

Supplementary Information for

**Dynamic characteristics of GMP reductase complexes revealed
by high resolution ³¹P field cycling NMR relaxometry**

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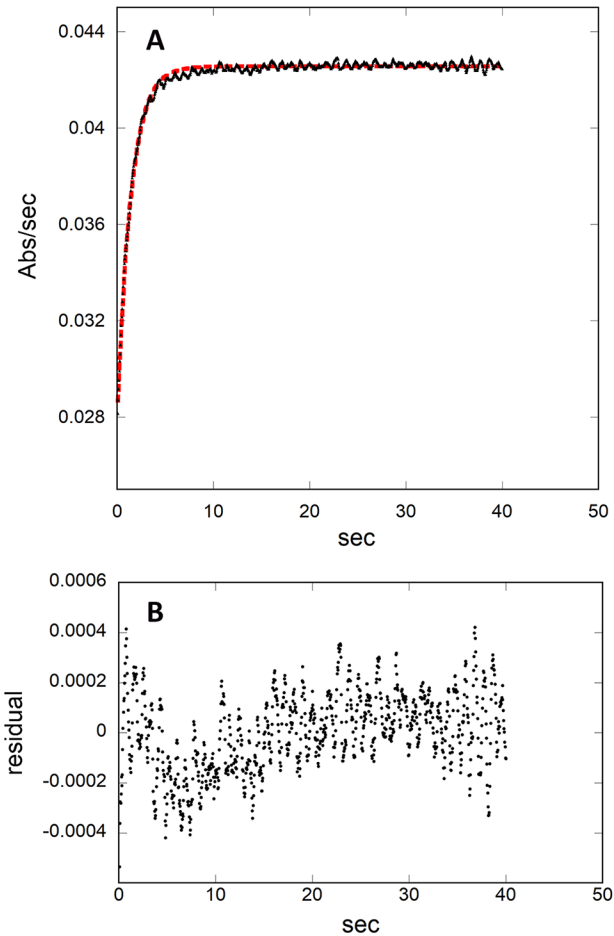
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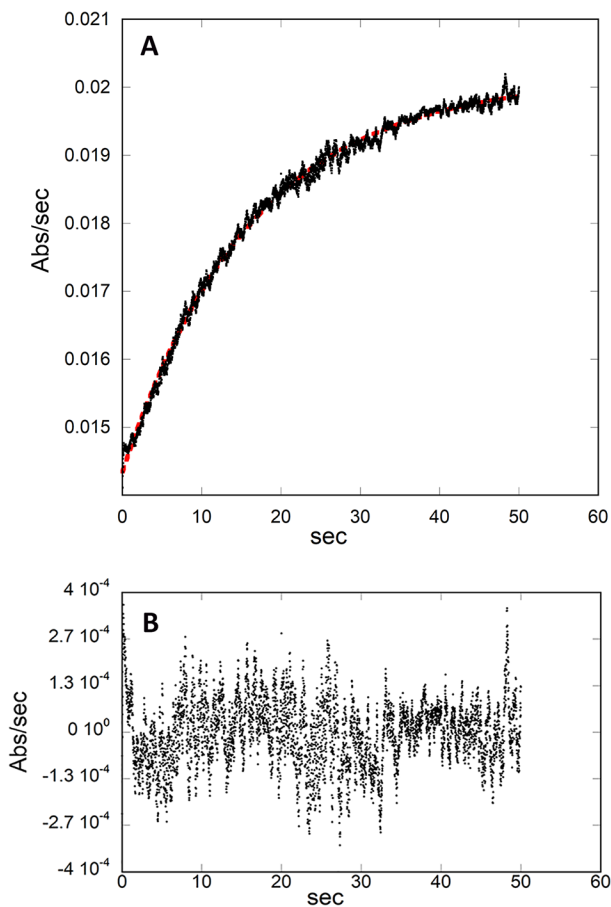
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Supporting Table S1. Extrapolated ^{31}P - ^1H distances.

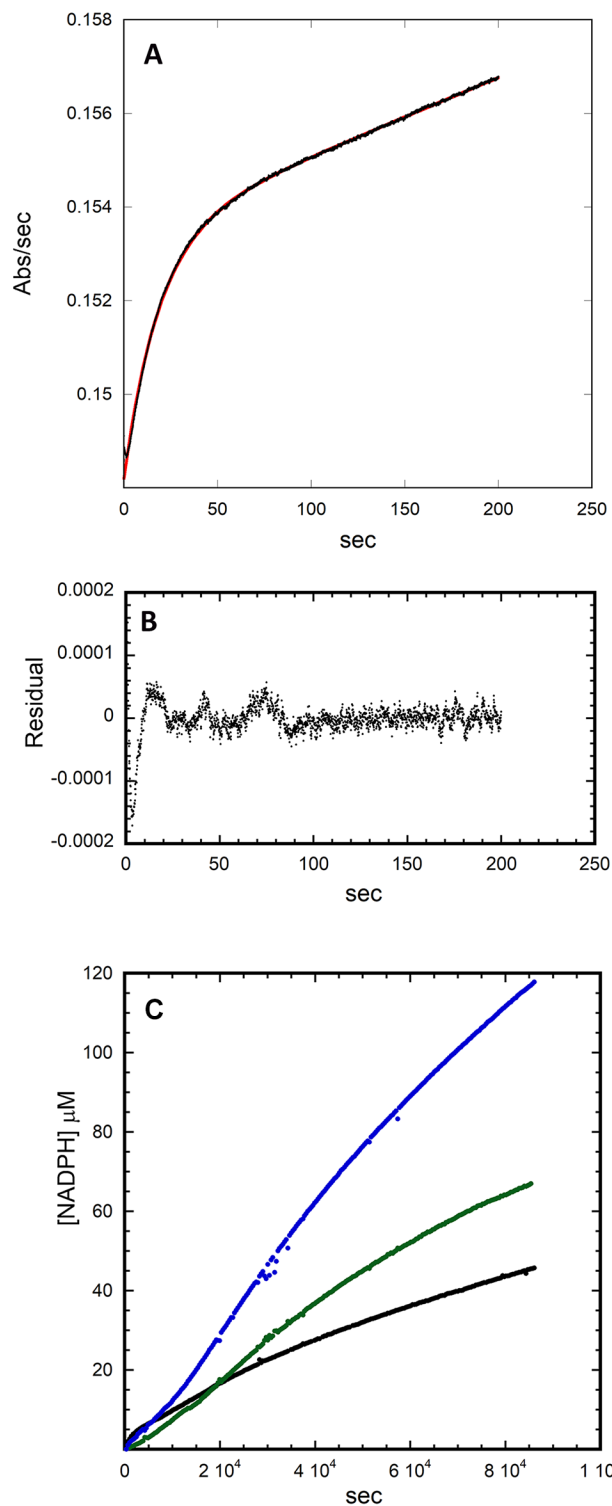
^{31}P distance	Exp.	IMP/GMP monoP	NADP ⁺ monoP	NADP ⁺ diP
WT•IMP •NADP ⁺	H ₂ O	2.9 ± 0.2	2.8 ± 0.2	3.0 ± 0.1
WT•GMP •NADP ⁺	H ₂ O	4.1 ± 0.2	3.1 ± 0.1	3.2 ± 0.1
WT•IMP •NADP ⁺	D ₂ O	3.4 ± 1.2	3.4 ± 1.6	3.6 ± 1.9
				3.5 ± 1.7
WT•GMP •NADP ⁺	D ₂ O	4.1 ± 2.0	3.7 ± 1.4	3.8 ± 0.8
				3.5 ± 0.9
WT•dIMP •NADP ⁺	H ₂ O	3.5 ± 1.8	3.6 ± 1.1	3.9 ± 1.0
				4.0 ± 1.0
WT•dGMP •NADP ⁺	H ₂ O	3.6 ± 1.8	3.1 ± 1.0	3.3 ± 1.0
				3.2 ± 1.0
D219A•IMP •NADP ⁺	H ₂ O	2.6 ± 0.9	2.8 ± 0.2	3.3 ± 0.2
				3.3 ± 0.3
D219A•GMP •NADP ⁺	H ₂ O	2.9 ± 1.2	3.2 ± 0.5	3.8 ± 0.6
				3.7 ± 0.7



Supplementary Figure S1. The partial reaction of IMP and NADP⁺ catalyzed by GMPR WT. Enzyme (10 μ M) was mixed with IMP (500 μ M) and NADP⁺ (500 μ M) and the formation of NADPH is monitored by the change in absorbance at $\lambda = 340$ nm. The amount of NADPH formed is ~16% of the active sites. **A.** A representative progress curve comprised of the average absorbance readings from at least 5 “shots”. The red line shows the fit to a single exponential equation. **B.** The residual errors on the fit in A.



Supplementary Figure S2. The partial reaction of dIMP and NADP⁺ catalyzed by GMPR WT. Enzyme (10 μM) was mixed with dIMP (500 μM) and NADP⁺ (500 μM) and the formation of NADPH is monitored by the change in absorbance at $\lambda = 340 \text{ nm}$. The amount of NADPH formed is $\sim 10\%$ of the active sites. **A.** A representative progress curve comprised of the average absorbance readings from at least 5 “shots”. The red line shows the fit to a single exponential equation. **B.** The residual errors on the fit in A.



Supplementary Figure S3. The partial reaction of IMP and NADP⁺ catalyzed by GMPR mutant D219A. **A.** A representative progress curve of the presteady state reaction comprised of the average absorbance readings from at least 5 “shots”. The red line shows the fit to a single exponential and a steady state. Enzyme (30 μ M) was mixed with IMP (10 mM) and NADP⁺ (3 mM) and the formation of NADPH is monitored by the change in absorbance at $\lambda =$

340 nm. The amount of NADPH formed is ~3% of the active sites after the burst phase. The apparent steady state rate is 0.001 s^{-1} . **B.** The residual errors on the fit in A. **C.** The partial reaction of IMP and NADP^+ catalyzed by D219A monitored for 24 hours. Formation of NADPH is monitored by the change in absorbance at $\lambda = 340 \text{ nm}$. Initial rates were determined over the first 10 min. *Black line*: conditions of the pre-steady state experiment: enzyme ($30 \text{ }\mu\text{M}$) was mixed with saturating IMP (10 mM) and NADP^+ (3 mM), initial rate = $6.3 \times 10^{-5} \text{ s}^{-1}$; *Green line*: conditions of the field cycling experiment: enzyme ($400 \text{ }\mu\text{M}$) was mixed with IMP (1.6 mM) and NADP^+ (1.6 mM), initial rate = $1.4 \times 10^{-5} \text{ s}^{-1}$; *Blue line*: enzyme ($400 \text{ }\mu\text{M}$) was mixed with saturating IMP (10 mM) and NADP^+ (3 mM), initial rate = $3.5 \times 10^{-6} \text{ s}^{-1}$. These observations suggest that the enzyme is 41% saturated with IMP and NADP^+ under the conditions of the field cycling experiment.