#### Figure S1. Models of early axonogenesis in vivo and in vitro. Related to Figure 1

(A) Schematic of *in vivo* generation of cortical excitatory neurons. Modified from Sakakibara and Hatanaka 2015. MZ: marginal zone. CP: cortical plate. IZ: intermediate zone. SVZ: subventricular zone. VZ: ventricular zone. (B) Representative images of immature pre-axonogenesis neurons (DIV 1) and axon-generating neurons (DIV 3) in cultures.

#### Figure S2. RNA-Seq analysis of DIV 1 and DIV 3 neurons. Related to Figure 1 and Figure 2

Two biological replicates of immature pre-axonongeneis neurons (DIV1\_1 and DIV1\_2) and two biological replicates of axon-generating neurons (DIV3\_1 and DIV3\_2) are cross compared. X and Y axes show the value of log<sub>2</sub>(TPM+1). The red lines depict fitted regression. R<sup>2</sup> = 0.995 (A), 0.994 (B), 0.978 (C), 0.976 (D), 0.977 (E), and 0.978 (F). (G) Axonogenesis-associated gene lists identified via transcriptome comparison are assessed against gene ontology terms enriched in previously published axonogenesis-controlling genes. A scatter plot shows the Bonferroni-corrected P values of enriched Gene Ontology terms for axonogenesis-associated gene groups on the y axis vs previously published axonogenesis-controlling genes on the x axis. Dashed lines demark P value equal to 0.05 on the y axis. Group 1 (grey dots): 252 genes regulated at the gene and isoform level. Group 2 (red dots): genes regulated exclusively at the isoform level. Amino acids encoded by axonogenesis-associated exons in (H) CLIP2, (I) CAMSAP1, (J) ABI2, and (K) PPP3CA. For CAMSAP1, inclusion of the exon disrupts the domain. For others, inclusion of the exon is required for the domain.

#### Figure S3. SHTN1L promotes actin polymerization. Related to Figure 4

(A) A representative Coomassie blue-stained gel image demonstrating the purity of FLAG-tagged target proteins. The protein concentrations were obtained by densitometric analysis compared with 1:2 serial dilutions of recombinant BSA. (B-C) Cosedimentation experiments were performed using 0.15 mg/ml actin alone or with 1:2 serial dilutions of purified FLAG-SHTN1L and FLAG-SHTN1L<sup>RRR>GGG</sup> proteins starting from 286 nM for both. Mutation of three conserved arginine residues to glycine (R482G, R483G, and R484G; SHTN1L<sup>RRR>GGG</sup> mutant) disrupted F-actin binding.

#### Figure S4. PTBP2 regulate axonogenesis-associated exons. Related to Figure 5

(A) Enriched motif identified for down-regulated exons during early axonogenesis. (B) Axonogenesis-associated exons, control non-regulated exons, and constitutive exons are examined for PTBP2 CLIP-Seq signals at each nucleotide position. The percent of exons with PTBP2 CLIP signals are plotted. (C) PTBP2-regulated exons significantly overlap with axongenesis-associated exons. P value =  $4.4 \times 10^{-78}$ , hypergeometric test. (D) A 2x2 table shows the distribution of the overlapping exons regarding their responses to PTBP2 deletion and during early axonogenesis. (E) PTBP2-regulated exons determined by PTBP2 overexpression significantly overlap with axongenesis-associated exons. P value =  $3.23 \times 10^{-82}$ , hypergeometric test. (F) Splicing changes (increase or decrease) from pre-axonogenesis neurons (DIV 1) to axon-generating neurons (DIV 3) are mostly decelerated by gain of PTBP2. (G) PSI values of PTBP2-regulated exons genes at DIV 1, DIV3, and DIV 5 are determined by quantitative capillary electrophoresis. Data are represented as mean  $\pm$  SEM of 3-4 animals. \*\*, P-value < 0.01; \*, P-value < 0.05; Student's t test.

#### Figure S5. Ptbp2 expression in various tissues profiled in the ENCODE project. Related to Figure 5

TPM expression values of *Ptbp2* transcripts in the developing forebrain, hindbrain, and midbrain at E10.5, E11.5, E12.5, E13.5, E14.5, E15.5, E16.5, PND0, and in adult cerebellum, cortical plate, frontal cortex, colon, duodenum, gonadal fat pad, heart, kidney, intestine, liver, lung, mammary gland, small intestine, stomach, subcutaneous adipose tissue, and thymus. Each sample has two replicates.

#### Figure S6. PTBP2 direct targets are confirmed by PTBP2 CLIP-Seq. Related to Figure 5

(A-J) Browser tracks of PTBP2 CLIP-Seq in embryonic day 18 brains overlay the gene structure of PTBP2 targets. Red boxes indicate exons mis-regulated in  $Ptbp2^{-/-}$  cortical neurons.

#### Figure S7. Mini Gene analysis of PTBP2-controlled *Clip2* Exon 9 and *Plekha5* Exon 10. Related to Figure 5

(A, E) Diagram of mini gene structures for *Clip2*-E9-pFlareA and *Plekha5*-E10-pFlareA. Blue box indicates the interested exon; bold blue horizontal lines flanking the blue box represent the flanking targeted introns; vertical lines within introns indicate the deletions of PTBP2 binding motifs in mutant minigenes for *Clip2* Exon 9 and *Plekha5* Exon 10. Exon inclusion results in red fluorescence and exon skipping leads to green fluorescence in transfected neurons. (B-D) Expression of WT or mutant *Clip2* Exon 9 mini genes when transfected in primary cortical neurons. (F-H) Expression of WT or mutant *Plekha5* Exon 10 mini genes when transfected in primary cortical neurons. Neurons were transfected at DIV 0 (after plating), fixed and imaged at DIV 2. Images were acquired under the same settings across experiments. Scale bar: 50 µm, applied to all images.

#### Figure S8. PTBP2 expression rescues axon growth of *Ptbp2<sup>-/-</sup>* neurons. Related to Figure 6

(A) Immunostaining images of WT and  $Ptbp2^{-/-}$  (KO) neurons infected with GFP-expressing lentivirus (GFP LV) or GFP- and PTBP2-coexpressing lentivirus (PTBP2 LV). GFP indicates the infected neurons. (B) PTBP2 expression rescue the shortened axon phenotype in  $Ptbp2^{-/-}$  neurons. The longest protrusion of each neuron was measured by GFP. N = 82 (WT + GFP LV), 82 (KO + GFP LV), and 131 (KO + PTBP2 LV). Neurons were infected at DIV0 and fixed at DIV3. Data are represented as mean  $\pm$  SD of all neurons from 2 biological replicates (litters). \*p<0.05, \*\*p<0.01, Student's t-test.

#### Figure S9. Axonal defects in *Ptbp2<sup>-/-</sup>* cortices. Related to Figure 7

(A) Representative images of single neurons from E17.5 WT and  $Ptbp2^{-/-}$  (KO) neocortices after *in utero* electroporation (IUE) with CAG-GFP plasmid at E13.5. Arrows point to primary trailing processes (axons) of the migrating neurons. Scale bar: 20 µm. (B)  $Ptbp2^{-/-}$  cortical neurons contain a higher percentage of surplus axons than WT neurons. P value = 0.0078, Fisher's exact test. Error bars represent 95% confidence intervals. N = 86 (WT) and 103 (KO) from 2 litters for each genotype. Only clearly traceable neurons were included and primary trailing processes longer than 10 µm were considered. (C-D) Representative images of the reconstructed neurons show normal neuronal migration and dendritic orientation with defective axon morphology in  $Ptbp2^{-/-}$ . CP: cortical plate. IZ: intermediate zone.

# Figure S10. Normal neurogenesis, cortical lamination, and neuronal numbers in *Ptbp2<sup>-/-</sup>* cortices. Related to Figure 7

(A-D) Cortical lamination of (A, C) WT control and (B, D) Emx1-Ptbp2cKO brains shown by layer markers (A, B) SATB2, TBR1, (C, D) CUX1, and CTIP2 at E17.5. A double arrow marks the gap between the cortical plate and the ventricle zone in each case. Scale bars are 50 $\mu$ m. (E) Quantification shows cortical lamination of Emx1-Ptbp2cKO brains is normal. 25-30 slices from 2 brains of each genotype were analyzed. Data are represented as the mean  $\pm$  SEM from all slices. n.s, p>0.05, Student's t-test.

Figure S1













Figure S2







### Figure S4





Figure S6



## Figure S7





Figure S9



## Figure S10

