, with a heating rate of 0.5 °C/5 sec. Fluorescence intensity was measured within the excitation/emission ranges 470–505/540–700 nm. All the experiments were performed in triplicate, to calculate average T_m values and associated standard errors.

Suramin (ΔT_m of about -13.5°C), NF023 and its analogs **5a** and **5b** (ΔT_m of about -7°C), and compounds **6** and **7** (ΔT_m of about -3°C) caused a significant effect on the protein conformational stability.

Figure S1. The V86E mutation is compatible with NF023 predicted binding. The structure of XIAP-BIR1 V86E mutant (in magenta) superimposed to XIAP-BIR1/NF023 docking prediction (light blue cartoons and green sticks) reveals that the steric hindrance of E86 side chain is fairly compatible with NF023 predicted binding, as confirmed by MST experiments, where the measured affinity of NF023 vs XIAP-BIR1 V86E is not drastically affected compared to the value observed vs the wild type protein domain.

