

Repurposing the selective estrogen receptor modulator *bazedoxifene* to suppress gastrointestinal cancer growth

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Appendix Figure S1 SERM-selective of suppression of IL11-mediated STAT3 signaling activity.

(A) Effect of *bazedoxifene* (BZA) and *tamoxifen* (TAM) on IL11-induced and STAT3-dependent pAPRE luciferase reporter activity in HEK293T cells expressing human IL11R α . Cells were co-transfected with a non-responsive renilla-luciferase plasmid. Cultures were stimulated with IL11 (30 ng/ml) and the indicated concentrations of BZA or TAM for 18 h. Results are expressed as relative luciferase units (RLU), i.e. firefly luciferase activity normalized against renilla luciferase activity in each individual culture. Data is presented as fold change to values obtained for the IL11 alone treatment wells; Mean \pm SEM, n=3 independent experiments, * p = 0.023, ** p = 0.003, both values compared to IL11 treatment; *** p = 0.0001 compared to no treatment; 2-way repeated measures ANOVA, Tukey's multiple comparison test.

(B) Effect of *tamoxifen* (TAM) on the proliferation of IL11-stimulated (30 ng/ml) BAF/03 murine B-cell lines expressing human IL11R α , as determined by MTS-assay. Measurements were carried out 48 h after treatments. Data are mean \pm SEM, n=3 independent experiments, *** p = 0.0001 compared to no treatment, ANOVA, Dunnett's comparison test.

Appendix Figure S2 Models of *bazedoxifene* interactions with Site III residues of the hexameric gp130 complex.

(A) Overlay of the two proposed *bazedoxifene* binding modes. The indole and azepanyl rings for each binding mode are indicated. *In silico* modeling identified two binding modes of *bazedoxifene* with gp130, shown in (C) and (E) as a grey ribbon with some Site III side-chains shown as sticks. *Bazedoxifene* is shown as purple or green sticks.

(B) Overlay of *tamoxifen* (orange sticks) with one of the *bazedoxifene* binding modes (purple sticks) via their common phenoxypropylamine core to illustrate the structural differences between the two compounds. When docked in this manner, *tamoxifen* is unable to bind to the gp130 Site III surface due to steric clashes with gp130 residues L3, P5, N92 and Y94. Additionally, the two methyl substituents on the *tamoxifen* amine are unable to reach the hydrophobic pocket in which the *bazedoxifene* azepanyl ring binds. In addition, there are differences in the types of interactions that *tamoxifen* could potentially

make with gp130. The *bazedoxifene* indole and phenol rings are able to interact with gp130 residues via hydrogen bonds, in contrast to the phenyl rings and ethyl group of tamoxifen which can not hydrogen bond. Similar steric clashes with gp130 residues are observed when *tamoxifen* is overlaid with the alternative *bazedoxifene* binding mode (shown as green sticks in (A)).

(C) The *bazedoxifene* indole ring hydroxyl substituent can form hydrogen bonds to Q78 and both the indole and phenol rings can form $\pi - \pi$ interactions with Y94. L3 interacts with both the *bazedoxifene* indole ring phenol substituent and the pendant phenyl ring via hydrophobic interactions. The *bazedoxifene* azepanyl ring is located in a largely hydrophobic pocket surrounded by C6, C32, F36, I83 and Q91.

(D) Gp130 depicted as a grey molecular surface in the same view as in (C). The locations of W157 and L57 in IL6 (brown), which interact with gp130, illustrate how the *bazedoxifene* indole ring and azepanyl ring (green, purple) mimic W157 and L57, respectively.

(E) The *bazedoxifene* indole ring can form $\pi - \pi$ interactions with Y94. The phenol substituent of *bazedoxifene* is able to form hydrogen bonds to N92. The *bazedoxifene* indole ring hydroxyl substituent can form hydrogen bonds with either the hydroxyl group of Y94 or the side-chain of E12. L3 can form hydrophobic interactions with the pendant phenyl substituent of *bazedoxifene*. The *bazedoxifene* azepanyl ring is located in the same pocket of gp130 as shown in (C).

(F) Gp130 depicted as a grey molecular surface in the same view as in (E). The locations of W157 and L57 in IL6 (brown), which interact with gp130, illustrate how the *bazedoxifene* indole ring and azepanyl ring (green, purple) mimic W157 and L57 interactions in this orientation.

Amino acid codes: asparagine (N), cysteine (C), glutamic acid (E), glutamine (Q), isoleucine (I), Leucine (L), phenylalanine (F), tyrosine (Y), tryptophan (W).

Appendix Figure S3 Identification of tumor-associated immune infiltrates by flow cytometry.

Flow cytometric analysis of immune cells isolated from colon tissue collected from wildtype mice, and colonic tumours derived from *Cdx2^{CreERT2};Apc^{flox}* mice treated with vehicle or *bazedoxifene* (BZA). Representative FACS plots showing the gating strategy to exclude

epithelial cells (EpCAM⁺) and isolate immune cells (EpCAM⁻,CD45⁺), T cells (CD4⁺, CD8⁺), and myeloid cell lineages with Ly6C, Ly6G, MHCII and F480 as markers.

Appendix Figure S4 Tumor infiltrating T-cells populations are unaffected in *bazedoxifene* (BZA) or vehicle-treated *Cdx2*^{CreERT2};*Apc*^{flox} mice.

Distribution of immune cell types identified by flow cytometry analysis in colon sections containing tumors from *Cdx2*^{CreERT2};*Apc*^{flox} mice treated with vehicle or BZA. Distal colon tissue of untreated wild-type mice (WT) were included as a reference.

(A) Representative flow cytometry plot showing EpCAM⁻,CD45⁺,TCRβ⁺,CD8⁺ and CD8⁺,PD1⁺ T cell populations, and quantitation from three experimental cohorts.

(B) Representative flow cytometry plot showing EpCAM⁻,CD45⁺,TCRβ⁺,CD4⁺ and CD4⁺,PD1⁺ T cell populations, and quantitation from three experimental cohorts.

Data are mean ± SEM, with n=3 mice per cohort, *p* value derived from unpaired T-test.

Appendix Figure S5 Tumor infiltrating myeloid and B cell populations are unaffected in *bazedoxifene* (BZA) or vehicle-treated *Cdx2*^{CreERT2};*Apc*^{flox} mice.

Distribution of immune cell types identified by flow cytometry analysis in colon sections containing tumors from *Cdx2*^{CreERT2};*Apc*^{flox} mice treated with vehicle or BZA. Distal colon tissue of untreated wild-type mice (WT) were included as a reference. Representative flow cytometry plots for specific immune cell subsets and quantitation in the three experimental cohorts are shown as follows:

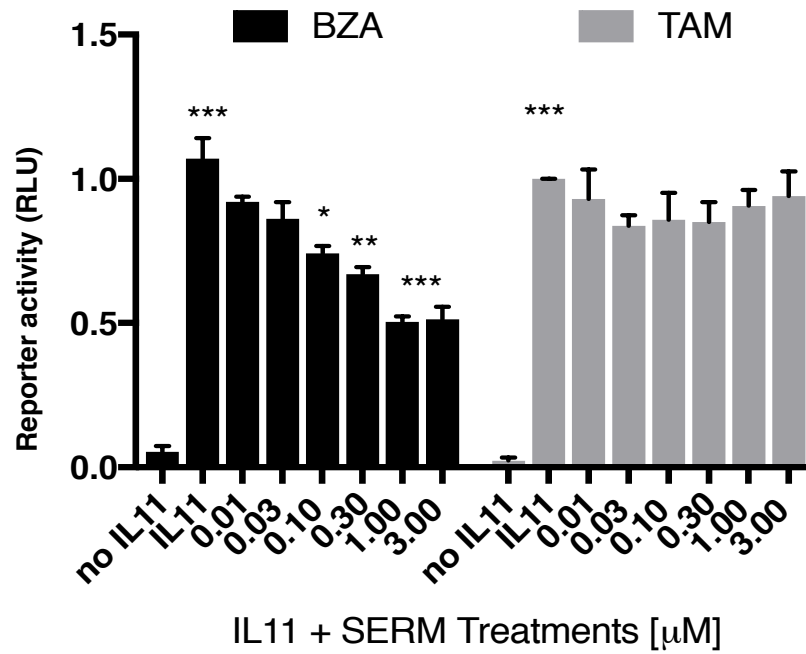
(A) EpCAM⁻,CD45⁺,Cd11b⁺,Ly6G^{lo}, MHCII⁺, F480⁺ macrophages,

(B) EpCAM⁻,CD45⁺,Ly6C^{hi},Ly6G^{lo} monocytic myeloid-derived suppressor cells (M-MDSCs) and CD45⁺,CD11b⁺,Ly6C^{lo},Ly6G^{hi} granulocytic MDSCs (G-MDSCs) populations, and

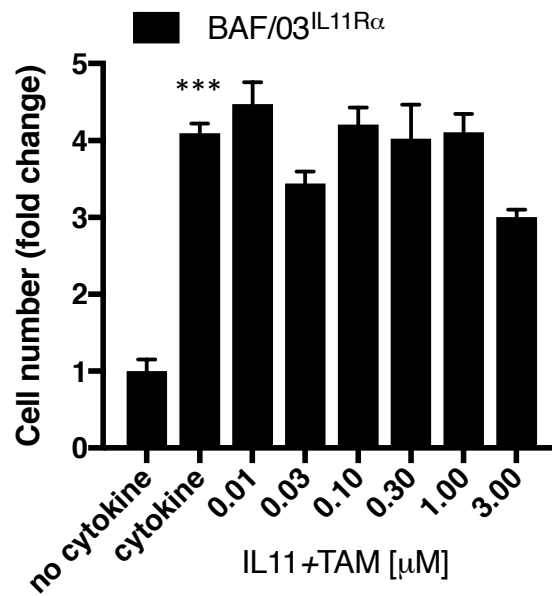
(C) EpCAM⁻,CD45⁺,CD19⁺ B cell populations.

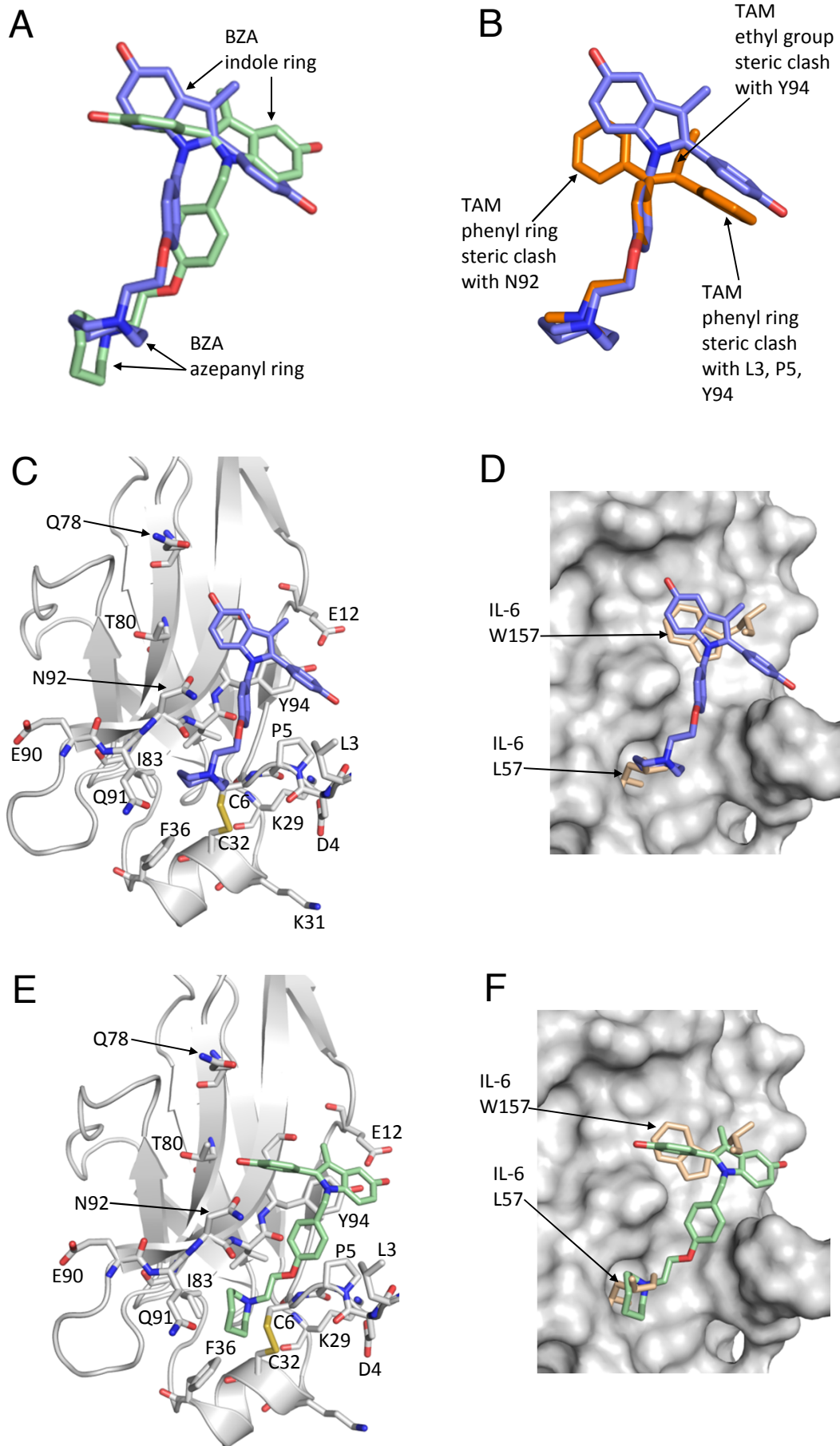
Data are mean ± SEM, with n=3 mice per cohort. Each symbol represents individual mice, *p* value derived from unpaired T-test.

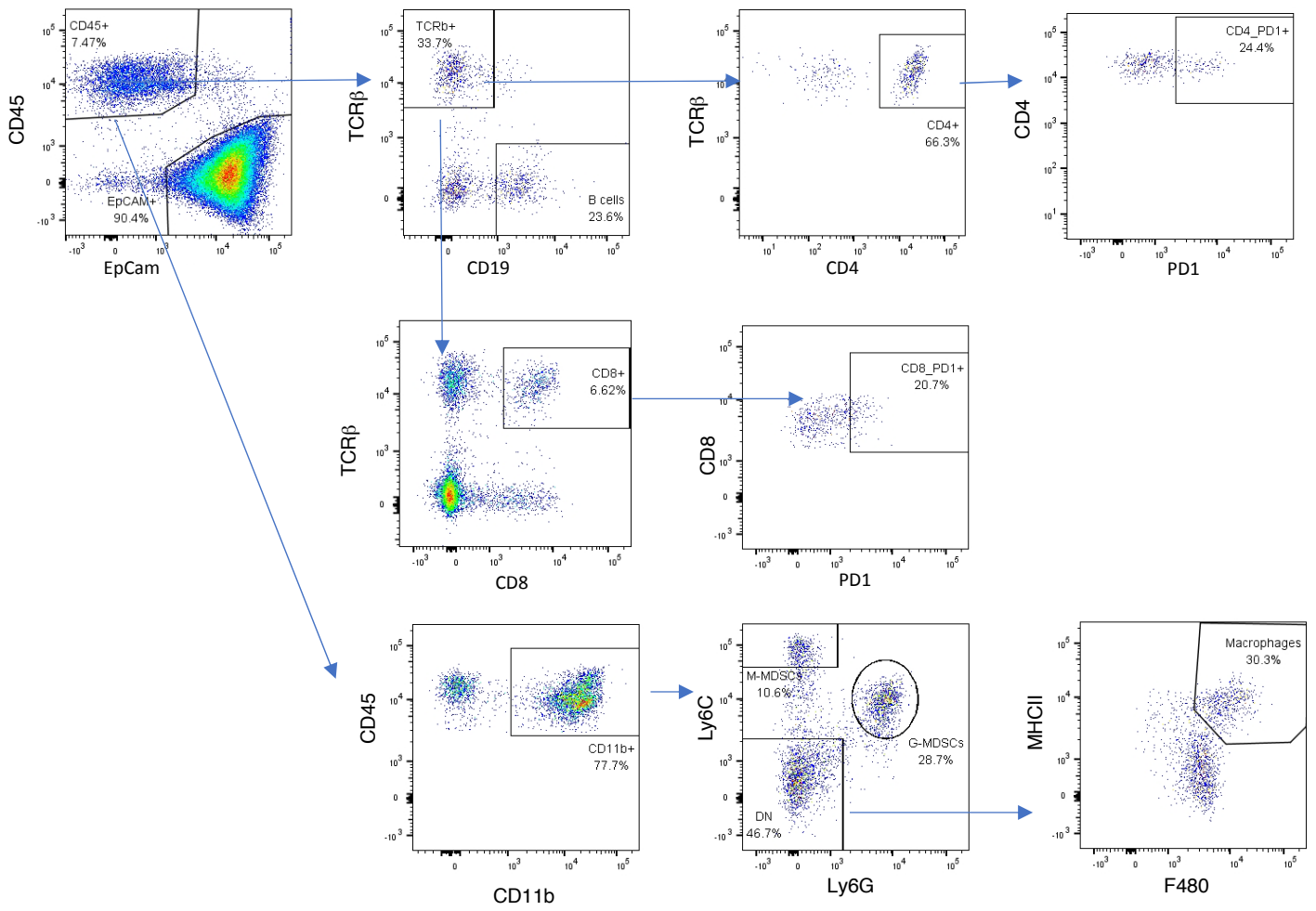
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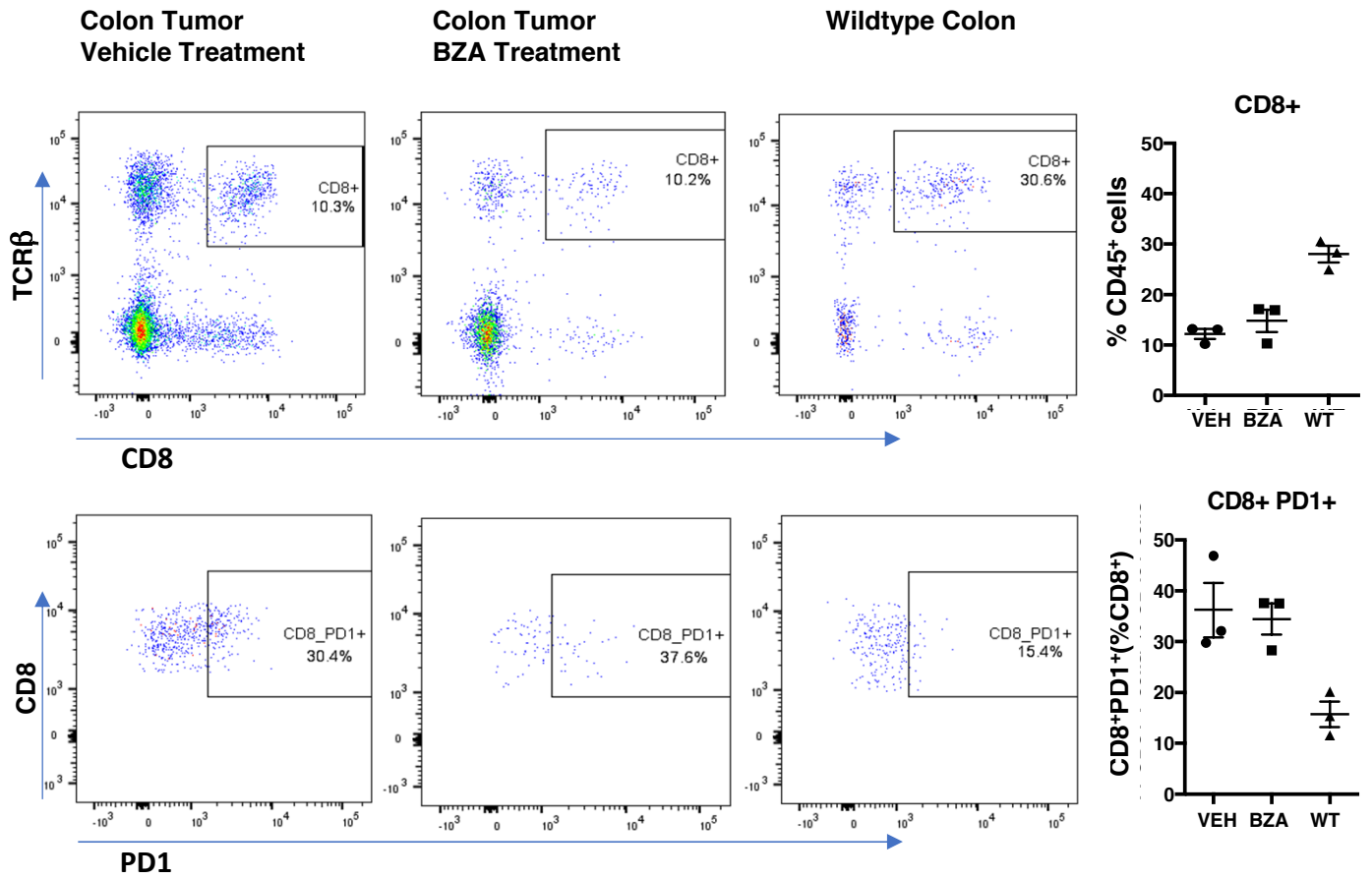
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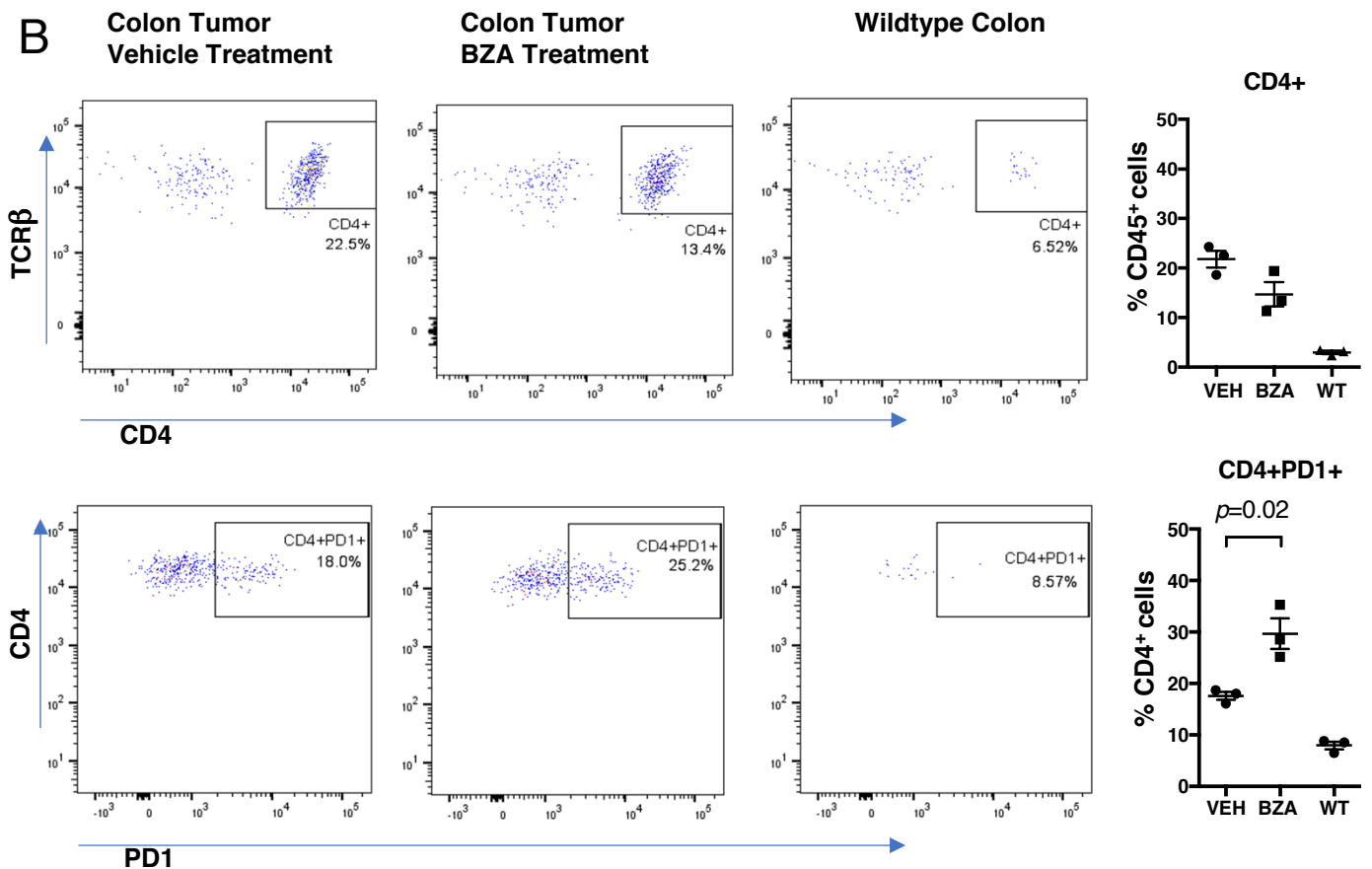


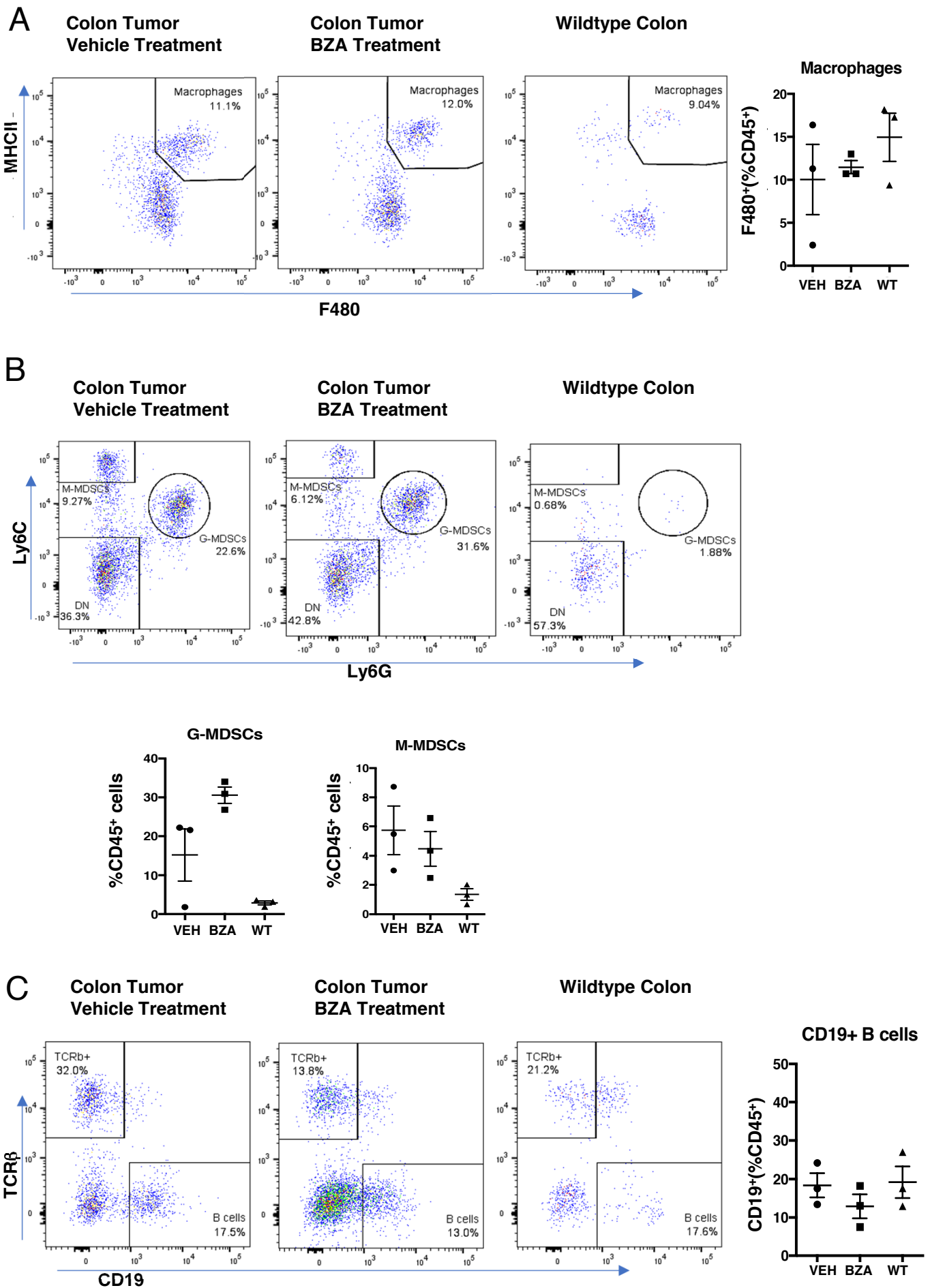


A



B





Appendix Table S1

Human Primer Sequences for QPCR	
Gene	Primer Sequence
<i>HPRT</i> Forward	CCCTGGCGTCGTGATTAGTG
<i>HPRT</i> Reverse	TGGGTCAGGGGTGGTTATTG
<i>Il11</i> Forward	ATGAACTGTGTTTGCCGCCT
<i>Il11</i> Reverse	GAATCCAGGTTGTGGTCCCC
<i>IL6</i> Forward	CAATGAGGAGACTTGCCTGGTG
<i>Il6</i> Reverse	TCTTGCCAGGTGACACTGAG
<i>IL6ST</i> Forward	AGATCCATCCCATACTCAAGGC
<i>IL6ST</i> Reverse	ACAGGGTGAGTAGCTTGAAAGTC

Mouse Primer Sequences for QPCR	
Gene	Primer Sequence
<i>Hprt</i> Forward	GGC CAG ACT TTG TTG GAT TTG
<i>Hprt</i> Reverse	CGC TCA TCT TAG GCT TTG TAT TTG
<i>Il11</i> Forward	ATG AAC TGT GTT TGT CGC CTG
<i>Il11</i> Reverse	CCA GGA AGC TGC AAA GAT CC
<i>Il6</i> Forward	TCT ATA CCA CTT CAC AAG TCG GA
<i>Il6</i> Reverse	GAA TTG CCA TTG CAC AAC TCT TT
<i>Icam1</i> Forward	CAA TTT CTC ATG CCG CAC AG
<i>Icam</i> Reverse	AGC GGA AGA TCG AAA GTC CG
<i>Reg3a</i> Forward	CTGGTC TGC CAG AAG AGA CC
<i>Reg3a</i> Reverse	GGT AGT TGT CCA CTC TGC CG
<i>Socs3</i> Forward	GCG GGC ACC TTT CTT ATC C
<i>Socs3</i> Reverse	TCC CCG ACT GGG TCT TGA C
<i>Axin2</i> Forward	AGC CTA AAG GTC TTA TGT GG
<i>Axin2</i> Reverse	ATG GAA TCG TCG GTC AGT
<i>Lgr5</i> Forward	CTT CAC TCG GTG CAG TGC T
<i>Lgr5</i> Reverse	GAT CAG CCA GCT ACC AAA TAG G
<i>Sox9</i> Forward	CCA CGG AAC AGA CTC ACA TCT CTC

<i>Sox9</i> Reverse	CTG CTC AGT TCA CCG ATG TCC ACG
<i>Gapdh</i> Forward	CGA CCC CTT CAT TGA CCT TA
<i>Gapdh</i> Reverse	GCC TTG ACT GTG CTG TTG AA
<i>Esr1</i> Forward	CCT CCC GCC TTC TAC AGG
<i>Esr1</i> Reverse	CAC ACG GCA CAG TAG CGA

Appendix Table S2

Antibodies used for Western Blot			
Antibody	Supplier	Catalog #	Dilutions Used
Phospho-STAT3 (Tyr705)	Cell Signaling	#9145	1:500
STAT3	Cell Signaling	#4904	1:1000
Phospho-p44/42 MAPK (Erk1/2)	Cell Signaling	#4370	1:2000
p44/42 (Erk1/2)	Cell Signaling	#4695	1:1000
Akt	Cell Signaling	#9272	1:1000
Phospho-Akt	Cell Signaling	#4058	1:1000
BIM	Cell Signaling	#2819	1:1000
Cyclin D1	Cell Signaling	#2978	1:1000
MCL-1	Cell Signaling	#5453	1:1000
Survivin	Cell Signaling	#2808	1:1000
BCL-xL	BD Bioscience	#551022	1:1000
β -actin	Sigma	#A1978	1:1000
GAPDH	Sigma	#G8795	1:2000
Antibodies used for Immunohistochemistry			
Phospho-STAT3	Santa Cruz	SC-7993	1:150
Ki67	ThermoFisher	MAS14520	1:150
β -Catenin	BD Biosciences	#610153	1:200

Appendix Table S3

Antibodies used for Flow Cytometry				
Marker	Label	Supplier	Catalog #	Dilutions Used
EpCAM (CD326)	FITC	MACS	130102214	1:300
EpCAM (CD326)	APC	e-Bioscience	EBS-17-5791-80	1:300
TCR β	PE	BioLegend	109208	1:500
PD1	PF710	INVITROGEN	46998582	1:300
CD8a	PECy7	BD Pharmingen	561097	1:300
CD45.2	A700	WEHI		1:500
F4/80	FITC	MACS	130-102-327	1:300
CD11b	PE	BD Pharmingen	553311	1:400
Ly-6G	PECy7	BD Pharmingen	560601	1:300
Ly-6C	eF450	e-Bioscience	48-5932-82	1:50