

Supplementary Materials

Ethanol extract of *Centipeda minima* exerts antioxidant and neuroprotective effects via activation of Nrf2 signaling

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Supplementary Materials and Methods

Hematoxylin-eosin (H&E) staining

At the end of animal experiments, mice were anesthetized and perfused with ice-cold PBS and 4% paraformaldehyde. In brief, sagittal sections at 10 μm were prepared by using a Leica ultra-thin semiautomatic microtome as previously described after tissue dehydration. Brain paraffin sections were washed in xylene and rehydrated through a graded series of ethanol and double-distilled water. Then the sections were dipped in hematoxylin staining (Beyotime, Shanghai, China) for 10 min and eosin staining (Beyotime, Shanghai, China) for 1 min at room temperature. Then slides were rinsed in double-distilled water and dehydrated through 80%, 90% and 100% alcohol, cleared in xylene. Images were analyzed by using a light microscope and LEICA QWin Plus (Leica Microsystems, Wetzlar, Germany).

Extraction and isolation of EBSC-26A–EBSC-26D

The air-dried and powdered *Centipeda minima* whole plants (60 kg) were extracted by refluxing with 70% EtOH ($3 \times 120 \text{ L} \times 2 \text{ h}$) to give a crude extract, which was suspended in water followed by extraction with EtOAc to afford an EtOAc soluble extract. The EtOAc extract was suspended in petroleum ether/MeOH (1:1) to afford the MeOH extract. The MeOH partition was applied to a silica gel (200-300 mesh) column, eluted with petroleum ether-EtOAc (10:1-0:1 gradient system), to give fractions A to F. Fraction B was divided into eight portions (B1–B8) by MCI gel CHP 20P eluted with gradient aqueous MeOH

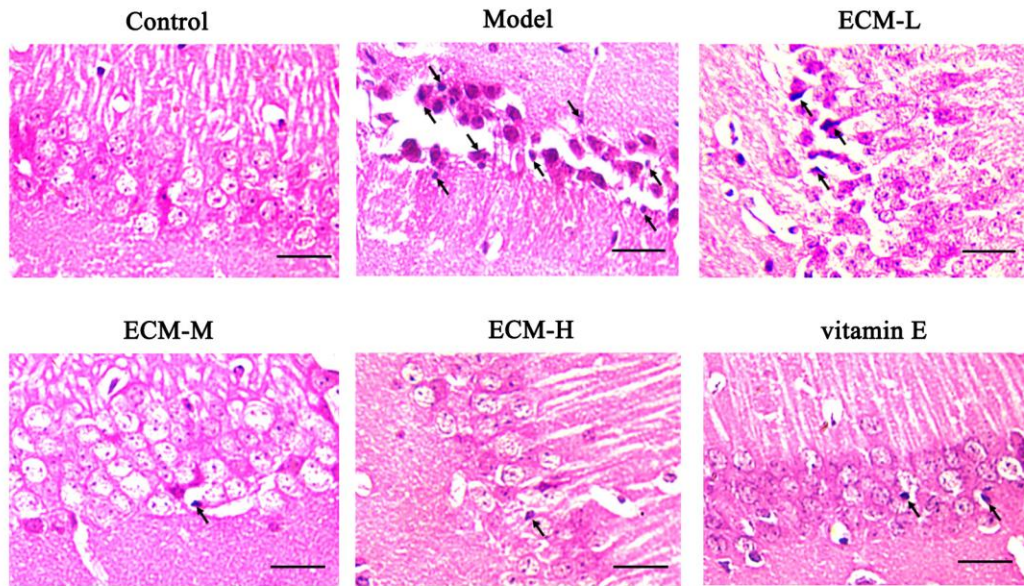
(30%–70%). Fraction B6 was subjected to a RP-18 column (MeOH/H₂O, 40%–70%) to provide six portions (B6.1–B6.6) monitored by TLC using previously purified EBSC-26A as a reference compound. Fraction B6.3 was recrystallized to afford EBSC-26A (30.0 g, content 2.5%). Fraction B6.4 was further purified using preparative HPLC (MeOH/H₂O, 65%) to yield EBSC-26A (3.0 g), EBSC-26B (2.0 g, content 0.7%), EBSC-26C (0.8 g), and EBSC-26D (0.6 g).

EBSC-26A, white solids, ESIMS: m/z 369 [M + Na]⁺; ¹H NMR (600 MHz, CDCl₃), see Supplementary Figure 2; ¹³C NMR (150 MHz, CDCl₃), see Supplementary Figure 3.

EBSC-26B, white solids, ESIMS: m/z 355 [M + Na]⁺; ¹H NMR (600 MHz, CDCl₃), see Supplementary Figure 4; ¹³C NMR (150 MHz, CDCl₃), see Supplementary Figure 5.

EBSC-26C, white solids, ESIMS: m/z 357 [M + Na]⁺; ¹H NMR (600 MHz, CDCl₃), see Supplementary Figure 6; ¹³C NMR (150 MHz, CDCl₃), see Supplementary Figure 7.

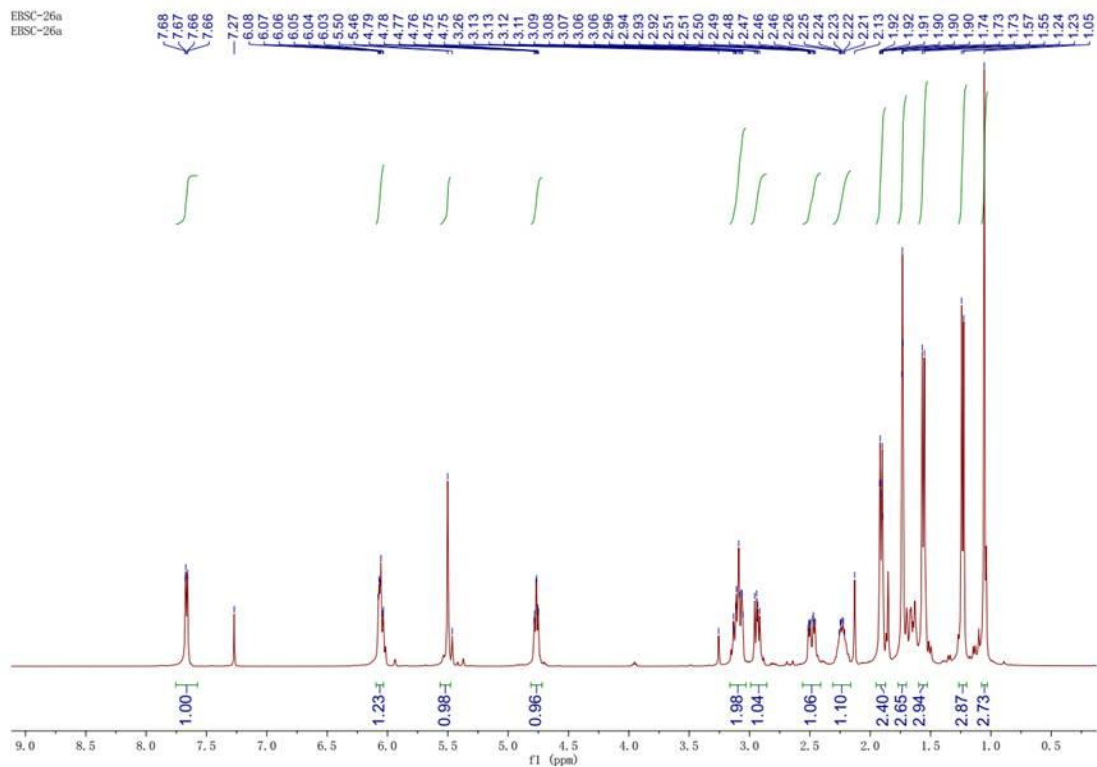
EBSC-26D, white solids, ESIMS: m/z 369 [M + Na]⁺; ¹H NMR (600 MHz, CDCl₃), see Supplementary Figure 8; ¹³C NMR (150 MHz, CDCl₃), see Supplementary Figure 9.



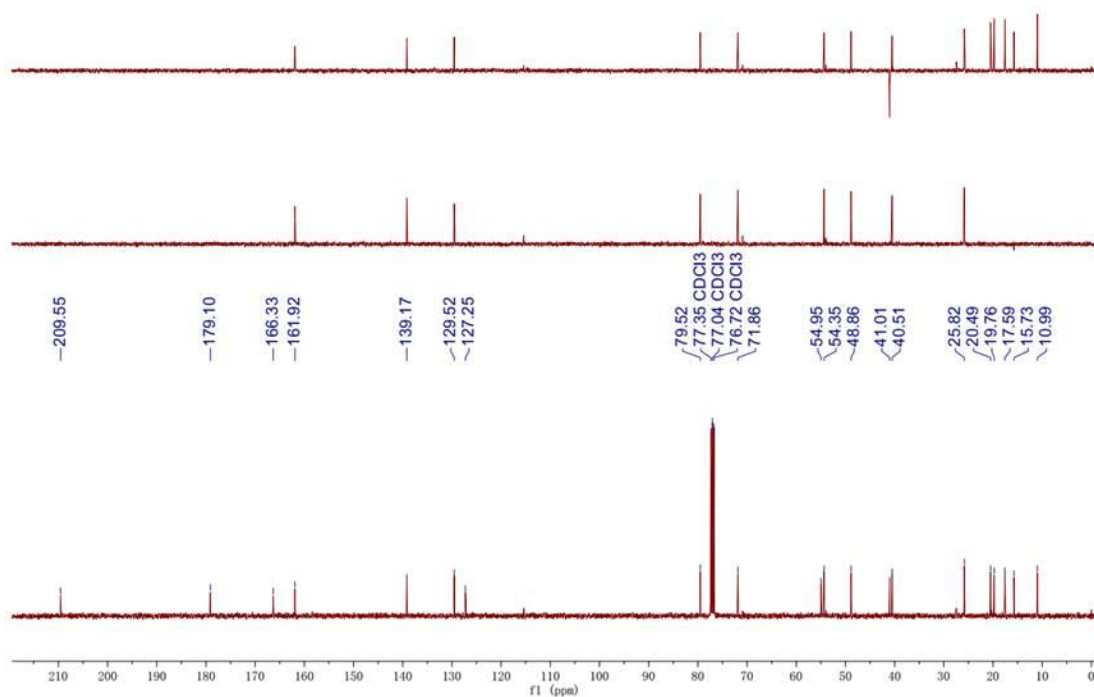
Supplementary Figure 1. ECM exerts neuroprotective effects in the hippocampus.

H&E staining show morphological changes of neurons in the hippocampus.

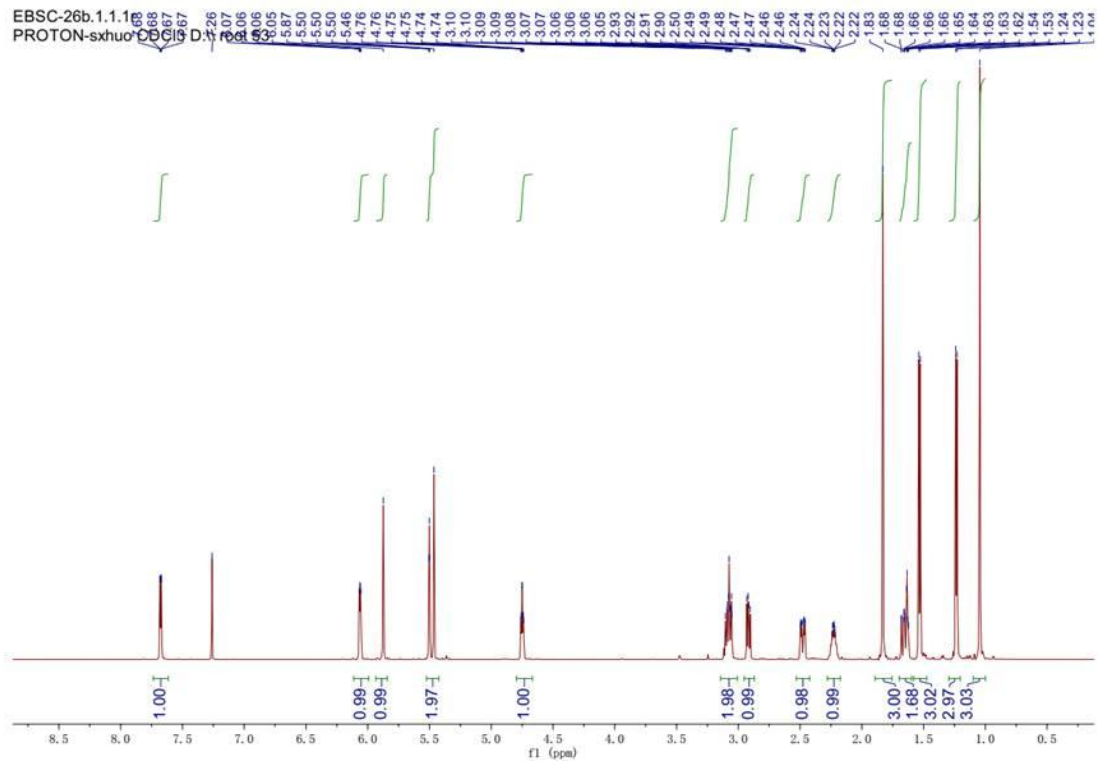
Scale bar, 25 μ m. Control: vehicle control; ECM-L: D-gal/ AlCl_3 + ECM (100 mg/kg/d); ECM-M: D-gal/ AlCl_3 + ECM (200 mg/kg/d); ECM-H: D-gal/ AlCl_3 + ECM (400 mg/kg/d); vitamin E: D-gal/ AlCl_3 + vitamin E (80 mg/kg/d).



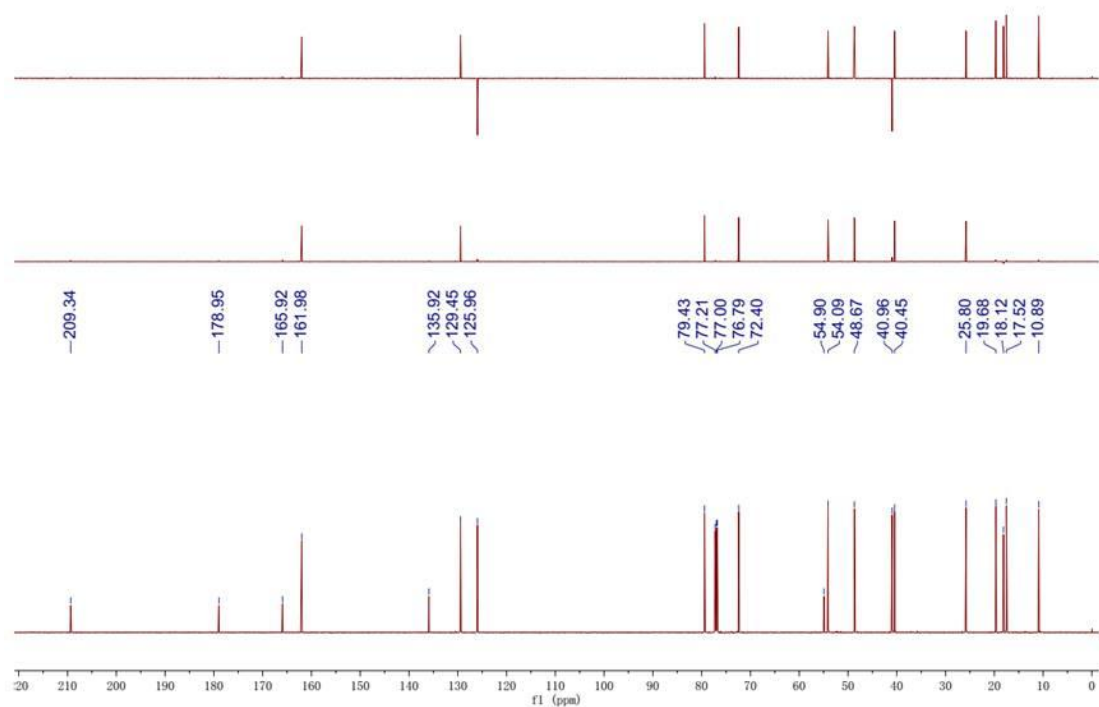
Supplementary Figure 2. ^1H NMR spectrum of EBSC-26A in CDCl_3 .



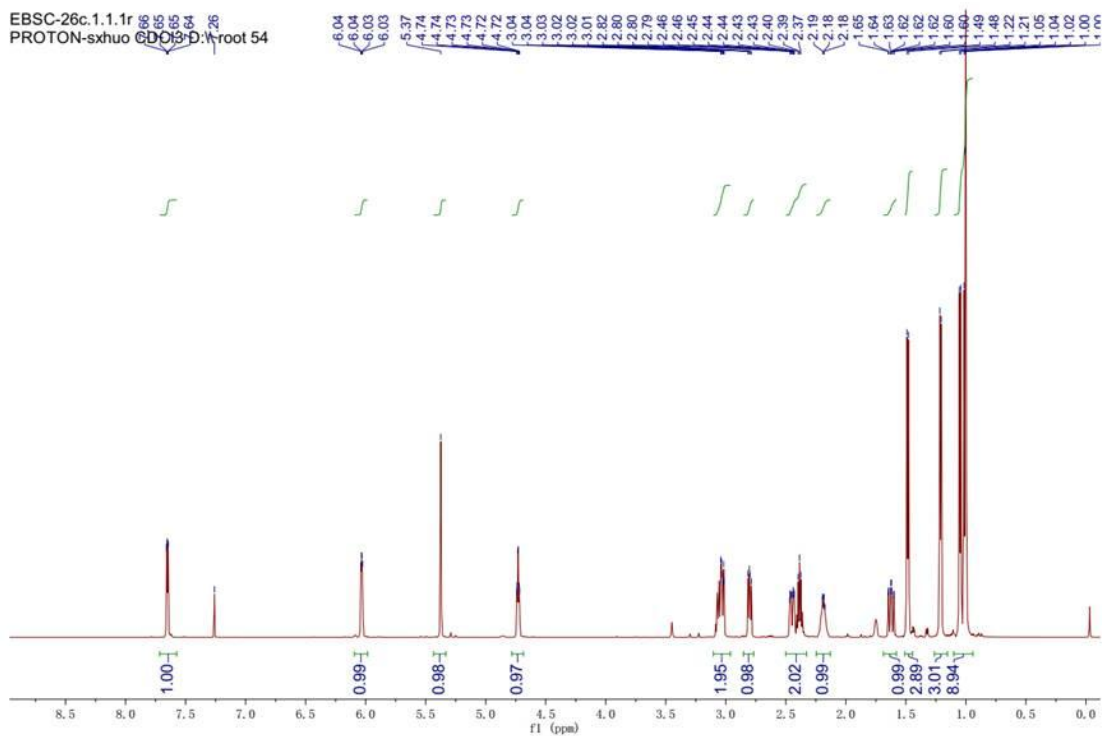
Supplementary Figure 3. ^{13}C NMR and DEPT spectra of EBSC-26A in CDCl_3 .



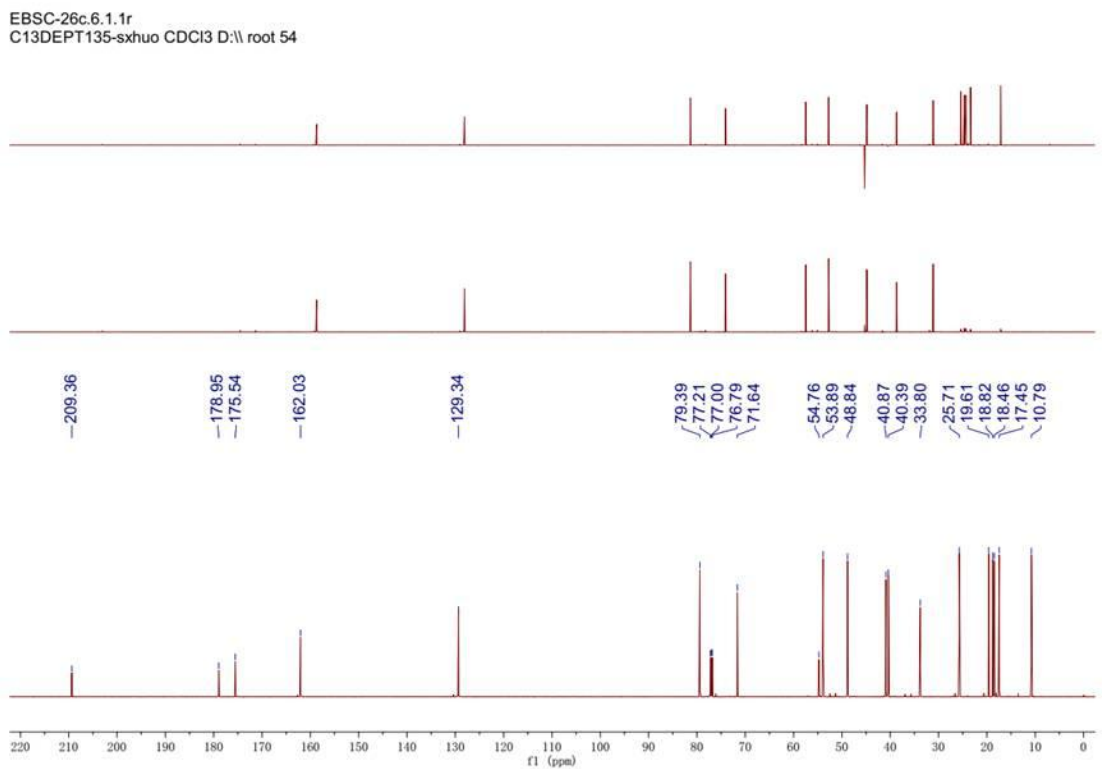
Supplementary Figure 4. ^1H NMR spectrum of **EBSC-26B** in CDCl_3 .



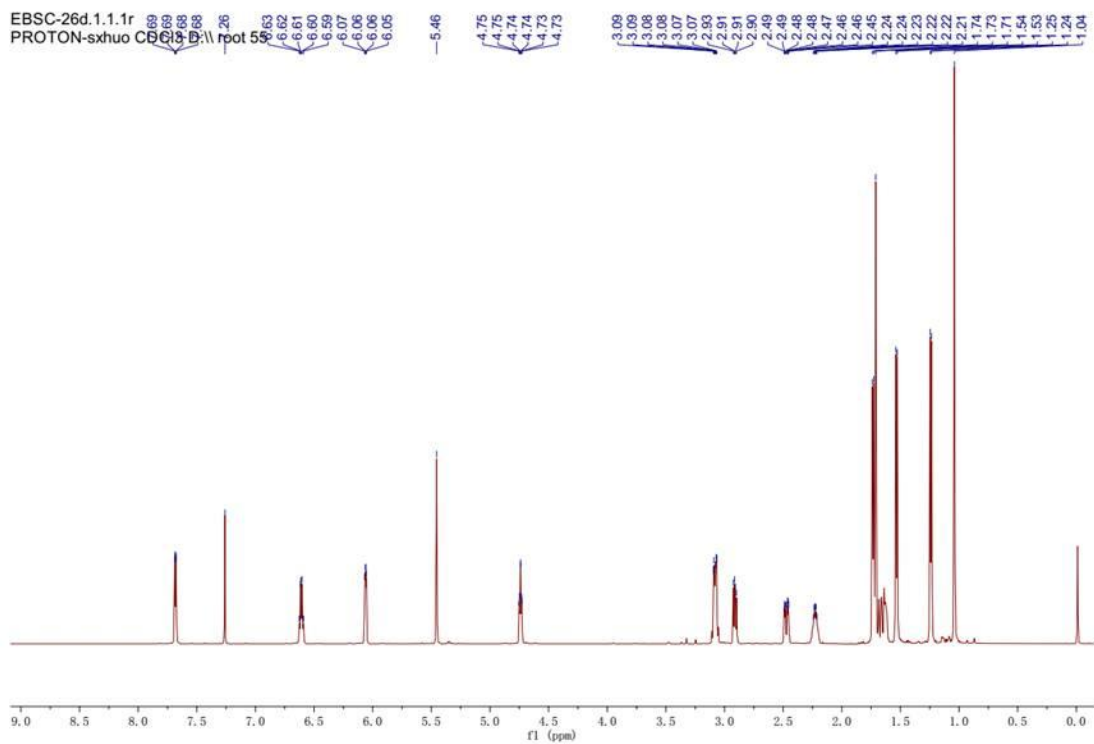
Supplementary Figure 5. ^{13}C NMR and DEPT spectra of **EBSC-26B** in CDCl_3 .



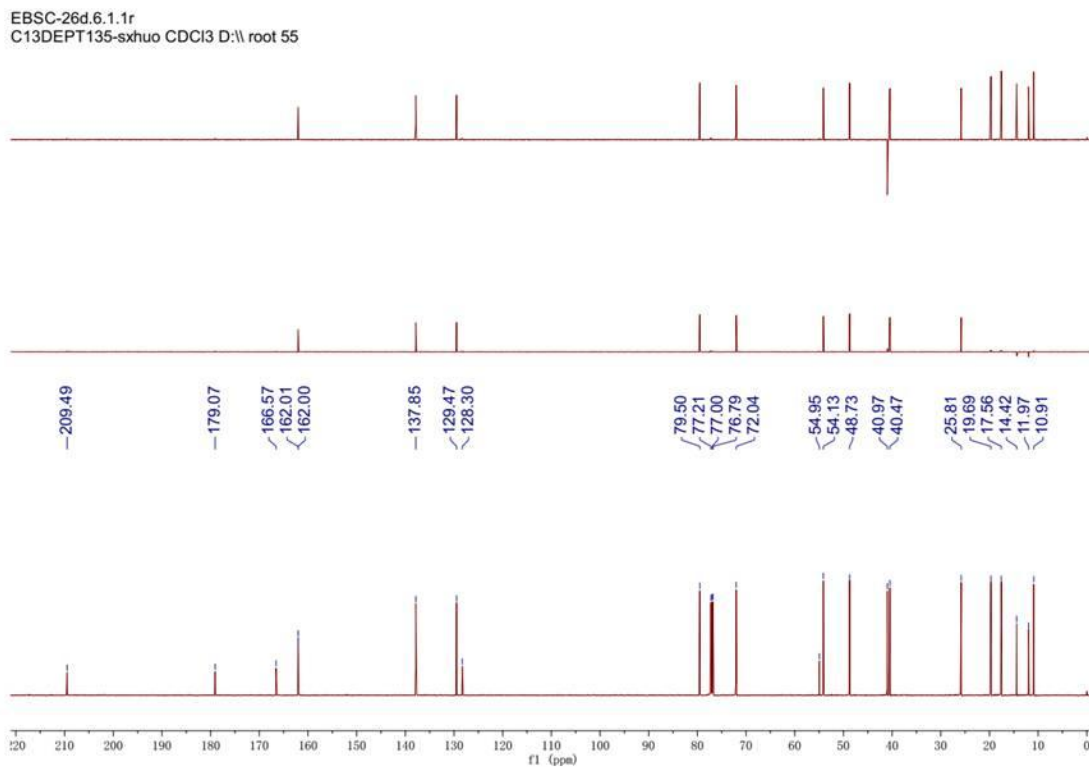
Supplementary Figure 6. ^1H NMR spectrum of EBSC-26C in CDCl_3 .



Supplementary Figure 7. ^{13}C NMR and DEPT spectra of EBSC-26C in CDCl_3 .



Supplementary Figure 8. ^1H NMR spectrum of **EBSC-26D** in CDCl_3 .



Supplementary Figure 9. ^{13}C NMR and DEPT spectra of **EBSC-26D** in CDCl_3 .