

Supplementary Information for

Secondary Amines as Coupling Partners in Direct Catalytic Asymmetric Reductive Amination

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I. General remarks

All reactions were performed in the nitrogen-filled glovebox or under nitrogen using standard Schlenk techniques unless otherwise noted. Column chromatography was performed using silica gel 60 (200 – 300 mesh). ¹H NMR, ¹³C NMR and ³¹P NMR spectral data were obtained from Bruker 500 MHz spectrometers. Chemical shifts are reported in ppm. Enantiomeric excess values were determined by chiral HPLC on an Agilent 1220 Series instrument or using ¹H NMR with chemical shift reagent to determine the enantiomeric excesses.. All new products were further characterized by HRMS. A positive ion mass spectrum of sample was acquired on a Thermo Scientific LTQ Orbitrap XL mass spectrometer or AB SCIEX LC-30A+SelexION+TripleTOF5600+ with an electrospray ionization source.

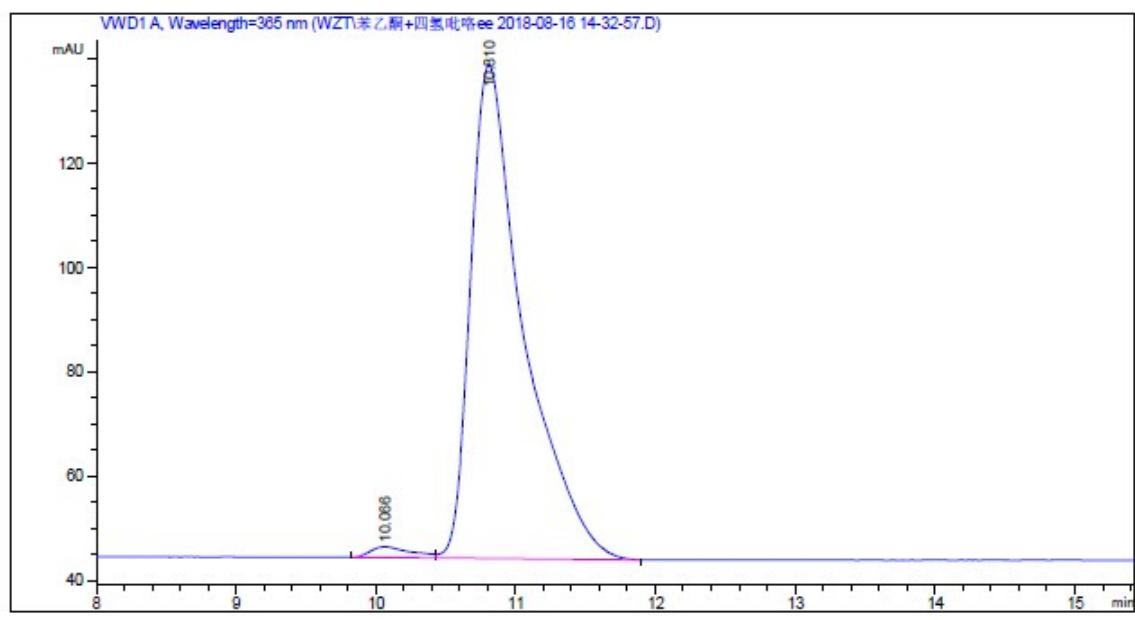
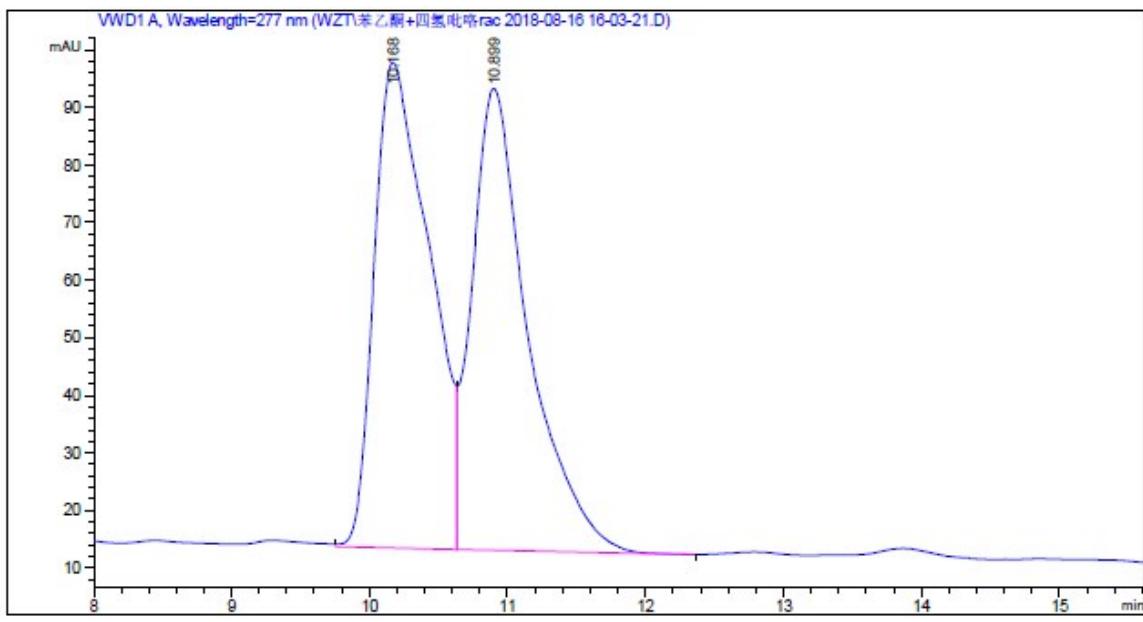
II. General Procedure for Asymmetric Reductive Amination

Chiral ligands **L5–L9** were prepared according to literatures.¹

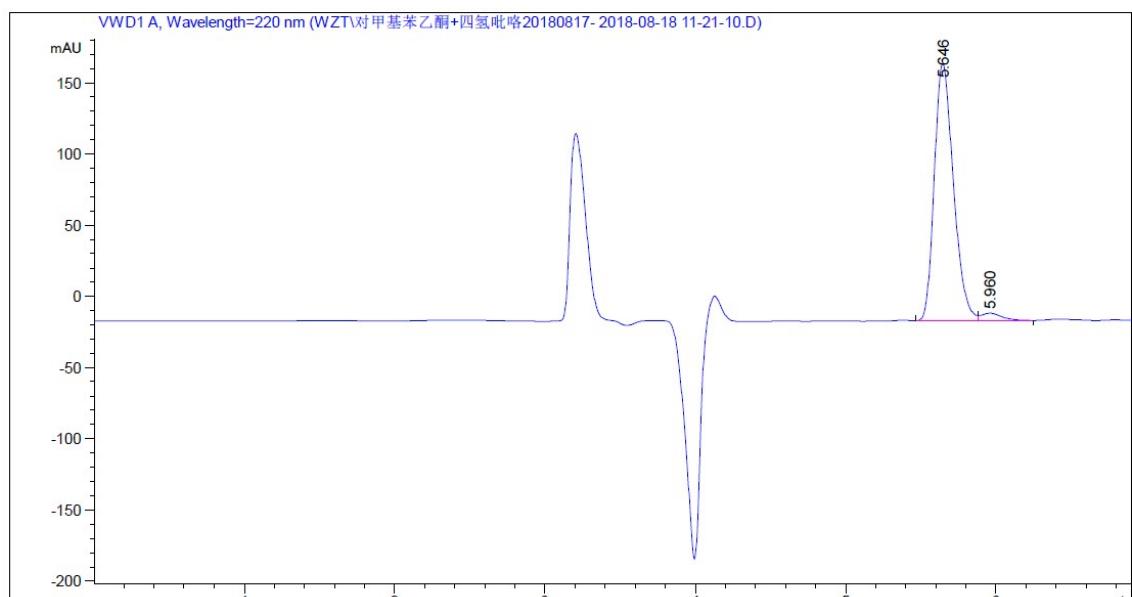
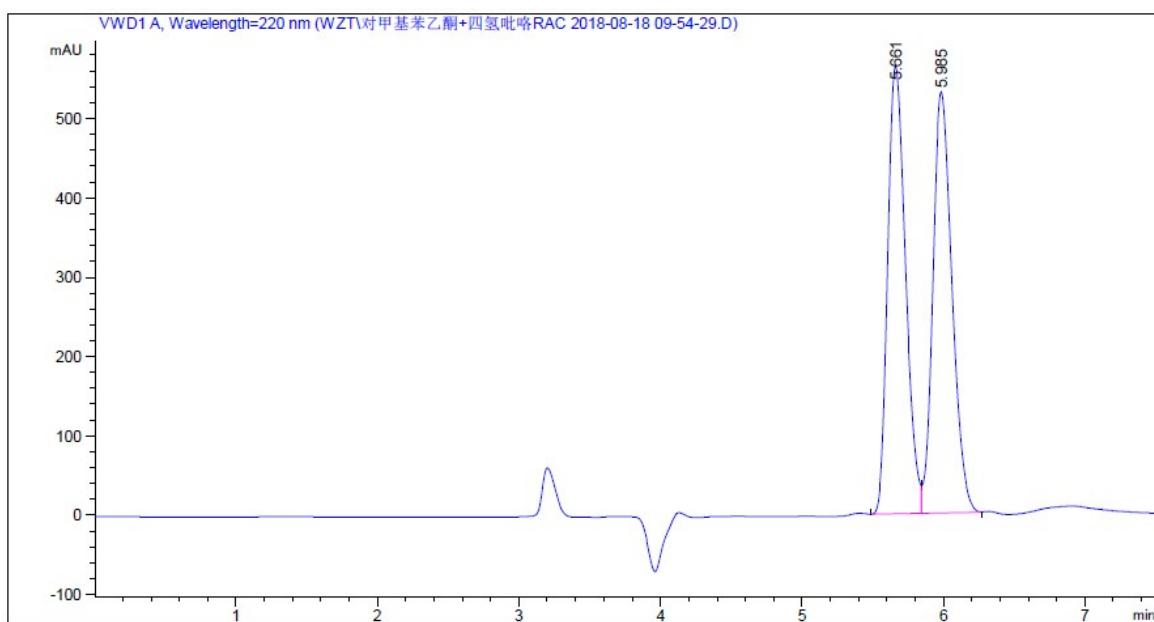
In a nitrogen-filled glovebox, [Ir(COD)Cl]₂ (10 µmol) and **L8a** (46 µmol) was dissolved in anhydrous CH₂Cl₂ (1.0 mL), stirred for 20 min, and equally divided into 10 vials charged with ketones (0.2 mmol) and pyrrolidine (0.22 mmol) in anhydrous CH₂Cl₂ solution (0.5 mL). Then Ti(O*i*-Pr)₄ (1.2 equiv.), 1,4-diazabicyclo[2.2.2]octane (DABCO) (0.1 eq), and I₂ (1 mol%) were added and the total solution was made to 2.0 mL (DCM/THF/DCE = 1:1:1.5) for each vial. The resulting vials were transferred to an autoclave, which was charged with 40 atm of H₂, and stirred at 4 °C for 20 h. The hydrogen gas was released slowly and the solution was quenched with aqueous sodium bicarbonate solution. The organic phase was concentrated and passed through a short column of silica gel to remove the metal complex to give the crude products, which were purified by column chromatography and then analyzed by chiral HPLC or using ¹H NMR with chemical shift reagent to determine the enantiomeric excesses.

¹H NMR method for determination the e.e. values: The amine product **3** was mixed with equal amount (mol/mol) of (*S* or *R*)-2-acetoxy-2-phenylacetic acid and dissolved in CDCl₃. Diastereoisomers are formed and the proton signal of amine β-methyl will be splitted. From integration ratio the e.e. value could be calculated.

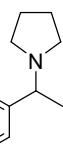
(S)-1-(1-phenylethyl)pyrrolidine (3a):² 91% yield, 97% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.24–7.42 (m, 5H, Ar-H), 3.20 (q, *J* = 6.3 Hz, 1H, CH), 2.60 (m, 2H, N-CH₂), 2.40 (m, 2H, N-CH₂), 1.81 (m, 4H, 2*CH₂), 1.46 (d, *J* = 6.4 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 145.3 (Ar-CC), 128.3, 127.3, 127.0, 66.1, 53.0, 23.4, 23.1. Enantiomeric excess was determined by chiral HPLC: Chiralpak OJ-3 column, Hex/IPA=97:3, 0.6 mL/min, 220 nm, 7.08 min, 7.53 min.

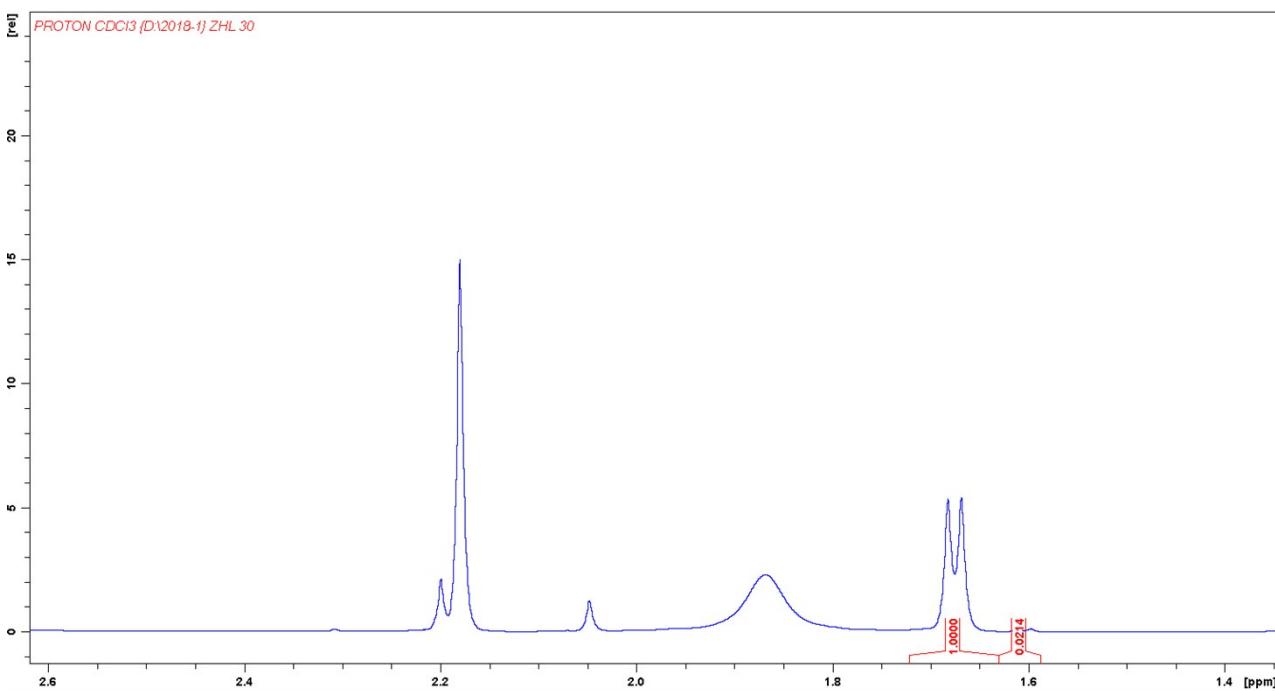
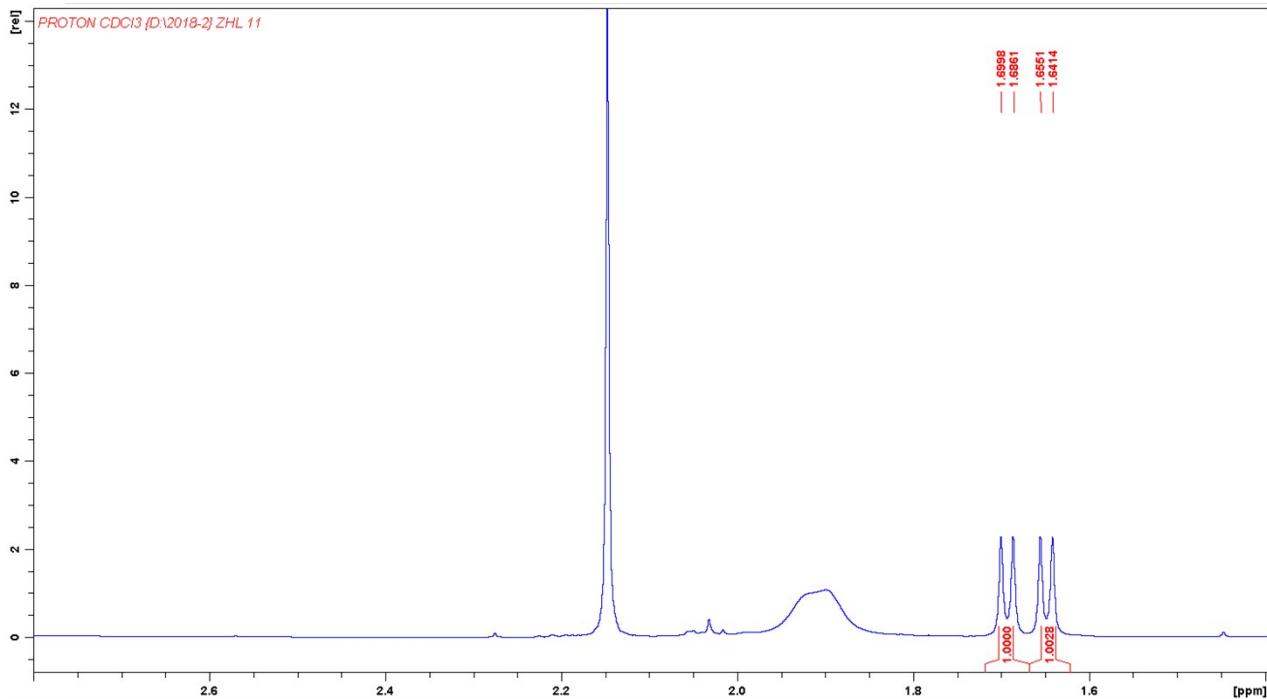


1-(1-(*p*-tolyl)ethyl)pyrrolidine (3b**):**³ 88% yield, 94% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.28 (d, *J* = 8.0 Hz, 2H, Ar-CH), 7.16 (d, *J* = 7.9 Hz, 2H, Ar-CH), 3.21 (q, *J* = 6.6 Hz, 1H, CH), 2.60 (m, 2H, CH₂), 2.42 (m, 2H, N-CH₂), 2.38 (s, 3H, N-CH₃), 1.80 (m, 4H, 2* CH₂), 1.44 (d, *J* = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 141.1, 136.9, 129.1, 127.3, 65.8, 52.9, 23.4, 22.7, 21.1. Enantiomeric excess was determined by chiral HPLC: Chiralpak OJ-H column, Hex/IPA=99:1, 1 mL/min, 220nm, 4.48 min, 5.67 min.

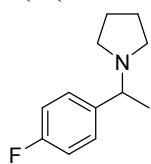


1-(1-(4-methoxyphenyl)ethyl)pyrrolidine (3c**):**² 93% yield, 95% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.28 (d,

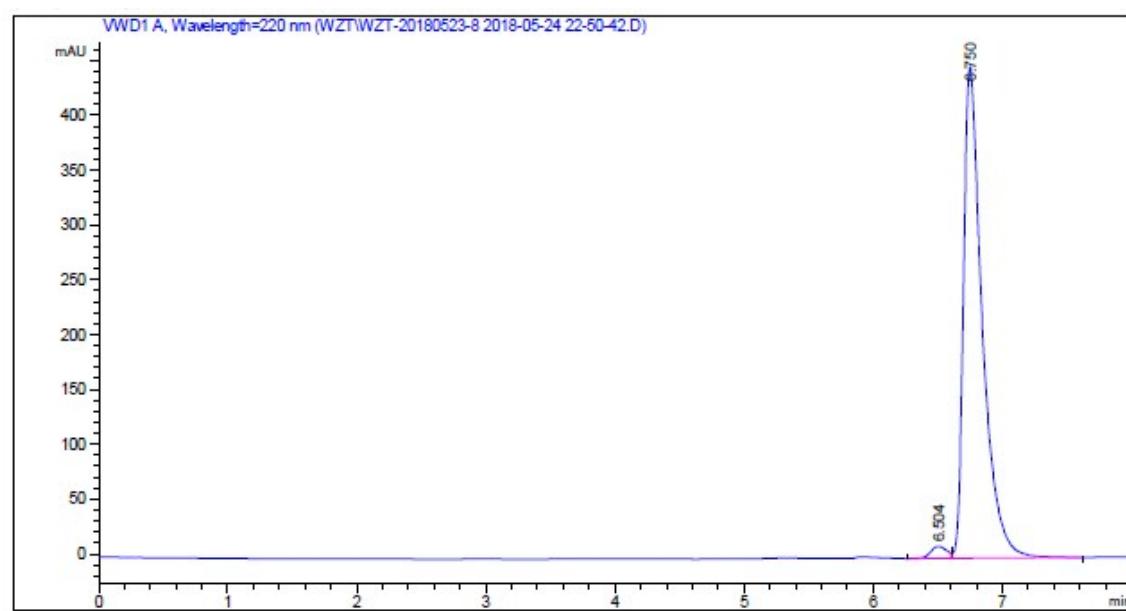
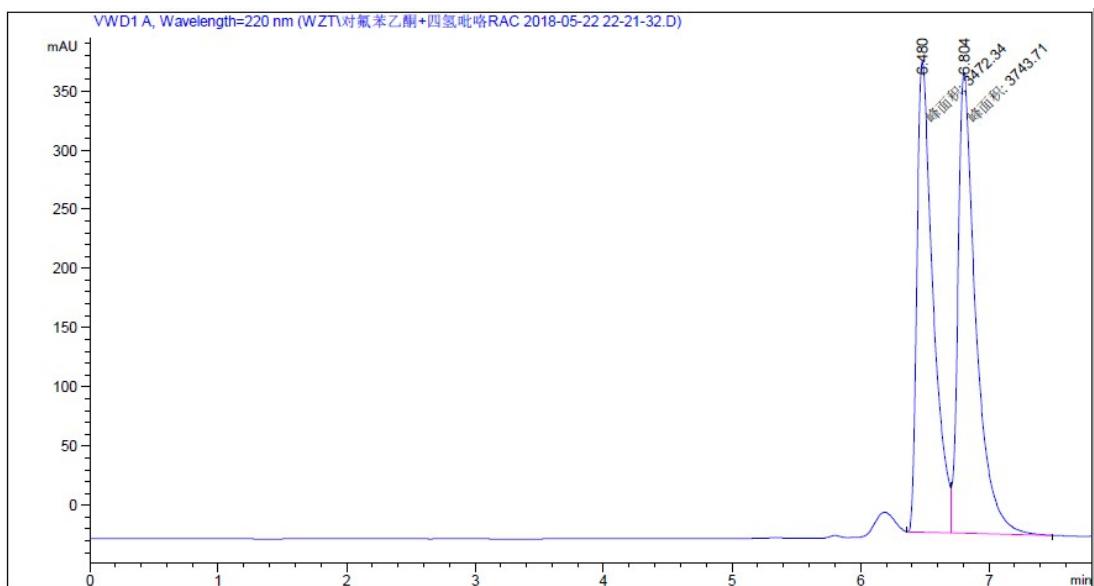

 $J = 8.7$ Hz, 2H, Ar-CH), 6.88 (d, $J = 8.7$ Hz, 2H, Ar-CH), 3.84 (s, 3H, O-CH₃), 3.18 (q, $J = 7.6$ Hz, 1H, CH), 2.58 (m, 2H, N-CH₂), 2.39 (m, 2H, N-CH₂), 1.79 (m, 4H, 2*CH₂), 1.42 (d, $J = 7.6$ Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 159.2 (Ar-CO), 132.1 (Ar-CC), 128.7, 113.8, 65.2, 55.0, 52.3, 23.0; 20.9. Enantiomeric excess was determined by ¹H NMR using (S)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



1-(1-(4-fluorophenyl)ethyl)pyrrolidine (3d):³ 90% yield, 96% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.33 (m, 2H, Ar-CH), 7.02 (m, 2H, Ar-CH), 3.21 (q, *J* = 6.6 Hz, 1H, CH), 2.57 (m, 2H, N-CH₂), 2.39 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.41 (d, *J* = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 162.9 (Ar-CF), 160.9 (Ar-CC), 128.8, 128.7, 115.2, 115.0, 65.3, 52.9, 23.4, 23.0. Enantiomeric excess was determined by chiral HPLC:

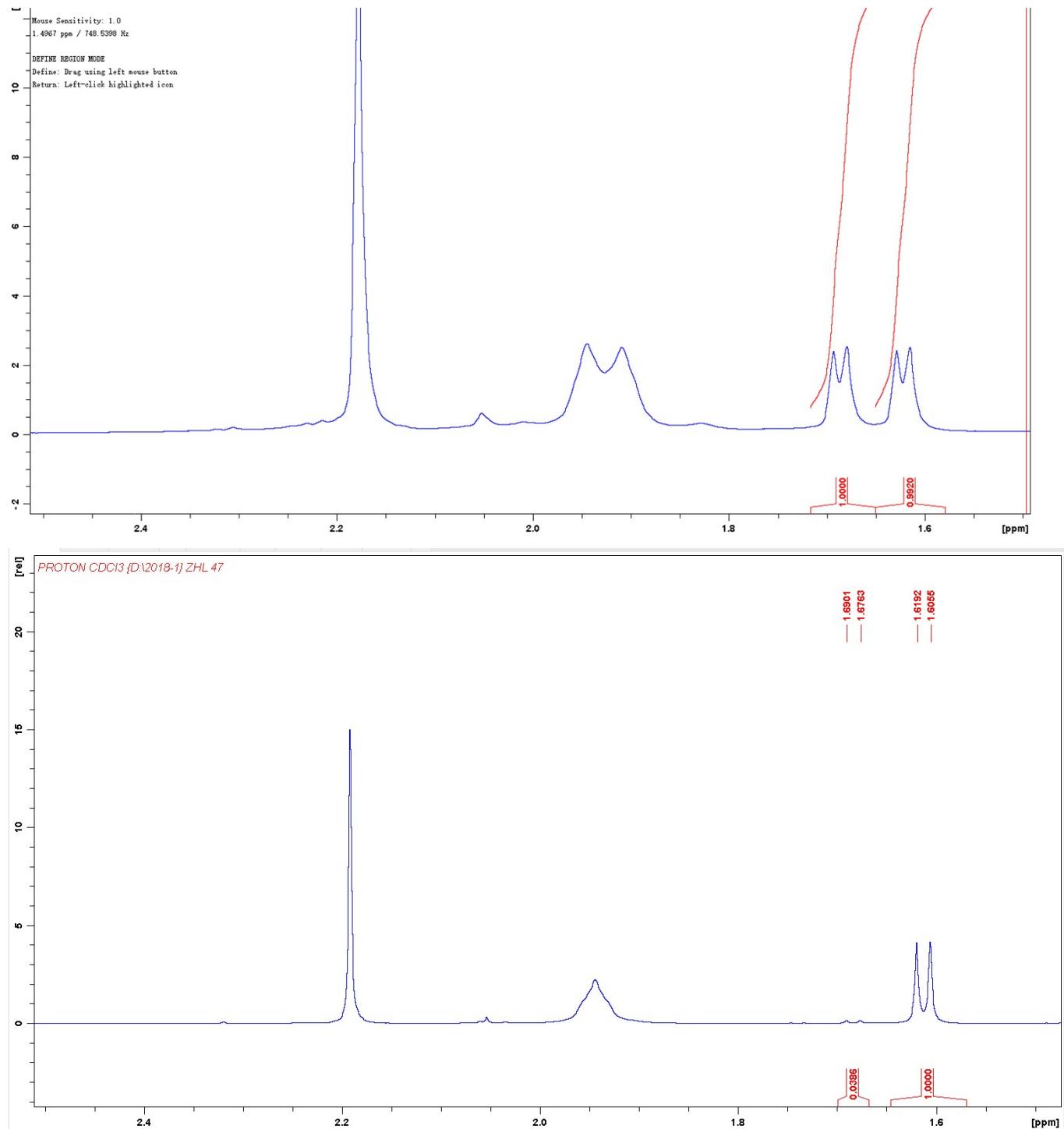


Chiraldak OJ-3 column, Hex/IPA=98:2, 0.6 mL/min, 220 nm, 6.48 min, 6.80 min.

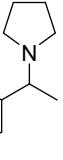


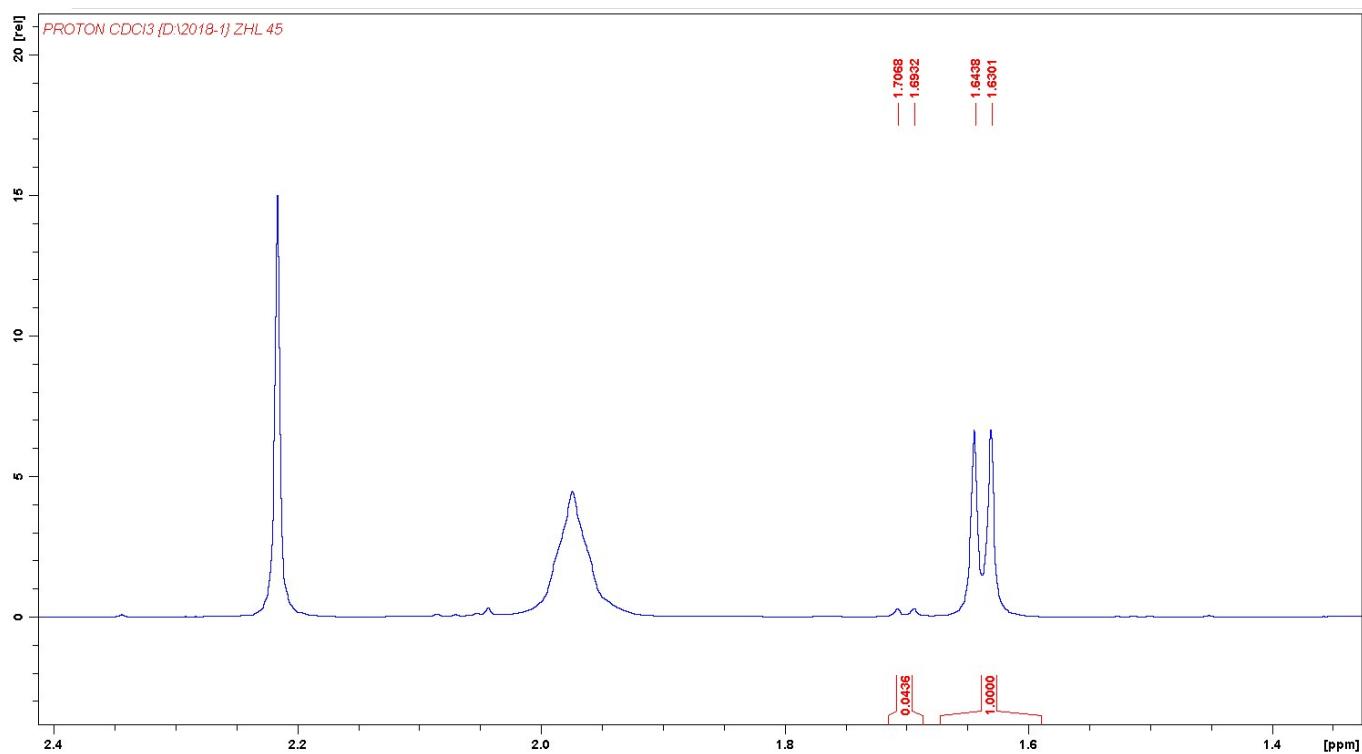
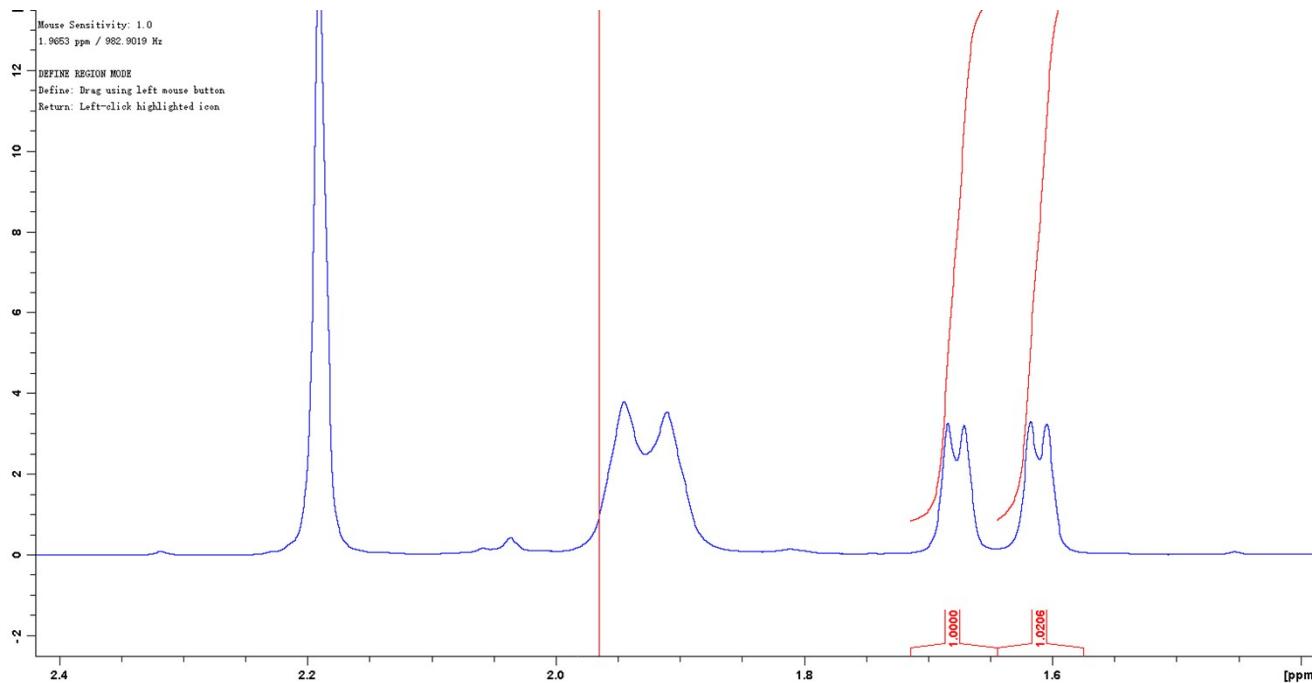
#	[min]	[min]	[mAU*s]	[mAU]	%	
1	6.504	BV	0.1360	95.05695	10.87021	2.0645
2	6.750	VB	0.1485	4509.29932	447.84070	97.9355

1-(1-(4-chlorophenyl)ethyl)pyrrolidine (3e):² 91% yield, 93% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.31 (s, 4H, Ar-CH), 3.21 (q, *J* = 6.5 Hz, 1H, CH), 2.57 (m, 2H, N-CH₂), 2.40 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.42 (d, *J* = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 143.3 (Ar-CCl), 132.7 (Ar-C), 128.7, 128.6, 65.4, 52.9, 23.4, 22.9. Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

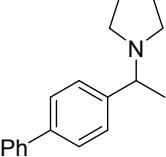


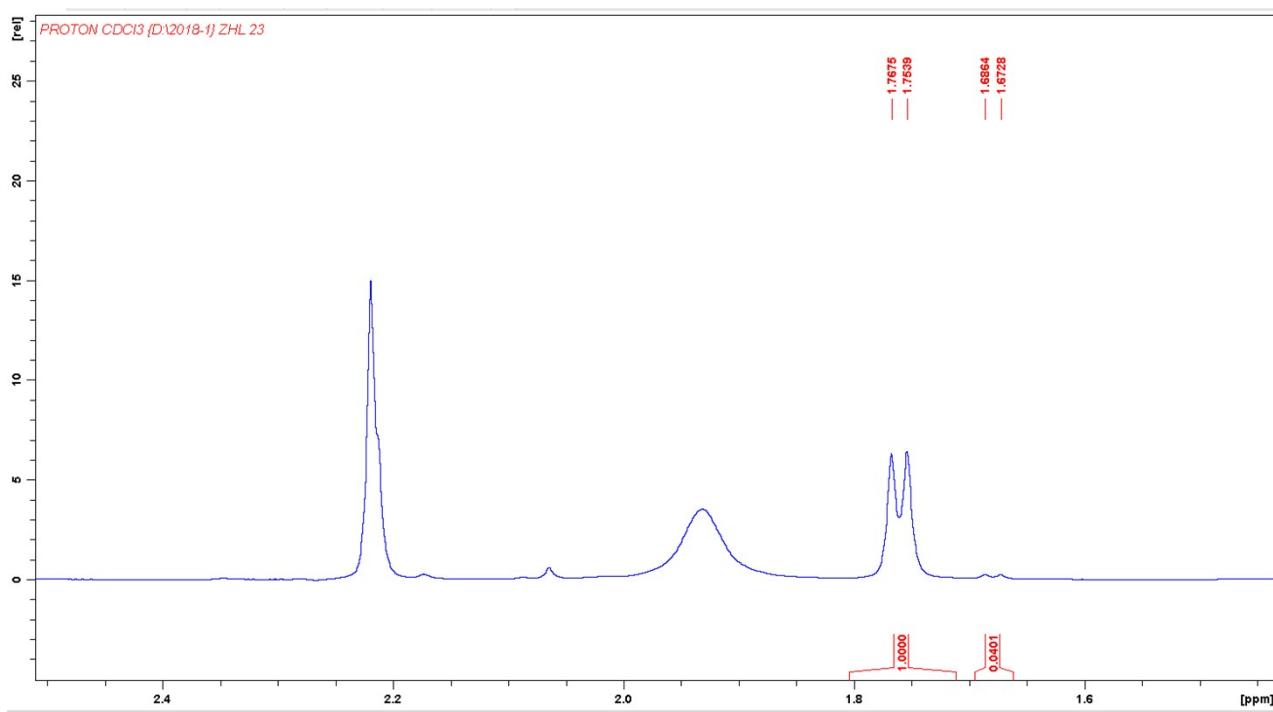
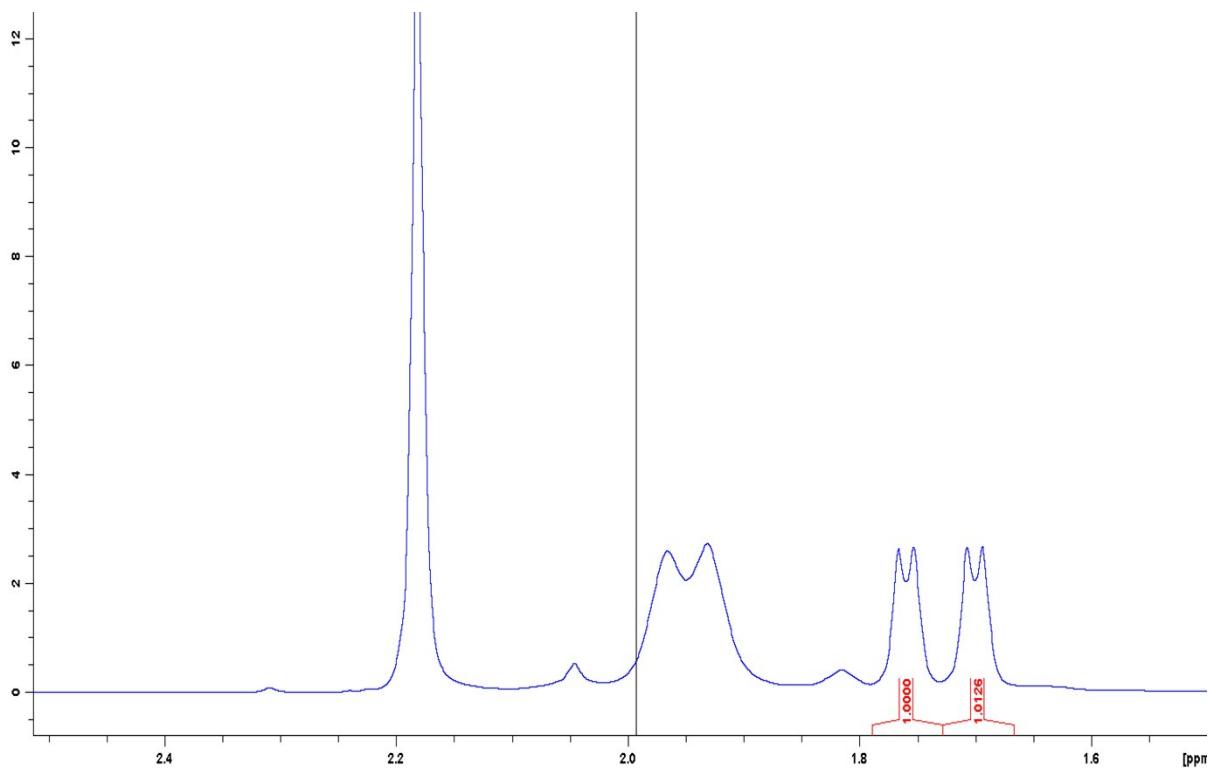
1-(1-(4-bromophenyl)ethyl)pyrrolidine (3f):⁴ 84% yield, 92% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.45 (d, *J* = 7.9

 Hz, 2H, Ar-CH), 7.25 (d, *J* = 7.8 Hz, 2H, Ar-CH), 3.18 (q, *J* = 6.2 Hz, 1H, CH), 2.55 (m, 2H, N-CH₂), 2.39 (m, 2H, N-CH₂), 1.79 (m, 4H, 2*CH₂), 1.40 (d, *J* = 6.3 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 144.5, 131.4, 129.0, 120.5, 65.4, 52.9, 23.4, 23.1. Enantiomeric excess was determined by ¹H NMR using (R)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

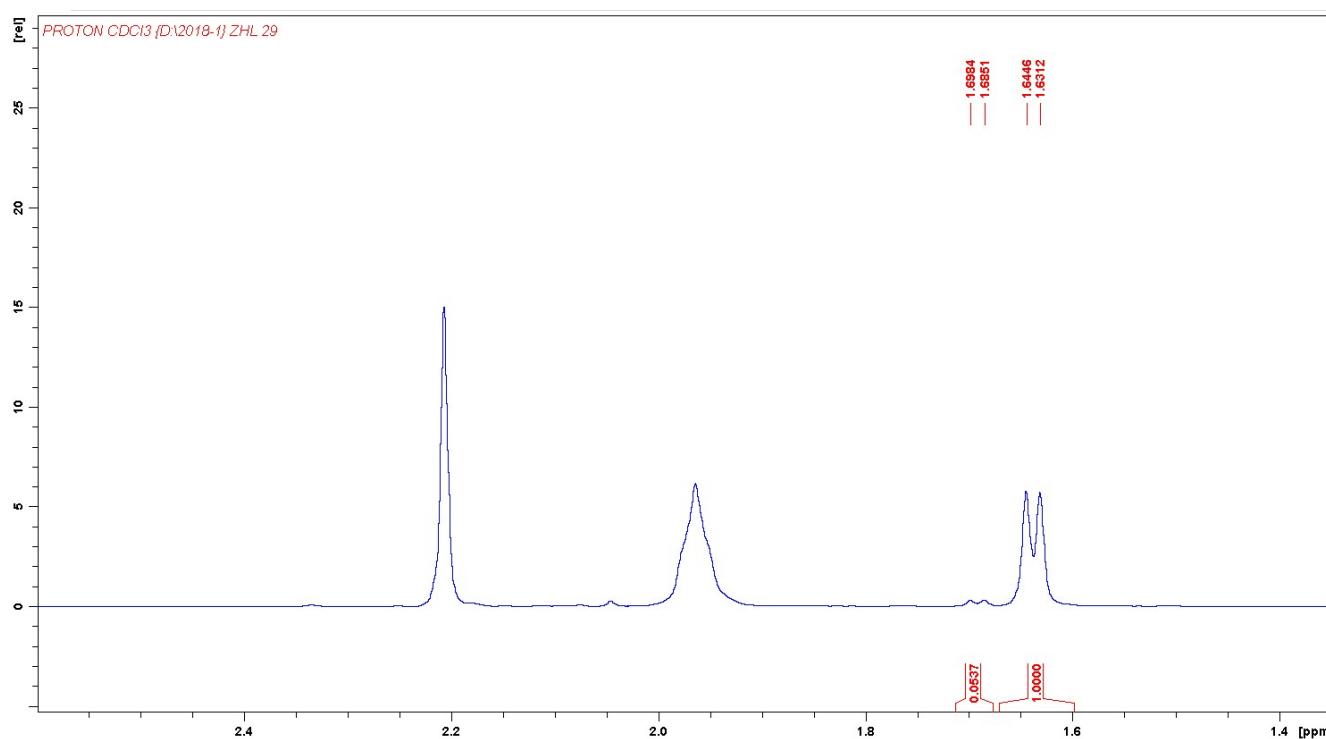
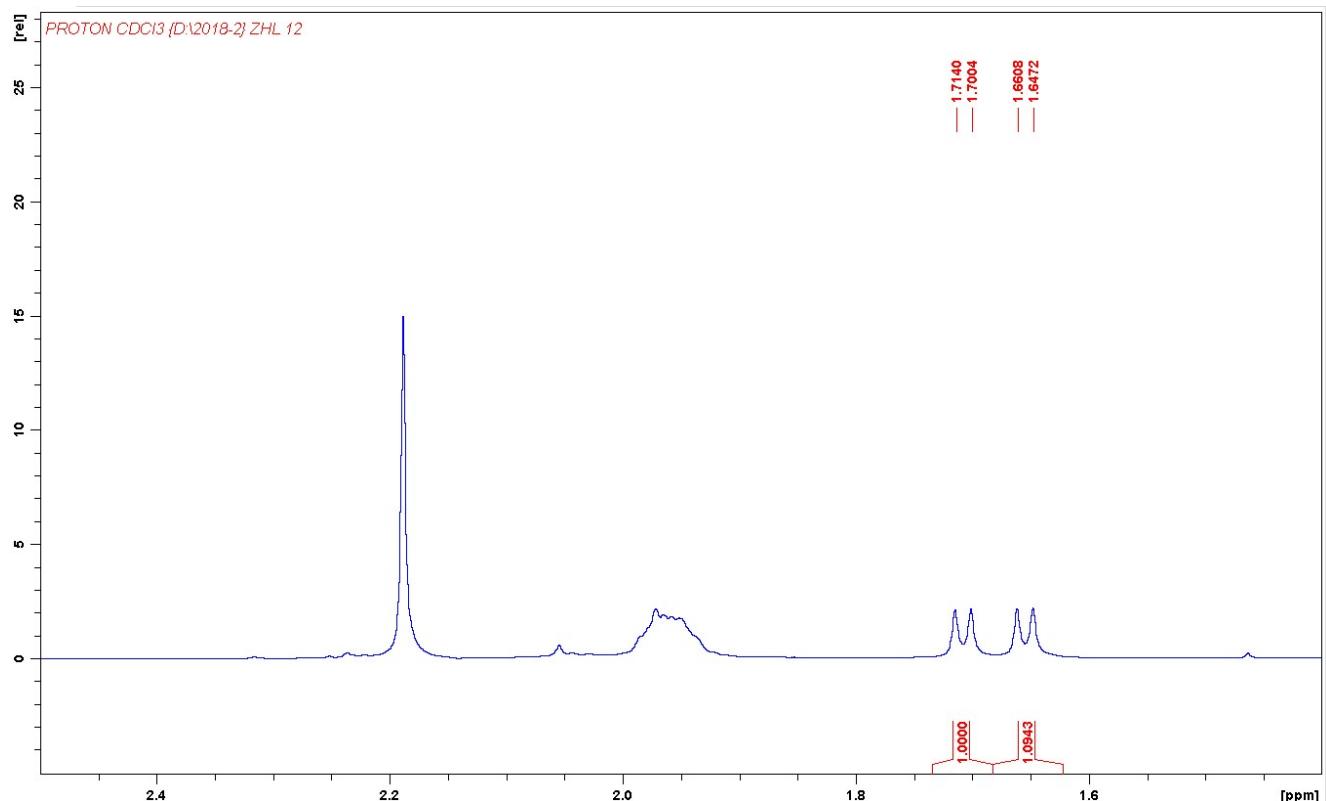


1-(1-([1,1'-biphenyl]-4-yl)ethyl)pyrrolidine (3g): 94% yield, 92% ee, brown oil, unknown compound. ^1H NMR (500 MHz,

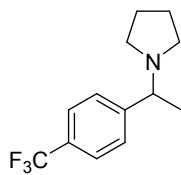
 CDCl₃): δ 7.65 (d, J = 7.3 Hz, 2H, Ar-CH), 7.59 (d, J = 8.1 Hz, 2H, Ar-CH), 7.47 (m, 4H, Ar-CH), 7.38 (m, 1H, Ar-CH), 3.28 (q, J = 6.5 Hz, 1H, CH), 2.63 (m, 2H, N-CH₂), 2.47 (m, 2H, N-CH₂), 1.83 (m, 4H, 2*CH₂), 1.49 (d, J = 6.6 Hz, 3H, CH₃); ^{13}C NMR (125 MHz, CDCl₃): δ 141.7, 140.2, 128.8, 128.5, 127.8, 127.7, 127.5, 66.1, 52.9, 23.4, 20.7. HRMS calcd for C₁₈H₂₂N [M+H]⁺: 252.1674, found: 252.1733. Enantiomeric excess was determined by ^1H NMR using (*S*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



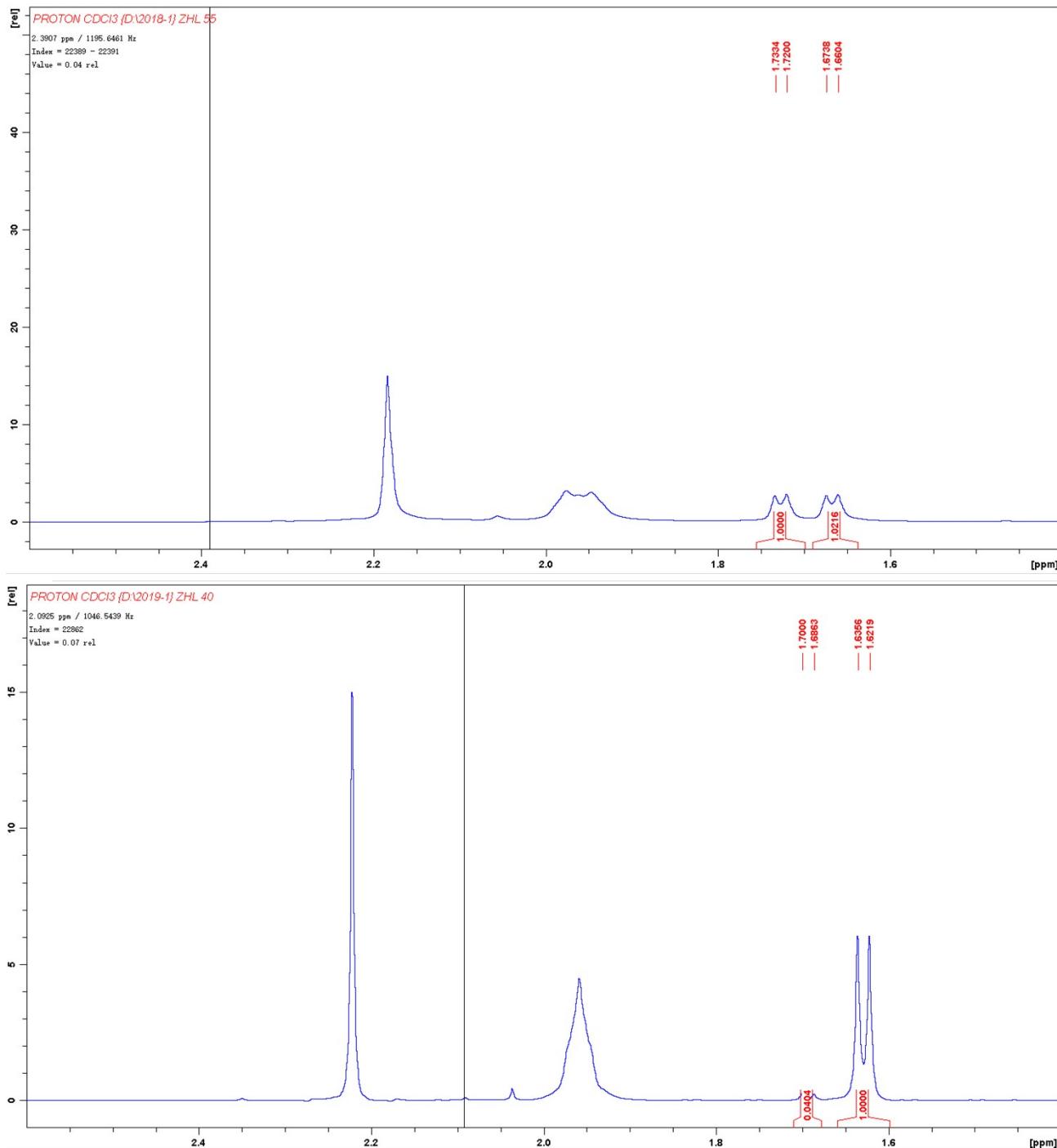
1-(1-(4-nitrophenyl)ethyl)pyrrolidine (3h):⁵ 90% yield, 90% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 8.20 (d, J = 8.7 Hz, 2H, Ar-CH), 7.55 (d, J = 8.6 Hz, 2H, Ar-CH), 3.34 (q, J = 6.6 Hz, 1H, CH), 2.57 (m, 2H, N-CH₂), 2.41 (m, 2H, N-CH₂), 1.81 (m, 4H, 2*CH₂), 1.42 (d, J = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 153.7, 146.9, 127.9, 123.7, 65.4, 52.9, 23.4, 23.3. Enantiomeric excess was determined by ¹H NMR using (R)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



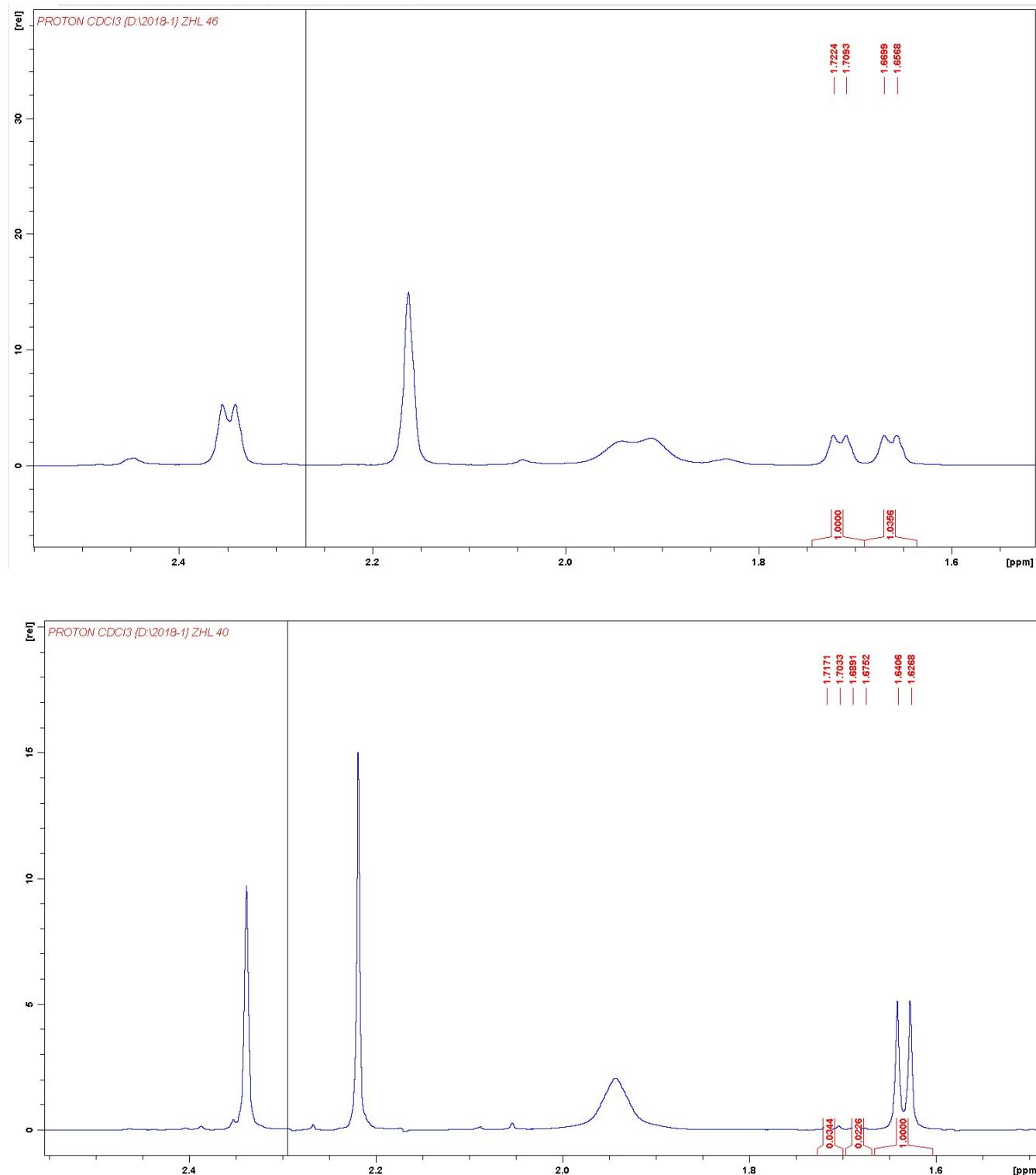
1-(1-(4-trifluoromethylphenyl)ethyl)pyrrolidine (3i**):**⁵ 94% yield, 90% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.59 (d,



J = 8.1 Hz, 2H, Ar-CH), 7.49 (d, *J* = 8.1 Hz, 2H, Ar-CH), 3.28 (q, *J* = 6.6 Hz, 1H, CH), 2.58 (m, 2H, N-CH₂), 2.40 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.42 (d, *J* = 7.1 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 150.0, 129.2, 127.5, 125.3, 125.2, 123.2, 66.3, 53.0, 23.4, 22.3, 21.5. Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

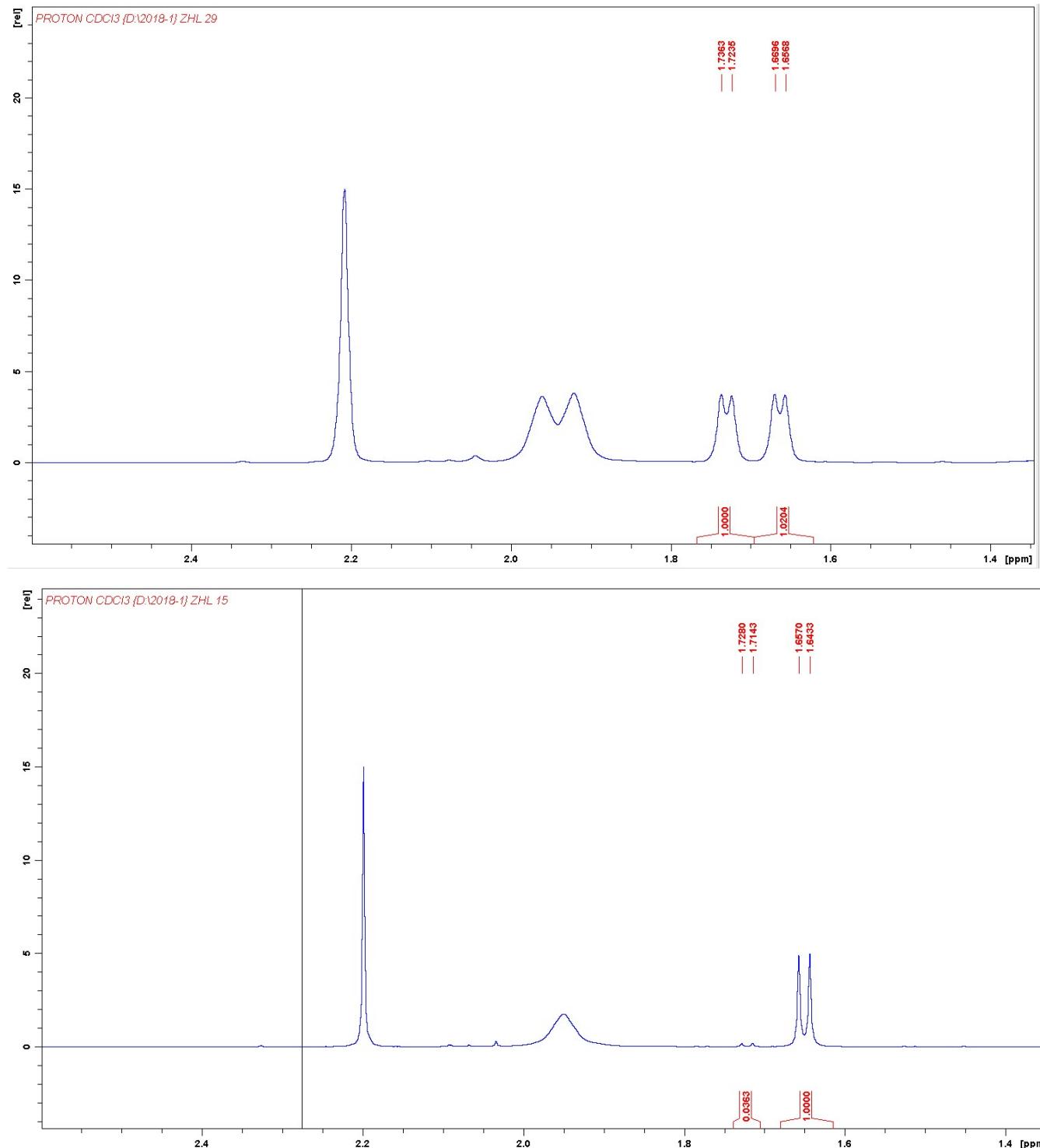


1-(1-(m-tolyl)ethyl)pyrrolidine (3j):³ 86% yield, 96% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.24 (m, 3H, Ar-CH), 7.11 (m, 1H, Ar-CH), 3.33 (m, 1H, CH), 2.71 (m, 2H, N-CH₂), 2.54 (m, 2H, N-CH₂), 2.38 (s, 3H, Ar-CH₃), 1.86 (m, 4H, 2*CH₂), 1.52 (d, *J* = 6.0 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 143.3, 138.2, 128.4, 128.3, 128.1, 124.6, 65.6, 52.9, 23.4, 23.3. Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

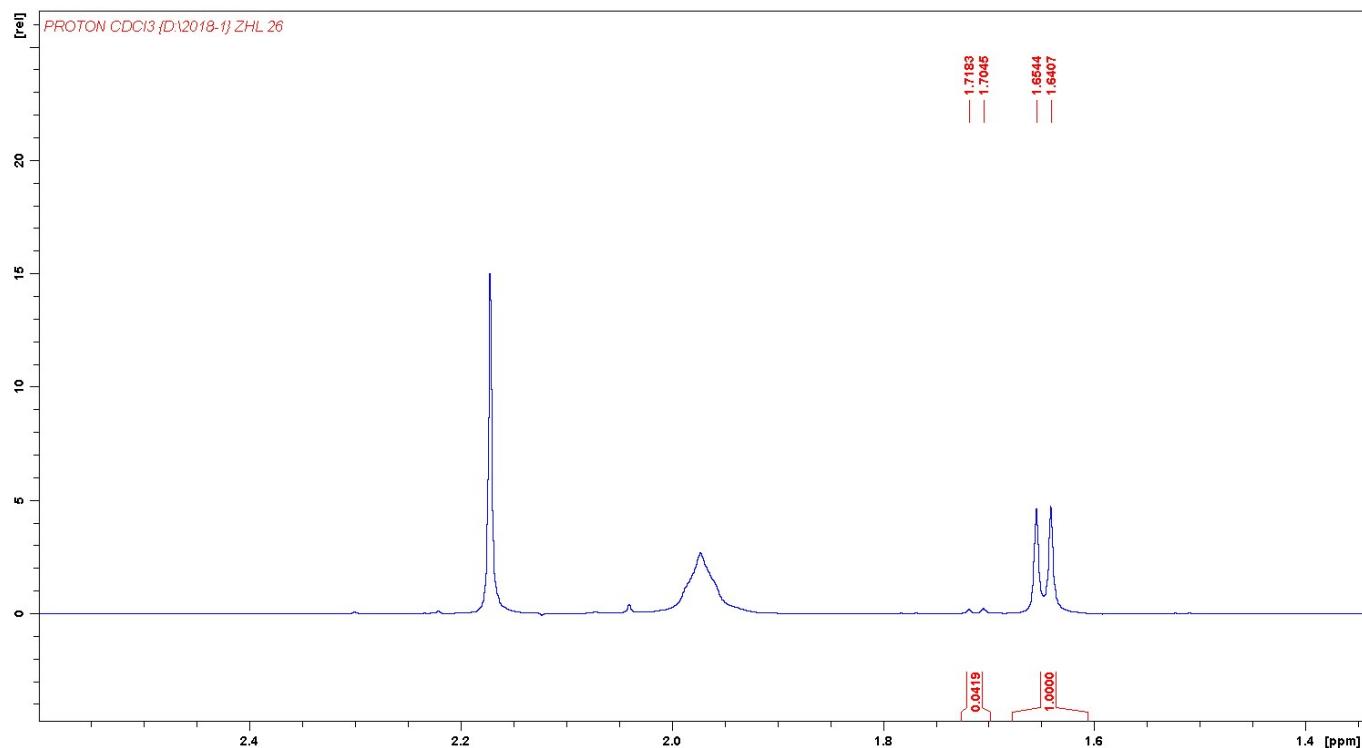
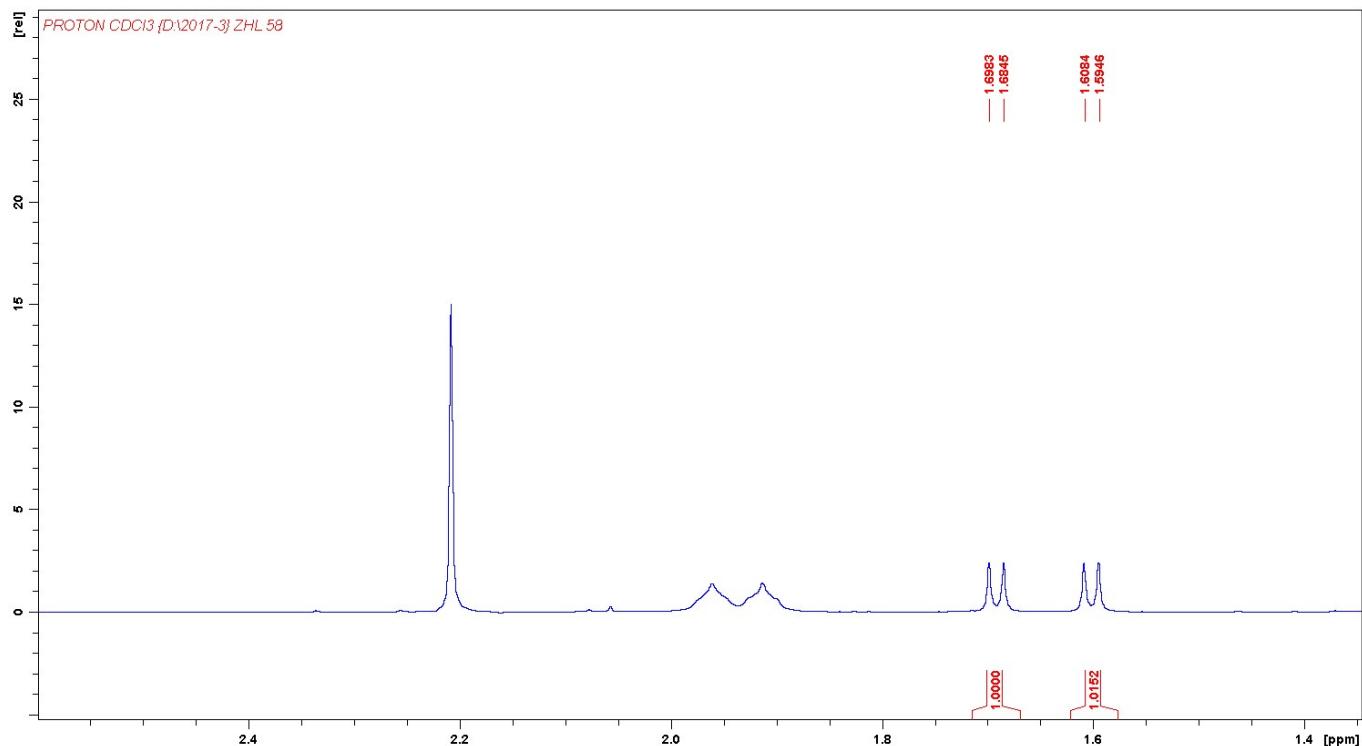
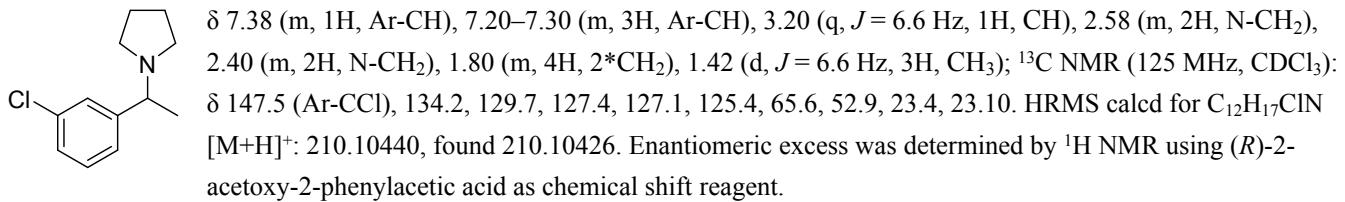


1-(1-(3-methoxyphenyl)ethyl)pyrrolidine (3k):³ 92% yield, 93% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.25 (m, 1H, Ar-CH), 6.96 (m, 2H, Ar-CH), 6.82 (m, 1H, Ar-CH), 3.84 (s, 3H, O-CH₃), 3.20 (q, *J* = 6.6 Hz, 1H, CH), 2.60 (m, 2H, N-CH₂), 2.42 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.44 (d, *J* = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 159.7, 147.5, 129.2, 119.7, 112.7, 112.3, 66.1, 55.3, 53.0, 23.5, 23.2.

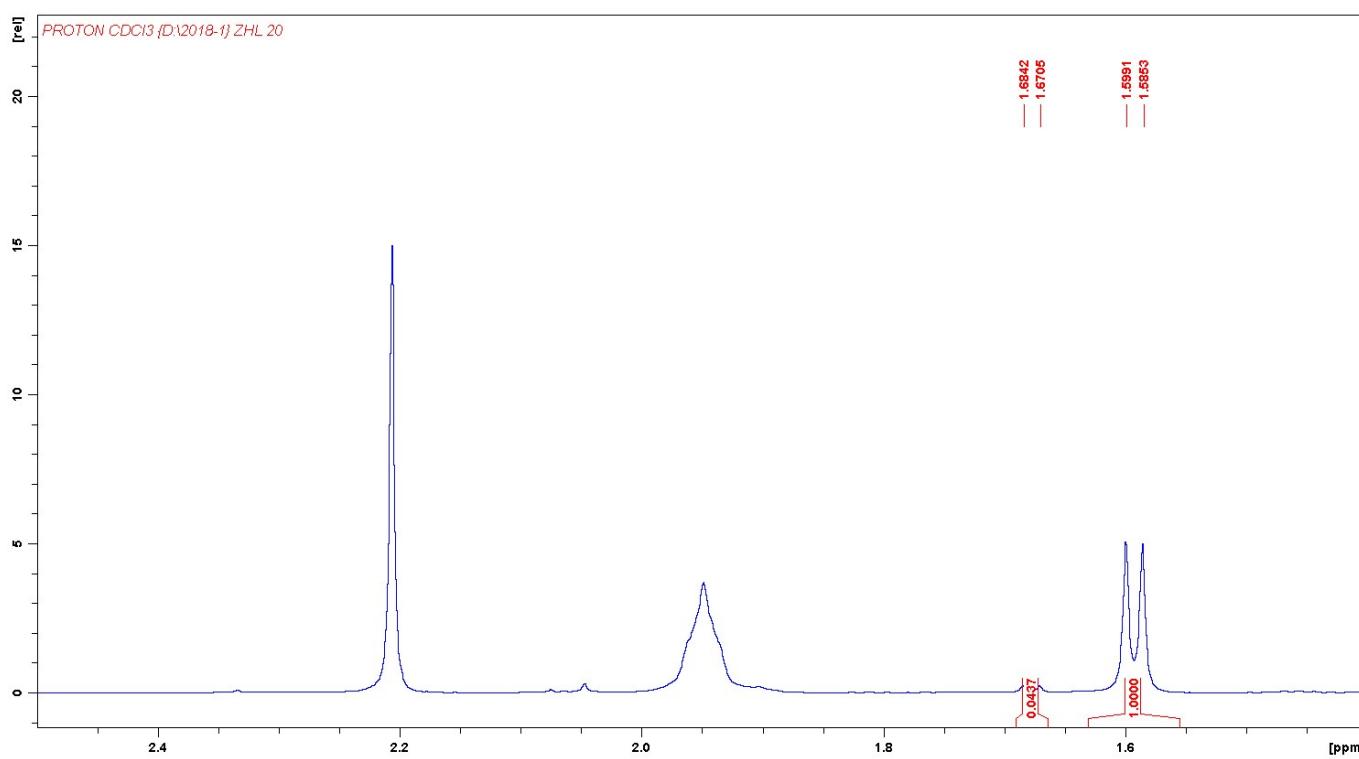
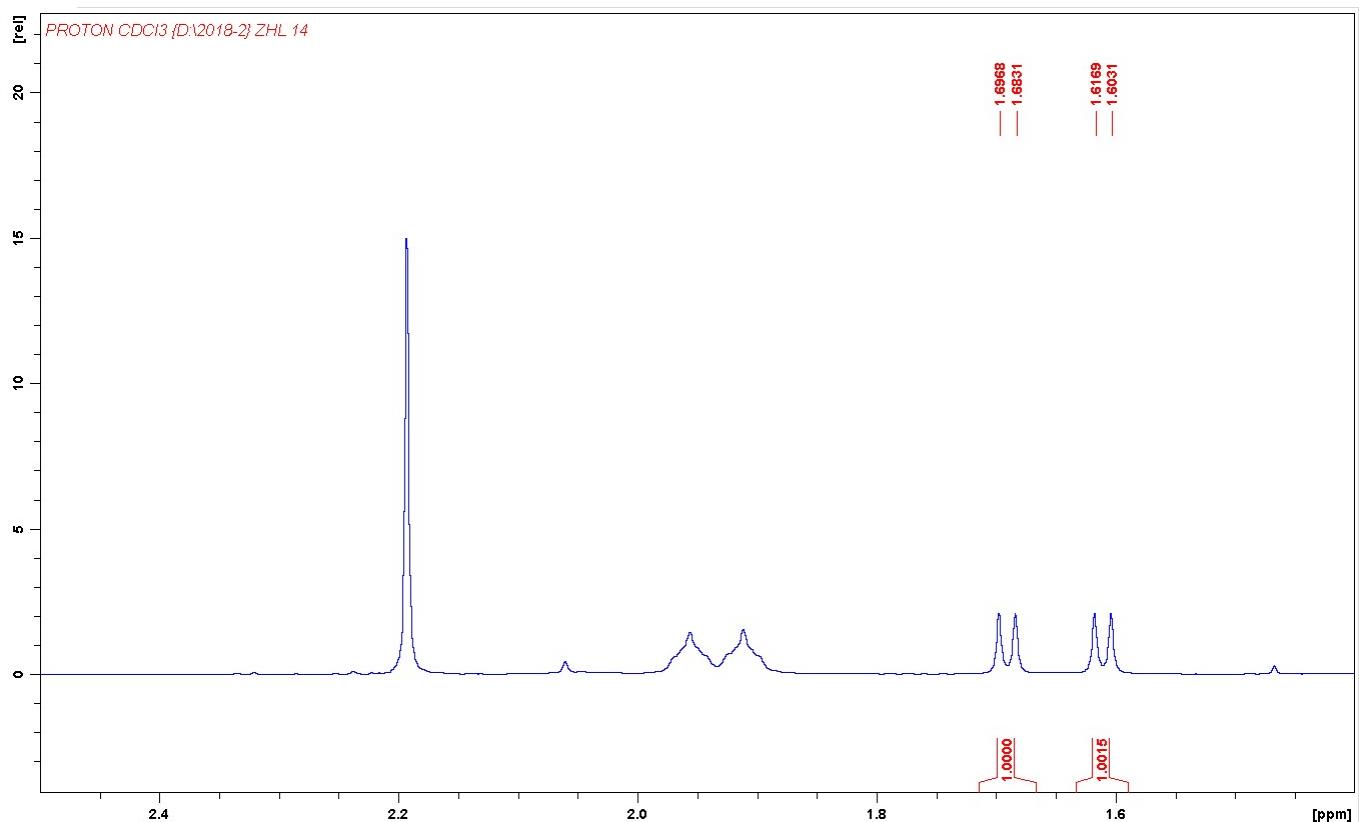
Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



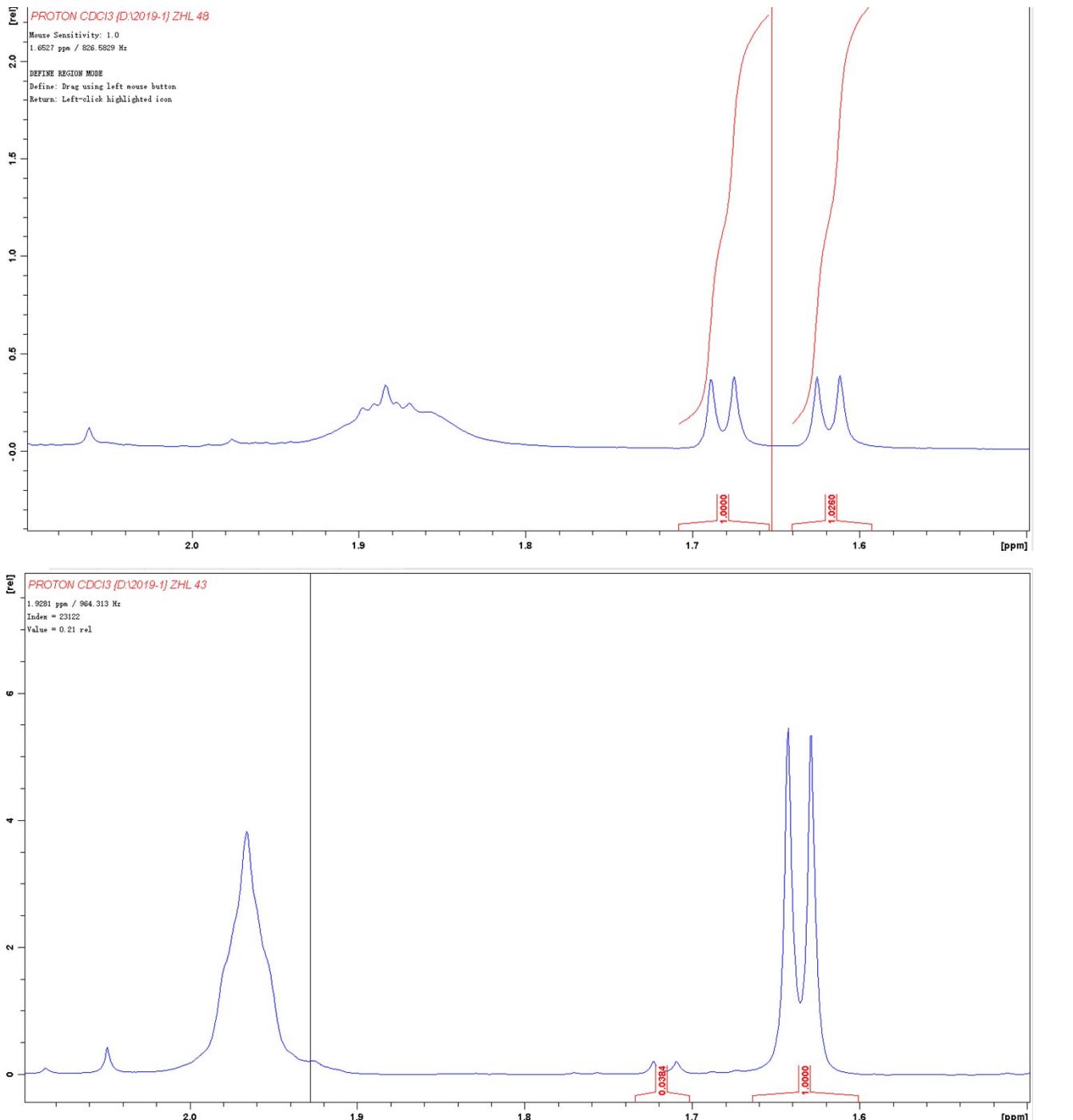
1-(1-(3-chlorophenyl)ethyl)pyrrolidine (3l): 91% yield, 92% ee, brown oil, unkown compound. ^1H NMR (500 MHz, CDCl_3):



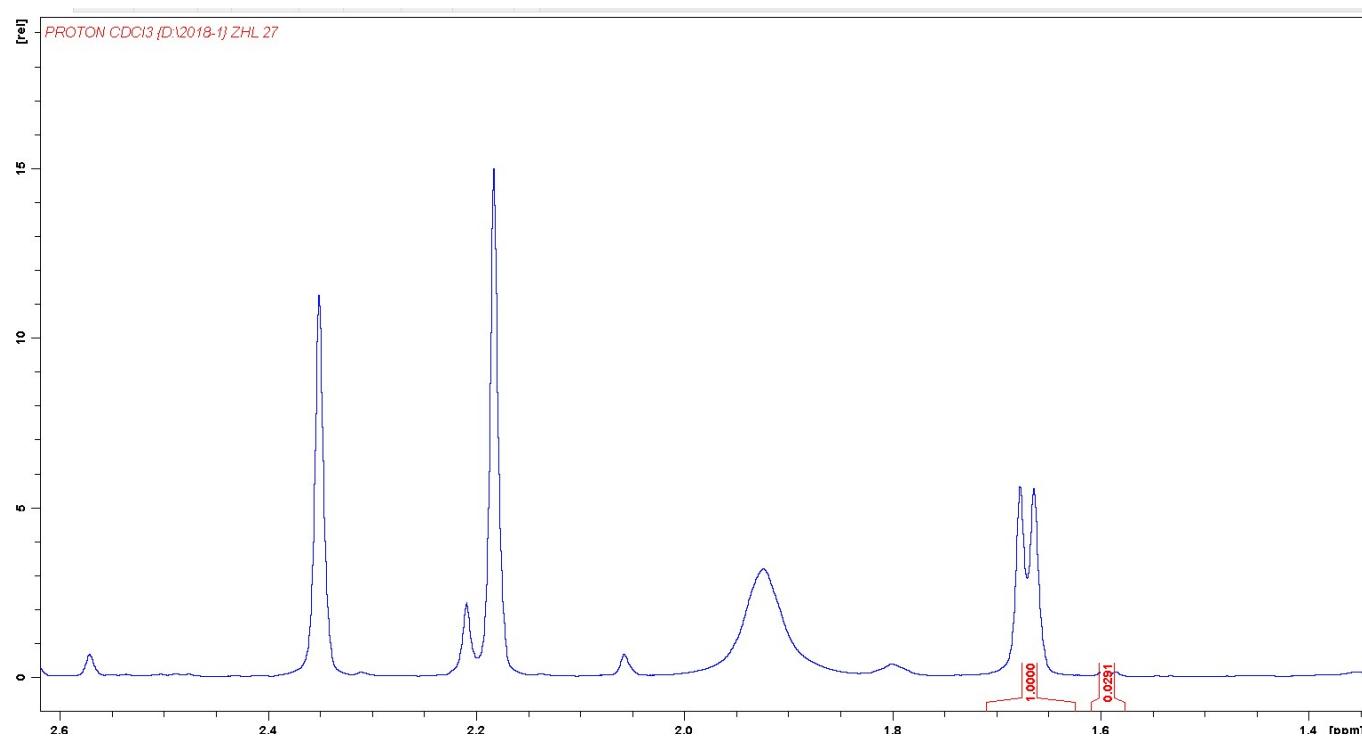
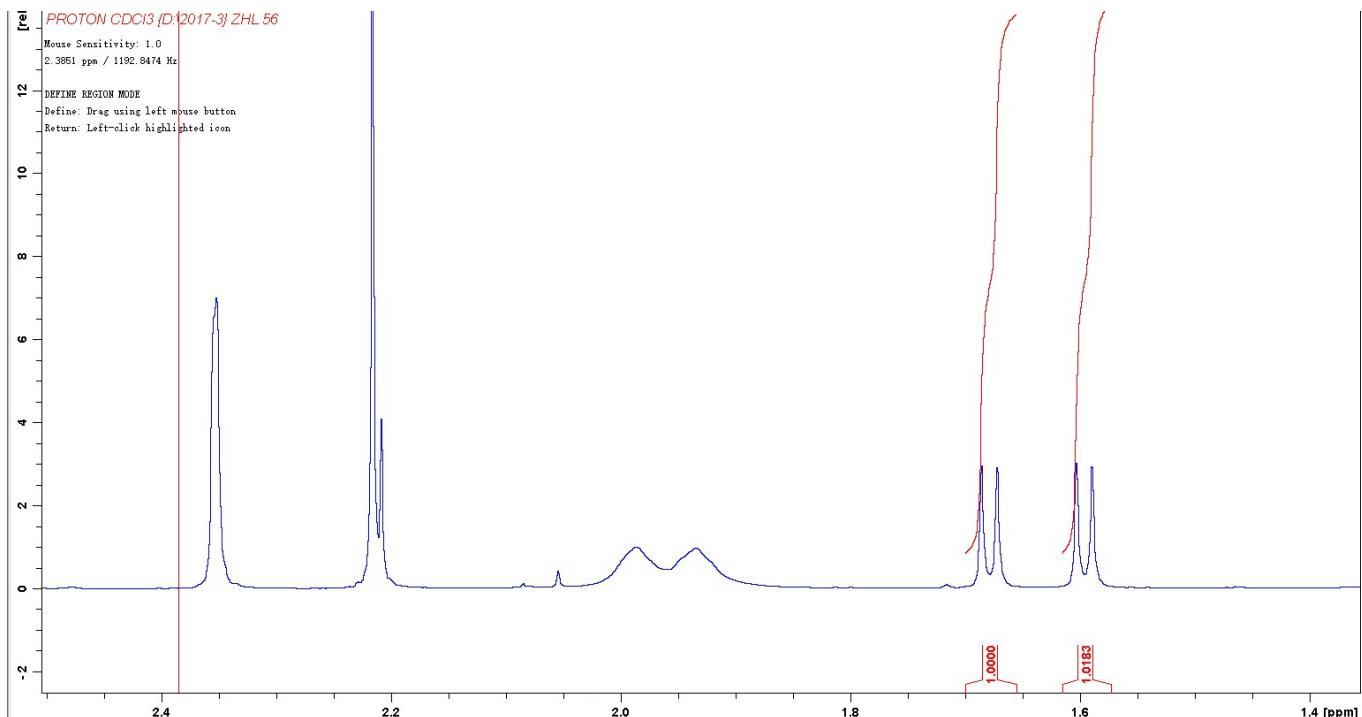
1-(1-(3-bromophenyl)ethyl)pyrrolidine (3m): 89% yield, 92% ee, brown oil. ^1H NMR (500 MHz, CDCl_3): δ 7.54 (s, 1H, Ar-CH), 7.39 (d, $J = 7.9$ Hz, 1H, Ar-CH), 7.39 (d, $J = 7.9$ Hz, 1H, Ar-CH), 7.20 (t, $J = 7.8$ Hz, 1H, Ar-CH), 3.18 (q, $J = 6.6$ Hz, 1H, CH), 2.57 (m, 2H, N-CH₂), 2.40 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.41 (d, $J = 6.6$ Hz, 3H, CH₃); ^{13}C NMR (125 MHz, CDCl_3): δ 148.3 (Ar-CBr), 130.2, 129.9, 129.9, 125.9, 122.4, 65.5, 52.9, 23.4, 23.2. HRMS calcd for $\text{C}_{12}\text{H}_{17}\text{BrN} [\text{M}+\text{H}]^+$: 254.0466, found: 254.0525. Enantiomeric excess was determined by ^1H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



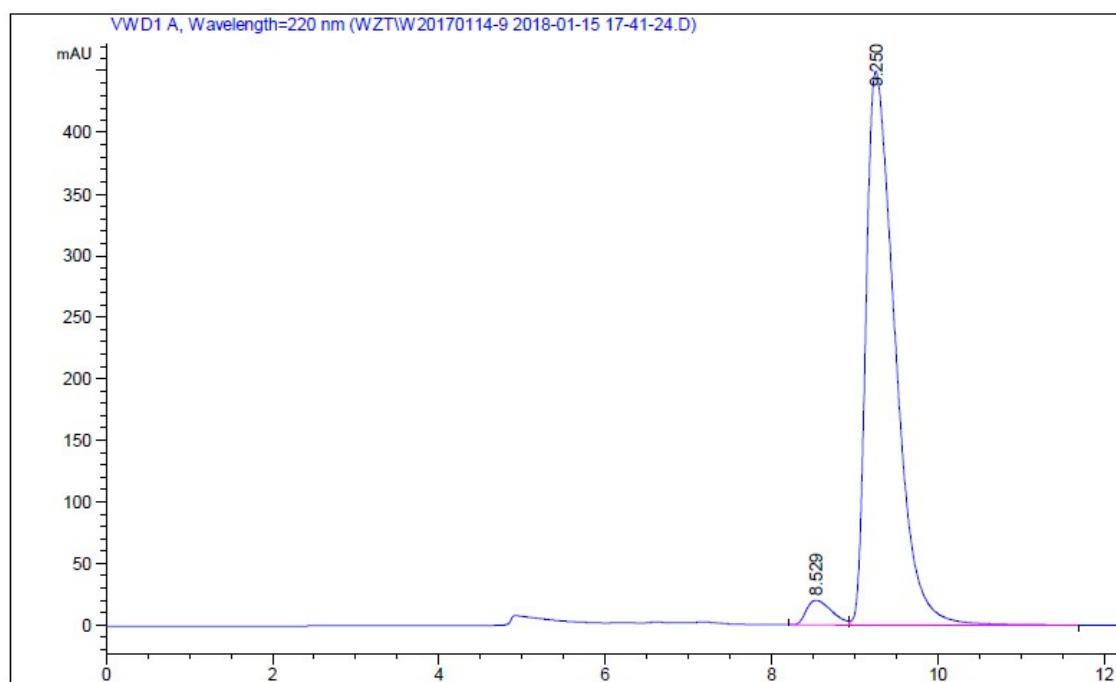
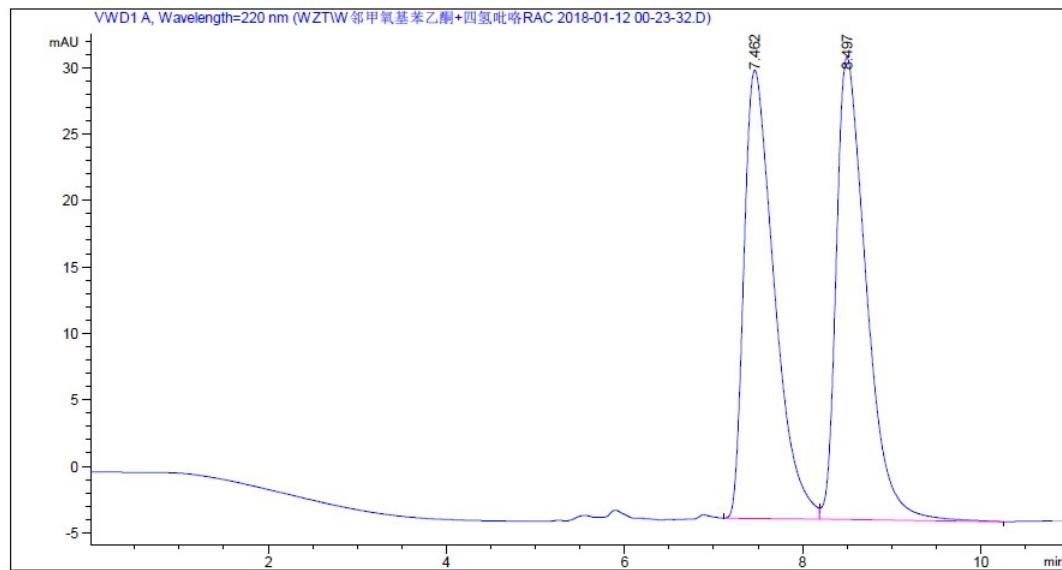
1-(1-(3-trifluoromethylphenyl)ethyl)pyrrolidine (3n**):** 91% yield, 93% ee, brown oil. ^1H NMR (500 MHz, CDCl_3): δ 7.64 (s, 1H, Ar-CH), 7.57 (d, J = 7.6 Hz, 1H, Ar-CH), 7.52 (d, J = 7.7 Hz, 1H, Ar-CH), 7.45 (t, J = 7.7 Hz, 1H, Ar-CH), 3.28 (q, J = 6.6 Hz, 1H, CH), 2.58 (m, 2H, N- CH_2), 2.40 (m, 2H, N- CH_2), 1.81 (m, 4H, 2* CH_2), 1.43 (d, J = 6.6 Hz, 3H, CH_3); ^{13}C NMR (125 MHz, CDCl_3): δ 146.9 (Ar-C- CF_3), 130.6, 128.8, 124.0, 123.7, 65.7, 53.0, 23.4, 23.3. HRMS calcd for $\text{C}_{12}\text{H}_{17}\text{BrN} [\text{M}+\text{H}]^+$: 254.0466, found: 254.0525. Enantiomeric excess was determined by ^1H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



1-(1-(o-tolyl)ethyl)pyrrolidine (3o):³ 89% yield, 94% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.58 (m, 1H, Ar-CH), 7.24 (m, 1H, Ar-CH), 7.15(m, 1H, Ar-CH), 3.52 (q, *J* = 6.5 Hz, 1H, CH), 2.58 (m, 2H, N-CH₂), 2.48 (m, 2H, N-CH₂), 2.42 (s, 3H, Ar-CH₃), 1.82 (m, 4H, 2*CH₂), 1.38 (d, *J* = 6.0 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 144.1, 134.5, 130.2, 126.9, 126.2, 126.1, 61.2, 53.0, 23.5, 22.4, 19.6. Enantiomeric excess was determined by ¹H NMR using (*S*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

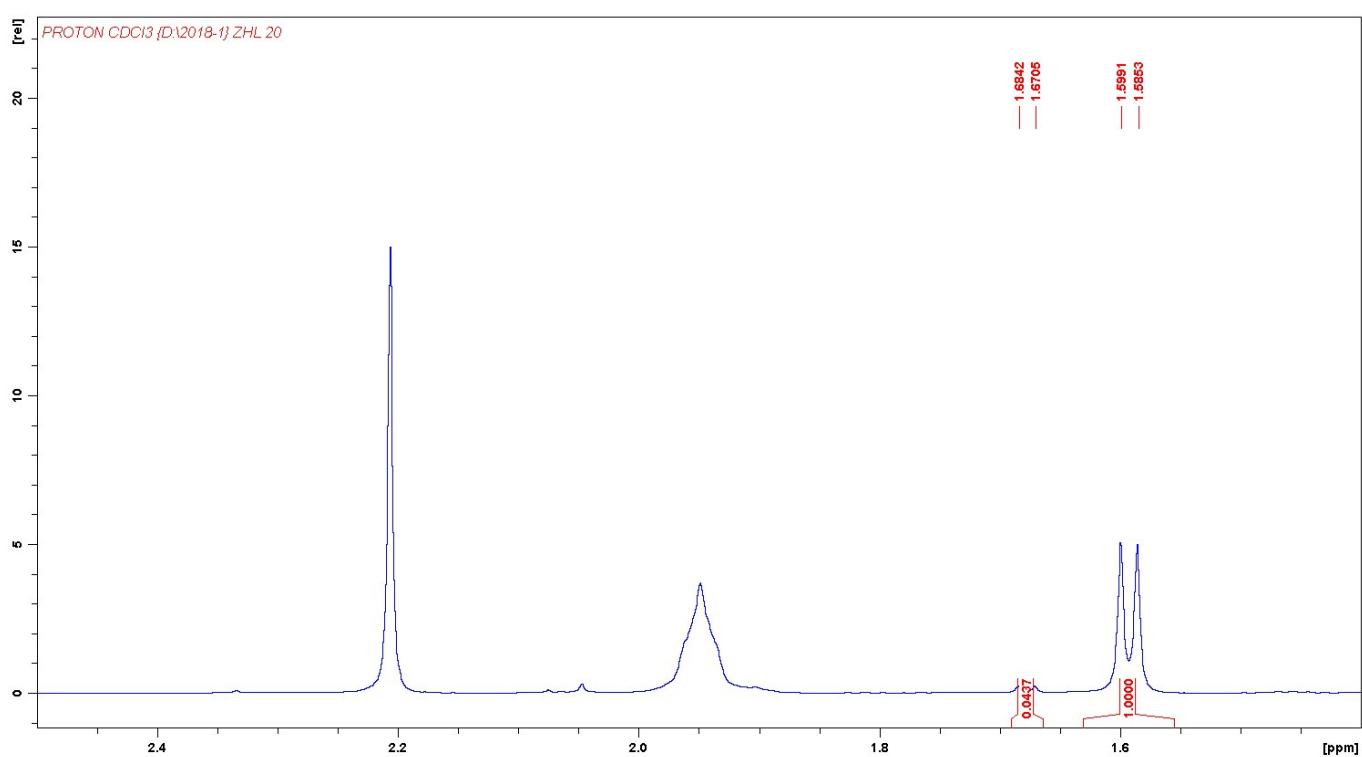
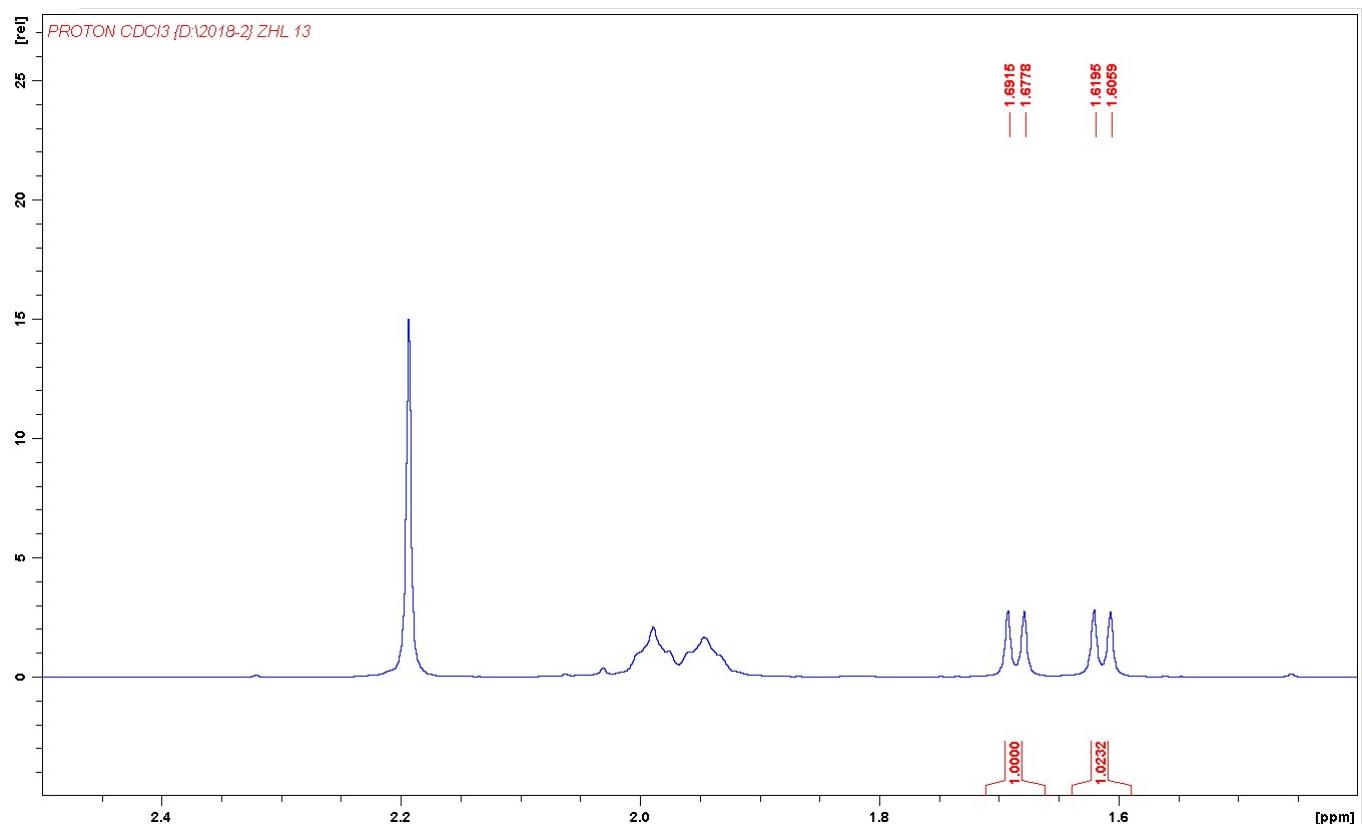


1-(1-(2-methoxyphenyl)ethyl)pyrrolidine (3p):³ 97% yield, 94% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.56 (m, 1H, Ar-CH), 7.23 (m, 1H, Ar-CH), 7.01 (m, 1H, Ar-CH), 6.90 (m, 1H, Ar-CH), 3.82-3.90 (m, 4H, O-CH₃+CH), 2.60 (m, 2H, N-CH₂), 2.51 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.39 (d, *J* = 6.6 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 156.2 (Ar-COMe), 130.0, 128.9, 121.6, 110.8, 57.8, 55.6, 52.5, 23.4, 19.6. Enantiomeric excess was determined by chiral HPLC: Chiralpak OD-H column, Hex/IPA=99.5:0.5, 0.6 mL/min, 220 nm, 7.46 min, 8.50 min.

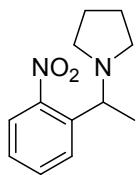


1-(1-(2-chlorophenyl)ethyl)pyrrolidine (3q): 91% yield, 92% ee, brown oil, unknown compound. ^1H NMR (500 MHz,

CC1(Cl)CCCC1c2ccccc2 CDCl₃): δ 7.38 (m, 1H, Ar-CH), 7.20–7.30 (m, 3H, Ar-CH), 3.20 (q, J = 6.6 Hz, 1H, CH), 2.58 (m, 2H, N-CH₂), 2.40 (m, 2H, N-CH₂), 1.80 (m, 4H, 2*CH₂), 1.42 (d, J = 6.6 Hz, 3H, CH₃); ^{13}C NMR (125 MHz, CDCl₃): δ 142.1, 132.6, 129.3, 128.8, 127.9, 127.2, 61.0, 52.8, 23.5, 21.9. HRMS calcd for C₁₂H₁₇ClN [M+H]⁺: 210.10440, found 210.10420. Enantiomeric excess was determined by ^1H NMR using (S)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

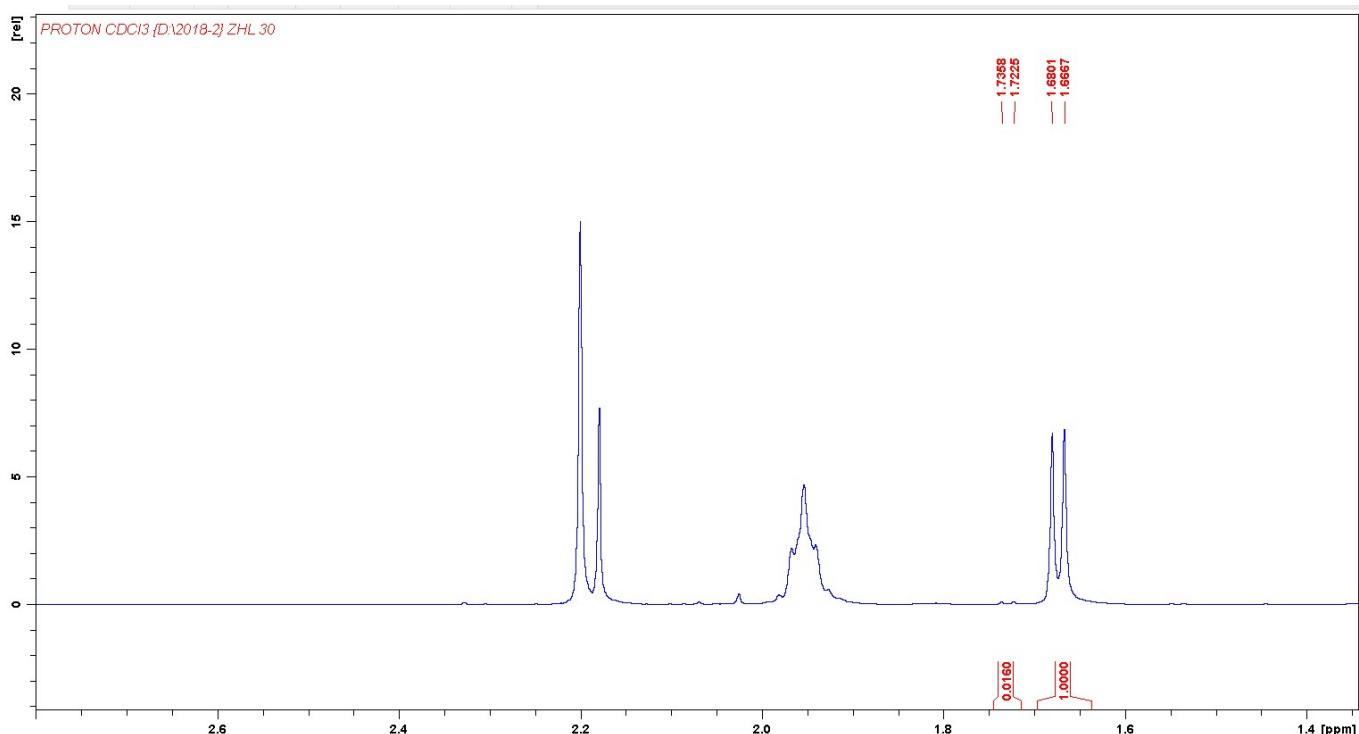
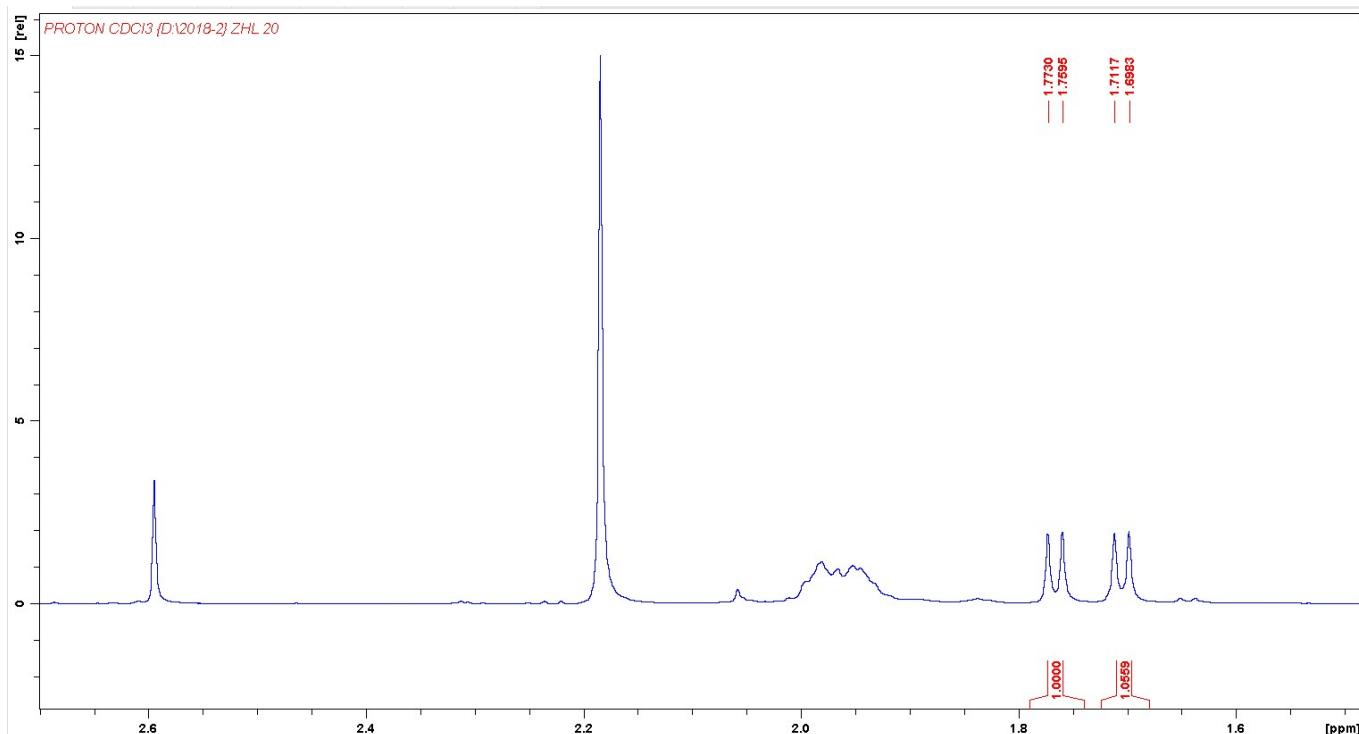


1-(1-(2-nitrophenyl)ethyl)pyrrolidine (3r): 94% yield, 97% ee, brown oil, unknown compound. ^1H NMR (500 MHz, CDCl_3):

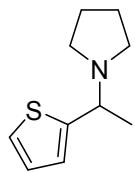


δ 7.90 (d, $J = 7.7$ Hz, 1H, Ar-CH), 7.72 (d, $J = 8.0$ Hz, 1H, Ar-CH), 7.59 (t, $J = 7.5$ Hz, 1H, Ar-CH), 7.37 (q, $J = 7.5$ Hz, 1H, Ar-CH), 3.71 (q, $J = 6.1$ Hz, 1H, CH), 2.54 (m, 2H, N- CH_2), 2.40 (m, 2H, N- CH_2), 1.79 (m, 4H, 2* CH_2), 1.50 (d, $J = 6.1$ Hz, 3H, CH_3); ^{13}C NMR (125 MHz, CDCl_3): δ 149.4, 140.0, 132.6, 129.3, 127.3, 123.3, 59.7, 52.7, 23.5, 22.7. HRMS (ESI) m/z calcd for $\text{C}_{12}\text{H}_{17}\text{N}_2\text{O}_2^+ [\text{M}+\text{H}]^+$ 221.12845, found 221.12848.

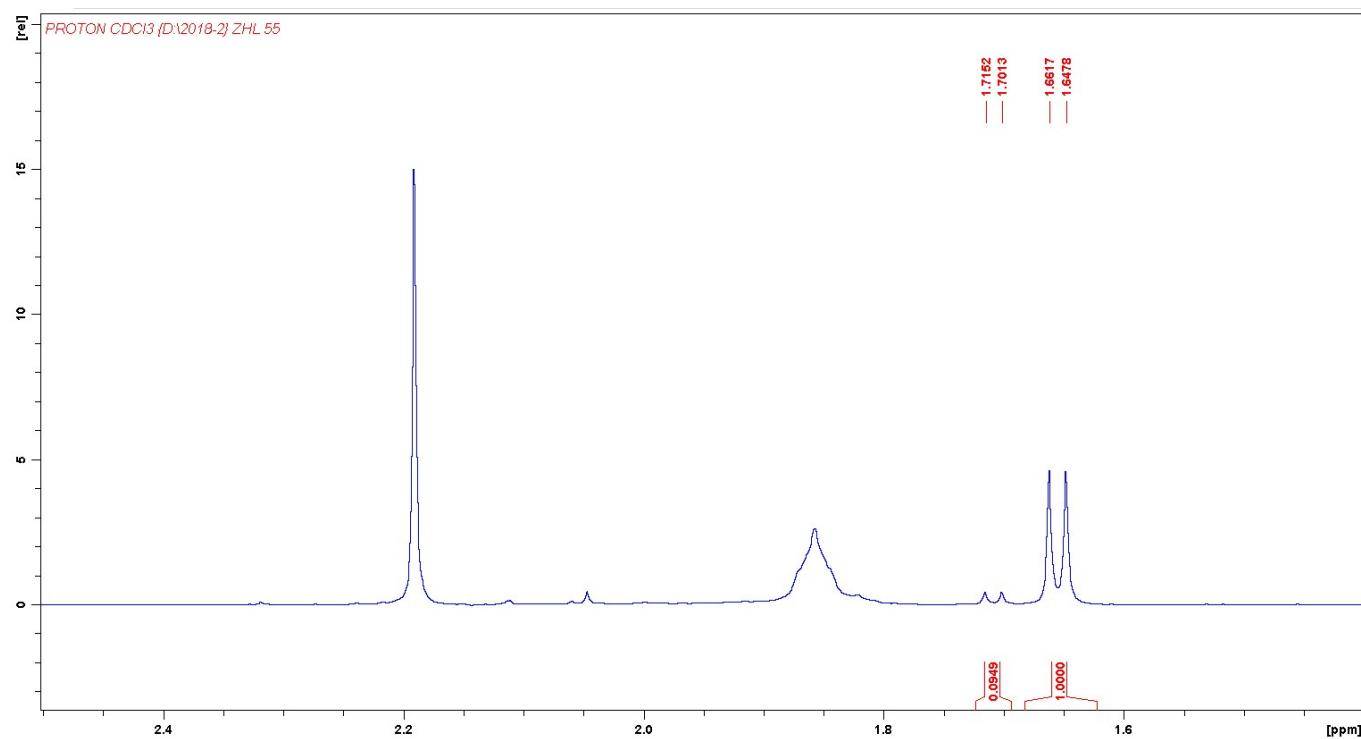
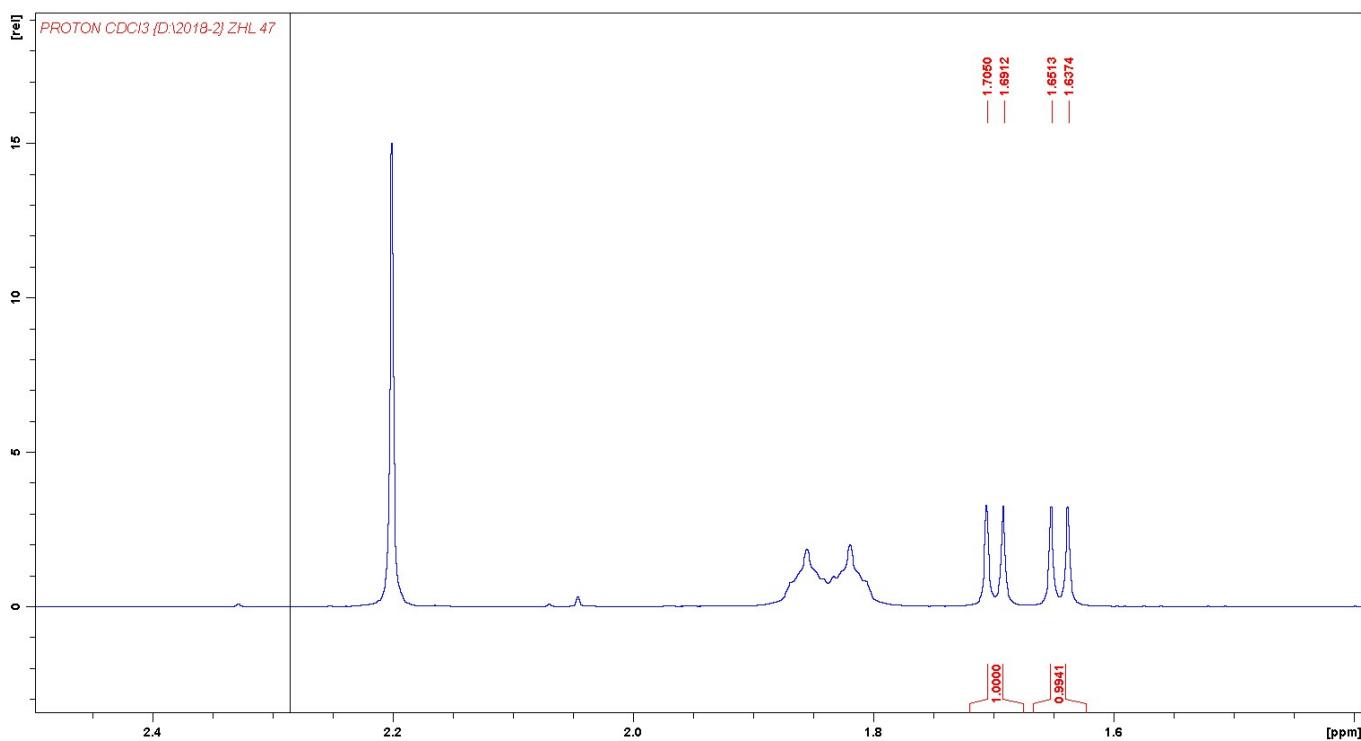
Enantiomeric excess was determined by ^1H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



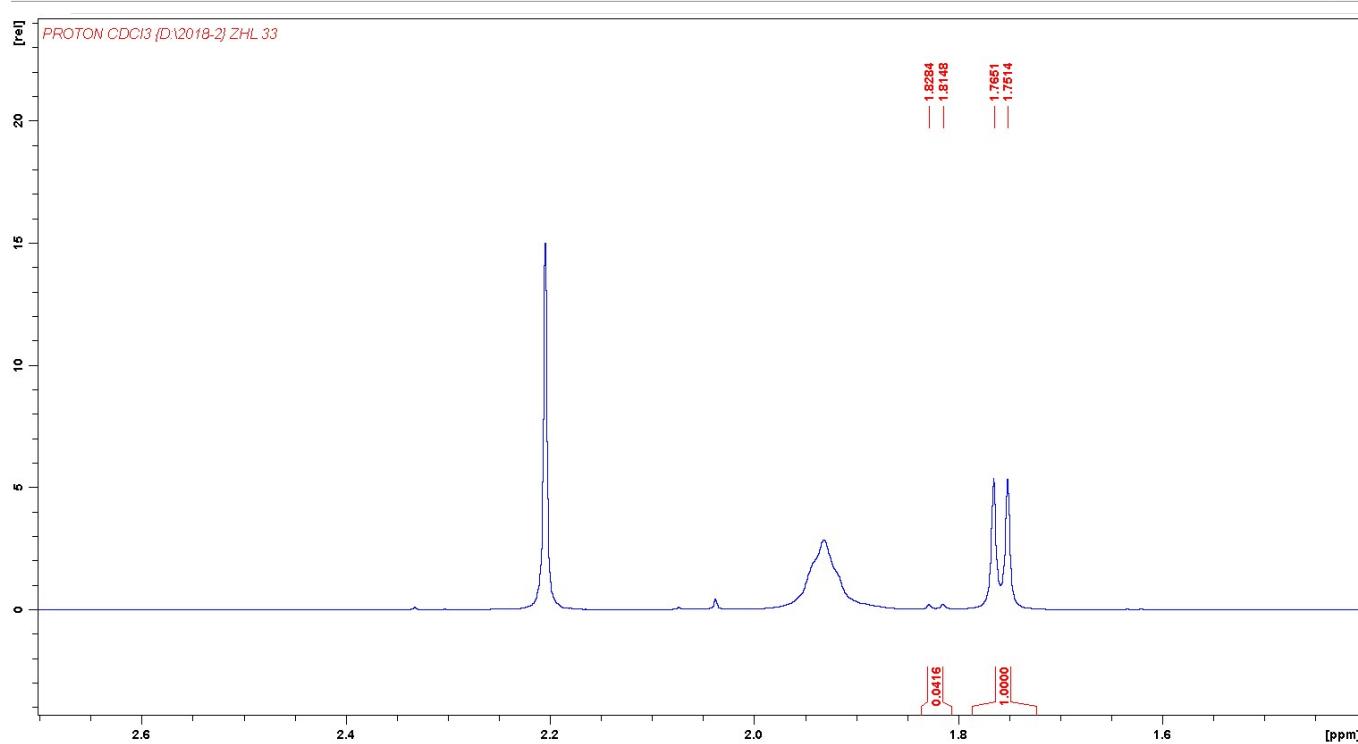
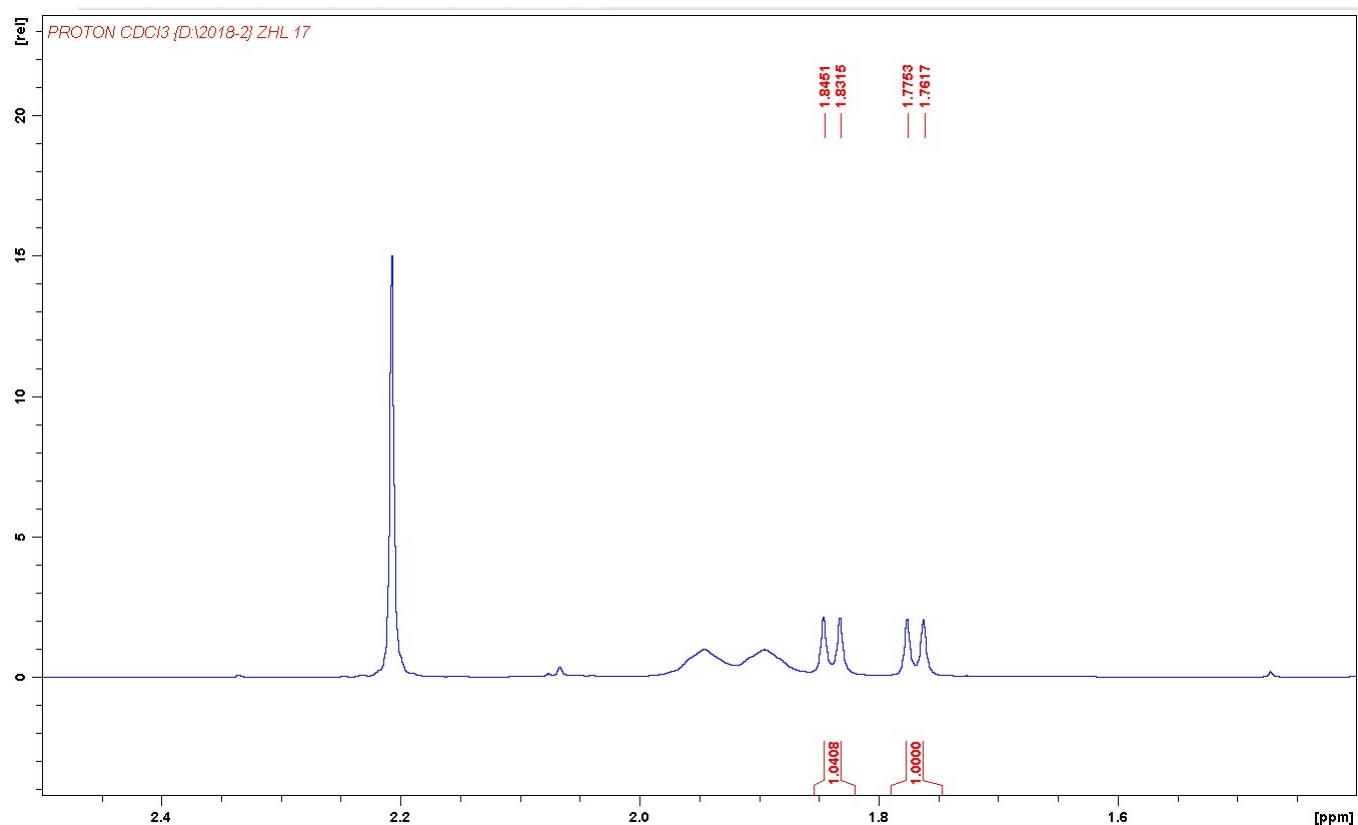
1-(1-(thiophen-2-yl)ethyl)pyrrolidine (3s): 91% yield, 82% ee, brown oil, unknown compound. ^1H NMR (500 MHz, CDCl_3):



δ 7.22 (m, 1H, Ar-CH), 6.95 (m, 3H, Ar-CH), 3.73 (q, $J = 6.6$ Hz, 1H, CH), 2.64 (m, 2H, N- CH_2), 2.53 (m, 2H, N- CH_2), 1.81 (m, 4H, 2* CH_2), 1.54 (d, $J = 6.7$ Hz, 3H, CH_3); ^{13}C NMR (125 MHz, CDCl_3): δ 126.0, 123.8, 123.7, 59.8, 52.0, 23.5. HRMS (ESI) m/z calcd for $\text{C}_{10}\text{H}_{16}\text{NS}^+ [\text{M}+\text{H}]^+$ 182.09980, found 182.09961. Enantiomeric excess was determined by ^1H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

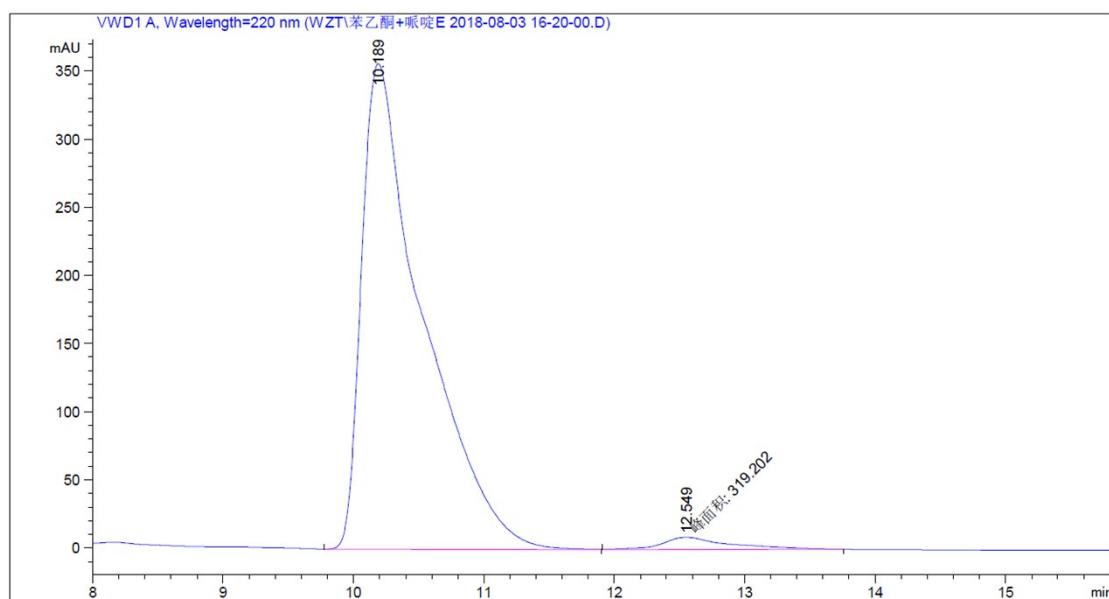
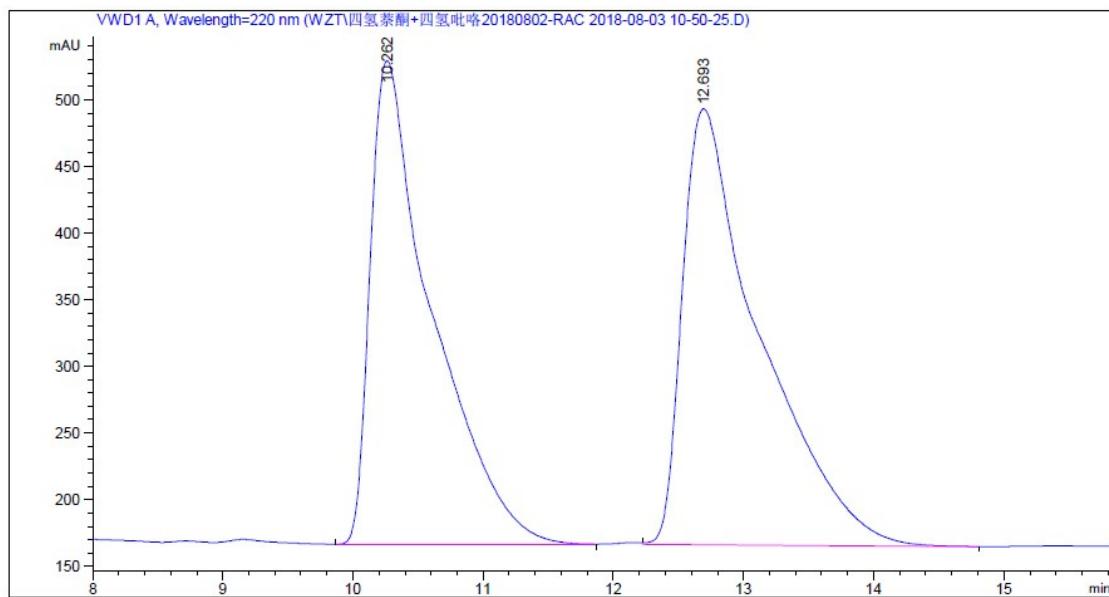


1-(1-(naphthalen-1-yl)ethyl)pyrrolidine (3t):⁶ 95% yield, 92% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 8.51 (d, *J* = 6.8 Hz, 1H, Ar-CH), 7.93 (d, *J* = 7.6 Hz, 1H, Ar-CH), 7.80 (m, 2H, Ar-CH), 7.54 (m, 3H), 4.01 (m, 1H, CH), 2.69 (m, 2H, N-CH₂), 2.57 (m, 2H, N-CH₂), 1.86 (m, 4H, 2*CH₂), 1.60 (d, *J* = 6.4 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 141.8, 134.0, 131.1, 128.9, 127.0, 125.7, 125.5, 125.2, 124.6, 123.7, 62.4, 53.1, 23.6, 22.9. Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



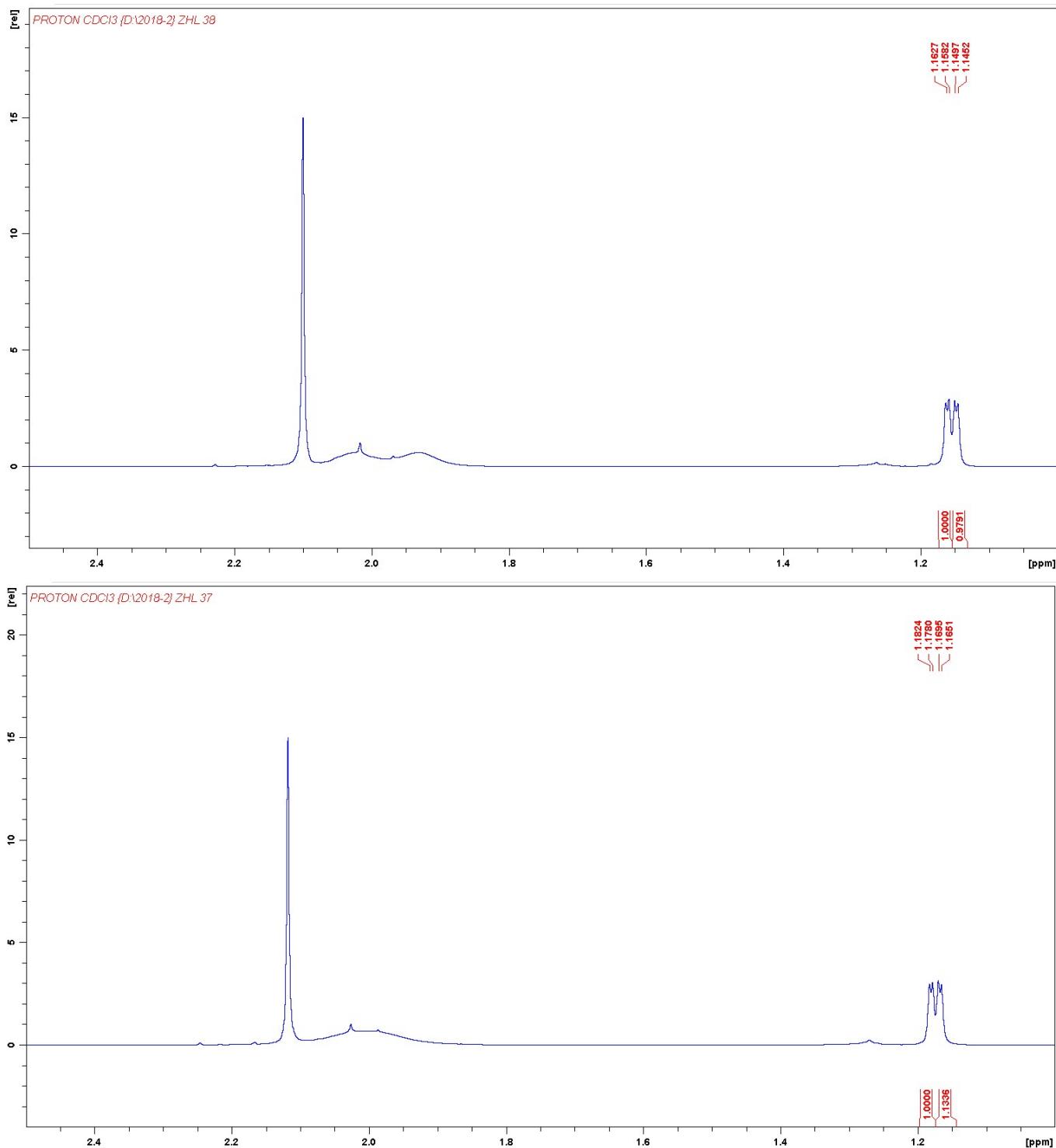
1-(1,2,3,4-tetrahydronaphthalen-1-yl)pyrrolidine (3u):⁷ 83% yield, 95% ee, yellow oil. ¹H NMR (500 MHz, CDCl₃): δ 7.42

(d, *J* = 7.3 Hz, 1H, Ar-CH), 7.17 (m, 3H, Ar-CH), 3.61 (t, *J* = 5.0 Hz, 1H, CH), 2.95 (m, 1H, CH₂), 2.80 (m, 1H, CH₂), 2.71 (m, 2H, N-CH₂), 2.54 (m, 2H, N-CH₂), 2.16 (m, 1H, CH₂), 2.00 (m, 1H, CH₂), 1.70–1.90 (m, 6H, 2*CH₂+CH₂); ¹³C NMR (125 MHz, CDCl₃): δ 138.8, 137.7, 129.4, 129.1, 126.6, 125.0, 60.7, 50.4, 29.1, 24.7, 23.8, 19.4. Enantiomeric excess was determined by chiral HPLC: Chiralpak OD-H column, Hex/IPA/TFA/DEA = 92:8:0.2:0.1, 0.5mL/min, 220 nm, 10.19 min, 12.55 min.



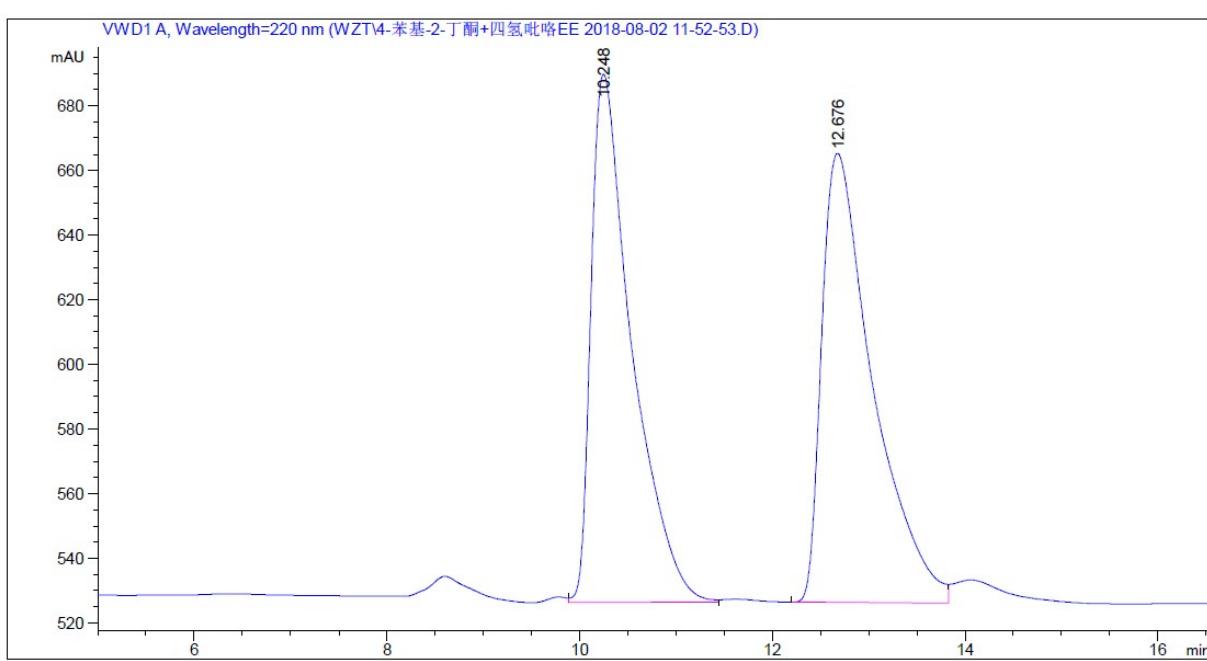
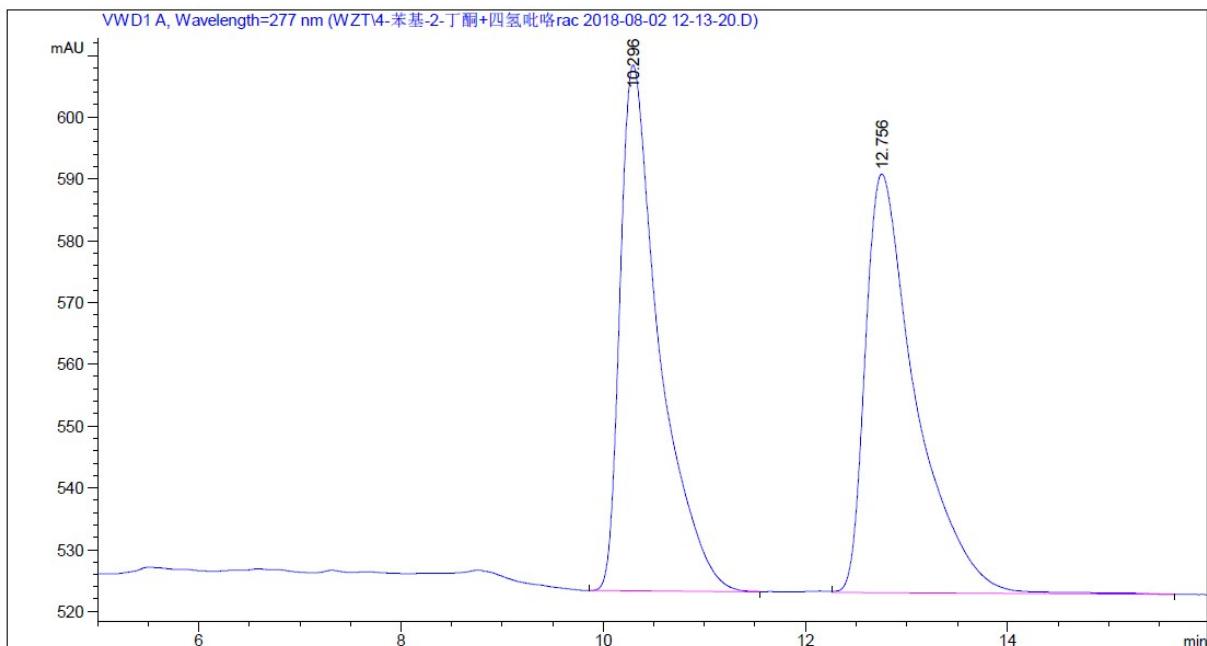
#	[min]	[min]	[mAU*s]	[mAU]	%
1	10.189	BV	0.4572	1.15778e4	356.22934
2	12.549	MM	0.5936	319.20221	8.96177
					2.6830

1-(1-(4-methoxyphenyl)propan-2-yl)pyrrolidine (3v**):**⁸ 97% yield, 6% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.12 (d, *J* = 8.6 Hz, 2H, Ar-CH), 6.86 (d, *J* = 8.6 Hz, 2H, Ar-CH), 3.83 (s, 3H, O-CH₃), 3.10 (dd, *J* = 8.6, 13.5 Hz, 1H, CH), 2.70 (m, 4H, 2*N-CH₂), 2.52 (m, 1H, CH₂), 2.41 (m, 1H, CH₂), 1.85 (m, 4H, 2*CH₂), 1.02 (d, *J* = 6.3 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 158.2, 130.7, 130.3, 113.9, 61.6, 55.2, 51.2, 39.9, 23.5, 16.9. Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



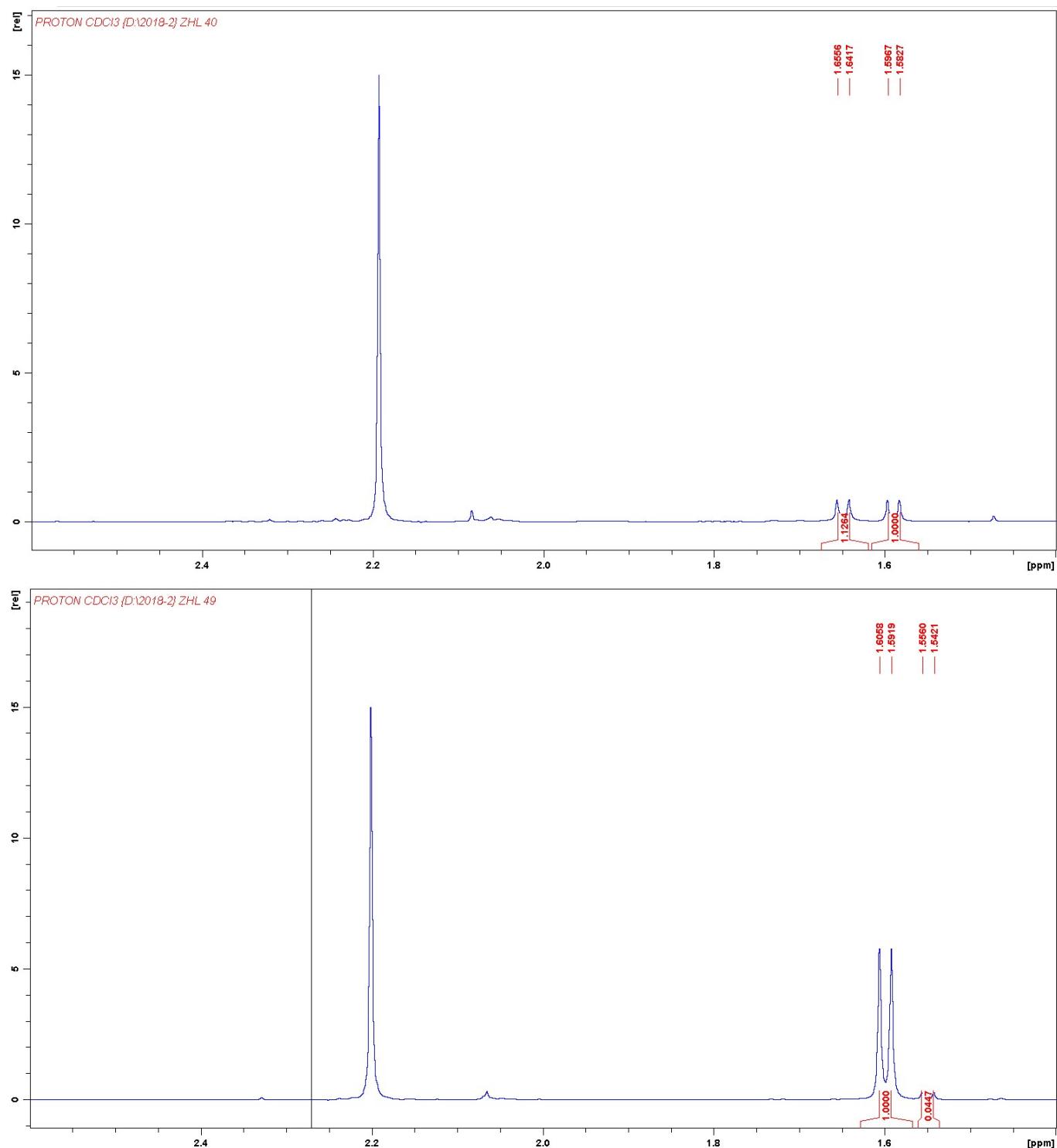
1-(4-phenylbutan-2-yl)pyrrolidine (3w):⁹ 95% yield, 3% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.32 (m, 2H, Ar-CH),

7.25 (m, 3H, Ar-CH), 2.78 (m, 1H, CH₂), 2.62 (m, 5H, 2*N-CH₂+CH₂), 2.41 (m, 1H, CH), 1.98 (m, 1H, CH₂), 1.82 (m, 4H, 2*CH₂), 1.74 (m, 1H, CH₂), 1.21 (d, *J* = 6.4 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 141.1, 128.6, 128.3, 126.2, 59.3, 50.8, 35.2, 32.0, 23.6, 16.4. Enantiomeric excess was determined by chiral HPLC: Chiralpak OD-H column, Hex/IPA/TFA/DEA = 92:8:0.2:0.1, 0.5mL/min, 220 nm, 10.26 min, 12.68 min.

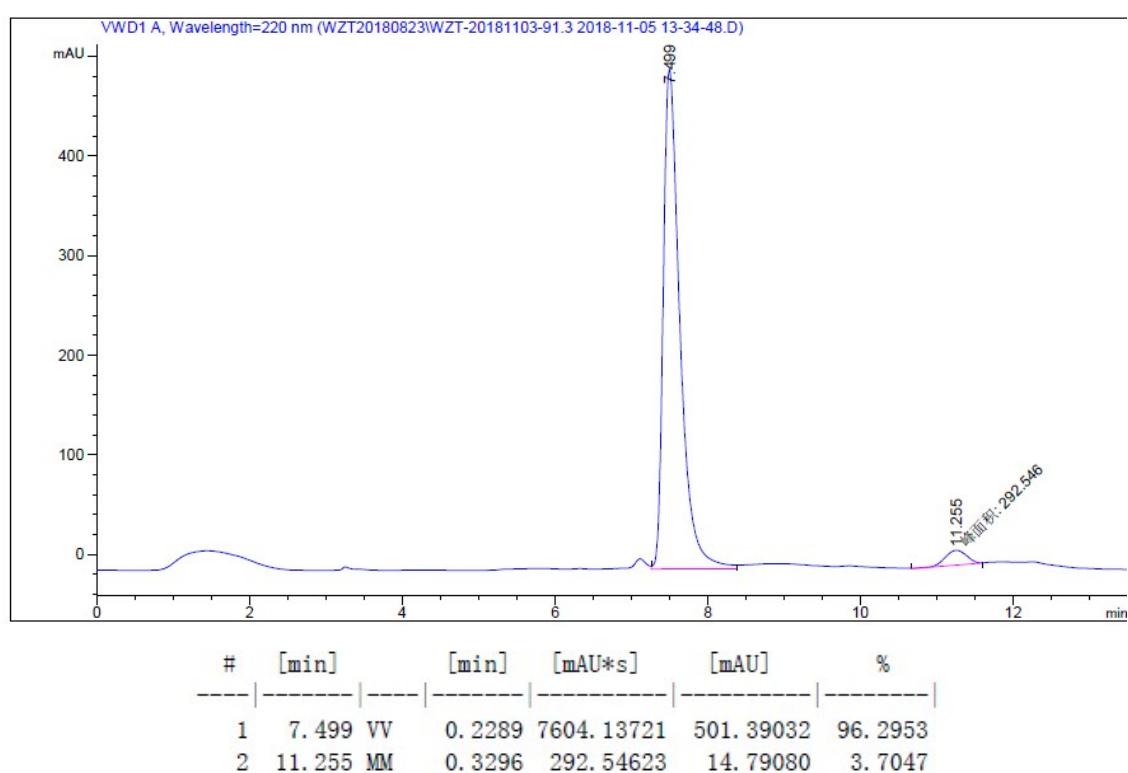
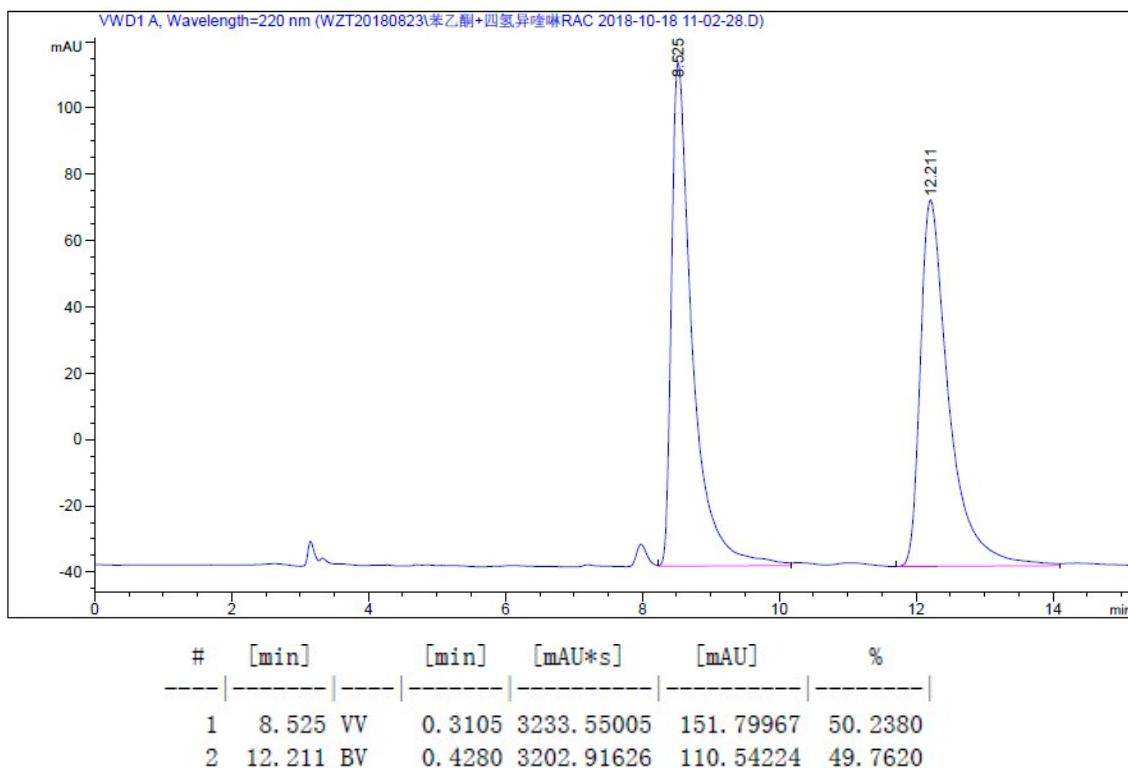


#	[min]	[min]	[mAU*s]	[mAU]	%
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2	12.676	BV	0.5411	5110.53809	138.94679 51.5315

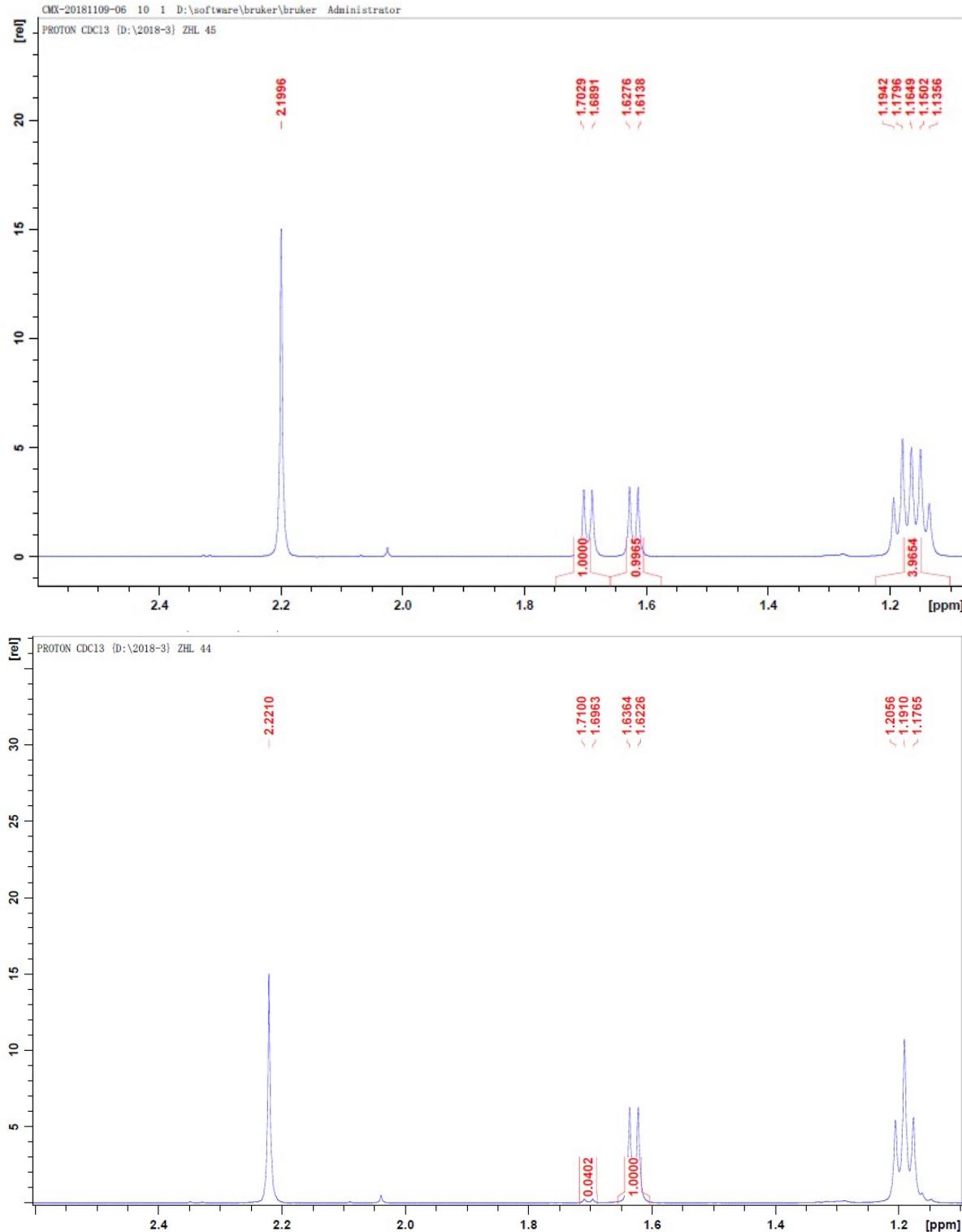
4-(1-phenylethyl)morpholine (3ab):² 89% yield, 92% *ee*, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.34(m, 4H, Ar-CH), 7.28 (m, 1H, Ar-CH), 3.72 (m, 4H, 2*O-CH₂), 3.34 (q, *J* = 6.7 Hz, 1H, CH), 2.52 (m, 2H, N-CH₂), 2.41 (m, 2H, N-CH₂), 1.40 (d, *J* = 6.7 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 144.0, 128.3, 127.7, 127.0, 67.3, 65.4, 51.3, 19.9. Enantiomeric excess was determined by ¹H NMR using (*R*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



2-(1-phenylethyl)-1,2,3,4-tetrahydroisoquinoline (3ac):¹⁰ 89% yield, 92% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.54 (d, *J* = 7.3 Hz, 2H, Ar-CH), 7.48 (t, *J* = 7.4 Hz, 2H, Ar-CH), 7.40 (m, 1H, Ar-CH), 7.25 (m, 3H, Ar-CH), 7.13 (m, 1H, Ar-CH), 3.98 (d, *J* = 14.8 Hz, 1H, N-CH₂-Ar), 3.72 (m, 2H, CH+N-CH₂-Ar), 2.90-3.10 (m, 3H, CH₂+CH₂), 2.77 (m, 1H, CH₂), 1.62 (d, *J* = 6.7 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 144.5, 135.4, 134.8, 128.8, 128.5, 127.7, 127.1, 126.9, 126.1, 125.7, 64.6, 53.8, 48.2, 29.5, 20.3. Enantiomeric excess was determined by chiral HPLC: Chiralpak OJ-H column, Hex/IPA = 95:5, 1 mL/min, 220 nm, 10.52 min, 12.21 min.

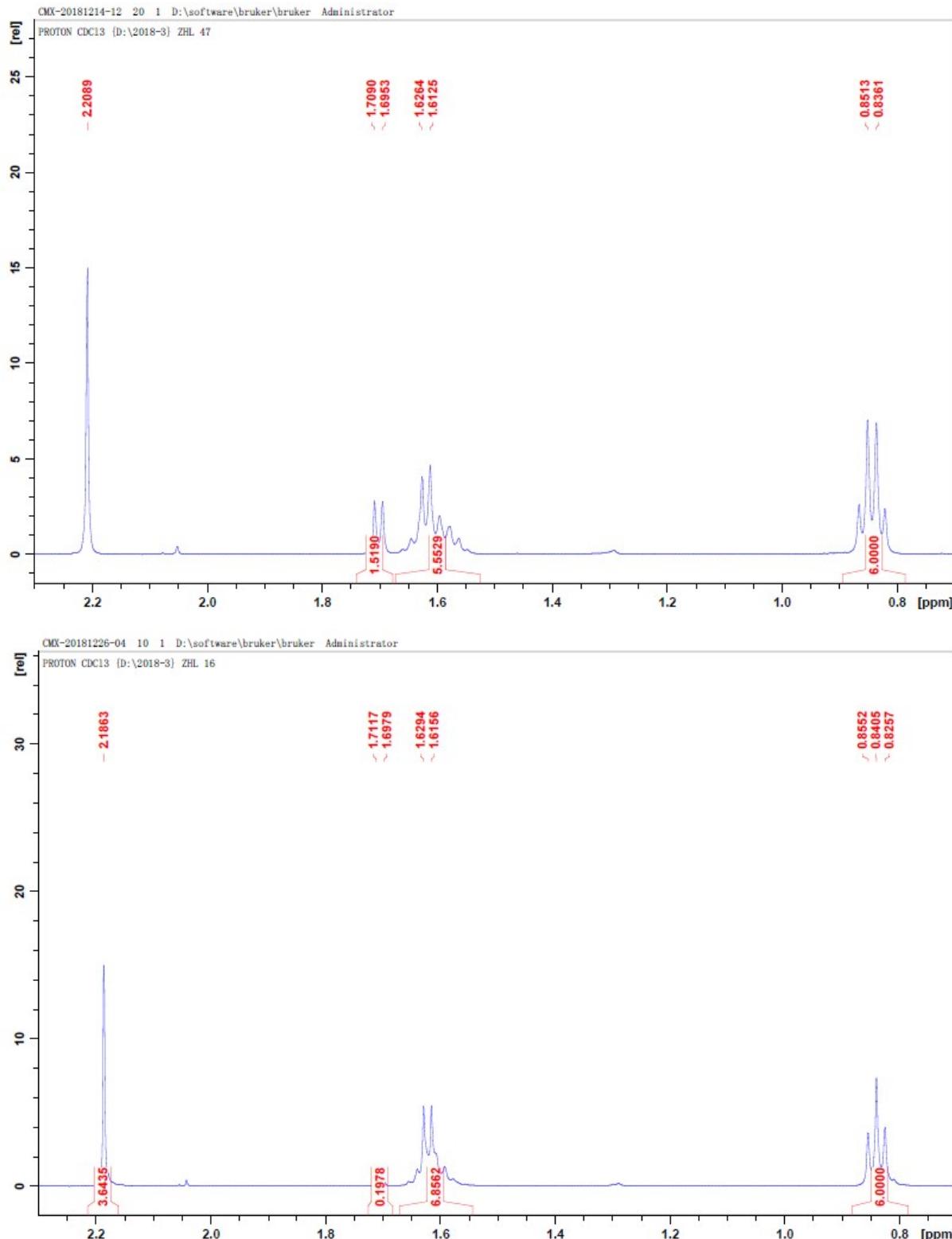


N,N-diethyl-1-phenylethan-1-amine (3ad):³ 91% yield, 92% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.40 (d, *J* = 7.4 Hz, 2H, Ar-CH), 7.35 (t, *J* = 7.4 Hz, 2H, Ar-CH), 7.25 (m, 1H, Ar-CH), 3.85 (q, *J* = 6.8 Hz, 1H, CH), 2.50–2.70 (m, 4H, 2*N-CH₂), 1.38 (d, *J* = 6.8 Hz, 3H, CH₃), 1.04 (t, *J* = 7.2 Hz, 6H, 2*CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 145.3, 128.1, 127.6, 126.5, 59.3, 42.9, 29.7, 18.4, 12.2. Enantiomeric excess was determined by ¹H NMR using (S)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.

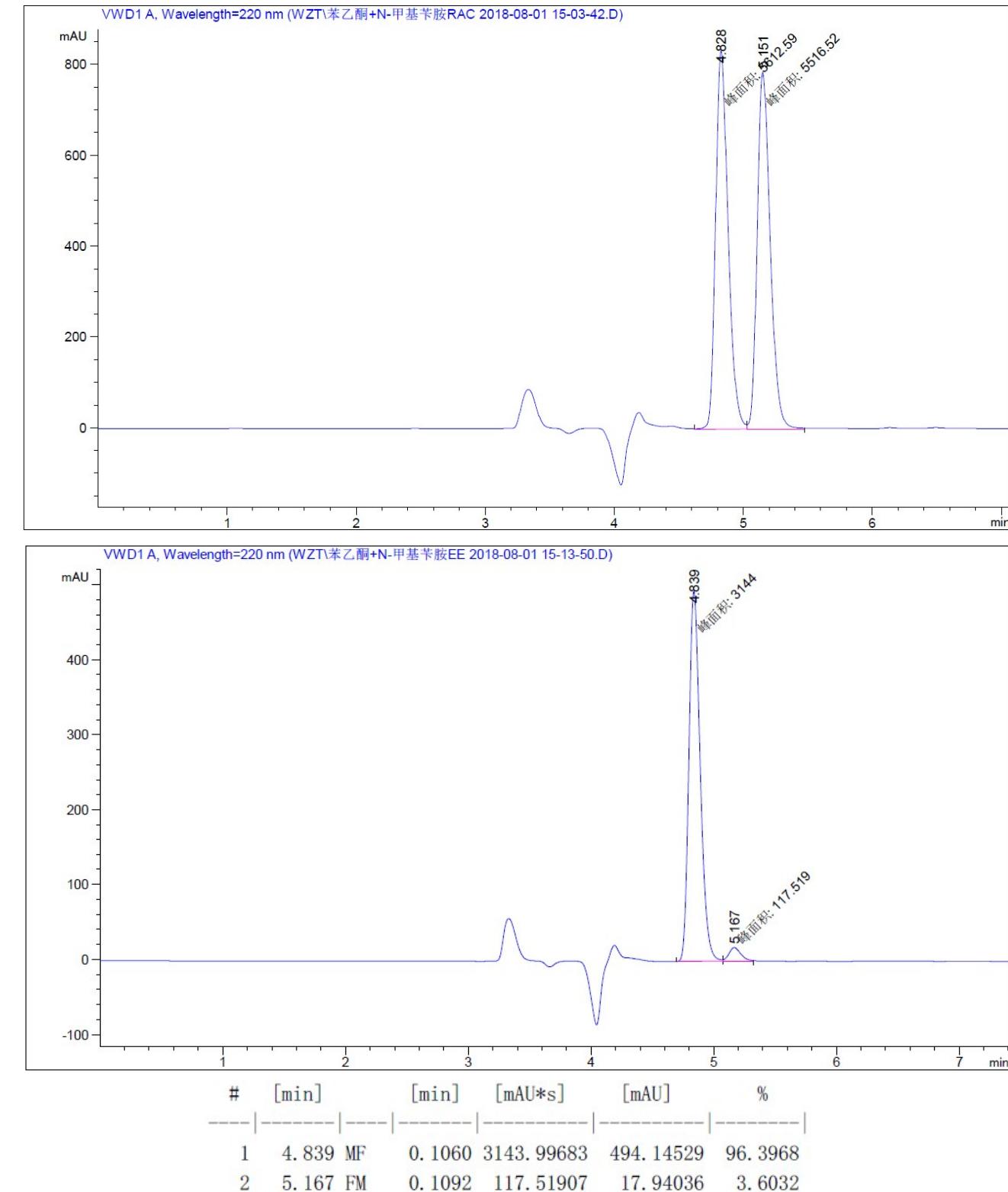


N,N-dipropyl-1-phenylethan-1-amine (3ae):¹¹ 89% yield, 87% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.40 (d, *J* = 7.5 Hz, 2H, Ar-CH), 7.34 (t, *J* = 7.4 Hz, 2H, Ar-CH), 7.25 (m, 1H, Ar-CH), 3.87 (q, *J* = 6.7 Hz, 1H, CH), 2.30–2.52 (m, 4H, 2*N-CH₂), 1.48 (m, *J* = 7.4 Hz, 4H, 2*CH₂), 1.36 (d, *J* = 6.8 Hz, 3H, CH₃), 0.86 (t, *J* = 7.2 Hz, 6H, 2*CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 145.3, 128.2, 127.9, 127.0, 59.9, 51.9, 20.2, 16.8, 11.8.

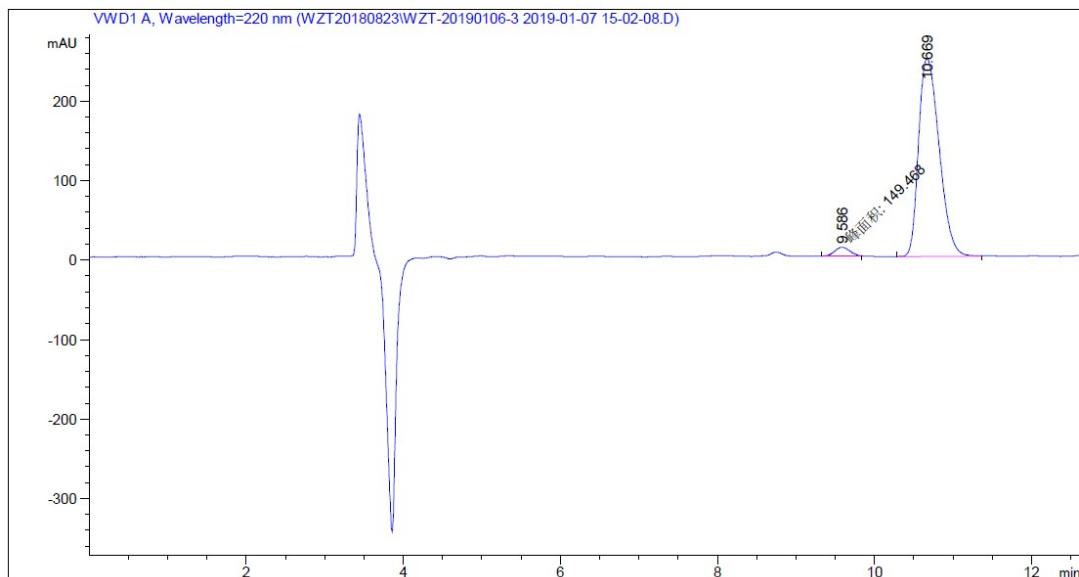
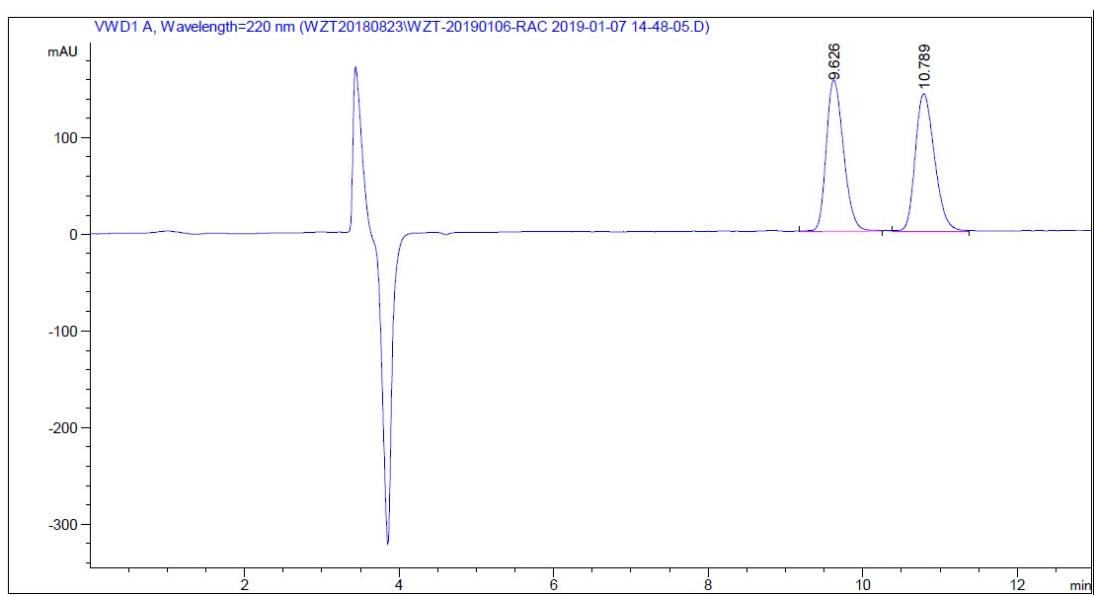
Enantiomeric excess was determined by ¹H NMR using (*S*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



N-benzyl-N-methyl-1-phenylethan-1-amine (3af).² 92% yield, 93% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.26–7.48 (m, 10H, Ar-CH), 3.68 (m, 1H, CH), 3.63 (m, 1H, CH₂), 3.37 (m, 1H, CH₂), 2.19 (m, 3H, N-CH₃), 1.48 (m, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 144.3, 140.2, 128.9, 128.3, 128.2, 127.8, 126.9, 126.8, 63.4, 59.0, 38.5, 18.5. Enantiomeric excess was determined by chiral HPLC: Chiralpak OJ-H column, Hex/IPA/TFA/DEA=90:10:0.2:0.1, 1 mL/min, 220 nm, 4.83 min, 5.15 min.

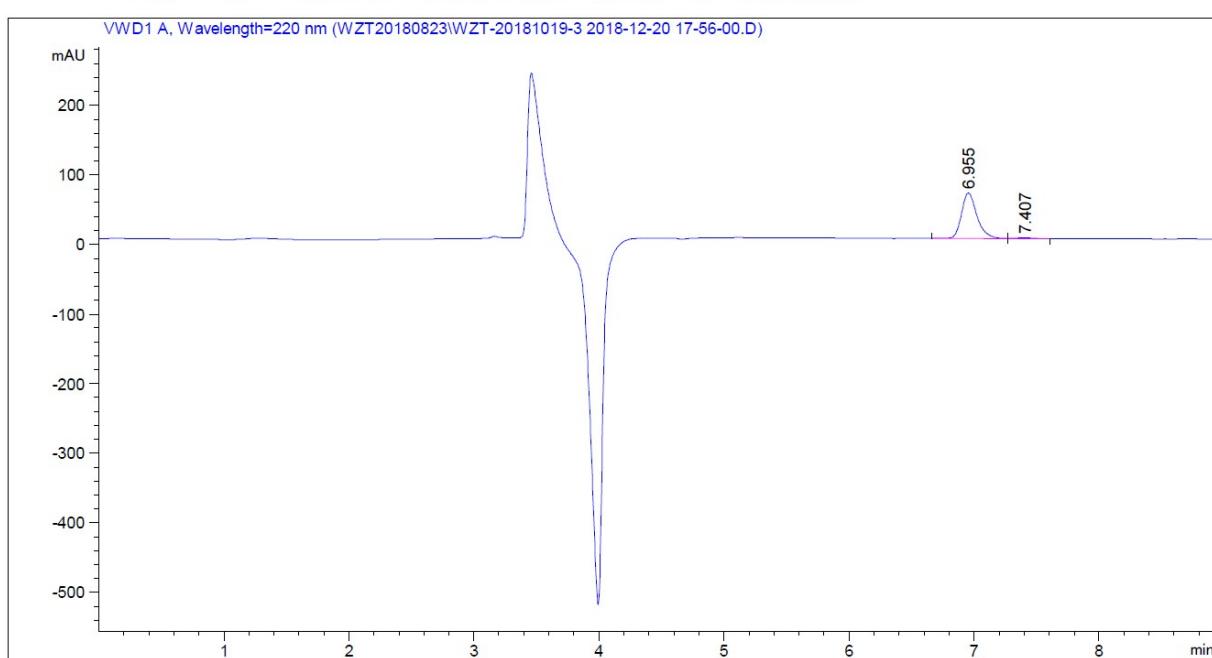
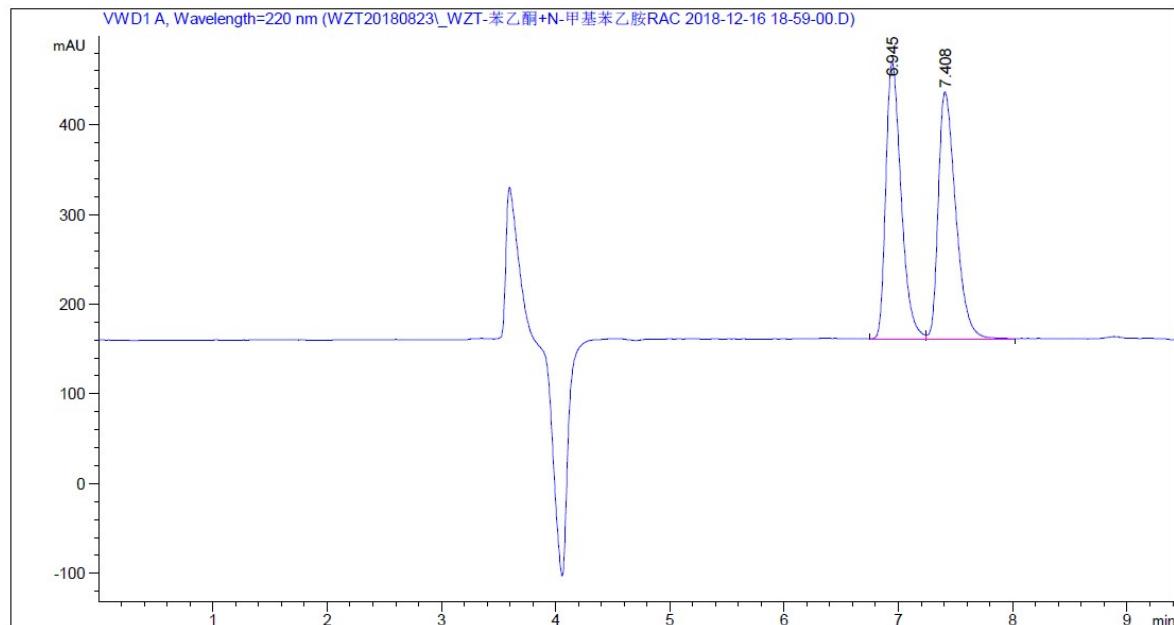


N-(4-methoxybenzyl)-N-methyl-1-phenylethan-1-amine (3ag):¹² 90% yield, 94% *ee*, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.46 (d, *J* = 7.3 Hz, 2H, Ar-CH), 7.38 (t, *J* = 7.0 Hz, 2H, Ar-CH), 7.28 (m, 3H, Ar-CH), 6.90 (d, *J* = 7.7 Hz, 2H, Ar-CH), 3.84 (s, 3H, O-CH₃), 3.68 (q, *J* = 6.1 Hz, 1H, CH), 3.56 (d, *J* = 13 Hz, 1H, CH₂), 3.30 (d, *J* = 13 Hz, 1H, CH₂), 2.17 (s, 3H, N-CH₃), 1.46 (d, *J* = 6.4 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 158.5, 144.3, 132.1, 129.9, 128.2, 127.7, 126.8, 113.6, 63.1, 58.2, 55.3, 38.2, 18.4. Enantiomeric excess was determined by chiral HPLC: Chiraldak OJ-H column, Hex/IPA/DEA=97:3:0.1, 1 mL/min, 220 nm, 9.63 min, 10.78 min.



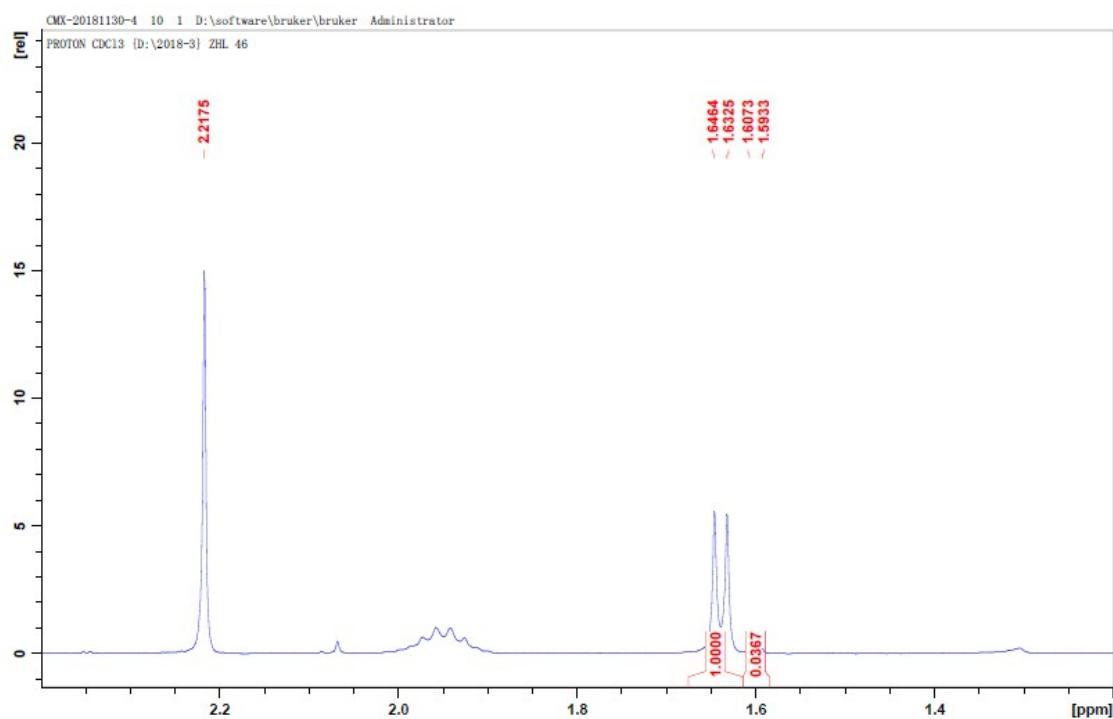
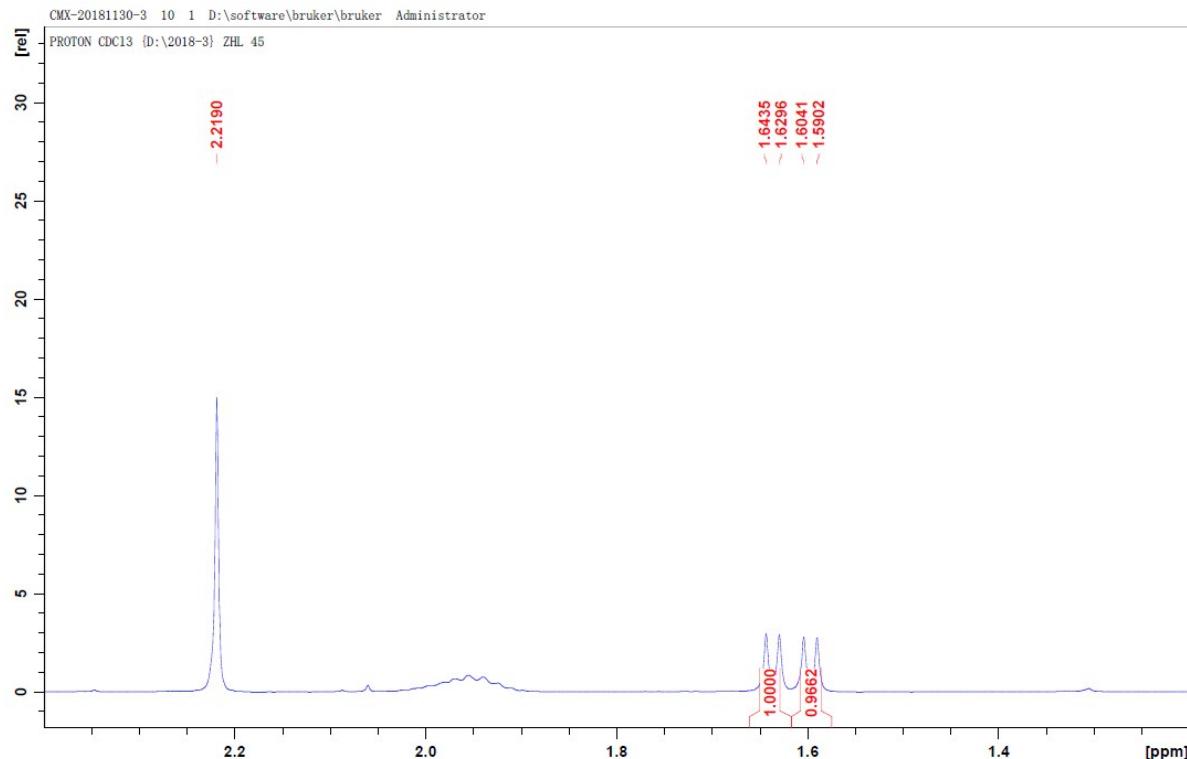
#	[min]	[min]	[mAU*s]	[mAU]	%	
1	9.586	MM	0.2221	149.46837	11.21587	3.1838
2	10.669	VV	0.2890	4545.12305	249.00073	96.8162

N-methyl-N-phenethyl-1-phenylethan-1-amine (3ah):¹³ 96% yield, 96% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.16-7.40 (m, 10H, Ar-CH), 3.70 (q, J = 6.7 Hz, 1H, CH), 2.60-2.90 (m, 4H, 2*CH₂), 2.37 (s, 3H, N-CH₃), 1.44 (d, J = 6.8 Hz, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃): δ 142.6, 140.1, 128.8, 128.4, 128.3, 127.9, 127.3, 126.1, 63.6, 56.2, 38.5, 33.3, 18.7. Enantiomeric excess was determined by chiral HPLC: Chiralpak OJ-3 column, Hex/IPA/DEA=97:3:0.1, 1 mL/min, 220 nm, 6.94 min, 7.41 min.

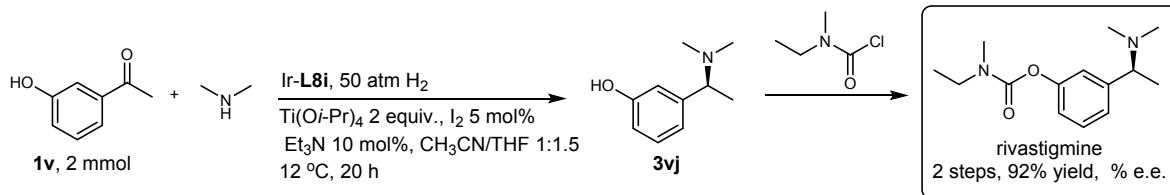


#	[min]	[min]	[mAU*s]	[mAU]	%
1	6.955	BV	0.1307	564.60931	65.42831 97.9282
2	7.407	VB	0.1385	11.94481	1.33325 2.0718

N-methyl-N-phenpropyl-1-phenylethan-1-amine (3ai): 96% yield, 93% *ee*, brown oil. ^1H NMR (500 MHz, CDCl_3): δ 7.16-7.40 (m, 10H), 3.62 (q, $J = 6.8$ Hz, 1H), 2.35-2.73 (m, 4H), 2.24 (s, 3H), 1.84 (m, $J = 7.6$ Hz, 2H), 1.40 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 144.12, 142.6, 128.4, 128.3, 128.1, 127.4, 126.7, 125.7, 63.3, 53.9, 38.4, 33.6, 29.0, 18.4. Enantiomeric excess was determined by ^1H NMR using (*S*)-2-acetoxy-2-phenylacetic acid as chemical shift reagent.



III Procedure for the Synthesis of rivastigmine



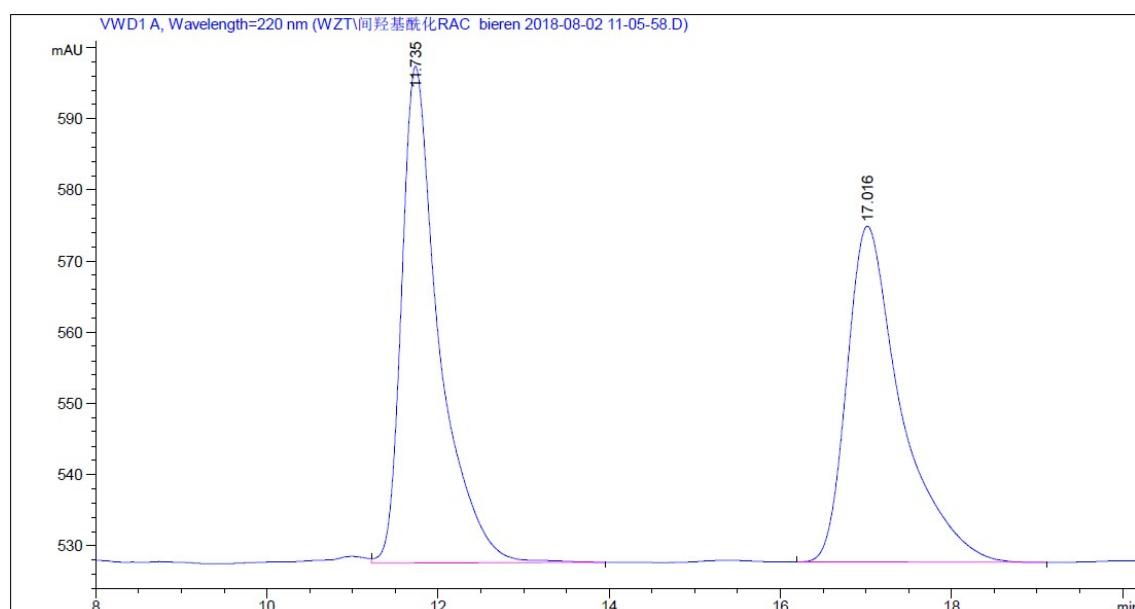
Step 1:

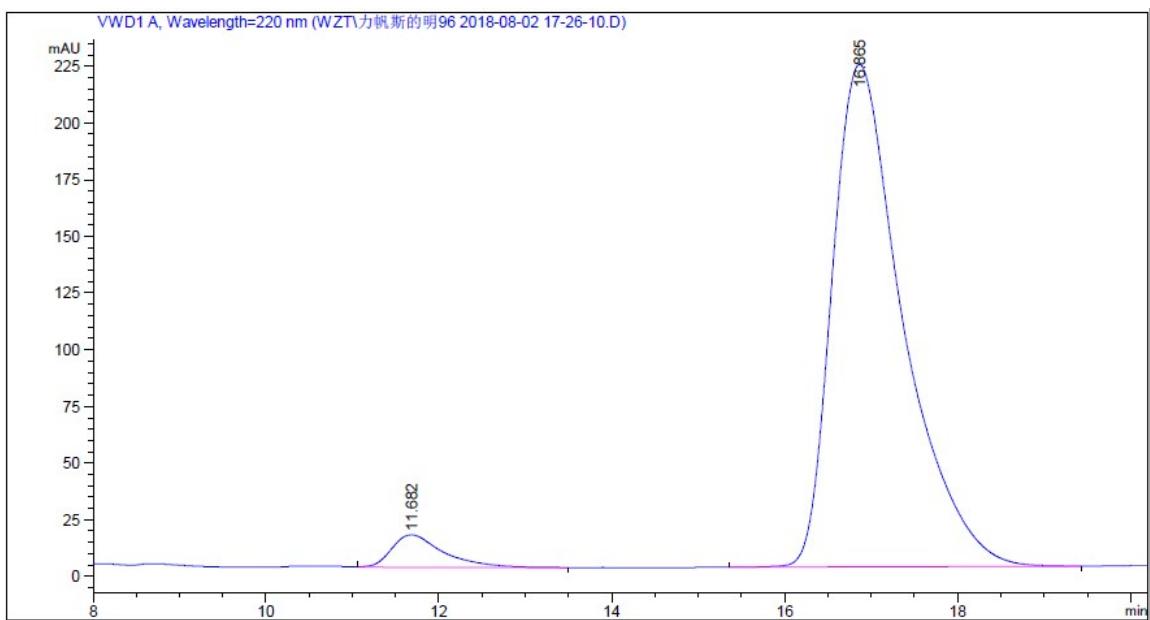
In a nitrogen-filled glovebox, the *in situ* generated complex Ir-L8i (20 μ mol) was added to the vial charged with ketones (2 mmol) and dimethylamine in THF (2 mol/L, 2 mL, 4 mmol) in anhydrous acetonitrile solution (5.0 mL). Ti(O*i*-Pr)₄ (2 equiv.), Et₃N (0.1 eq), and I₂ (5 mol%) were added and the total solution was made to 15 mL at a ratio CH₃CN/THF 1:1.5. The resulting vials were transferred to an autoclave, which was charged with 50 atm of H₂, and stirred at 12 °C for 20 h. The hydrogen gas was released slowly and the solution was quenched with aqueous sodium bicarbonate solution. The organic phase was concentrated and passed through a short column of silica gel to remove the metal complex to give the crude products **3vj**, which were used in next step without purification.

Step 2:

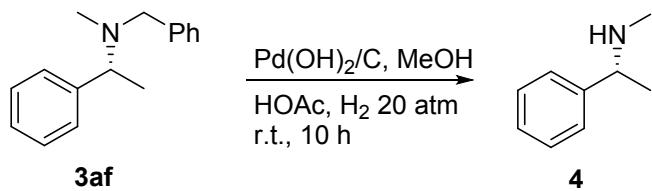
NaOH (2.5 mmol) was added to the solution of **3vj** in CH₃CN (15 mL). The above solution was stirred for 1 h at room temperature, then ethyl(methyl)carbamic chloride (2.2 mmol) was added. The reaction mixture was stirred for 12 h, quenched by aq. NH₄Cl solution. CH₃CN was removed under vacuum and the solution was extracted by EtOAc (15 mL*3). The organic phase was dried over anhydrous Na₂SO₄, concentrated and purified with column chromatography (EtOAc/PE) to give rivastigmine (461 mg, 92% yield for 2 steps).

(S)-3-[1-(Dimethylamino)ethyl]phenyl Ethyl-(methyl)carbamate (Rivastigmine):¹⁰ 93% yield for 2 steps, 91% ee, brown oil. ¹H NMR (500 MHz, CDCl₃): δ 7.32 (t, *J* = 7.9 Hz, 1H), 7.15 (d, *J* = 7.7 Hz, 1H), 7.10 (m, 1H), 7.05 (m, 1H), 3.50 (dq, *J* = 6.7 Hz, 31.2 Hz, 2H), 3.28 (t, *J* = 6.7 Hz, 1H), 3.05 (d, *J* = 37.2 Hz, 1H), 2.24 (s, 6H), 1.39 (d, *J* = 6.7 Hz, 3H), 1.27 (dt, *J* = 7.7 Hz, 26.9 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 154.6, 151.6, 145.8, 128.9, 124.2, 120.7, 120.3, 65.6, 44.0, 43.2, 34.2, 33.8, 20.1, 13.2, 12.5. HRMS (ESI) m/z calcd for C₁₄H₂₃N₂O₂⁺ [M+H]⁺ 251.17540, found 251.17538. Enantiomeric excess was determined by chiral HPLC: Chiralpak OD-H column, Hex/IPA/TFA/DEA = 92:8:0.2:0.1, 1 mL/min, 220 nm, 11.74 min, 17.02 min.





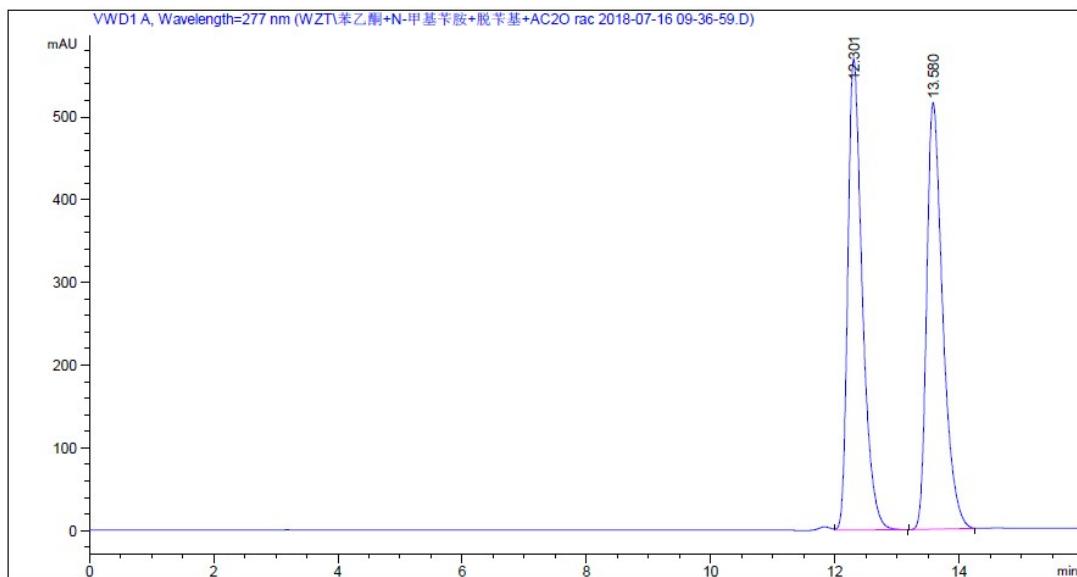
IV Debenzylation of 3ae



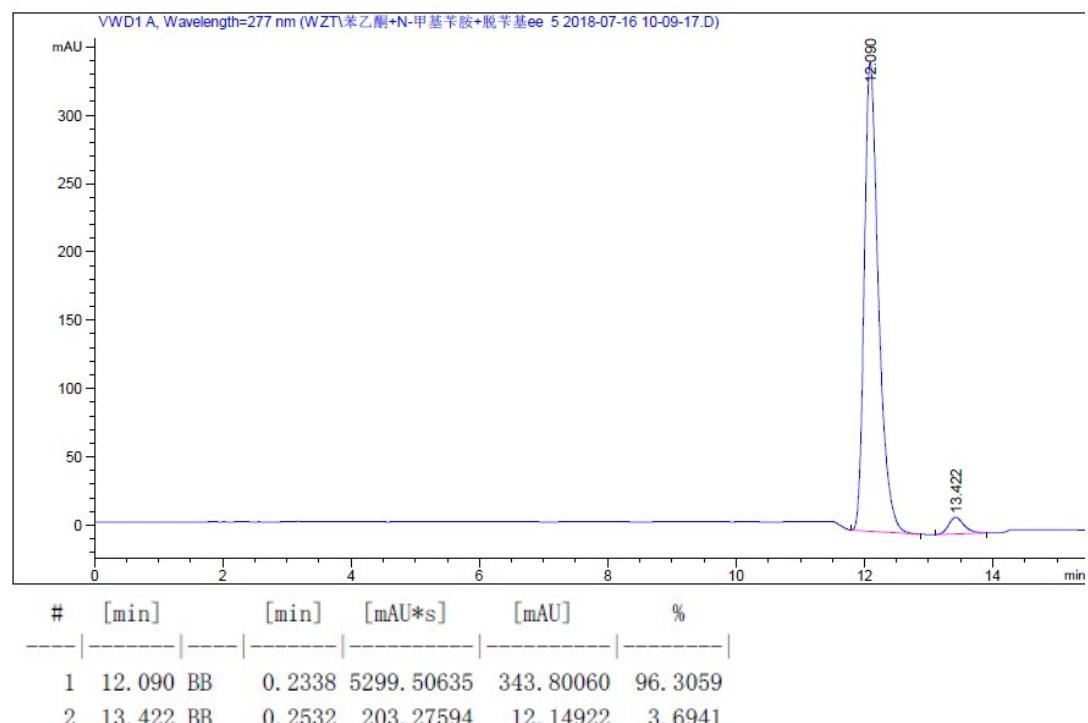
Pd(OH)_2 on carbon (10%, 4.5 mg) was added to the solution of (*R*)-3af (45 mg, 0.2 mmol) and acetic acid (1 drop) in MeOH (2 mL). The resulting mixture was transferred to an autoclave, which was charged with 20 atm of H_2 , and stirred at r.t. for 10 h. The hydrogen gas was released slowly and the solution was filtered and quenched with aqueous sodium bicarbonate solution. The organic phase was concentrated and passed through a short column of silica gel to remove the metal complex to give the crude product, which was purified with column chromatography to afford 4 (26 mg, 95% yield).

(*R*)-N-methyl-1-phenylethan-1-amine (4):¹¹ 26 mg, 95% yield, 93% *ee*, colorless oil. ^1H NMR (500 MHz, CDCl_3): δ 7.27–7.36 (m, 4H, Ar-CH), 7.23 (m, 1H, Ar-CH), 3.67 (q, $J = 6.6$ Hz, 1H, CH), 2.34 (s, 3H, N-CH₃), 1.86 (bs, 1H, NH), 1.39 (d, $J =$

6.7 Hz, 3H, CH₃). Enantiomeric excess was determined for the corresponding acetamide by chiral HPLC: Chiraldak AD-H column, Hex/IPA=97:3, 1 mL/min, 220 nm, 12.3 min (major), 13.6 min (minor).



#	[min]	[min]	[mAU*s]	[mAU]	%
1	12.301	VB	0.2451	9230.26953	569.60980 49.9640
2	13.580	BB	0.2712	9243.55957	515.57379 50.0360



#	[min]	[min]	[mAU*s]	[mAU]	%
1	12.090	BB	0.2338	5299.50635	343.80060 96.3059
2	13.422	BB	0.2532	203.27594	12.14922 3.6941

V Mechanistic study

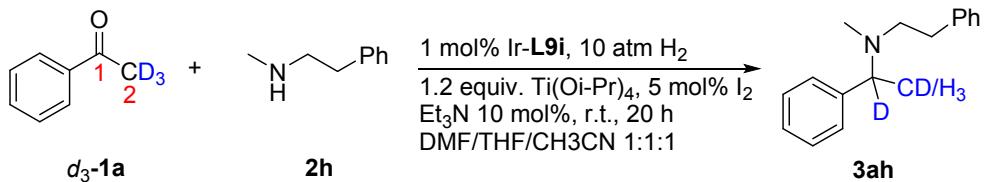
1. ^{31}P spectra.

^{31}P NMR for **L9i** (202 MHz, CDCl_3): δ 147.5;

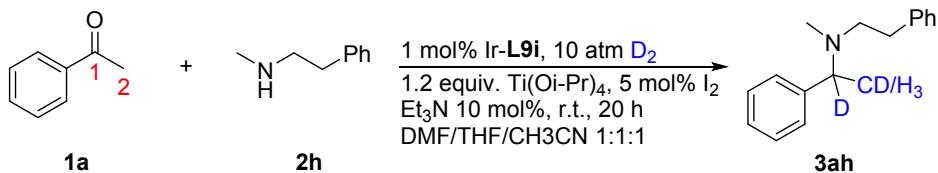
^{31}P NMR for Ir:**L9i** 1:1 (202 MHz, CDCl_3): δ 113.2;

^{31}P NMR for Ir:**L9i** 1:2 (202 MHz, CDCl_3): δ 147.5, 113.2;

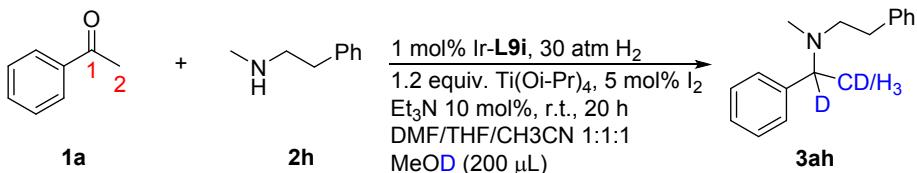
2. Deuterium incorporation study for the DARA of **1a** with **2h**.



N-methyl-N-phenethyl-1-phenylethan-1-amine-d (3ah):¹³ 82% yield, 94% ee, brown oil. ^1H NMR (500 MHz, CDCl_3): δ 7.16-7.40 (m, 10H), 3.70 (q, $J = 6.7$ Hz, 0.93H), 2.60-2.90 (m, 4H), 2.37 (s, 3H), 1.44 (m, $J = 6.8$ Hz, 1.76H).



N-methyl-N-phenethyl-1-phenylethan-1-amine-d (3ah):¹³ 82% yield, 94% ee, brown oil. ^1H NMR (500 MHz, CDCl_3): δ 7.16-7.40 (m, 10H), 3.70 (q, $J = 6.7$ Hz, 0.28H), 2.60-2.90 (m, 4H), 2.37 (s, 3H), 1.44 (m, $J = 6.8$ Hz, 1.16H).



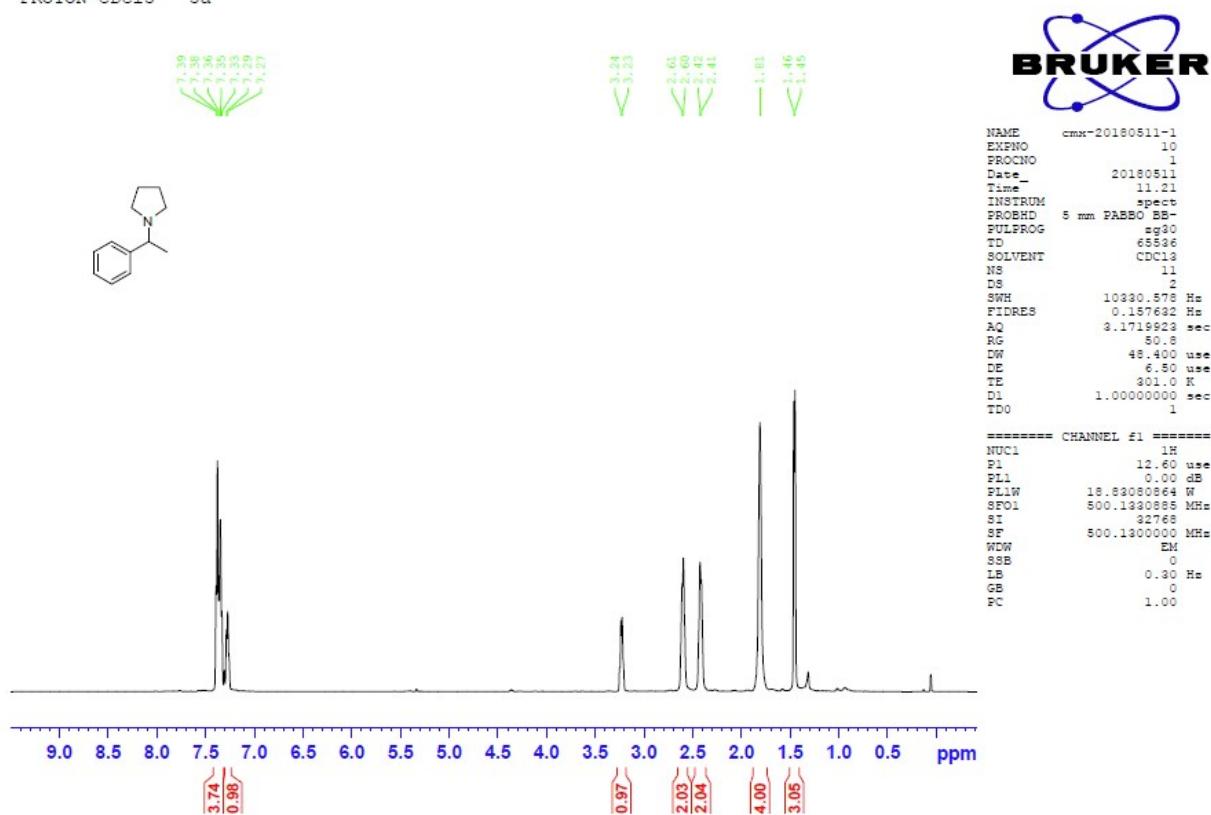
N-methyl-N-phenethyl-1-phenylethan-1-amine-d (3ah):¹³ 67% yield, 93% ee, brown oil. ^1H NMR (500 MHz, CDCl_3): δ 7.16-7.40 (m, 10H), 3.70 (q, $J = 6.7$ Hz, 0.81H), 2.60-2.90 (m, 4H), 2.37 (s, 3H), 1.44 (m, $J = 6.8$ Hz, 0.66 H).

VI References:

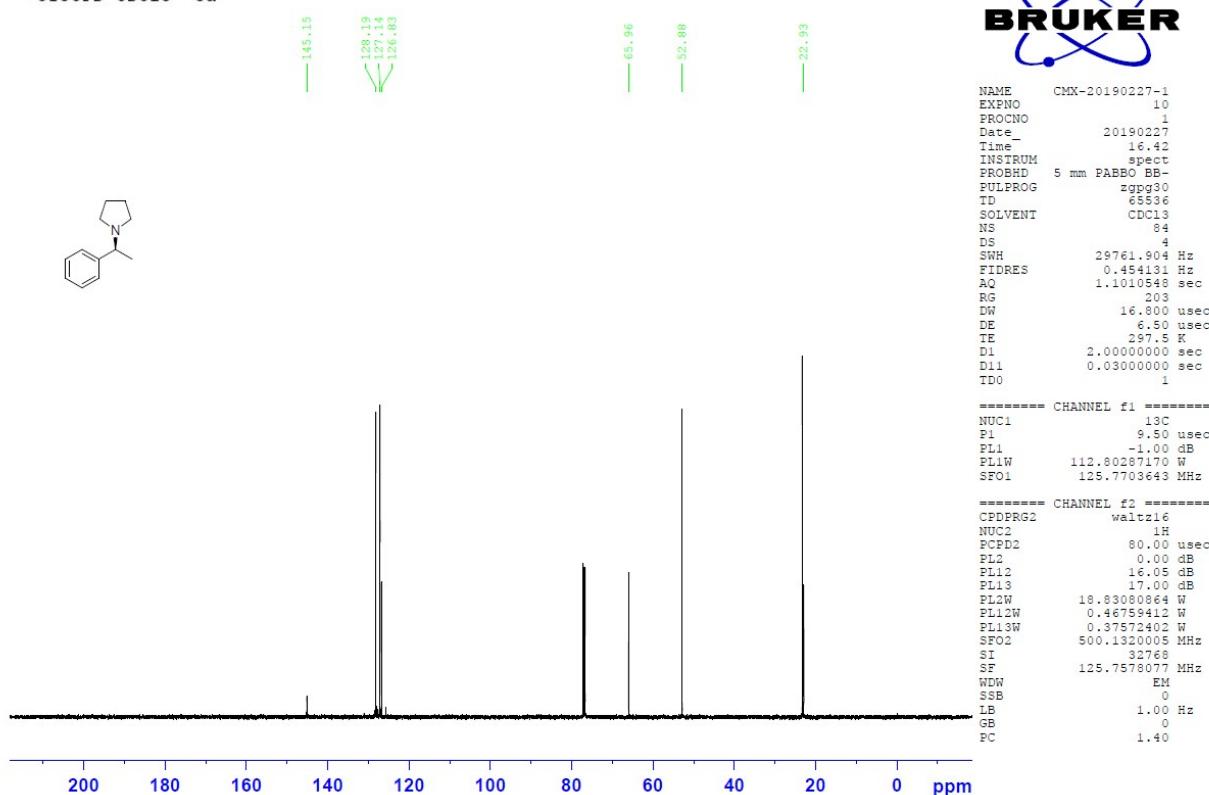
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VII NMR & HRMS Spectra

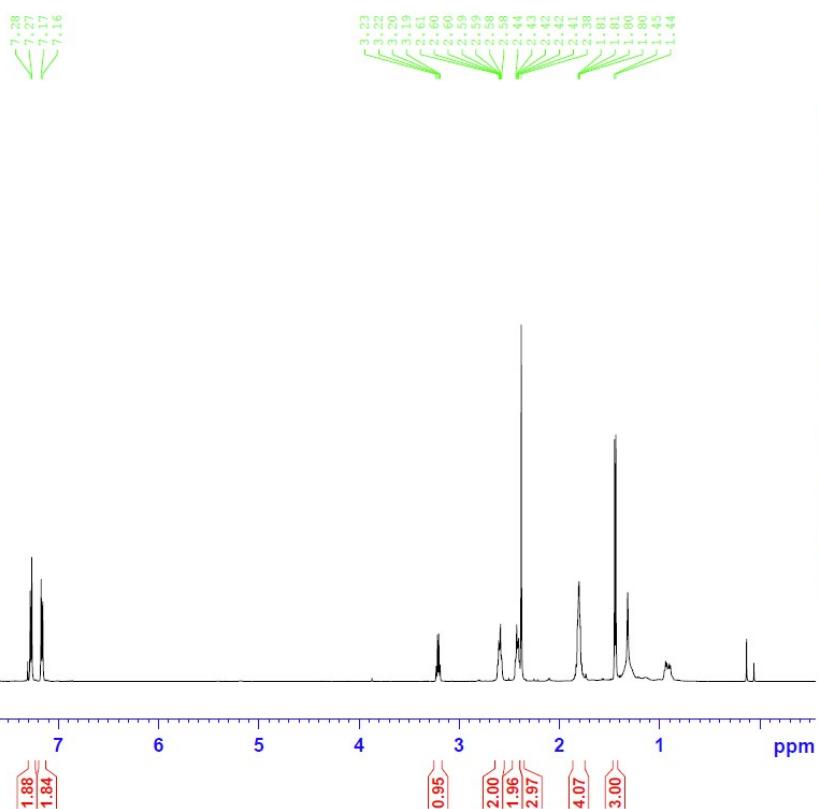
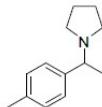
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C¹³CPD CDCl₃ 3a



PROTON CDC13 3b



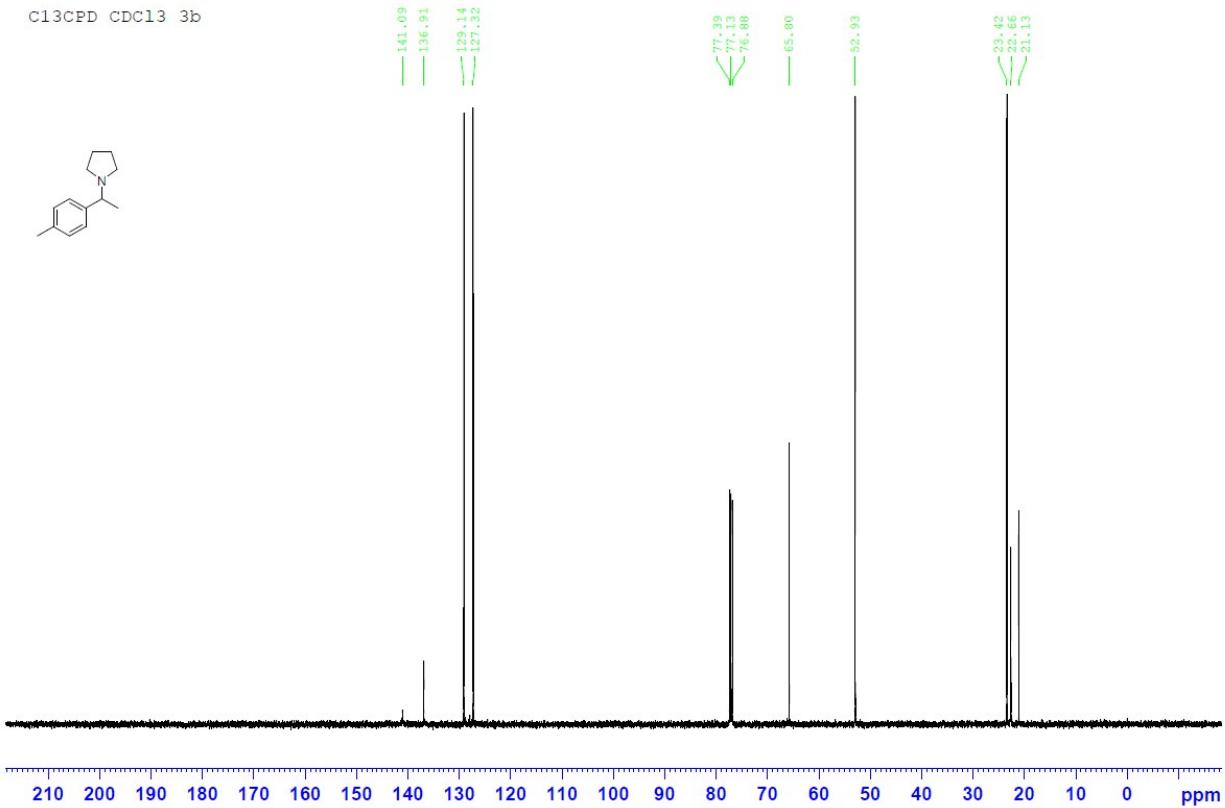
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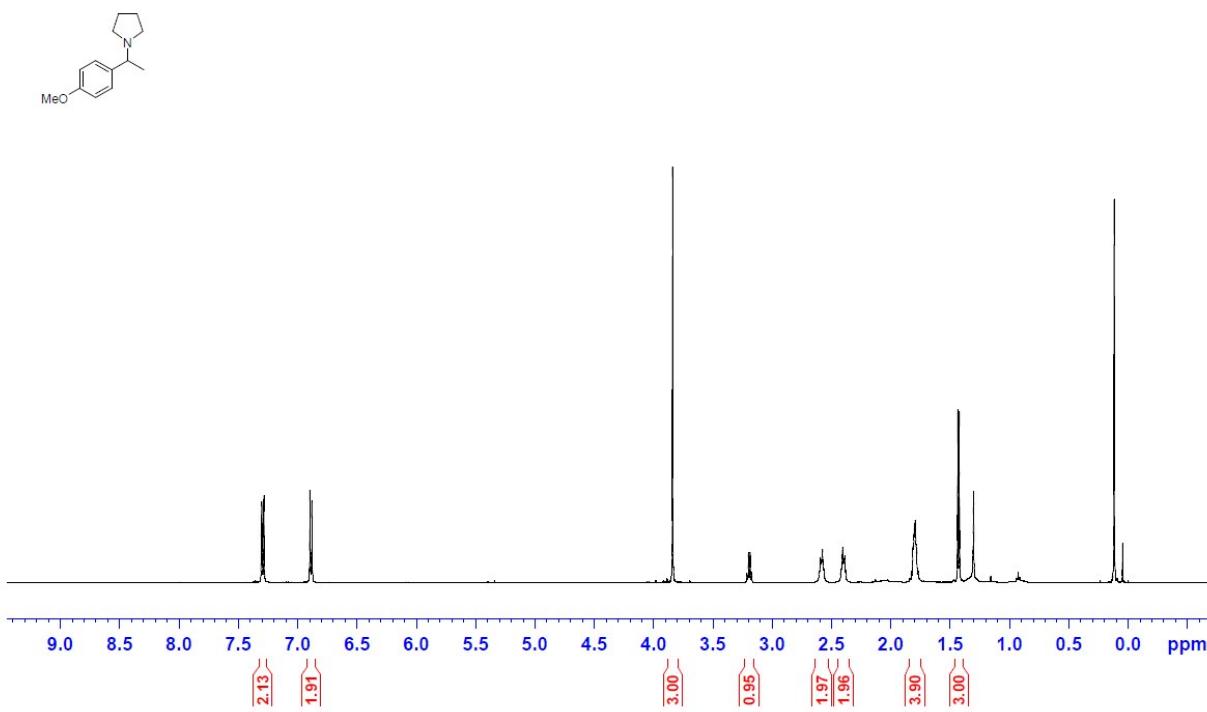
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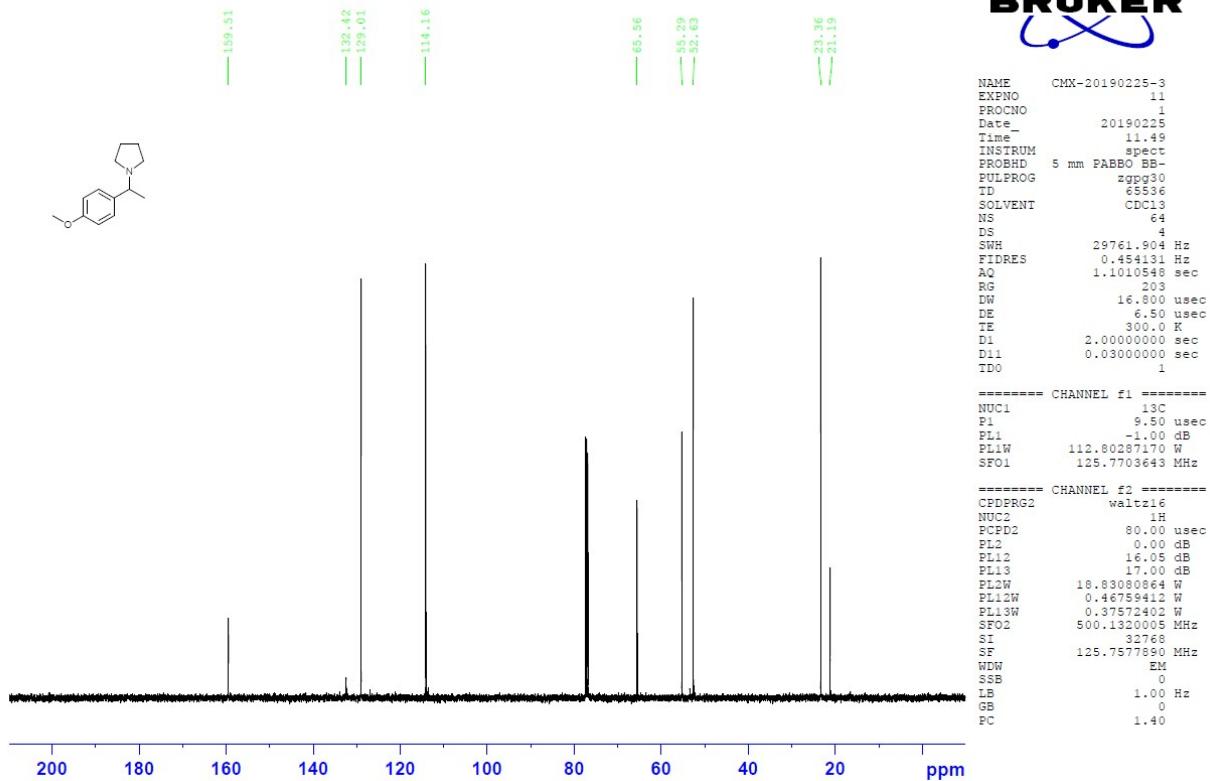
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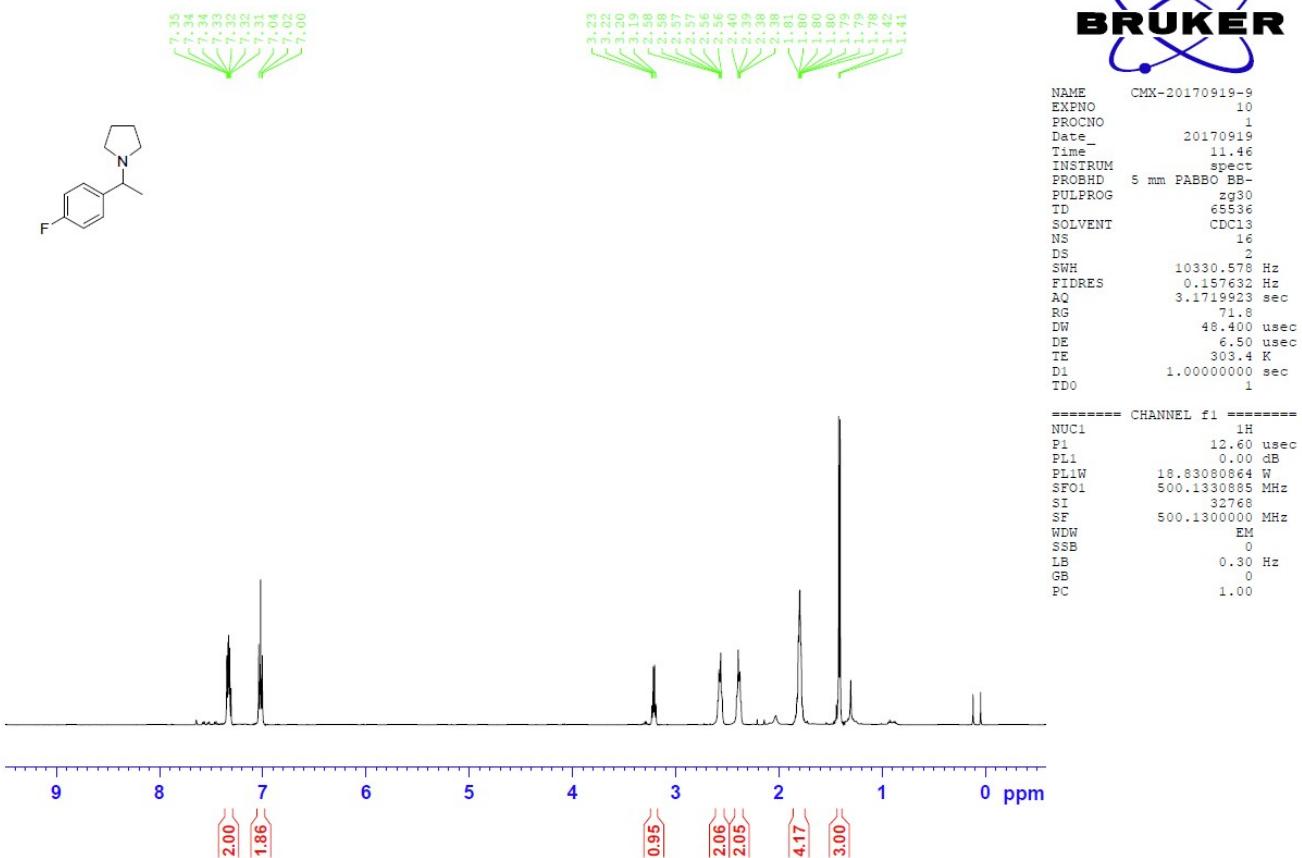
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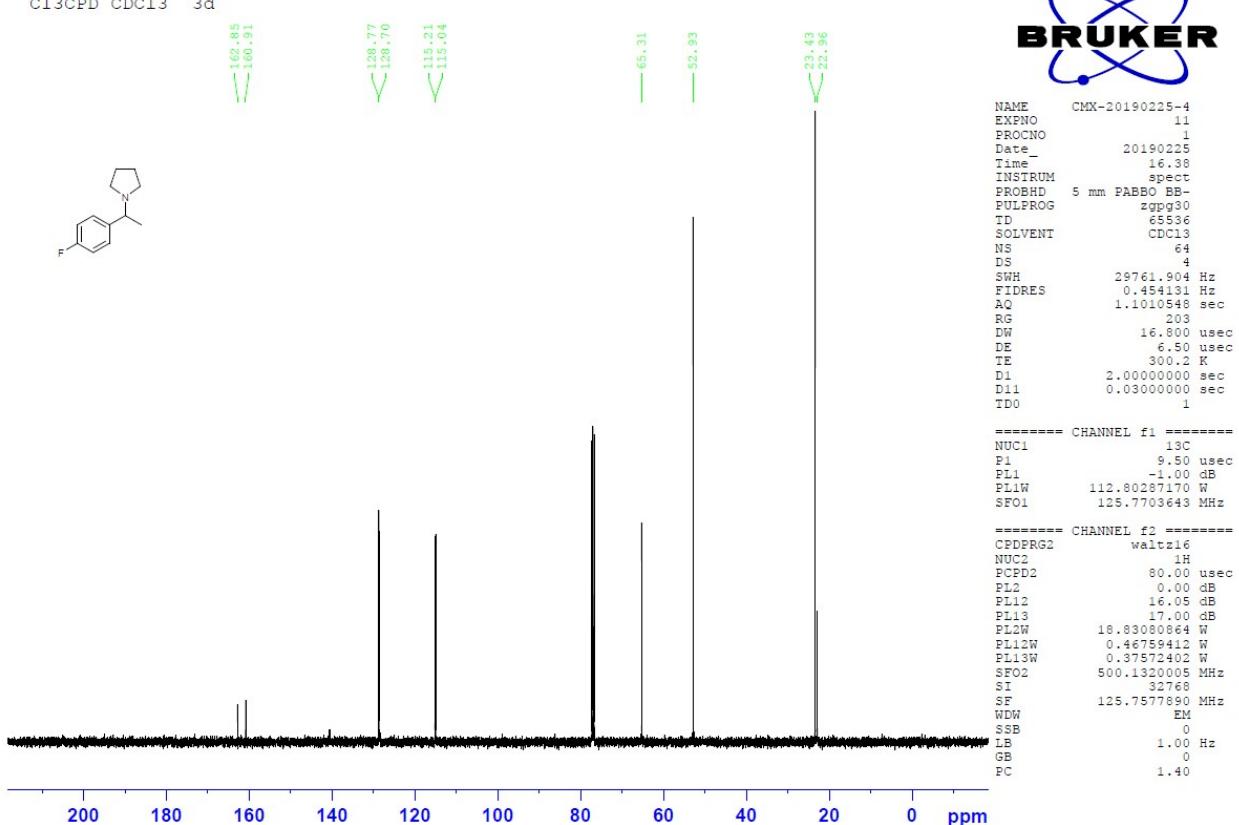
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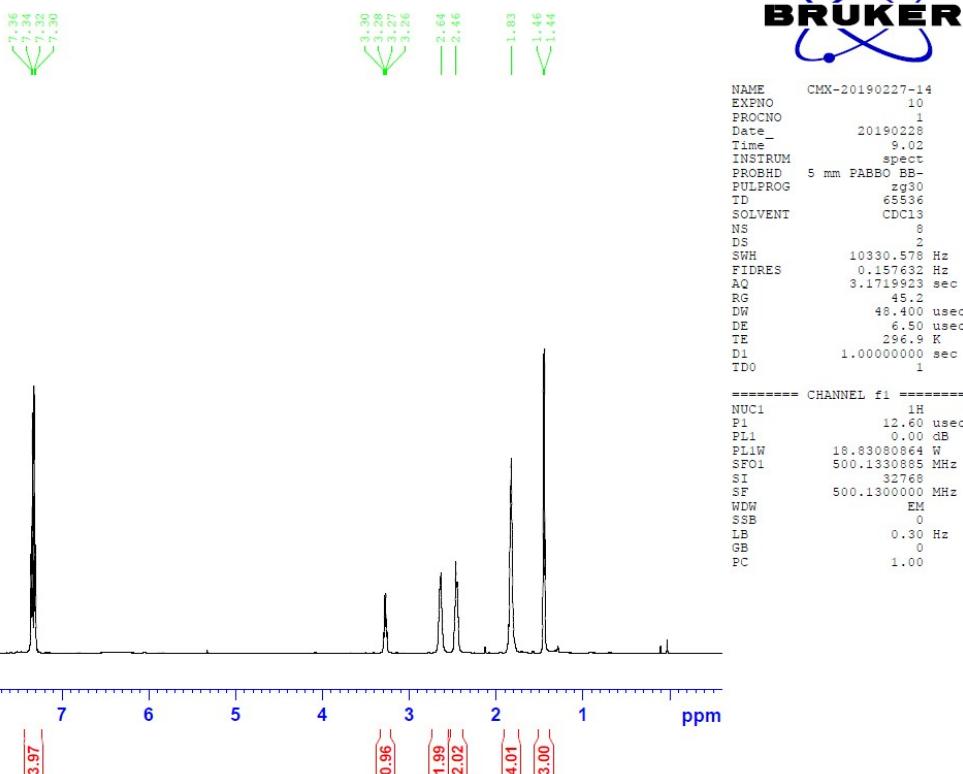
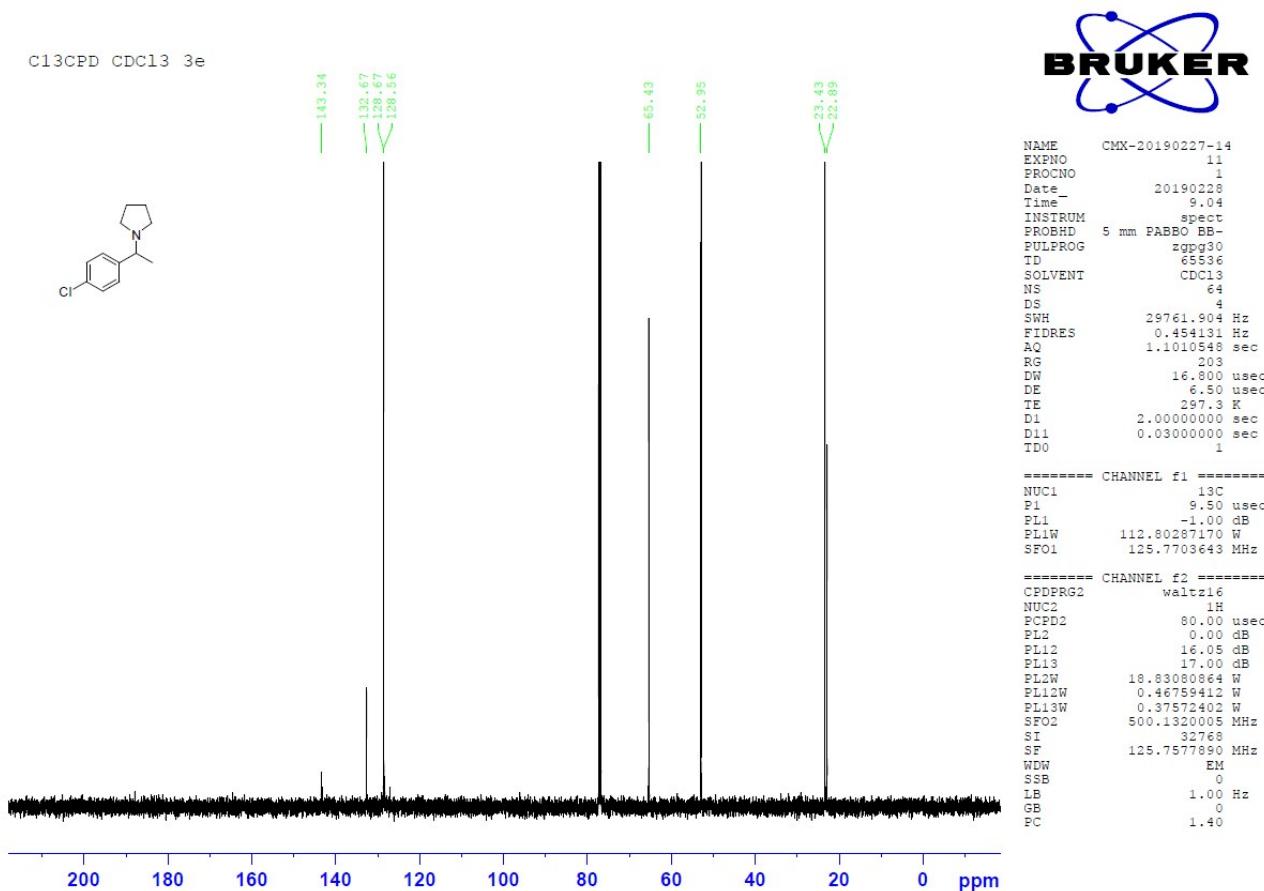


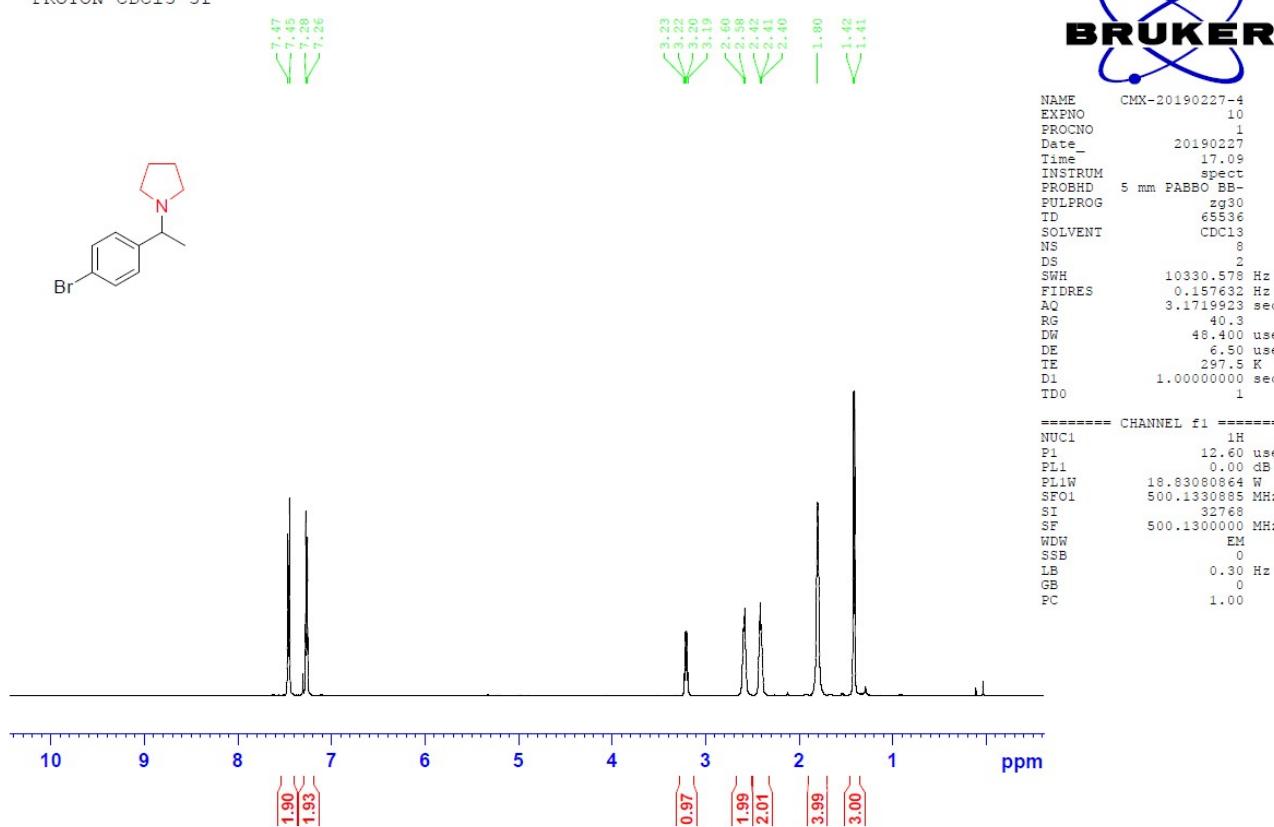
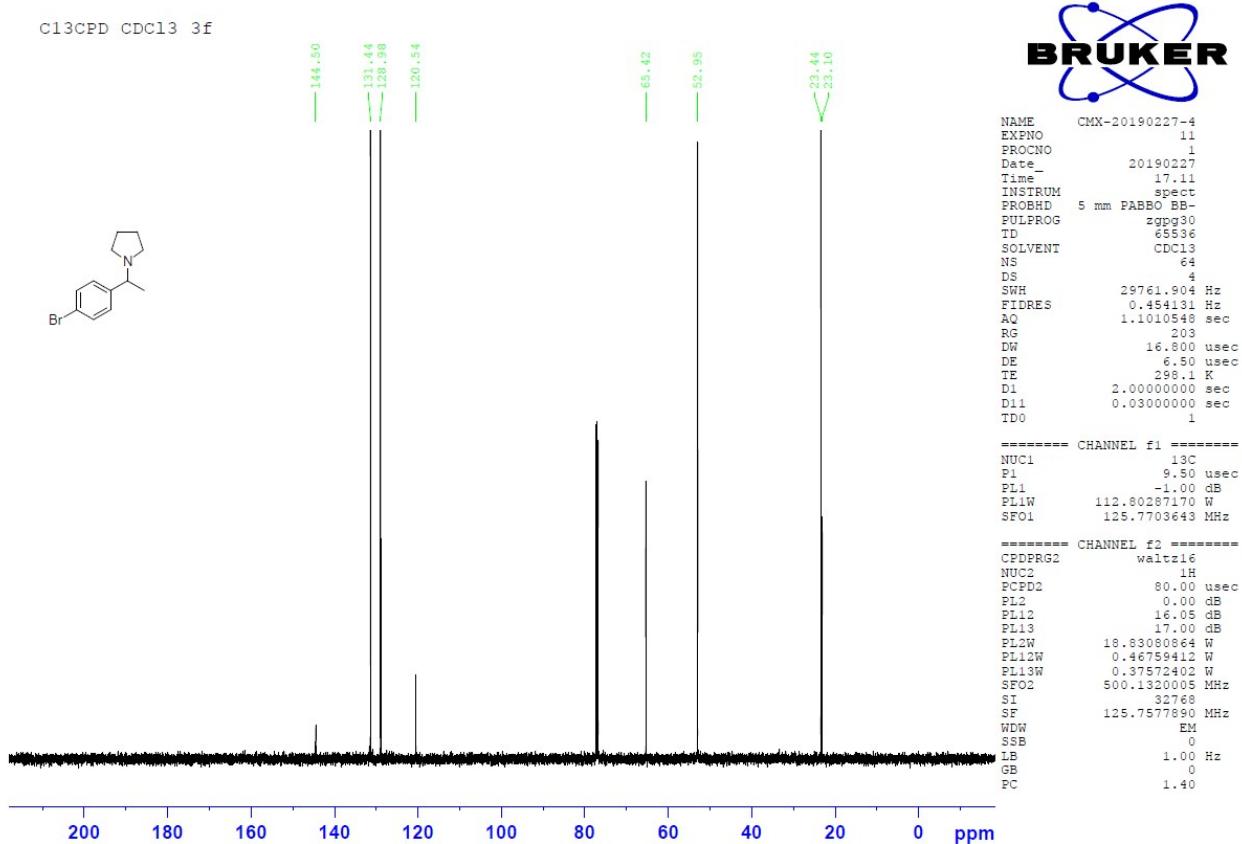
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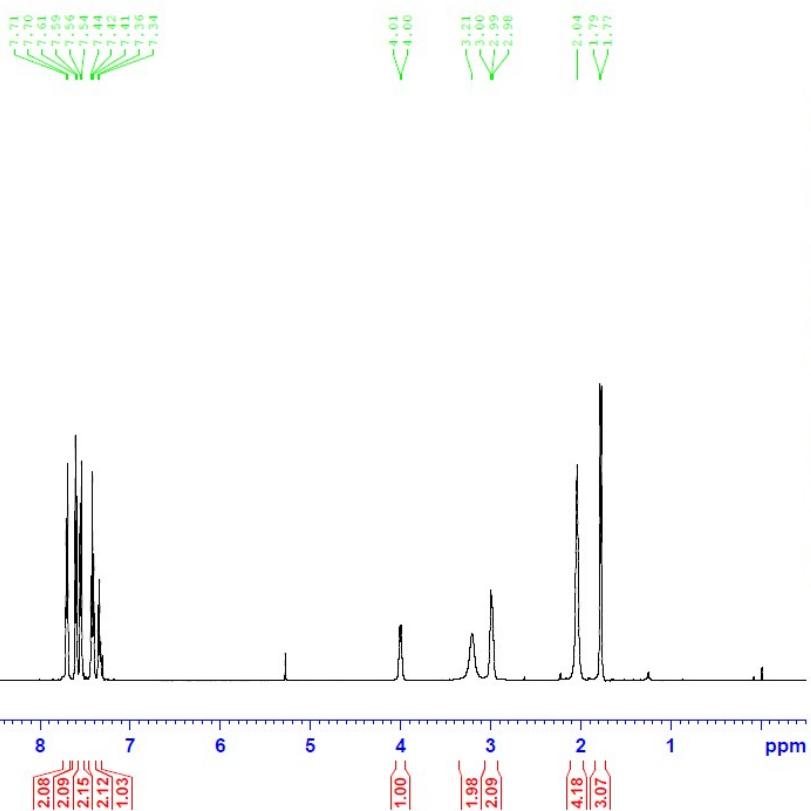


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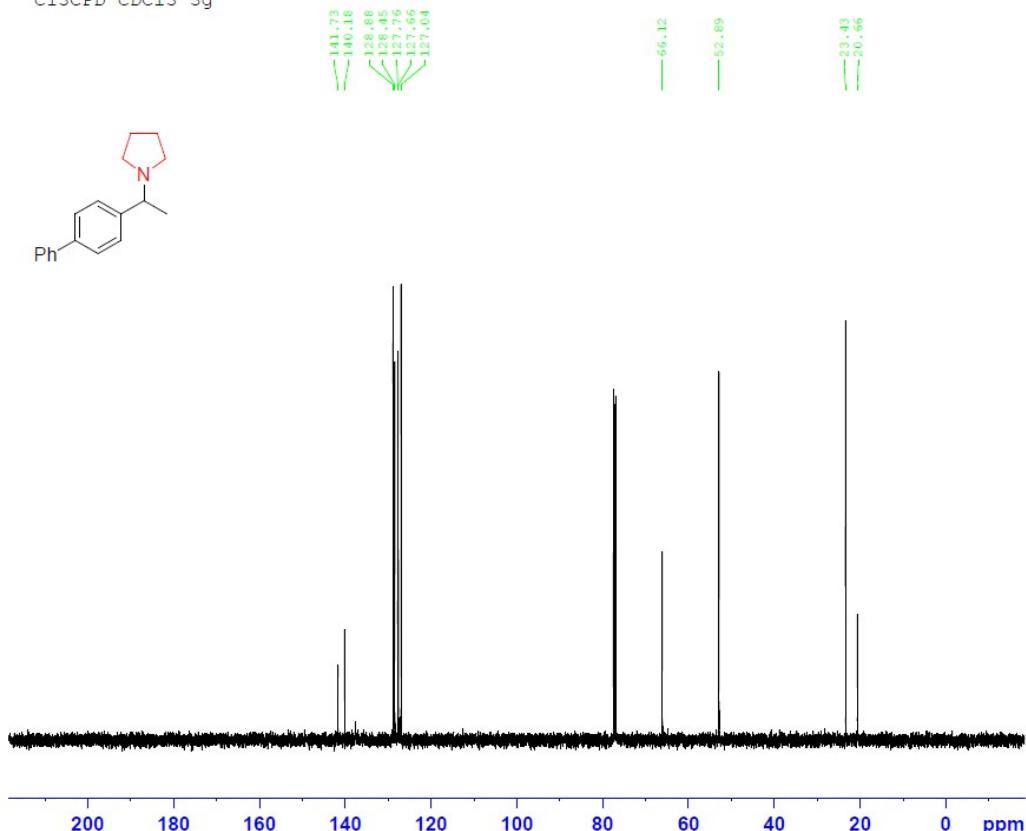
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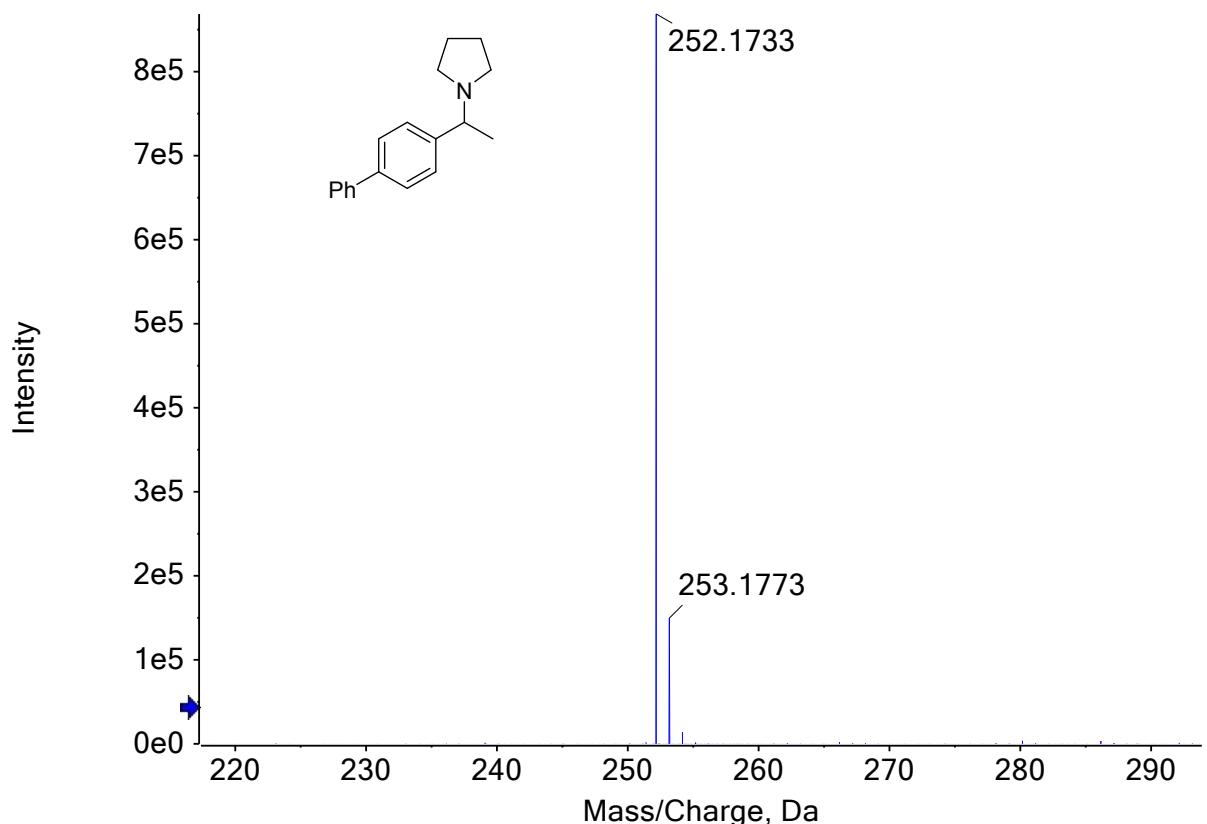
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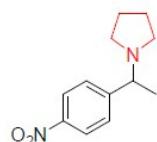
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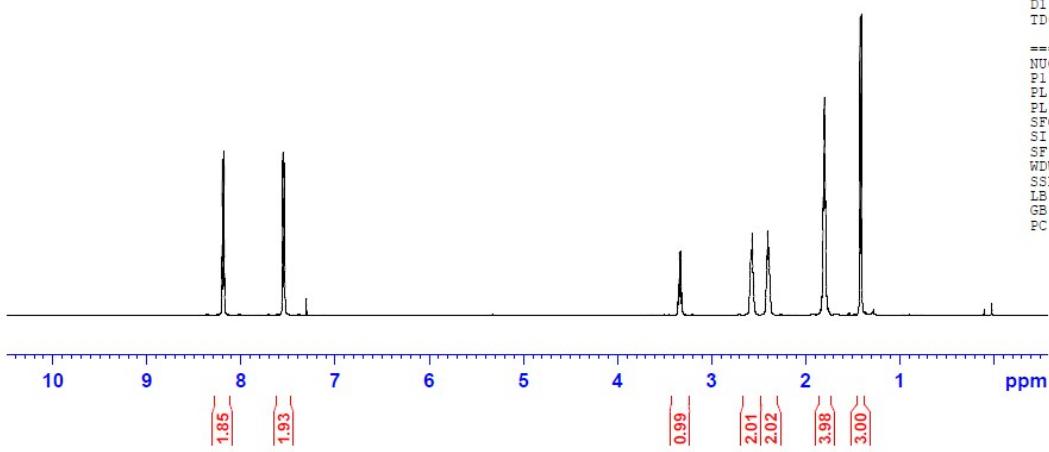
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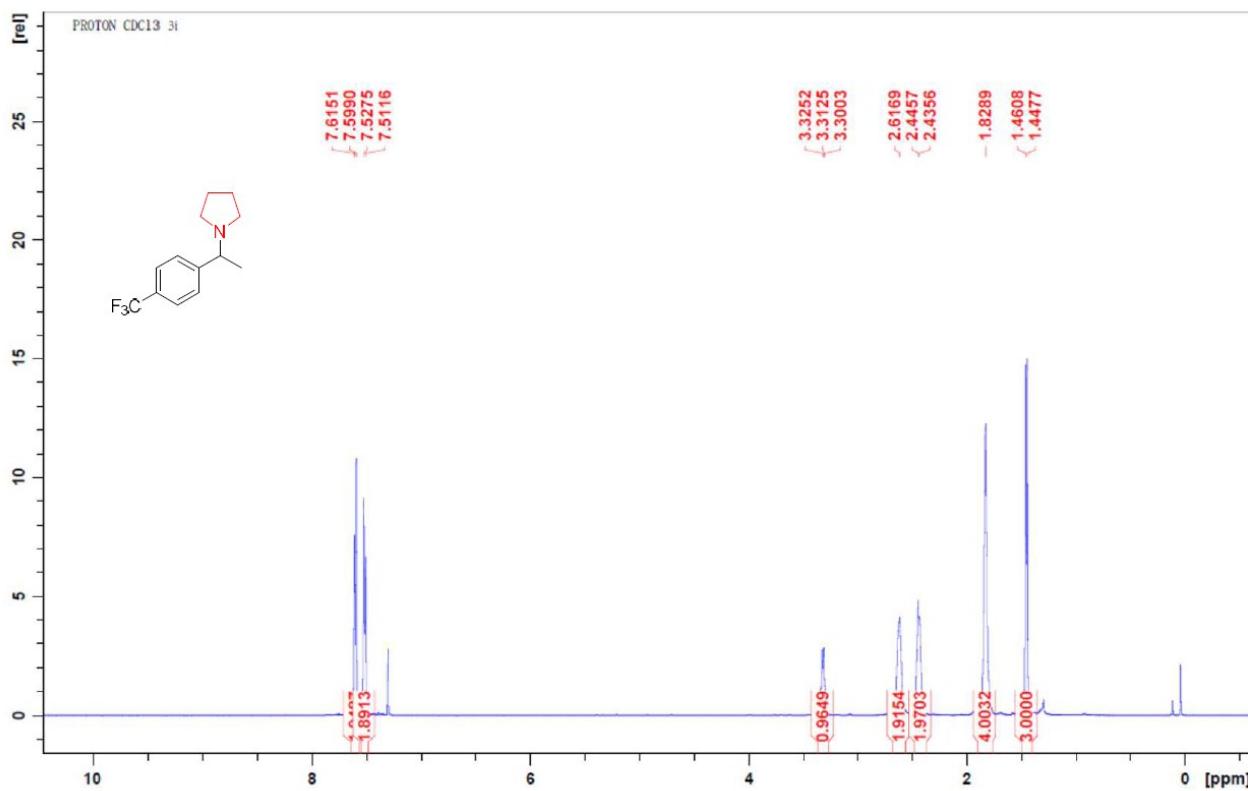
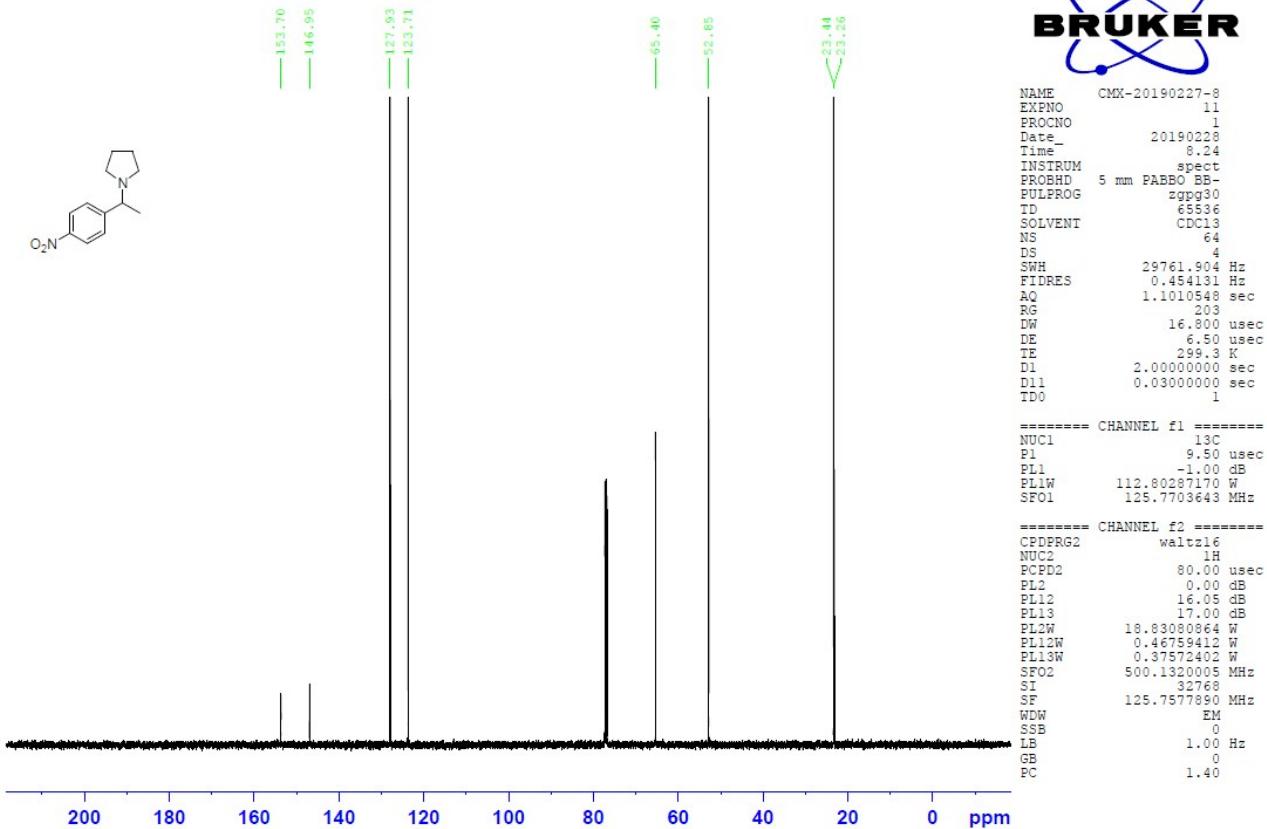


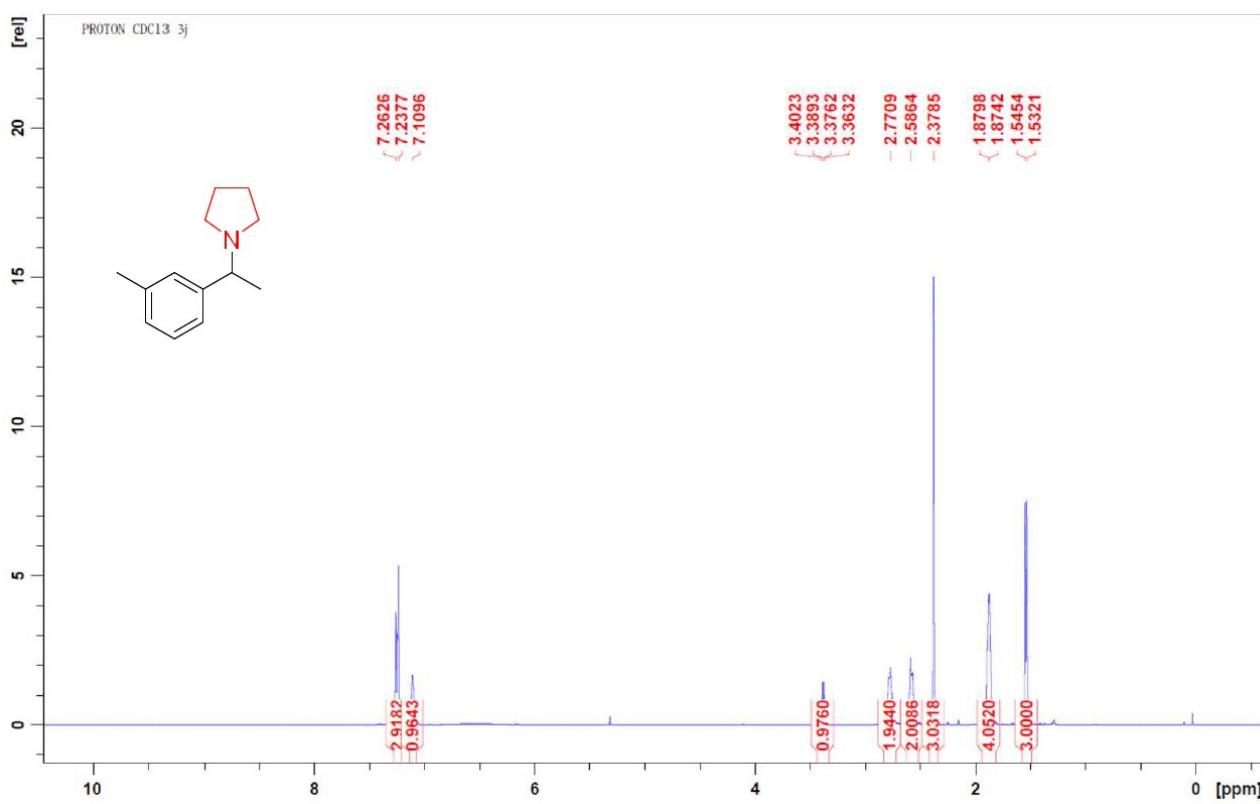
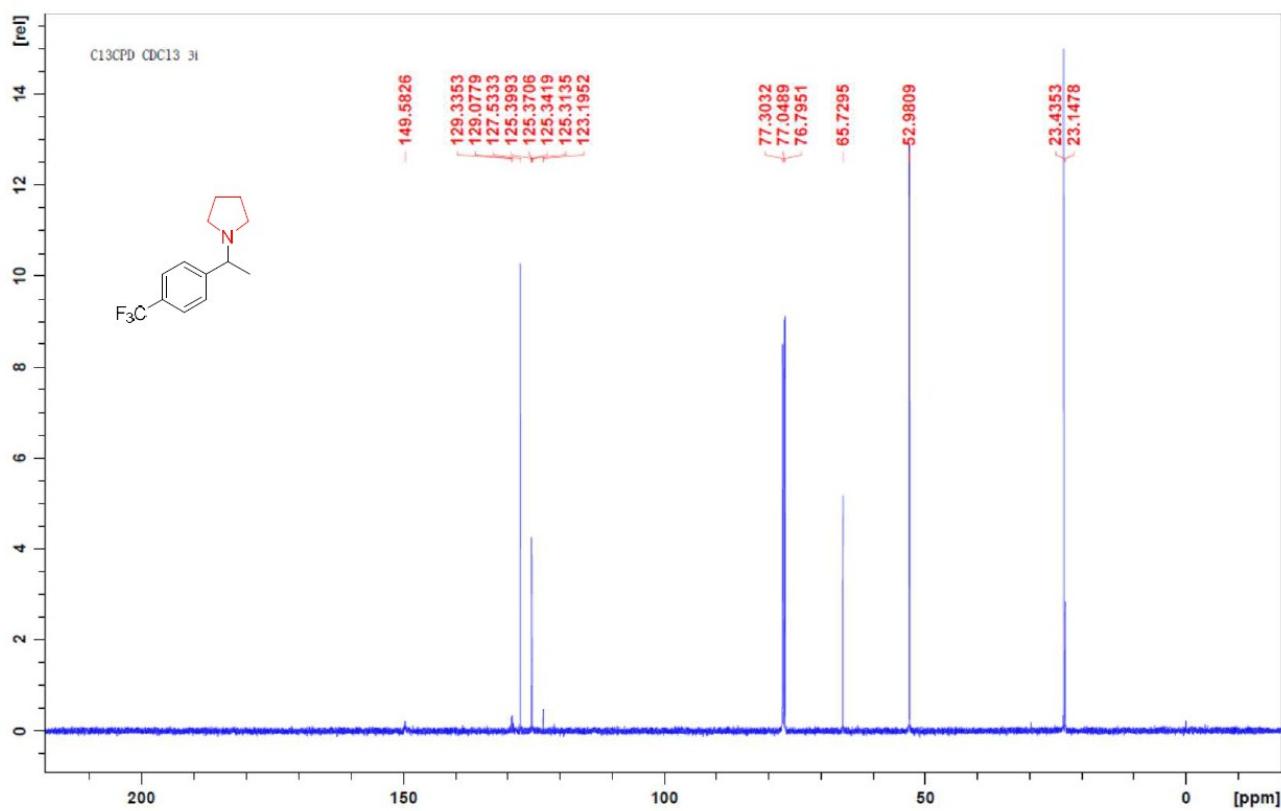
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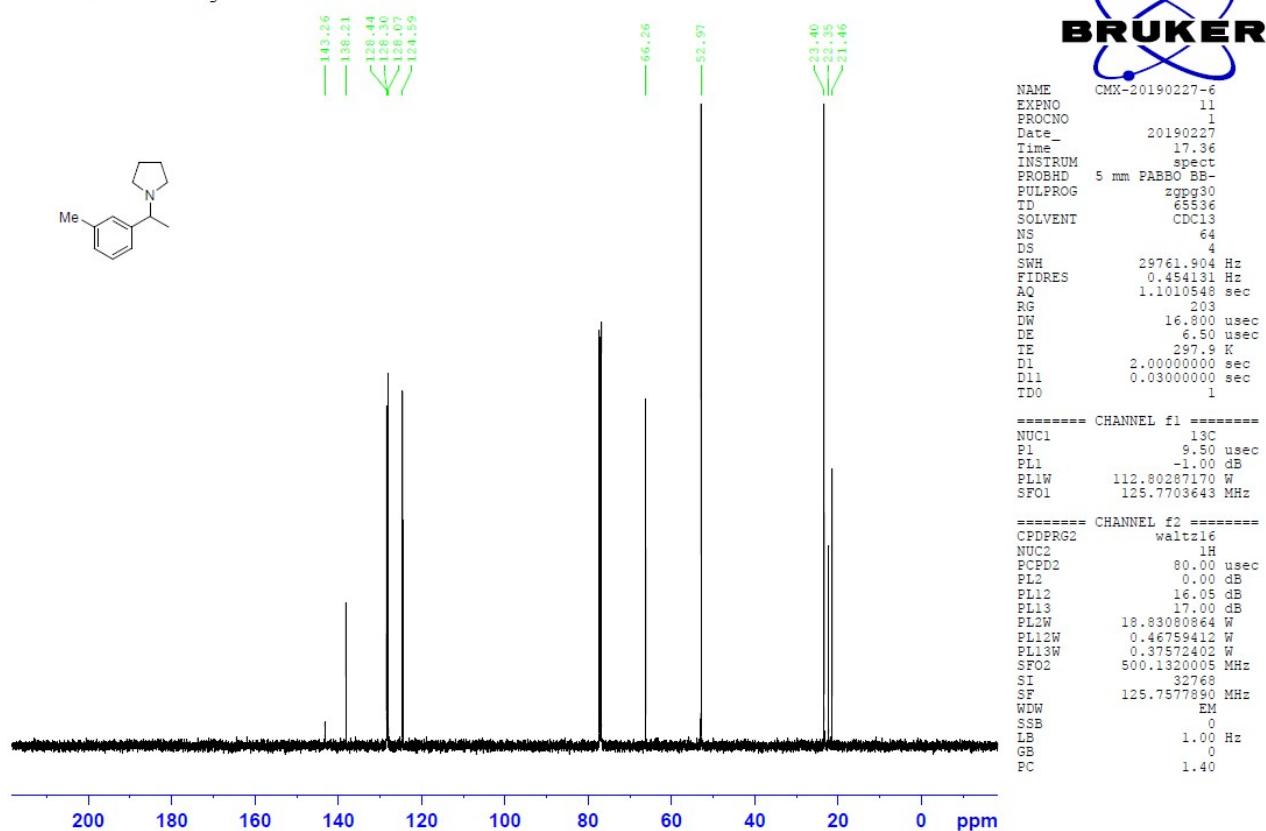
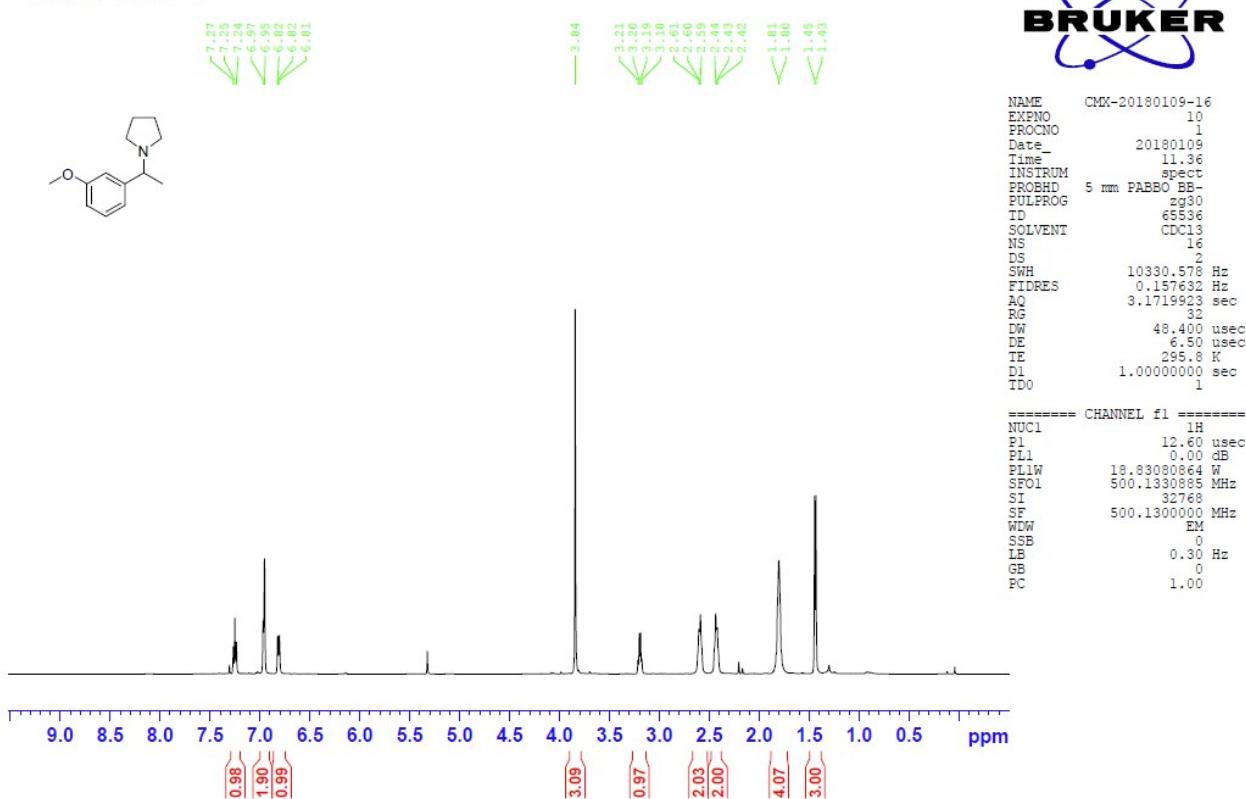
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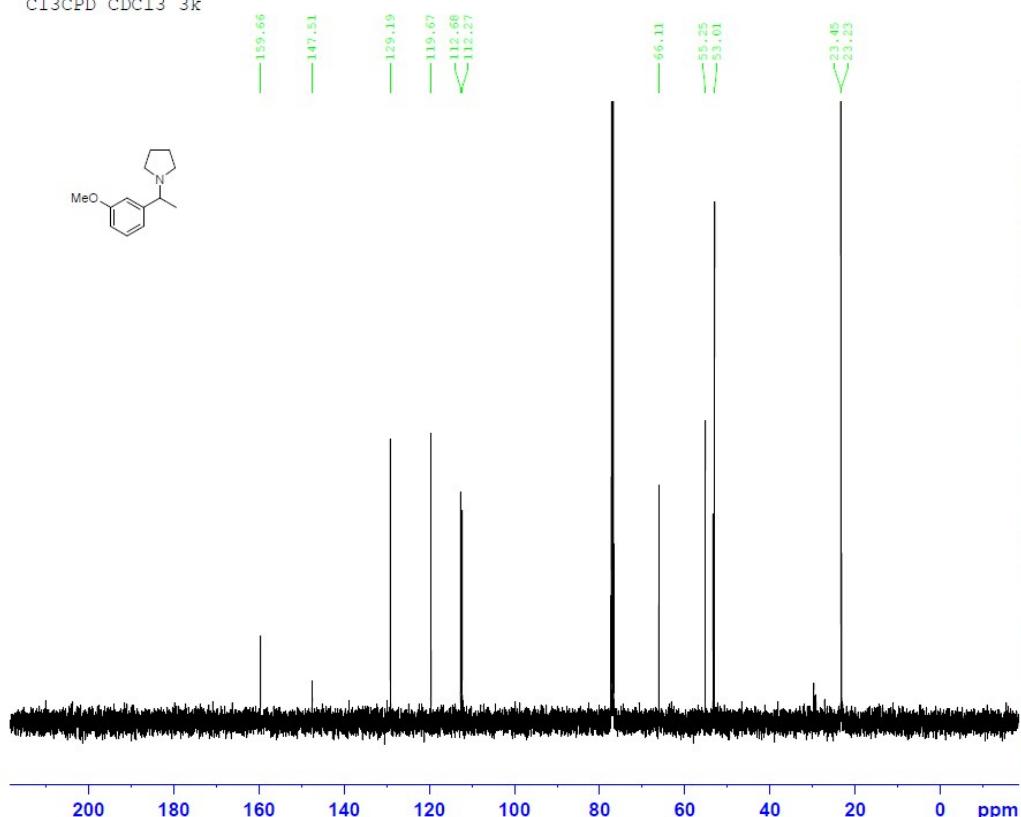
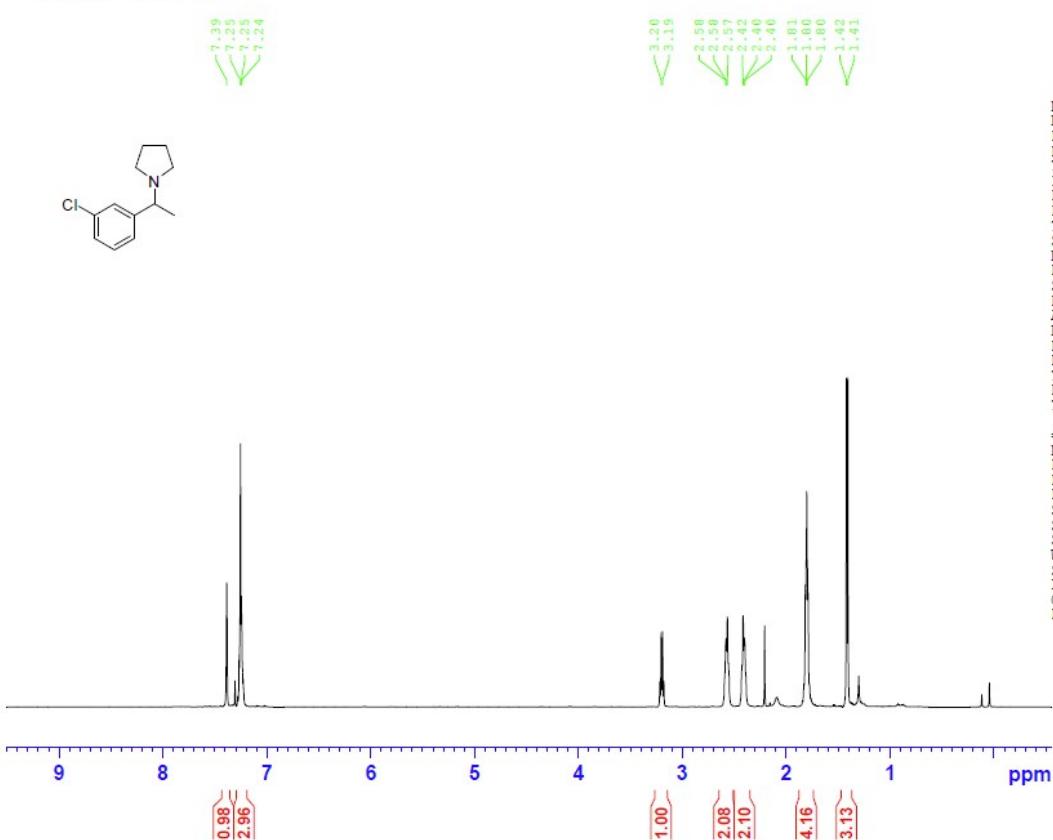


C13CPD CDCl₃ 3h





C13CPD CDCl₃ 3jPROTON CDCl₃ 3k

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C13CPD CDCl₃ 31

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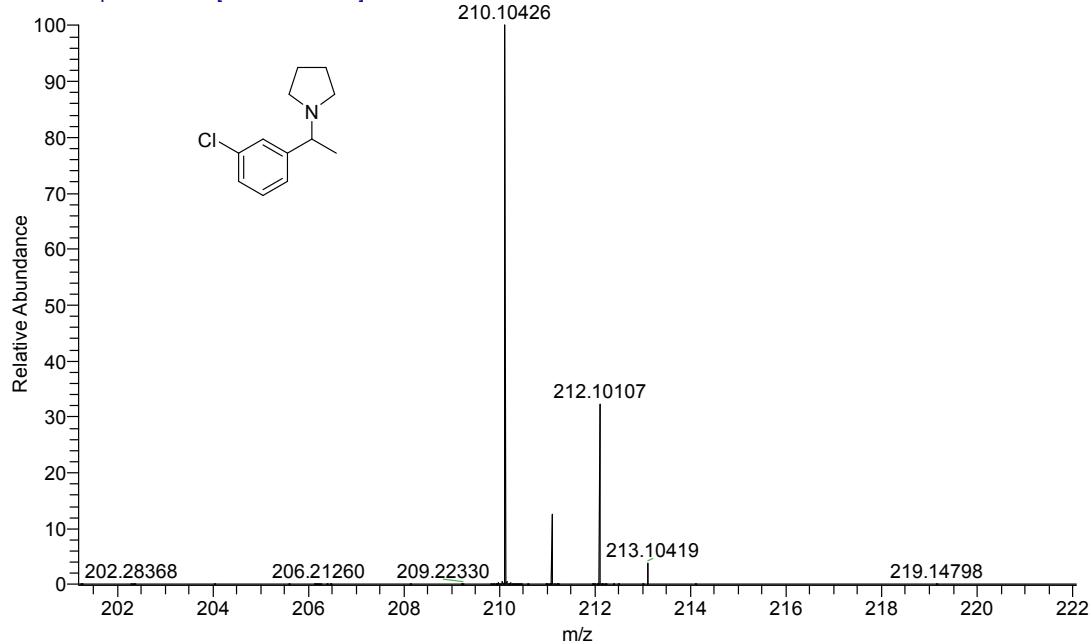
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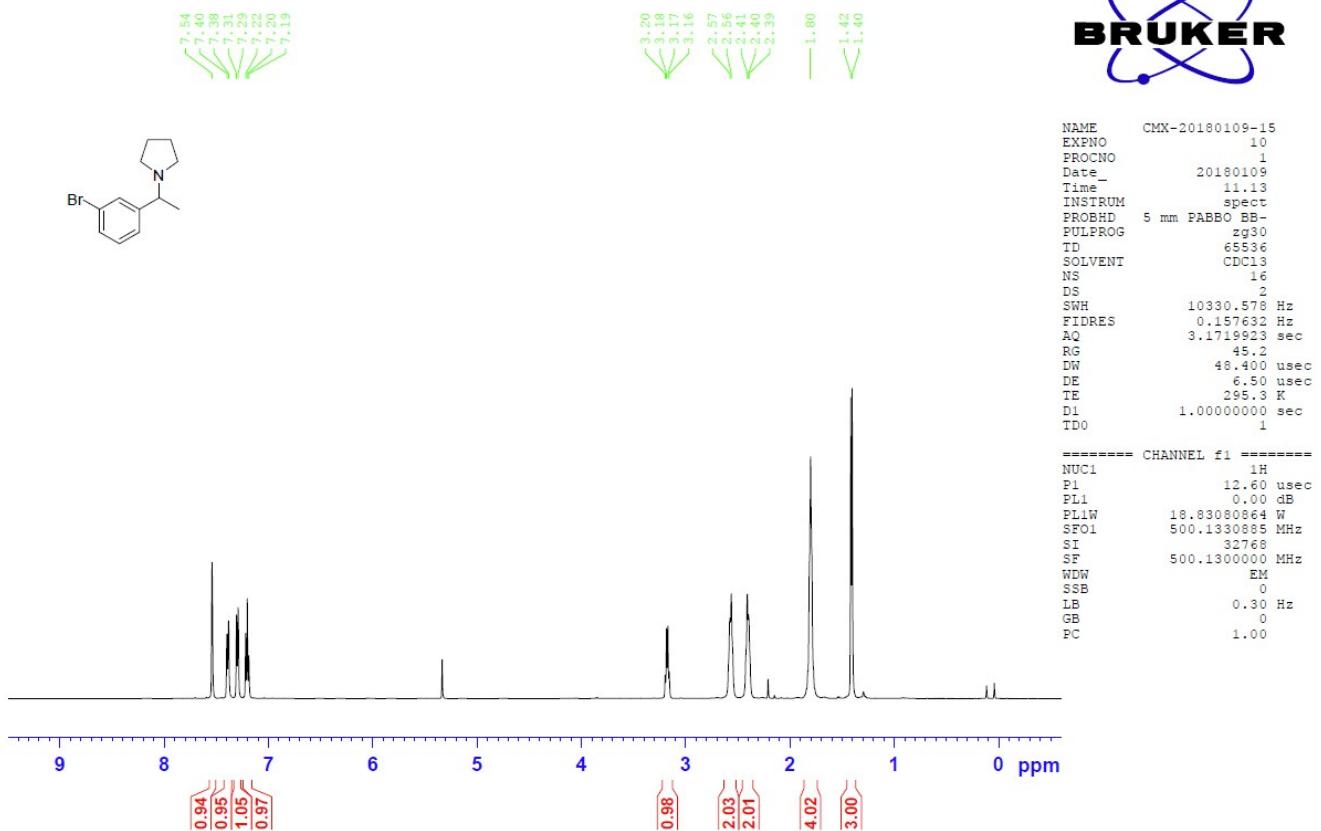
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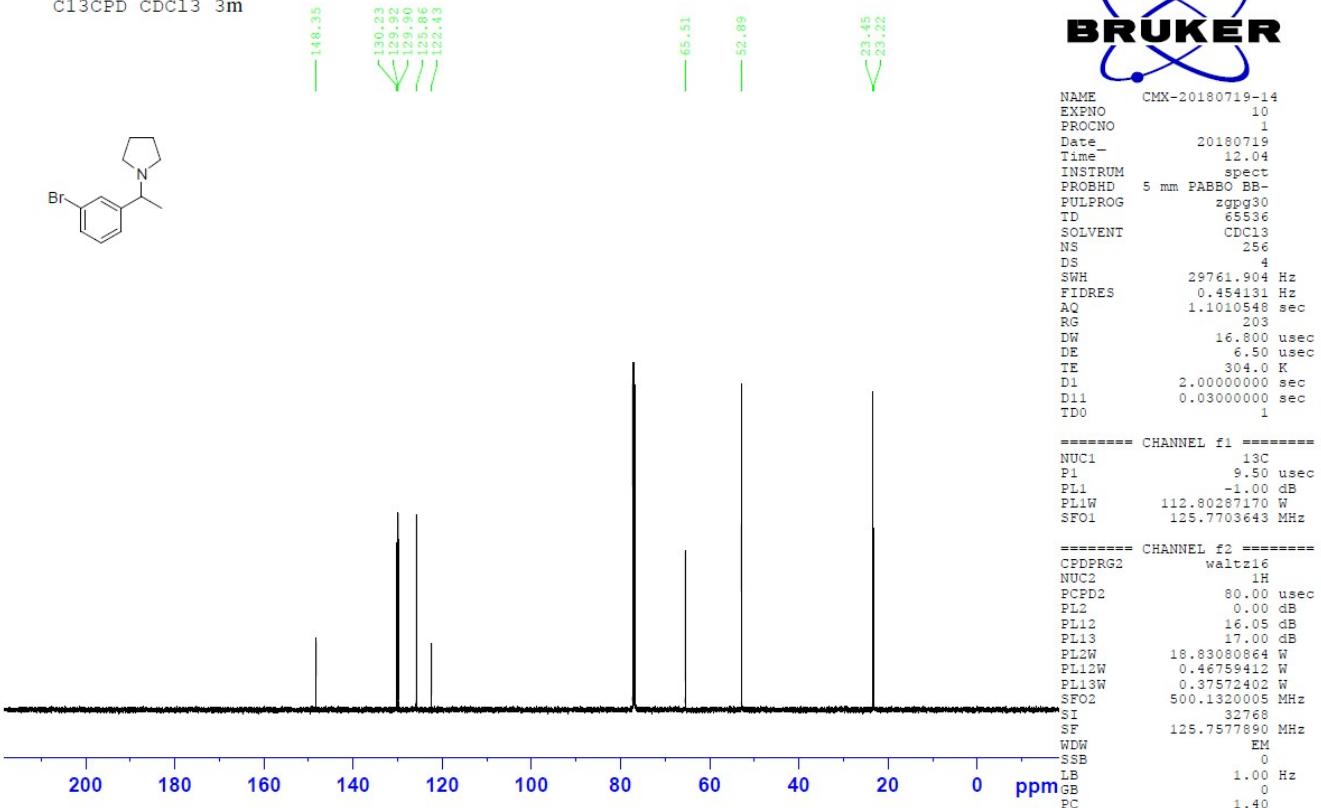
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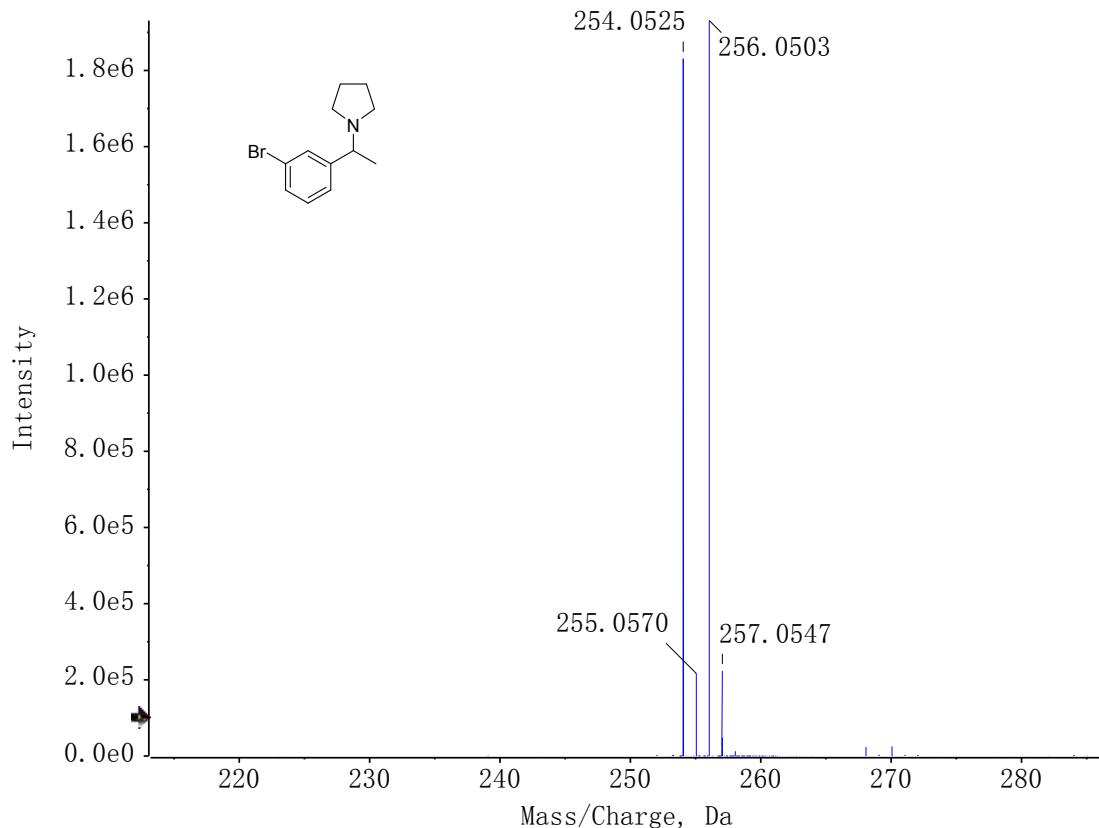
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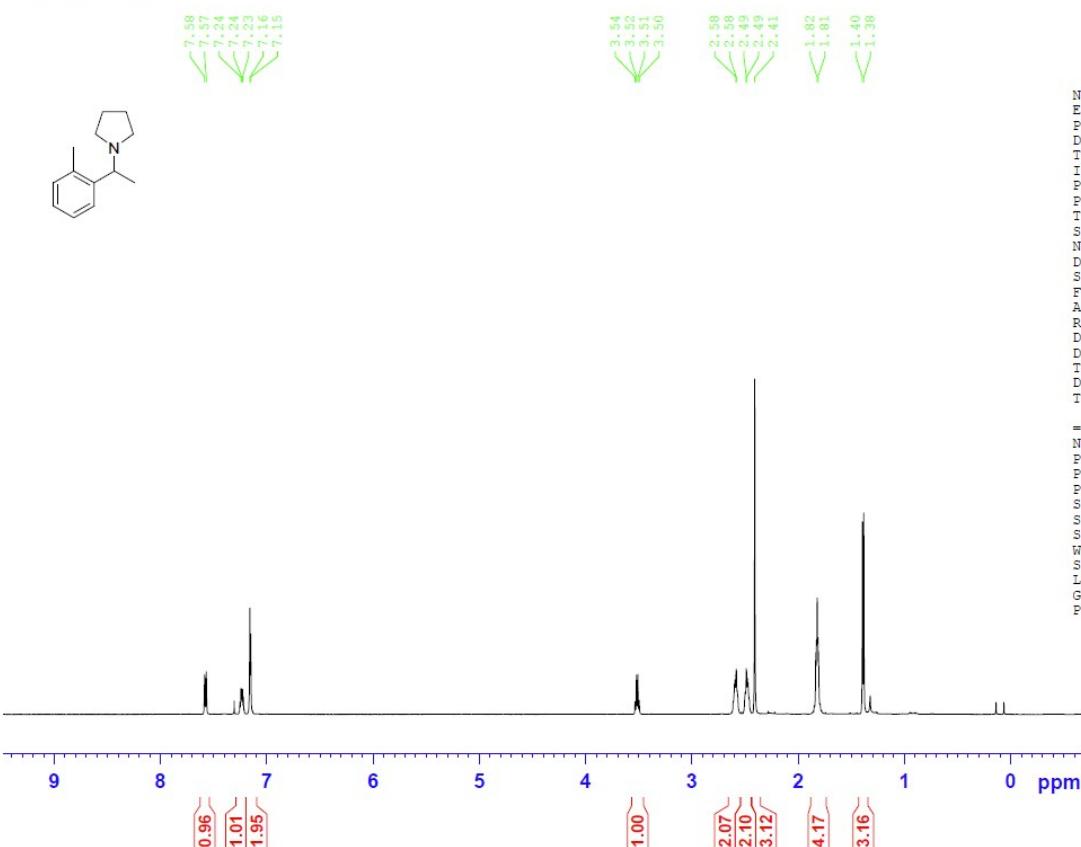
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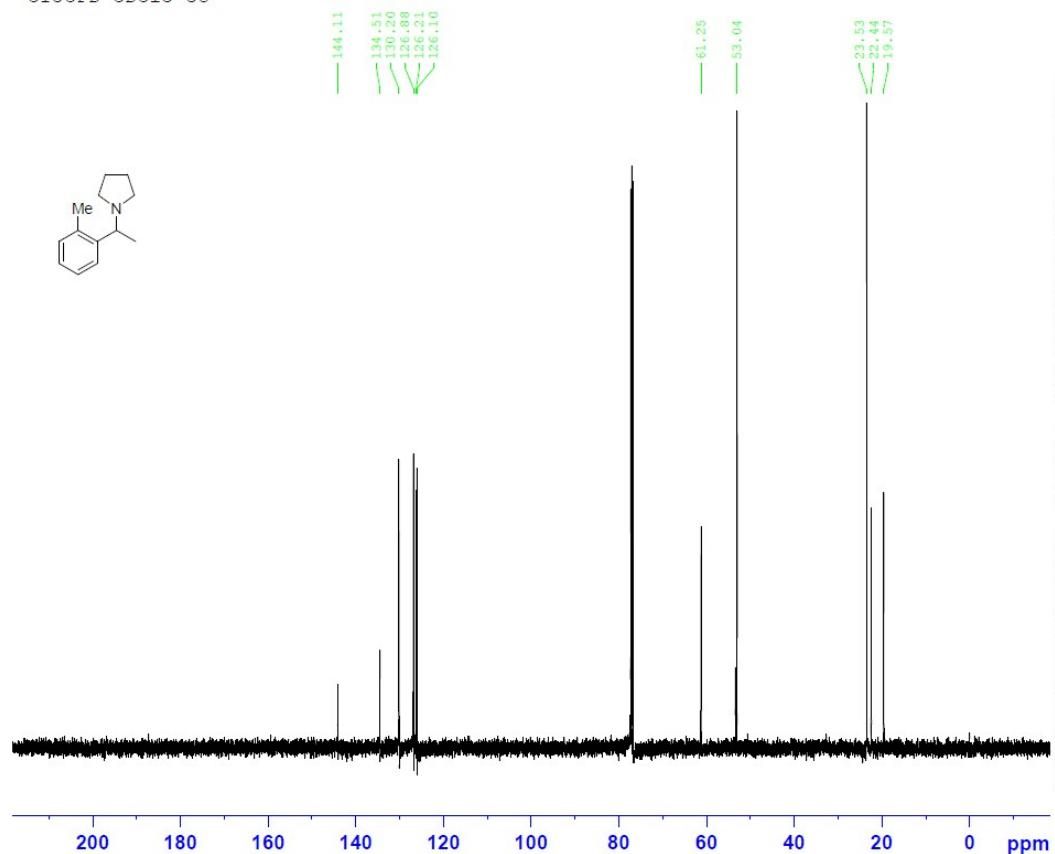
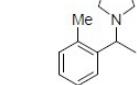


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PC       1.00

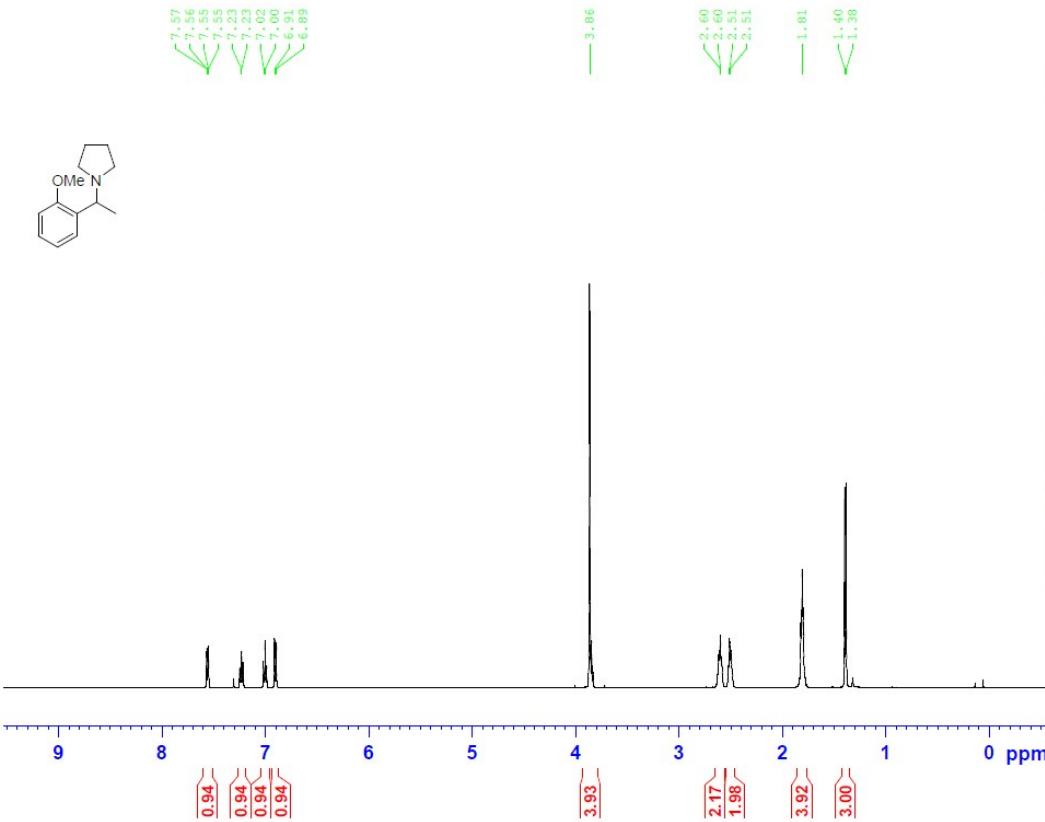
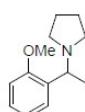
```

C13CPD CDCl₃ 30

```

NAME      CMX-20190225-9
EXPNO          11
PROCNO         1
Date_   20190225
Time_   12.35
INSTRUM spect
PROBHD  5 mm PABBO BB-
PULPROG zgpg30
TD      65536
SOLVENT   CDCl3
NS       64
DS        4
SWH     29761.904 Hz
FIDRES  0.454131 Hz
AQ      1.1010548 sec
RG      203
DW      16.800 usec
DE      6.500 usec
TE      300.0 K
D1      2.00000000 sec
D11     0.03000000 sec
TDO      1
===== CHANNEL f1 =====
NUC1      13C
P1      9.50 usec
PL1    -1.00 dB
PL1W   112.80287170 W
SF01    125.7703643 MHz
===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2      1H
PCPD2     80.00 usec
PL2      0.00 dB
PL12     16.05 dB
PL13     17.00 dB
PL2W    18.83080864 W
PL12W   0.46759412 W
PL13W   0.37572402 W
SF02    500.1320005 MHz
SI      32768
SF     125.7577890 MHz
WDW      EM
SSB      0
LB      1.00 Hz
GB      0
PC      1.40

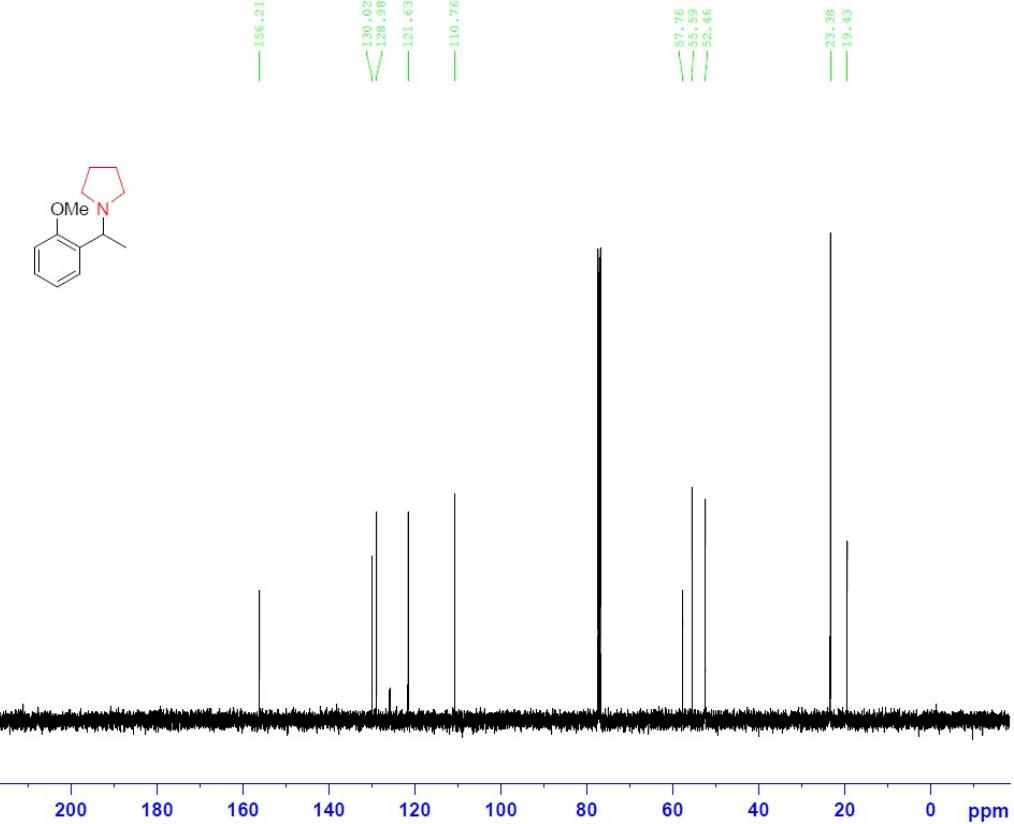
```

PROTON CDCl₃ 3p

```

NAME      CMX-20170919-5
EXPNO          10
PROCNO         1
Date_   20170919
Time_   11.27
INSTRUM spect
PROBHD  5 mm PABBO BB-
PULPROG zg30
TD      65536
SOLVENT   CDCl3
NS       16
DS        2
SWH     10330.578 Hz
FIDRES  0.157632 Hz
AQ      3.1719923 sec
RG      32
DW      48.400 usec
DE      6.500 usec
TE      303.4 K
D1      1.0000000 sec
TDO      1
===== CHANNEL f1 =====
NUC1      1H
P1      12.60 usec
PL1     0.00 dB
PL1W   18.83080864 W
SF01    500.1330865 MHz
SI      32768
SF     500.1300000 MHz
WDW      EM
SSB      0
LB      0.30 Hz
GB      0
PC      1.00

```

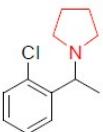
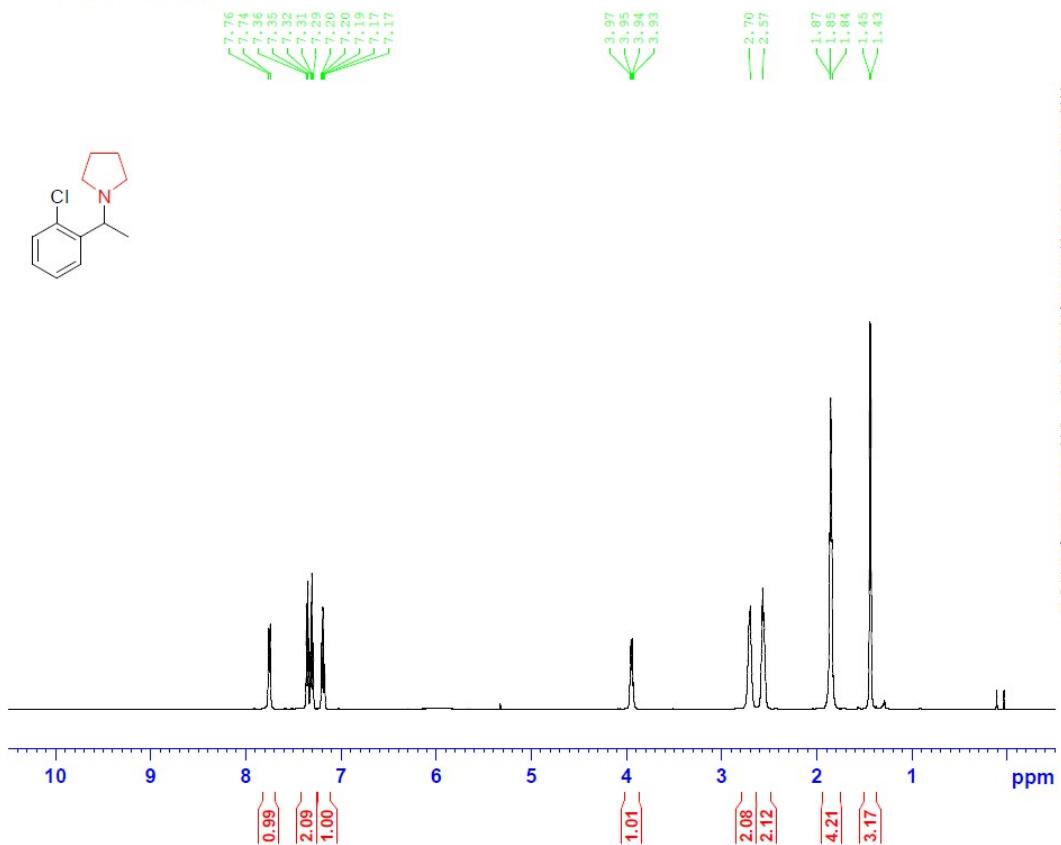
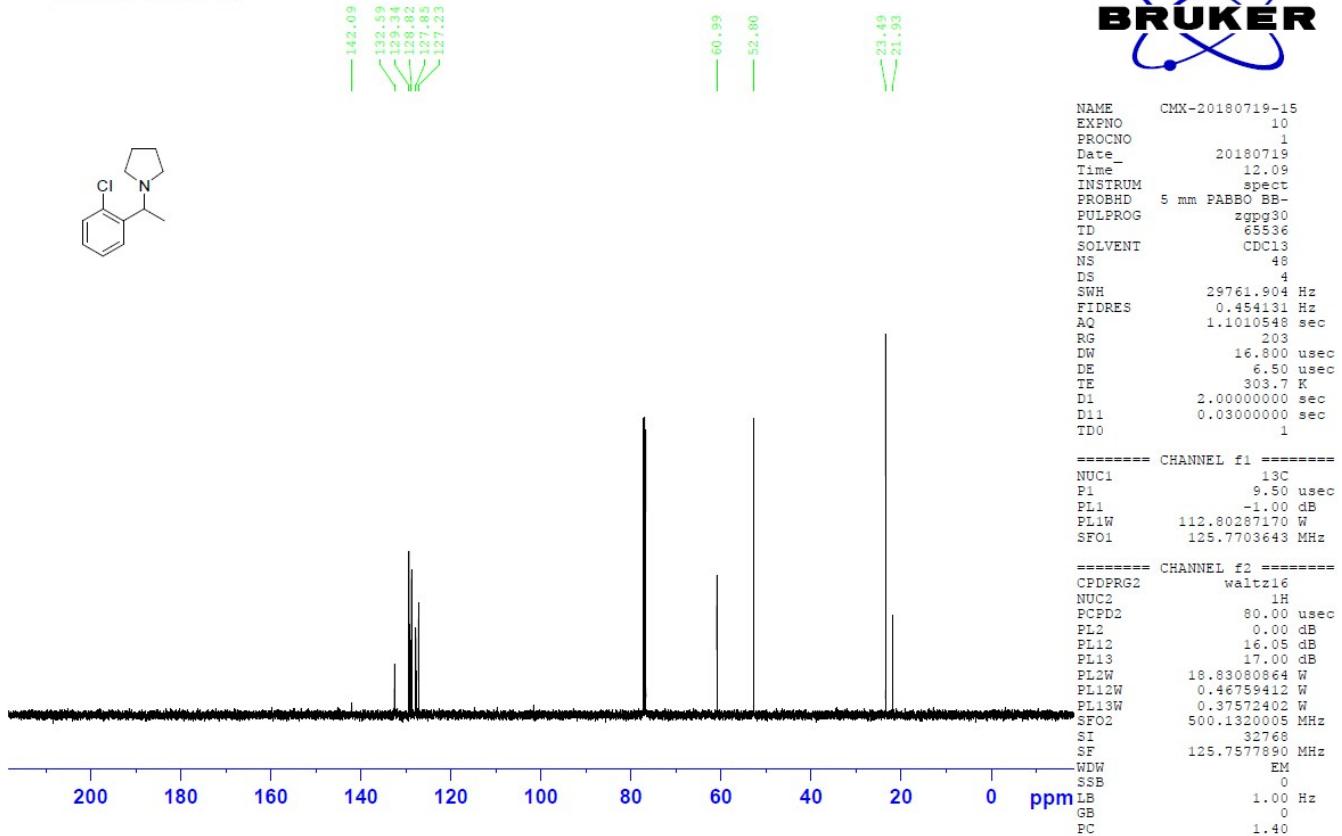
C13CPD CDCl₃ 3p

```

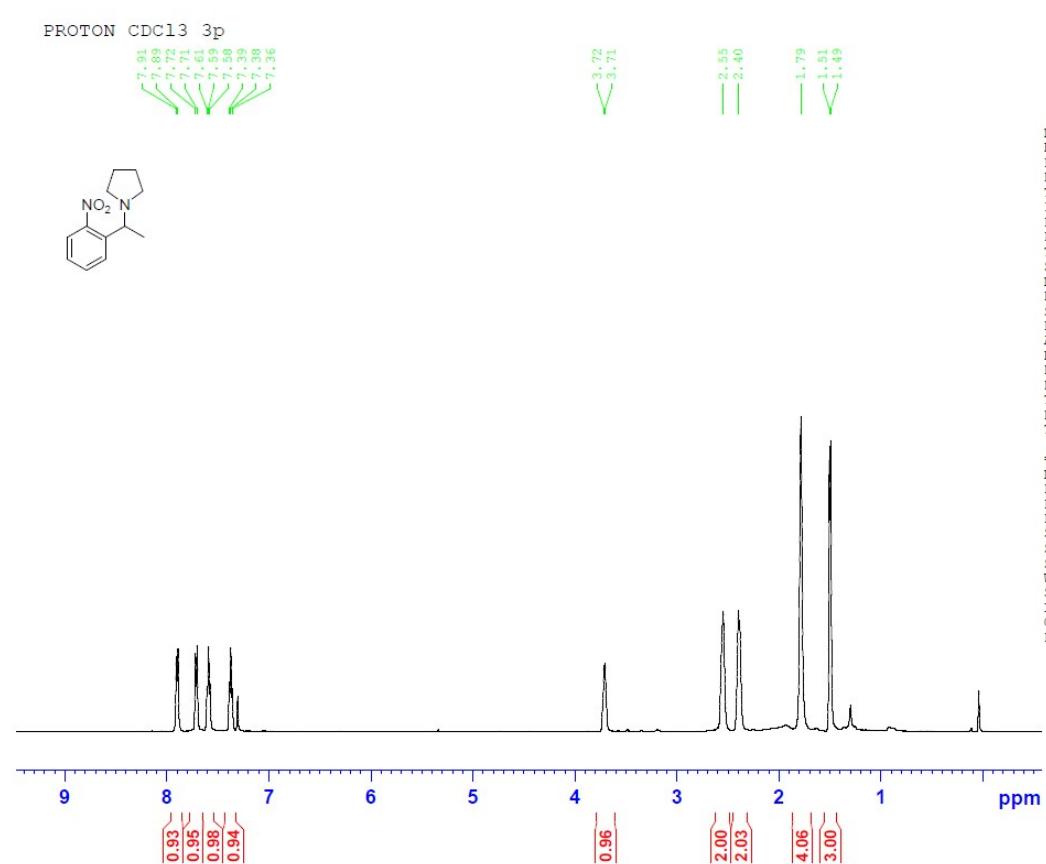
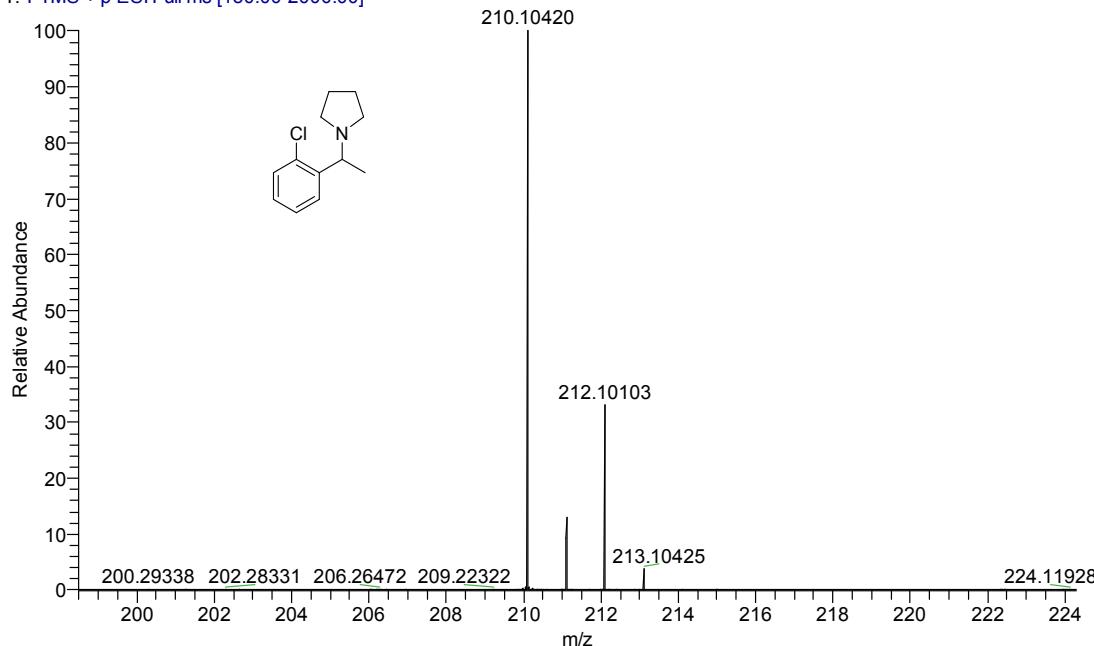
NAME      CMX-20190225-10
EXPNO           11
PROCNO          1
Date_        20190225
Time_        12.40
INSTRUM     spect
PROBHD    5 mm PABBO BB
PULPROG    zgpg30
TD        65536
SOLVENT    CDCl3
NS           21
DS            4
SWH       29761.904 Hz
FIDRES    0.454131 Hz
AQ        1.1010548 sec
RG          203
DW       16.800 usec
DE        6.50 usec
TE        300.0 K
D1        2.0000000 sec
D11       0.03000000 sec
TDO         1

===== CHANNEL f1 =====
NUC1        13C
P1          9.50 usec
PL1        -1.00 dB
PL1W      112.80287170 W
SF01      125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2   Waltz16
NUC2          1H
PCPD2        80.00 usec
PL2          0.00 dB
PL12        16.05 dB
PL13        17.00 dB
PL2W      18.83080864 W
PL12W      0.46759412 W
PL13W      0.37572402 W
SF02      500.1320005 MHz
SI          32768
SF      125.757790 MHz
WDW         EM
SSB          0
LB          1.00 Hz
GB          0
PC          1.40
  
```

PROTON CDCl₃ 3qC13CPD CDCl₃ 3q

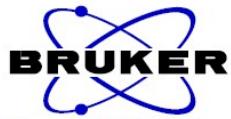
86 #11 RT: 0.15 AV: 1 NL: 8.84E8
T: FTMS + p ESI Full ms [150.00-2000.00]



NAME CMX-20180704-20
EXPNO 10
PROCNO 1
Date 20180704
Time 14.38
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 90.5
DW 48.400 usec
DE 6.50 usec
TE 302.0 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====

NUC1 1H
P1 12.60 usec
PL1 0.00 dB
PL1W 18.63080864 W
SF01 500.1330885 MHz
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

C13CPD CDCl₃ 3r

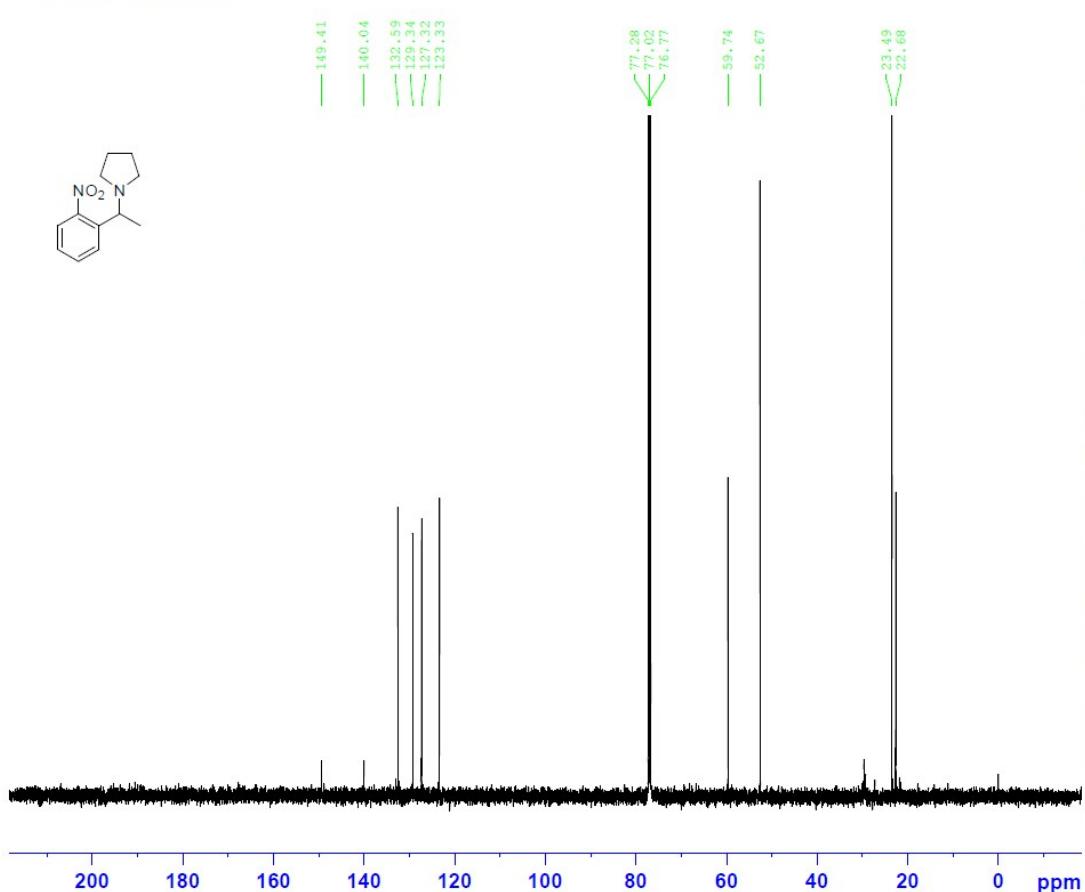
NAME CMX-20180801-2
 EXPNO 12
 PROCNO 1
 Date 20180801
 Time 10.59
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 1024
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010548 sec
 RG 203
 DW 16.800 usec
 DE 6.50 usec
 TE 304.7 K
 D1 0.6000002 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====

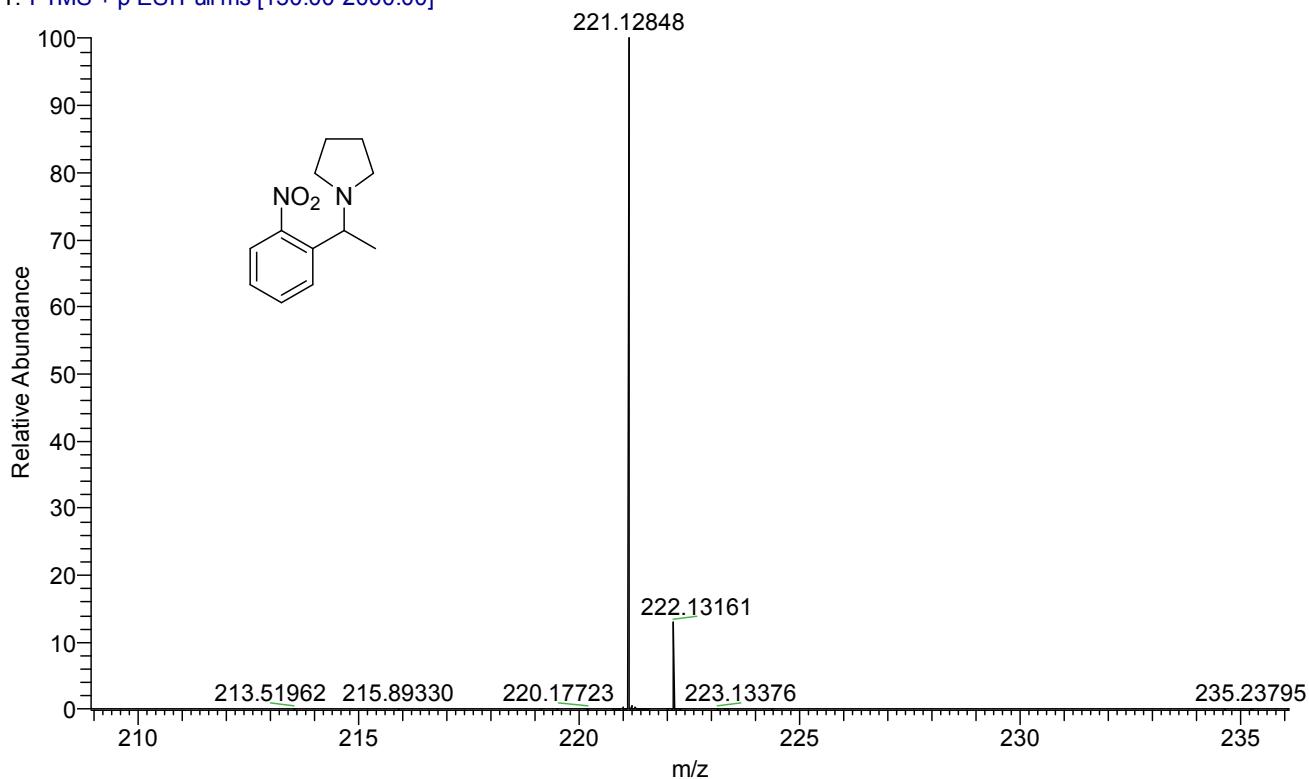
NUC1 ¹³C
 P1 9.50 usec
 PL1 -1.00 dB
 PL1W 112.80287170 W
 SF01 125.7703643 MHz

===== CHANNEL f2 =====

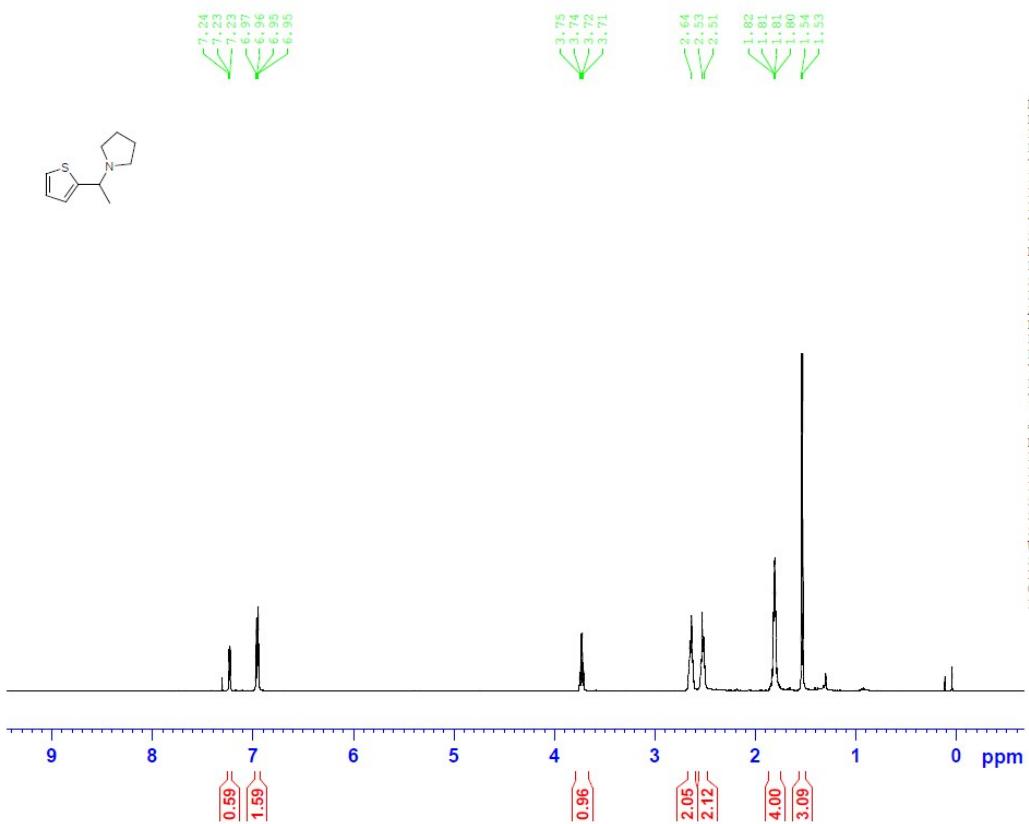
CPDPG2 waltz16
 NUC2 ¹H
 PCPD2 80.00 usec
 PL2 0.00 dB
 PL12 16.05 dB
 PL13 17.00 dB
 PL2W 18.83080864 W
 PL12W 0.46759412 W
 PL13W 0.37572402 W
 SF02 500.1320005 MHz
 SI 32768
 SF 125.7577890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



33 #10 RT: 0.12 AV: 1 NL: 6.16E8
 T: FTMS + p ESI Full ms [150.00-2000.00]



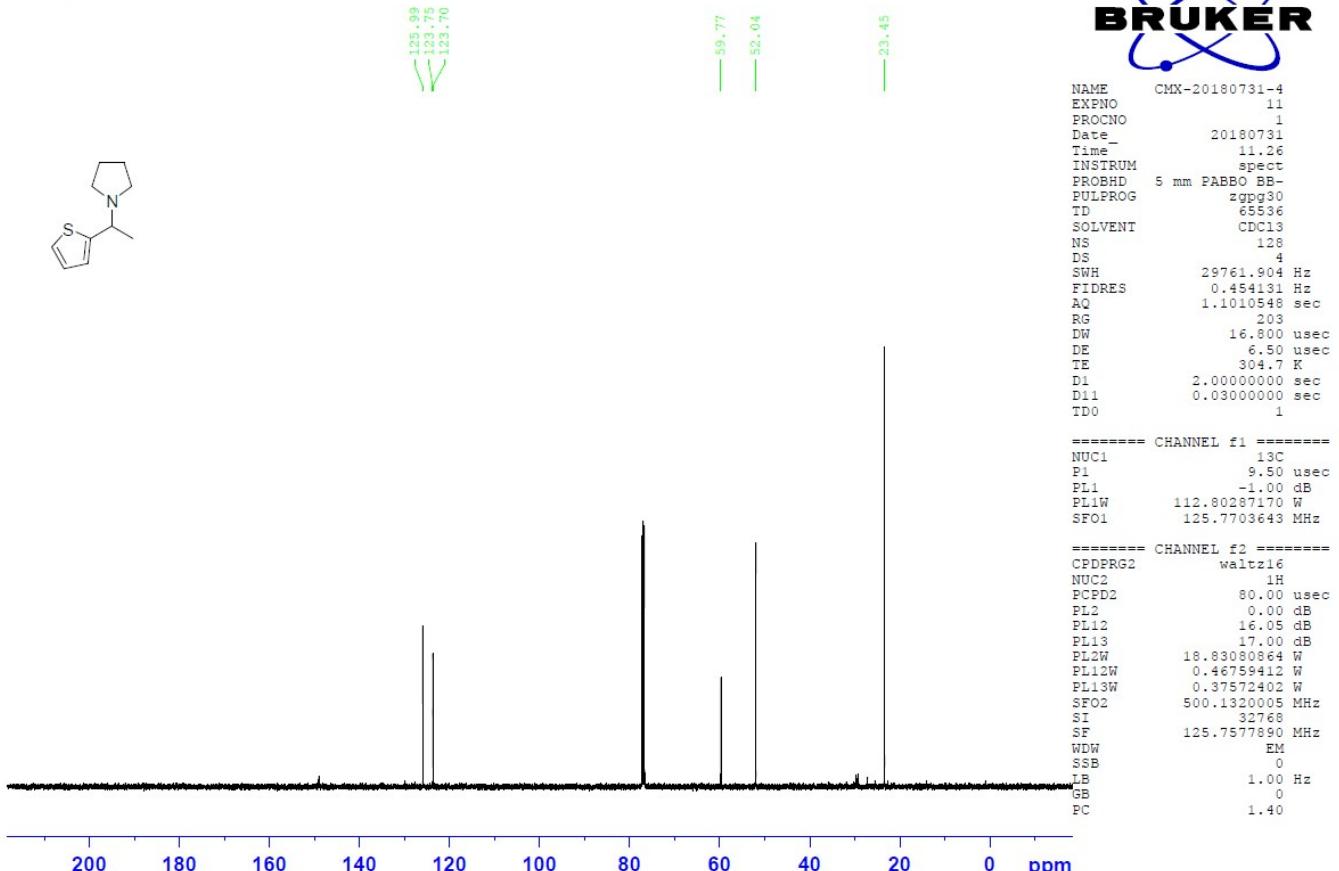
PROTON CDCl₃ 3s



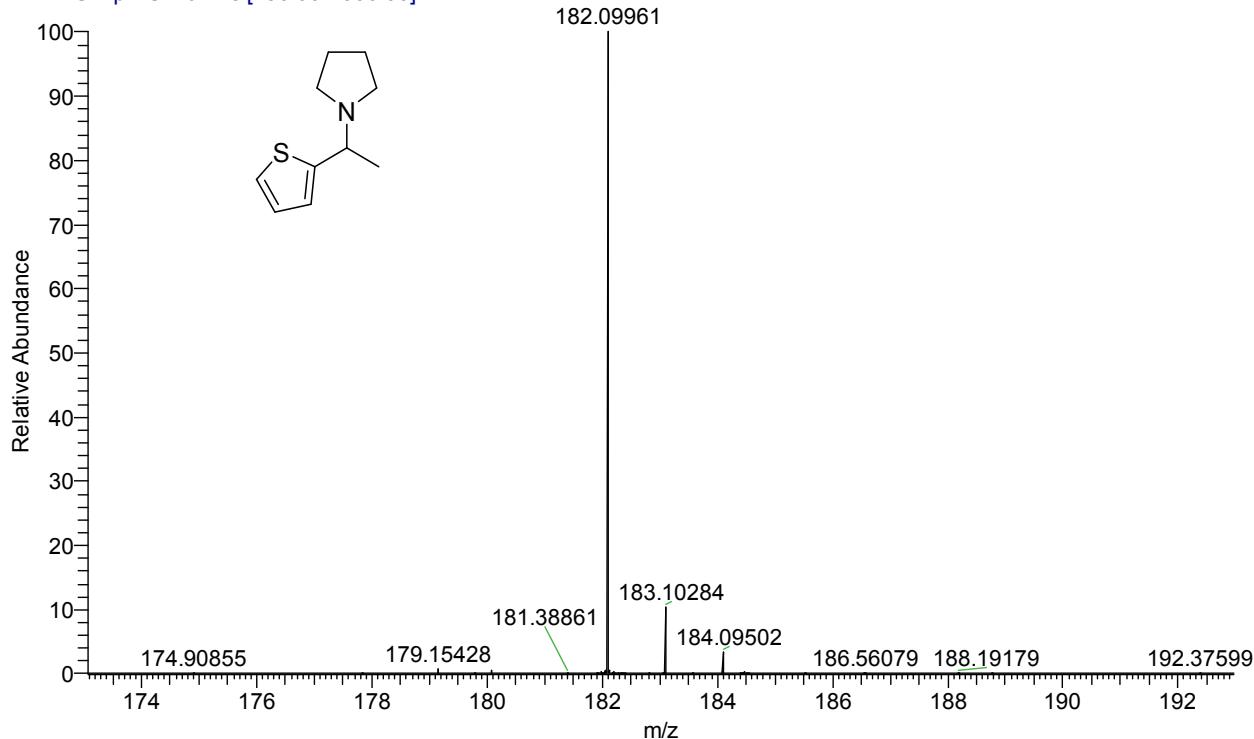
NAME CMX-20180725-9
EXPNO 10
PROCNO 1
Date 20180725
Time 14.14
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 71.8
DW 48.400 usec
DE 6.50 usec
TE 303.0 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.60 usec
PL1 0.00 dB
PL1W 18.83080864 W
SFO1 500.1330885 MHz
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

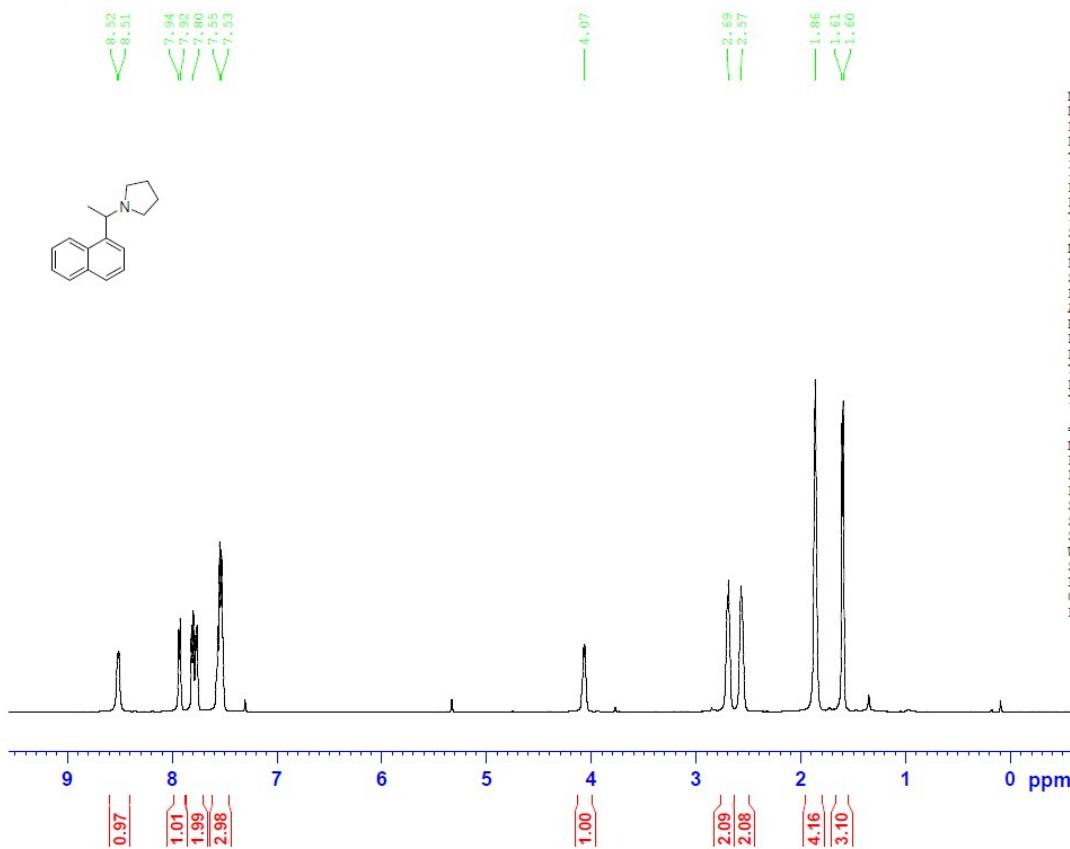
C13CPD CDCl₃ 3s



34 #12 RT: 0.16 AV: 1 NL: 2.04E8
T: FTMS + p ESI Full ms [150.00-2000.00]



PROTON CDCl₃ 3t



```

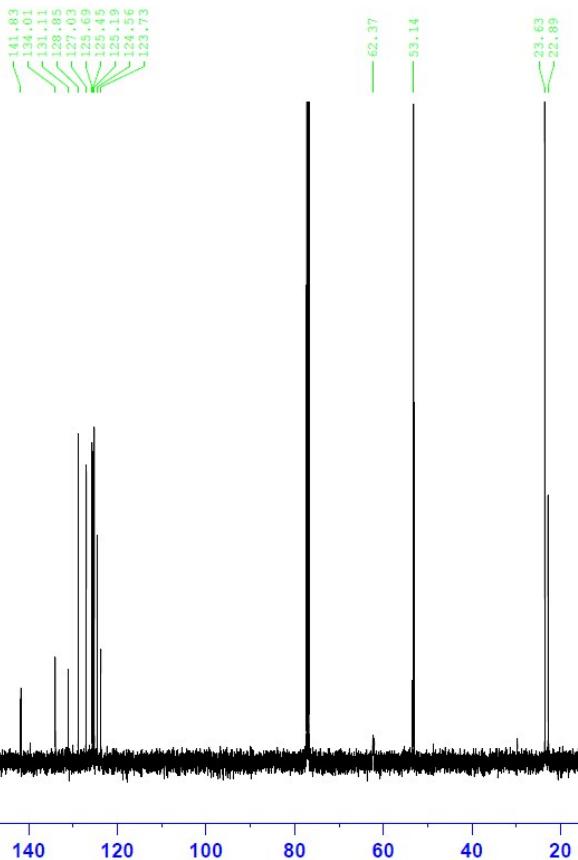
NAME      CMX-20180123-13
EXPNO          10
PROCNO         1
Date_   20180124
Time_   10.34
INSTRUM spect
PROBHD  5 mm PABBO BB-
PULPROG zg30
TD      65536
SOLVENT   CDCl3
NS       12
DS        2
SWH     10330.578 Hz
FIDRES  0.157632 Hz
AQ      3.1719923 sec
RG        32
DW      48.400 usec
DE       6.50 usec
TE      300.0 K
D1      1.0000000 sec
TD0           1

```

```

===== CHANNEL f1 =====
NUC1           1H
F1            12.60 usec
PL1            0.00 dB
PL1W          18.83080864 W
SFO1        500.13308855 MHz
SI            32768
SF          500.1300000 MHz
WDW               EM
SSB                 0
LB            0.30 Hz
GB                 0
PC            1.00

```

C13CPD CDCl₃ 3t

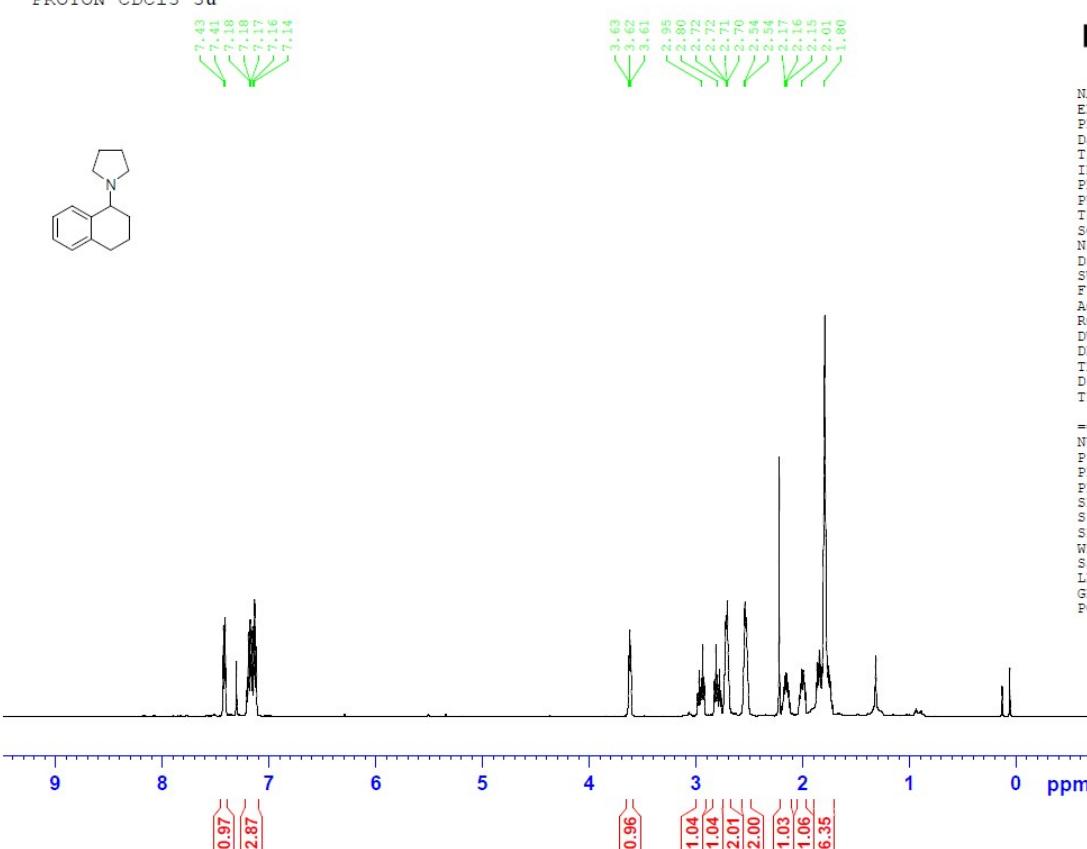
NAME CMX-20190225-11
EXPNO 11
PROCNO 1
Date_ 20190225
Time 12:46
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgppg30
TD 65536
SOLVENT CDCl₃
NS 64
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 203
DW 16.800 usec
DE 6.50 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

===== CHANNEL f1 ======

NUC1 13C
P1 9.50 usec
PL1 -1.00 dB
PL1W 112.80287170 W
SFO1 125.7703643 MHz

===== CHANNEL f2 ======

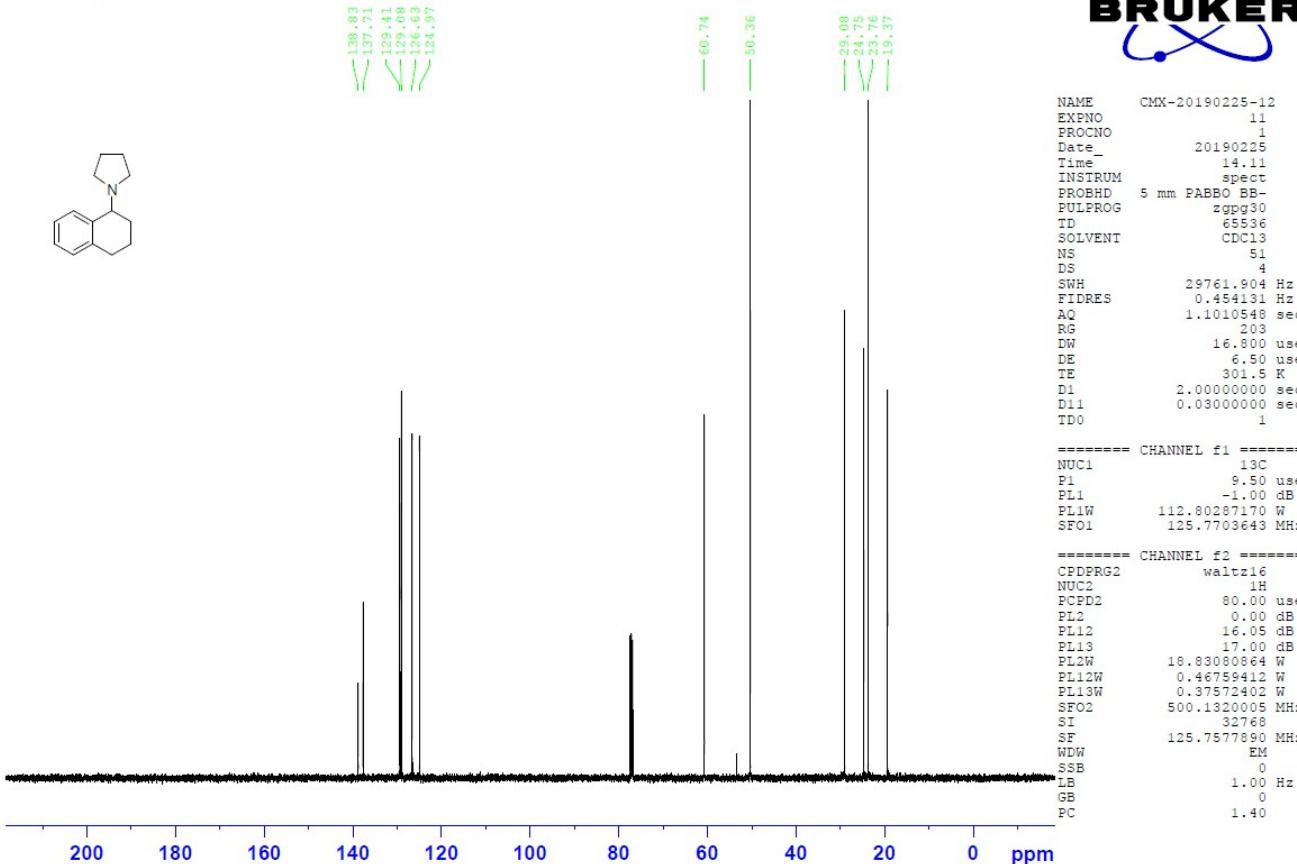
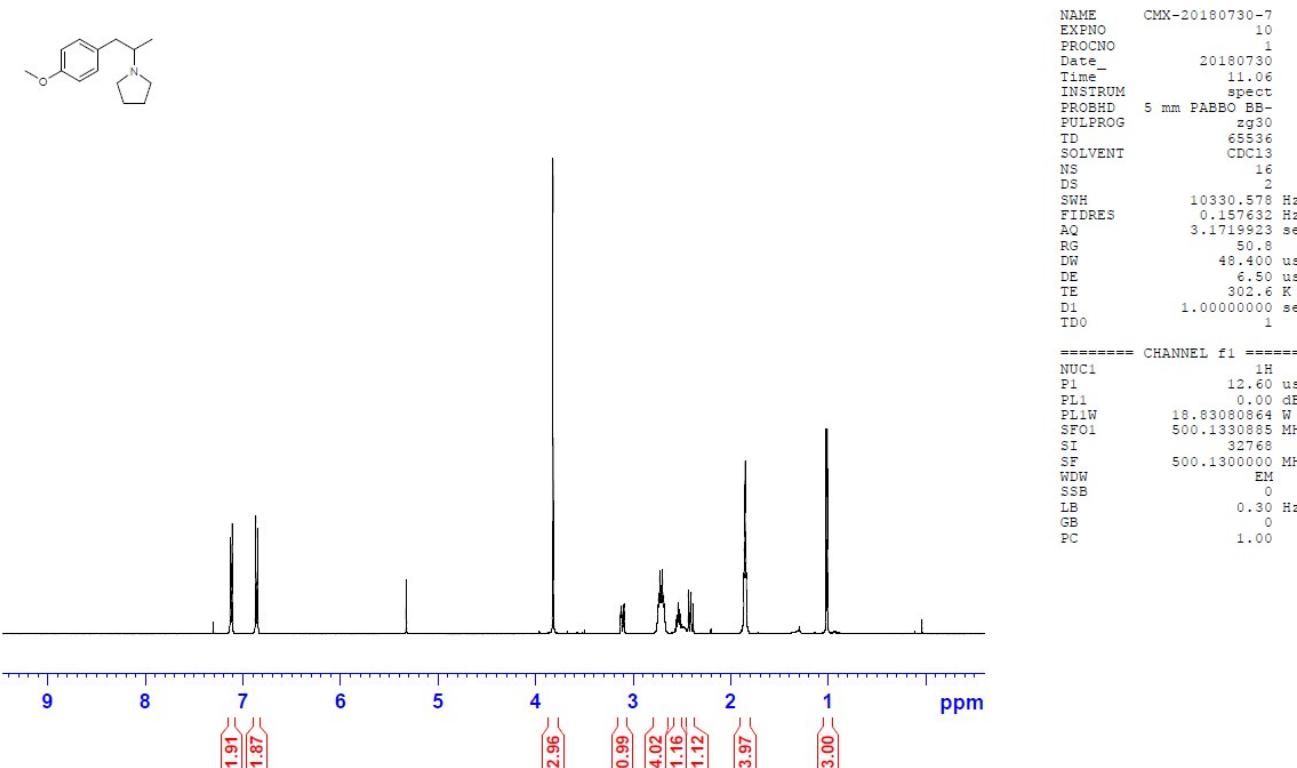
CPDPG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 16.05 dB
PL13 17.00 dB
PL2W 18.83080864 W
PL12W 0.46759412 W
PL13W 0.37572402 W
SFO2 500.1320005 MHz
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

PROTON CDCl₃ 3u

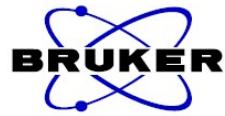
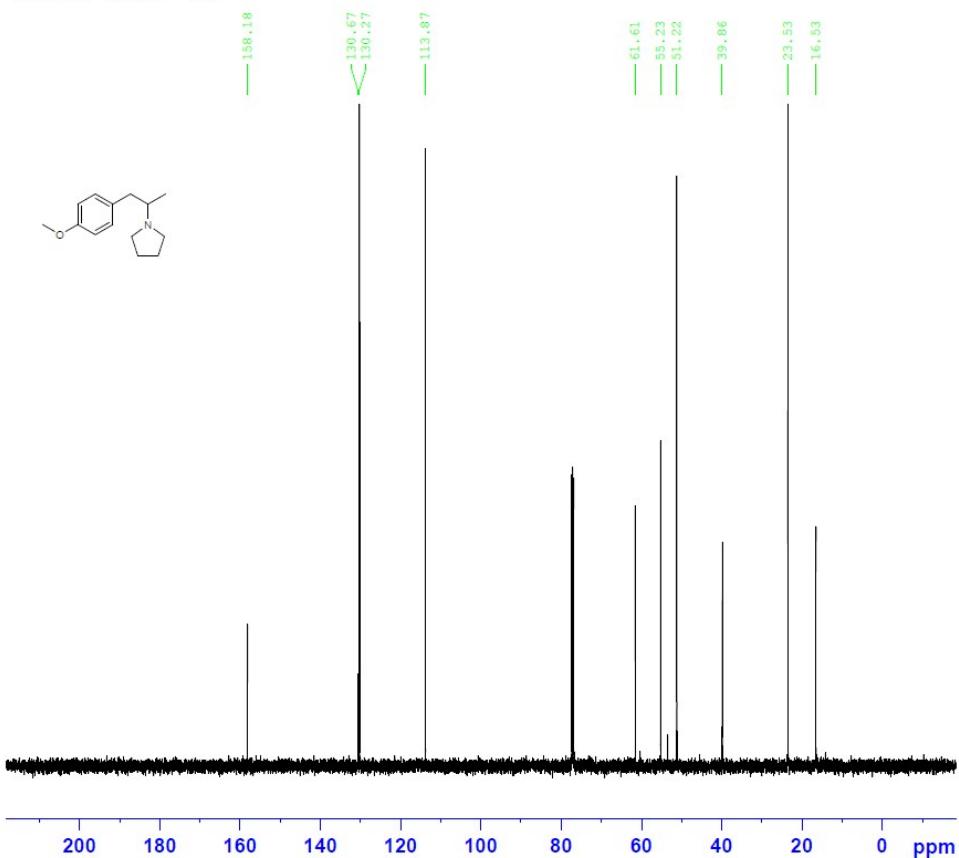
NAME CMX-20170925-6
EXPNO 10
PROCNO 1
Date_ 20170926
Time 9:15
INSTRUM spect
PROBHD 5 mm PASEI 1H/
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.171923 sec
RG 71.8
DW 48.400 usec
DE 6.50 usec
TE 301.9 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 ======

NUC1 1H
P1 6.80 usec
PL1 0.00 dB
PL1W 18.83080864 W
SFO1 500.1330895 MHz
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

C13CPD CDCl₃ 3uPROTON CDCl₃ 3t

C13CPD CDCl₃ 3v



```

NAME      CMX-20190225-13
EXPNO           11
PROCNO          1
Date_   20190225
Time       14.18
INSTRUM  spect
PROBHD  5 mm PABBO BB-
PULPROG zgpg30
TD        65536
SOLVENT   CDCl3
NS         40
DS            4
SWH      29761.904 Hz
FIDRES    0.454131 Hz
AQ        1.1010548 sec
RG          203
DW        16.800 usec
DE        6.50 usec
TE        300.8 K
DI     2.0000000 sec
D11    0.03000000 sec
TDO        1

```

```

===== CHANNEL f1 =====
NUC1      13C
P1        9.50 usec
PL1      -1.00 dB
PL1W    112.8028170 W
SFO1    125.7703643 MHz

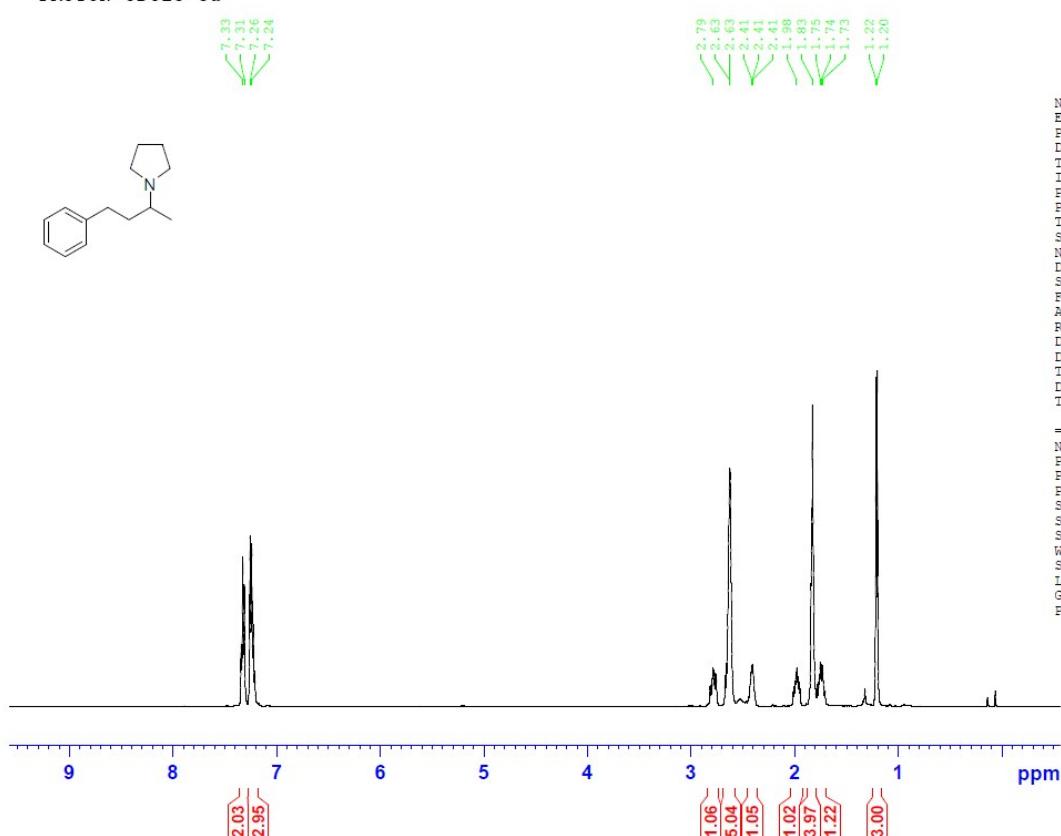
```

```

===== CHANNEL f2 =====
CPDPG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2        0.00 dB
PL12     16.05 dB
PL13     17.00 dB
PL2W    18.83080864 W
PL12W   0.46759412 W
PL13W   0.37572402 W
SFO2    500.1320005 MHz
SI        32768
SF      125.7577890 MHz
WDW        EM
SSB        0
LB        1.00 Hz
GB        0
PC        1.40

```

PROTON CDCl₃ 3u



```

NAME      CMX-20170919-6
EXPNO           10
PROCNO          1
Date_   20170919
Time       11.32
INSTRUM  spect
PROBHD  5 mm PABBO BB-
PULPROG zg30
TD        65536
SOLVENT   CDCl3
NS         16
DS            2
SWH      10330.578 Hz
FIDRES    0.157632 Hz
AQ        3.1719923 sec
RG          36
DW        48.400 usec
DE        6.50 usec
TE        303.4 K
DI     1.0000000 sec
TDO        1

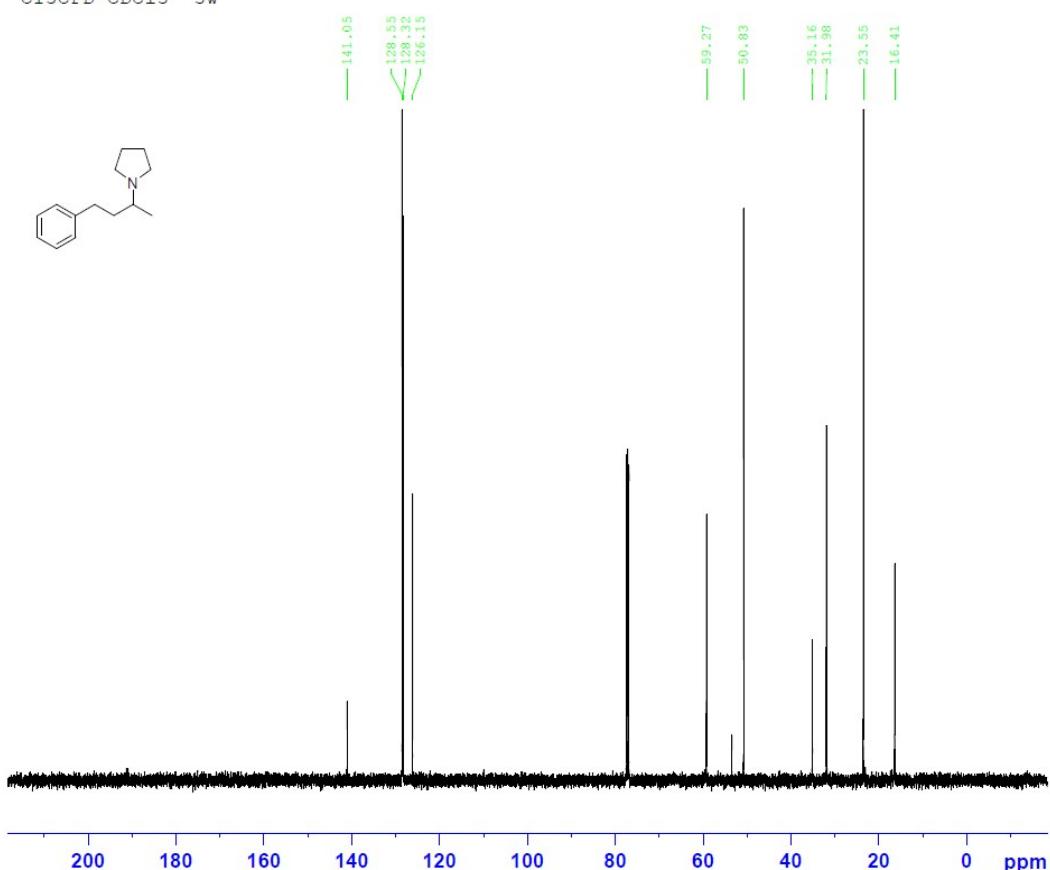
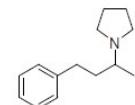
```

```

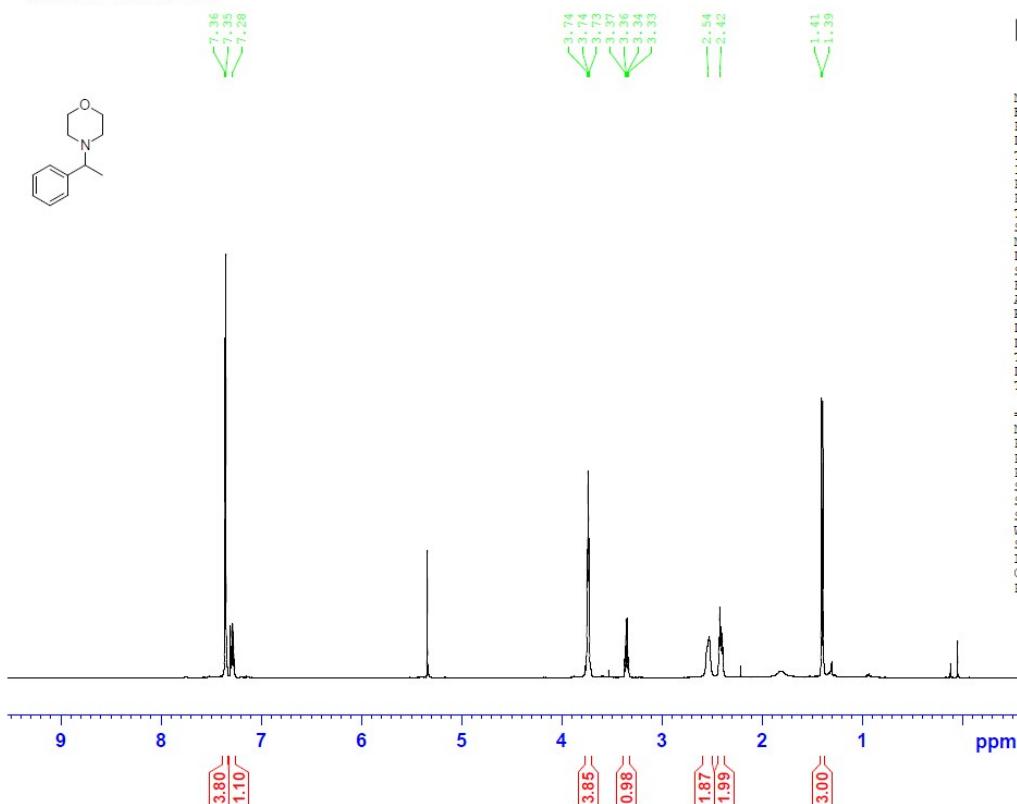
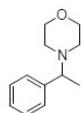
===== CHANNEL f1 =====
NUC1      1H
P1        12.60 usec
PL1      0.00 dB
PL1W    18.83080864 W
SFO1    500.13300885 MHz
SI        32768
SF      500.1300000 MHz
WDW        EM
SSB        0
LB        0.30 Hz
GB        0
PC        1.00

```

C13CPD CDC13 3w



PROTON CDC13 3ab

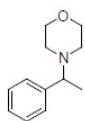
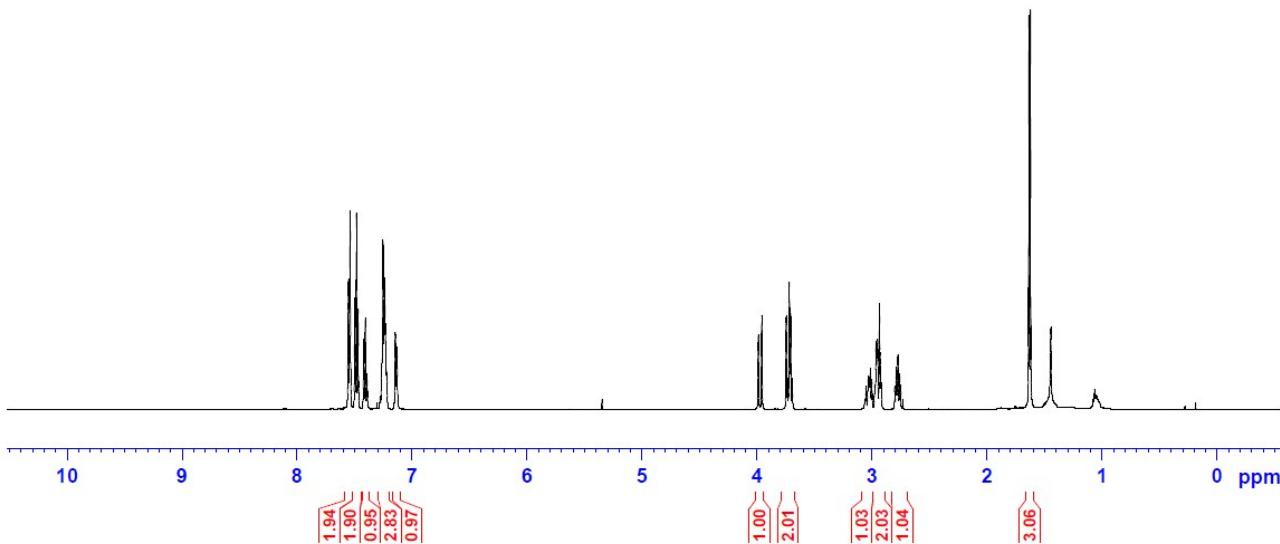
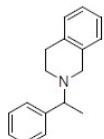


C13CPD CDCl₃ 3ab

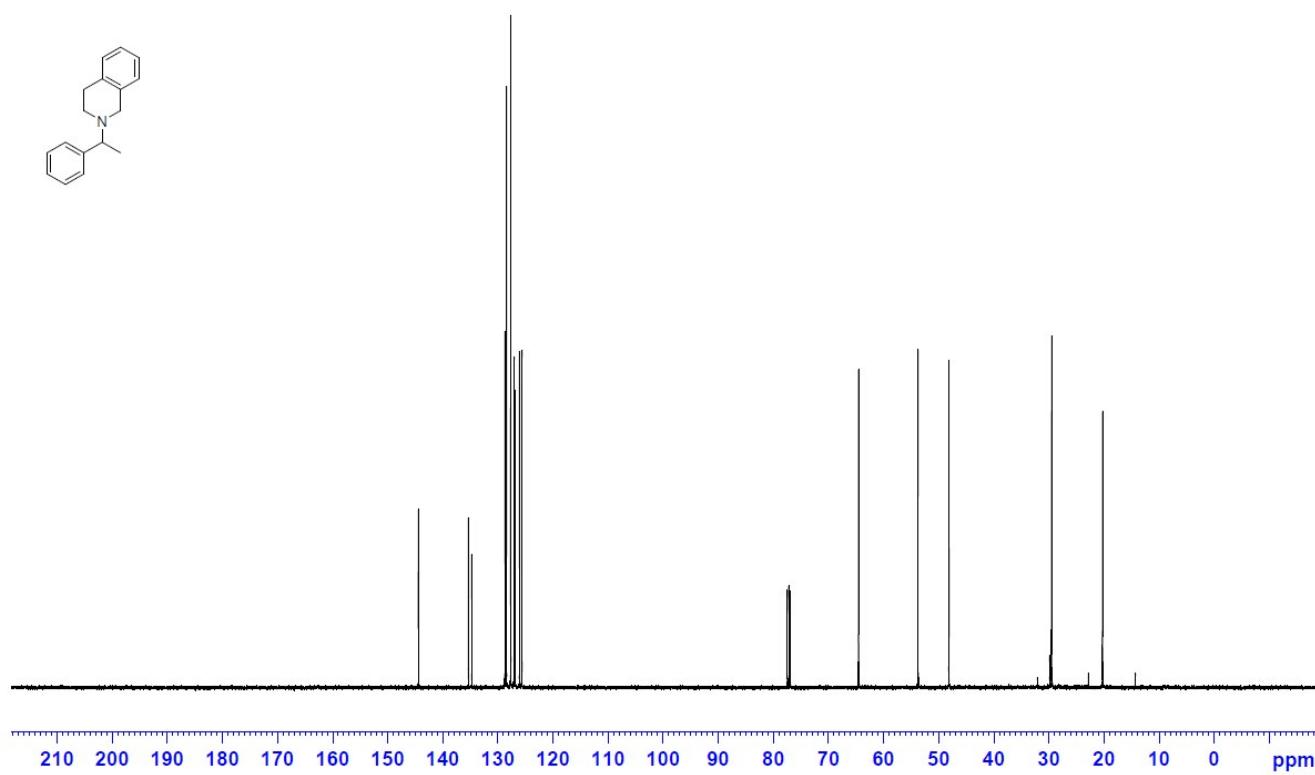
NAME CMX-20190225-21
 EXPNO 11
 PROCN0 1
 Date 20190225
 Time 15.41
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl₃
 NS 64
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010548 sec
 RG 203
 DW 16.800 usec
 DE 6.50 usec
 TE 300.4 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 ======
 NUC1 ¹³C
 P1 9.50 usec
 PL1 -1.00 dB
 PL1W 112.80287170 W
 SFO1 125.7703643 MHz

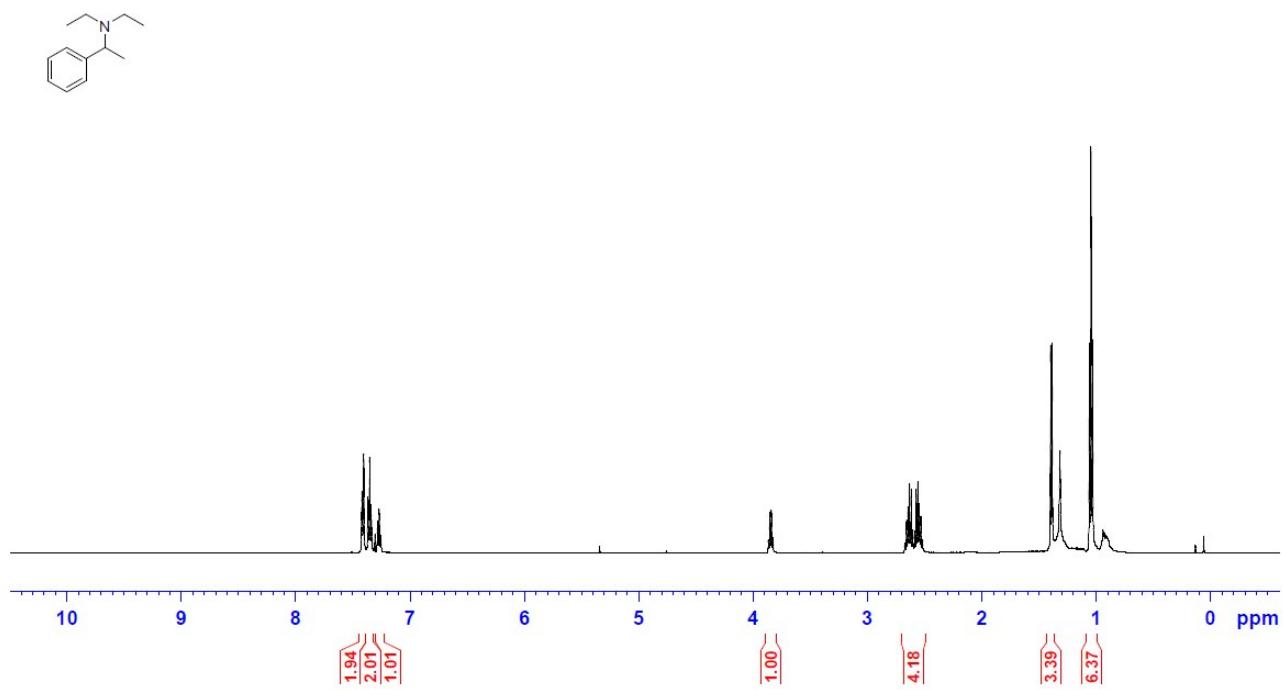
===== CHANNEL f2 ======
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 80.00 usec
 PL2 0.00 dB
 PL12 16.05 dB
 PL13 17.00 dB
 PL2W 18.83080864 W
 PL12W 0.46759412 W
 PL13W 0.37572402 W
 SFO2 500.1320005 MHz
 SI 32768
 SF 125.7577890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

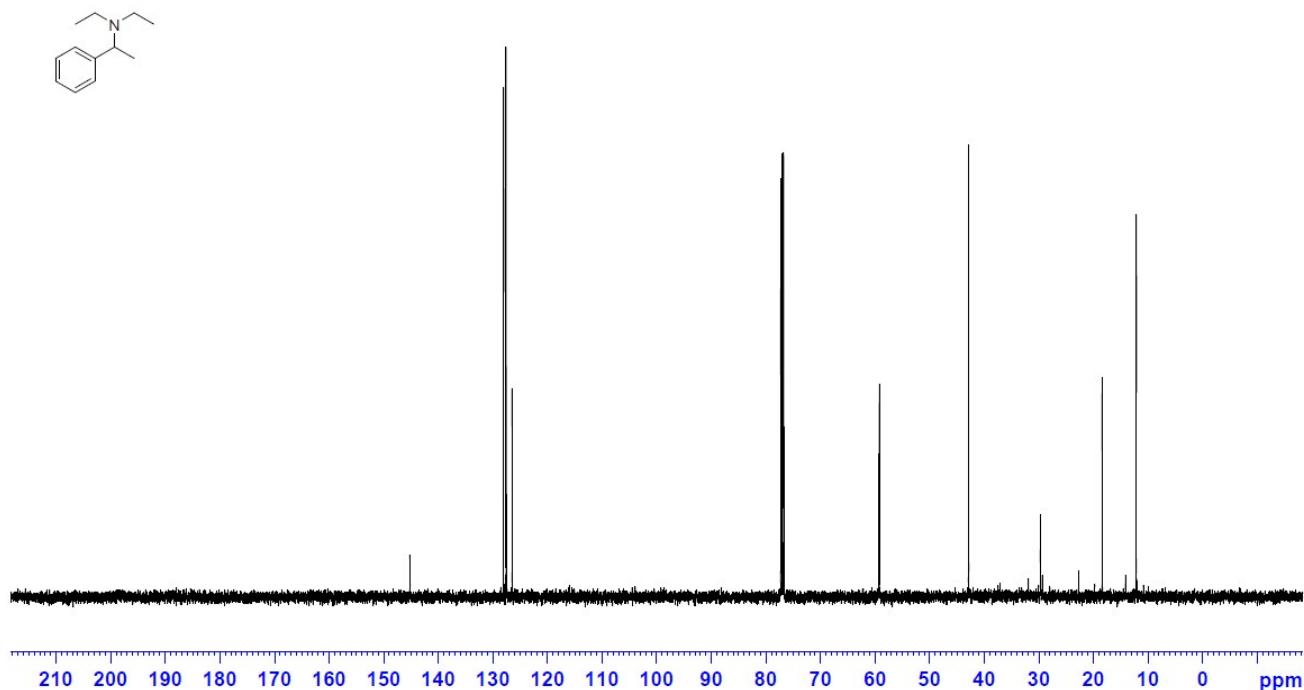
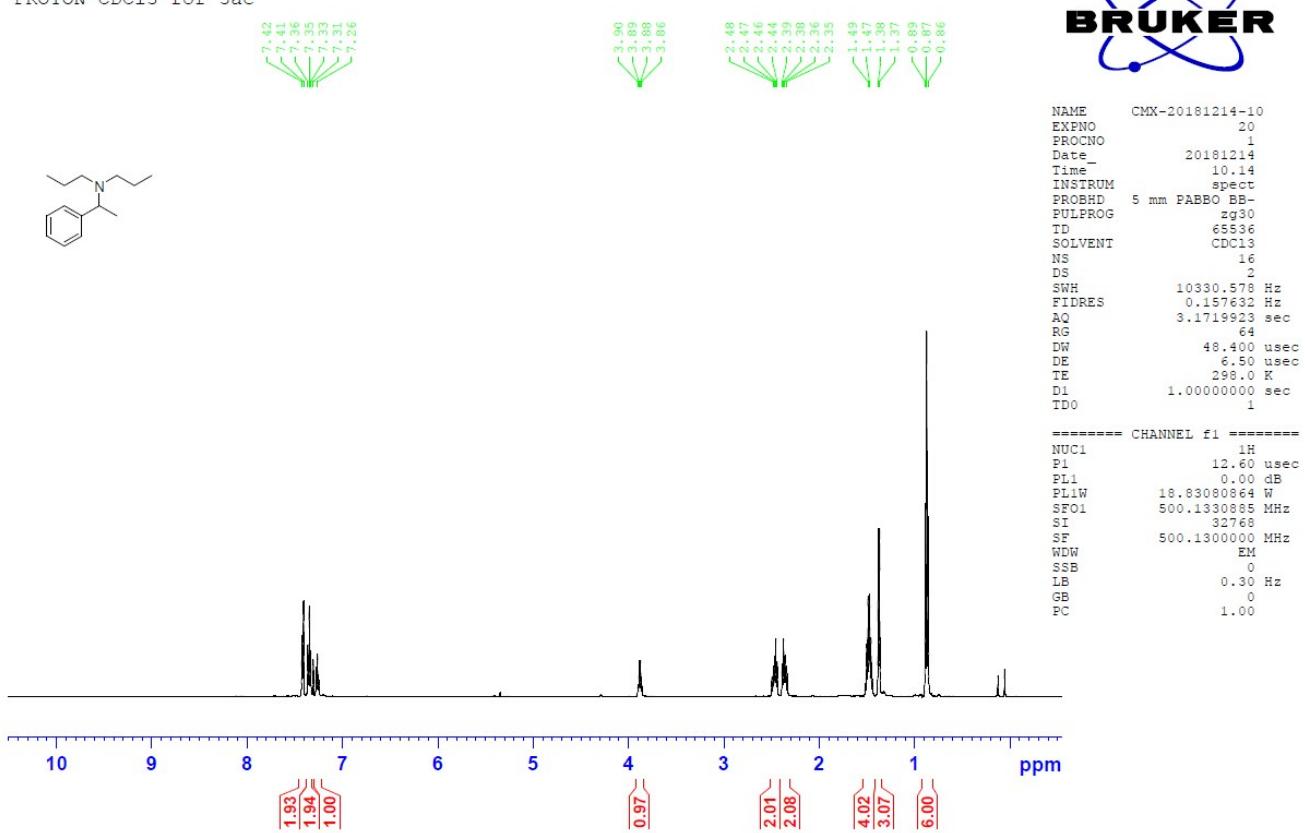
PROTON CDCl₃ for 3ac

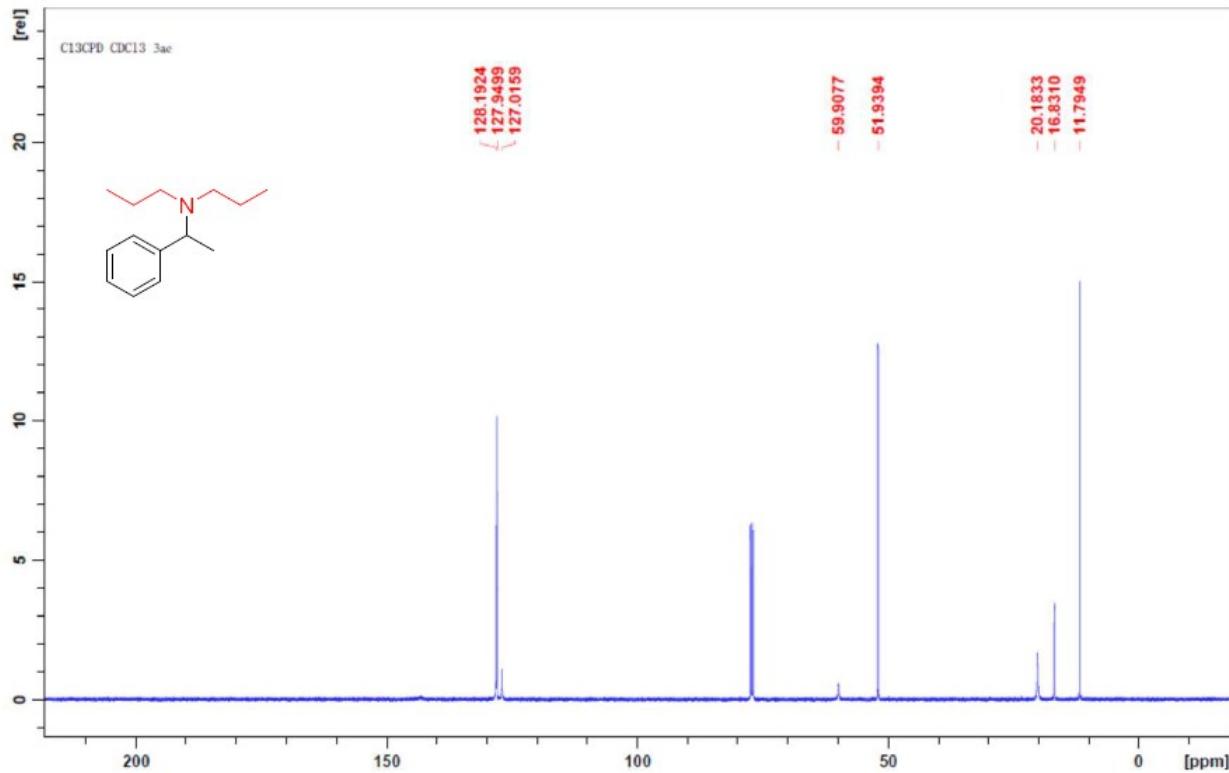
C13CPD CDCl₃ 3ac



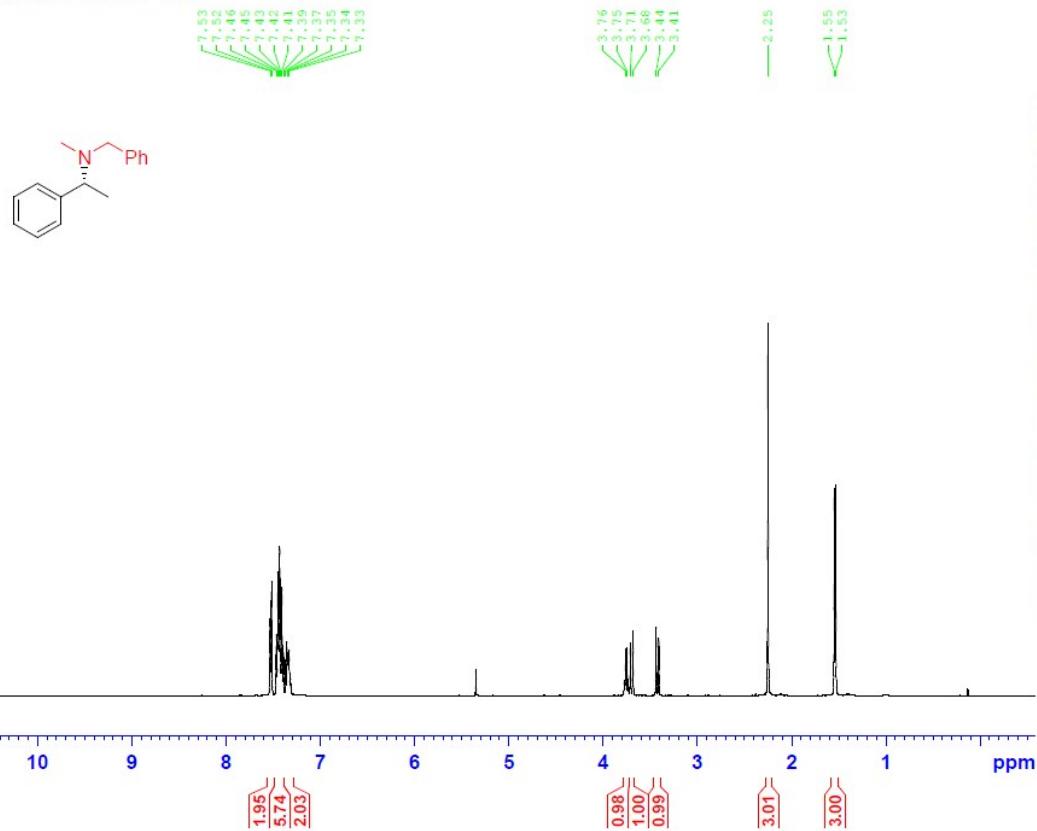
PROTON CDCl₃ for 3ad



PROTON CDCl₃ for 3ae



PROTON CDCl₃ 3af

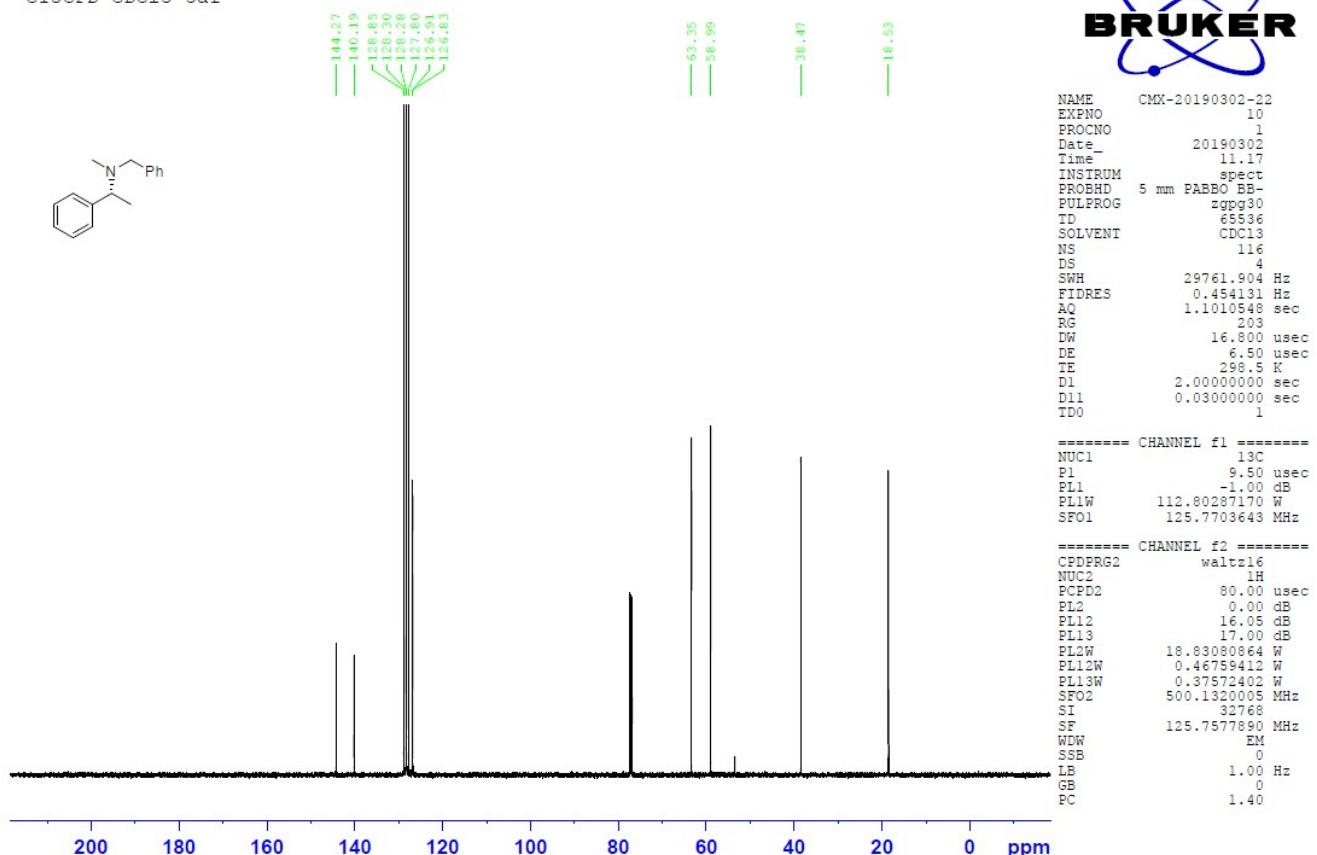
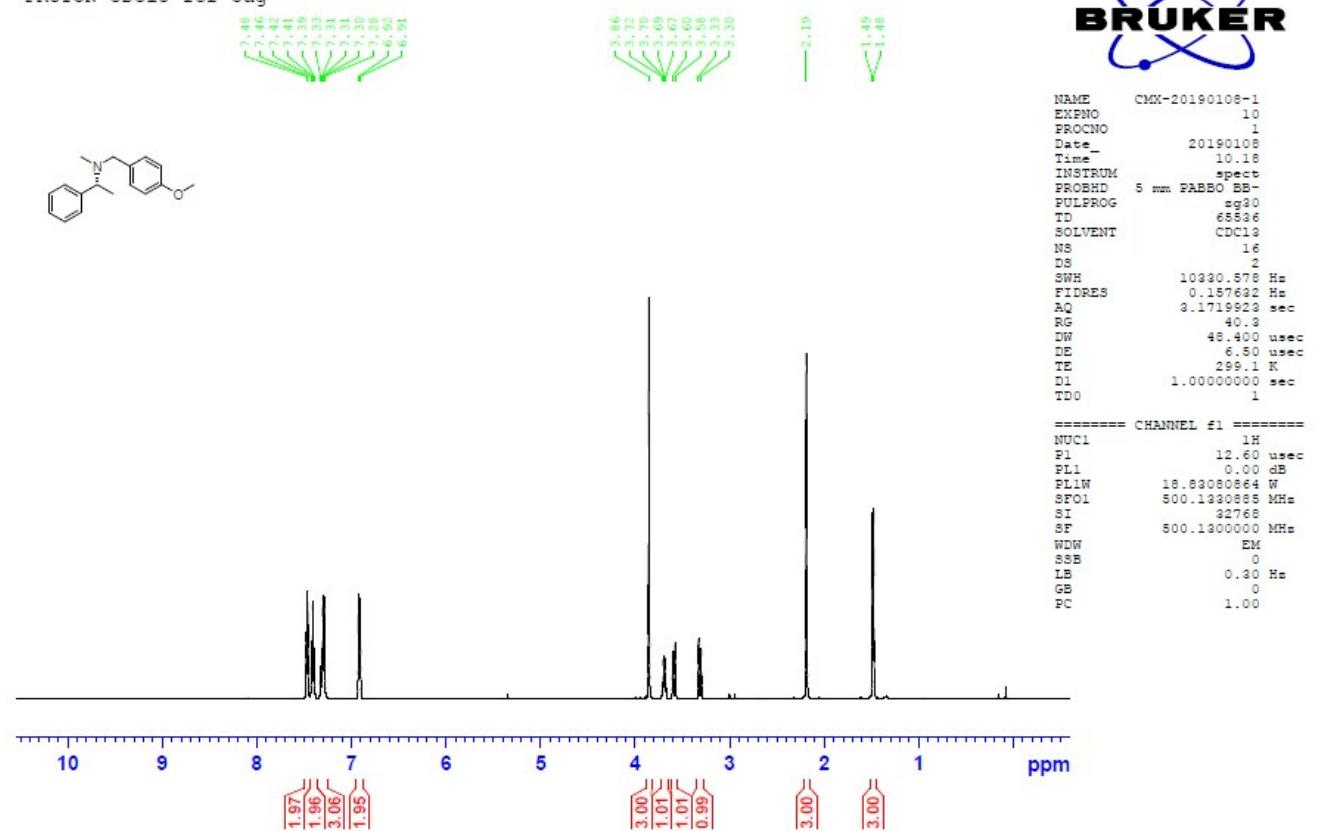


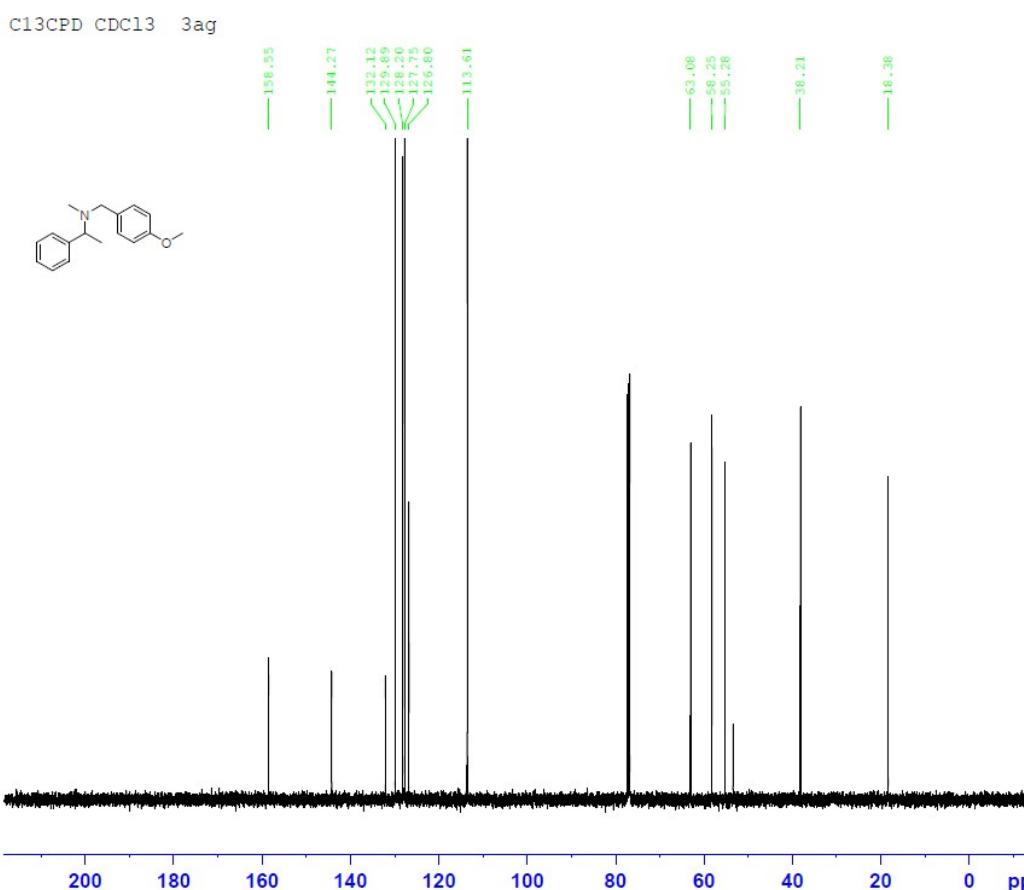
```

NAME      CMX-20190225-22
EXPNO     10
PROCNO    1
Date_     20190225
Time_     15.48
INSTRUM   spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD        65536
SOLVENT   CDCl3
NS        6
DS        2
SWH       10330.578 Hz
FIDRES   0.157632 Hz
AQ        3.1719923 sec
RG        28.5
DW        48.400 usec
DE        6.50 usec
TE        300.3 K
D1        1.0000000 sec
TDO      1

===== CHANNEL f1 =====
NUC1      1H
P1        12.60 usec
PLL      0.00 dB
PL1W     18.83080864 W
SFO1     500.1330865 MHz
SI        32768
SF        500.1300000 MHz
WDW      EM
SSB      0
LB        0.30 Hz
GB        0
PC        1.00

```

C13CPD CDCl₃ 3afPROTON CDCl₃ for 3ag





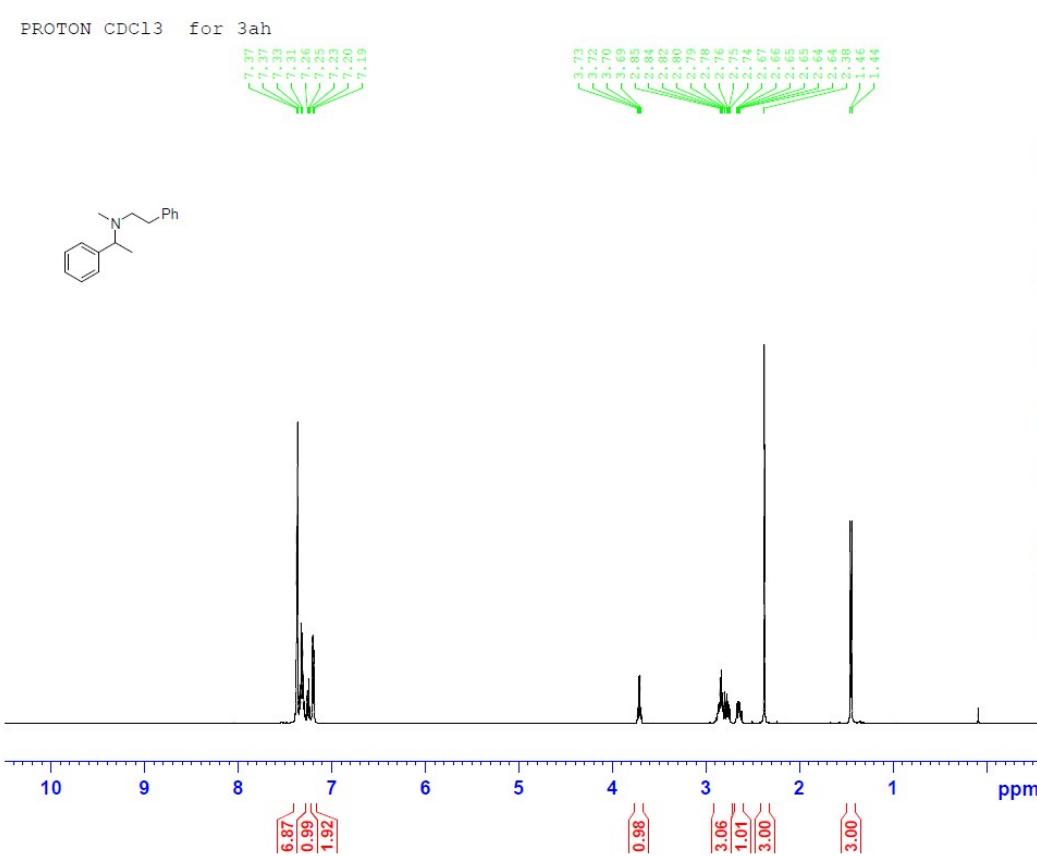
```

NAME CMX-20190225-15
EXPN 11
PROCNO 1
Date_ 20190225
Time 14.34
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgppg30
TD 65536
SOLVENT CDC13
NS 54
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 203
DW 16.800 usec
DE 6.50 usec
TE 301.0 K
D1 2.0000000 sec
D11 0.03000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.50 usec
PL1 -1.00 dB
PL1W 112.80287170 W
SF01 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 16.05 dB
PL13 17.00 dB
PL2W 18.83080864 W
PL12W 0.46759412 W
PL13W 0.37572402 W
SF02 500.1320005 MHz
SI 32768
SF 125.75778890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

```



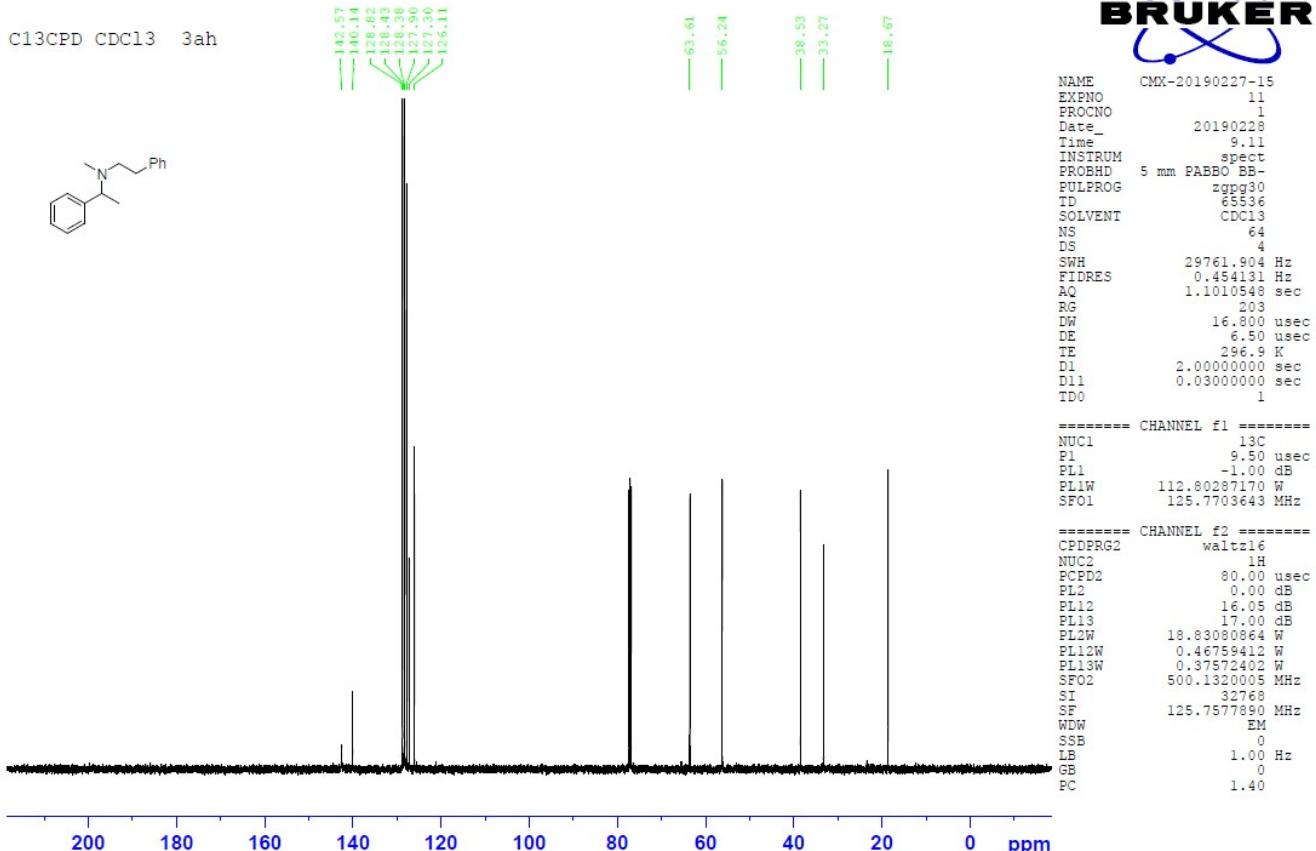
```

NAME      CMX-20181130-1
EXPN0     10
PROCNO   1
Date      20181130
Time_     22.34
INSTRUM  spect
PROBHD  5 mm PABBO BB-
PULPROG zg30
TD       65536
SOLVENT  CDC13
NS       8
DS       2
SWH     10330.578 Hz
FIDRES  0.157632 Hz
AQ      3.1719923 sec
RG      45.2
DW      48.400 usec
DE      6.50 usec
TE      301.4 K
D1      1.0000000 sec
TD0          1

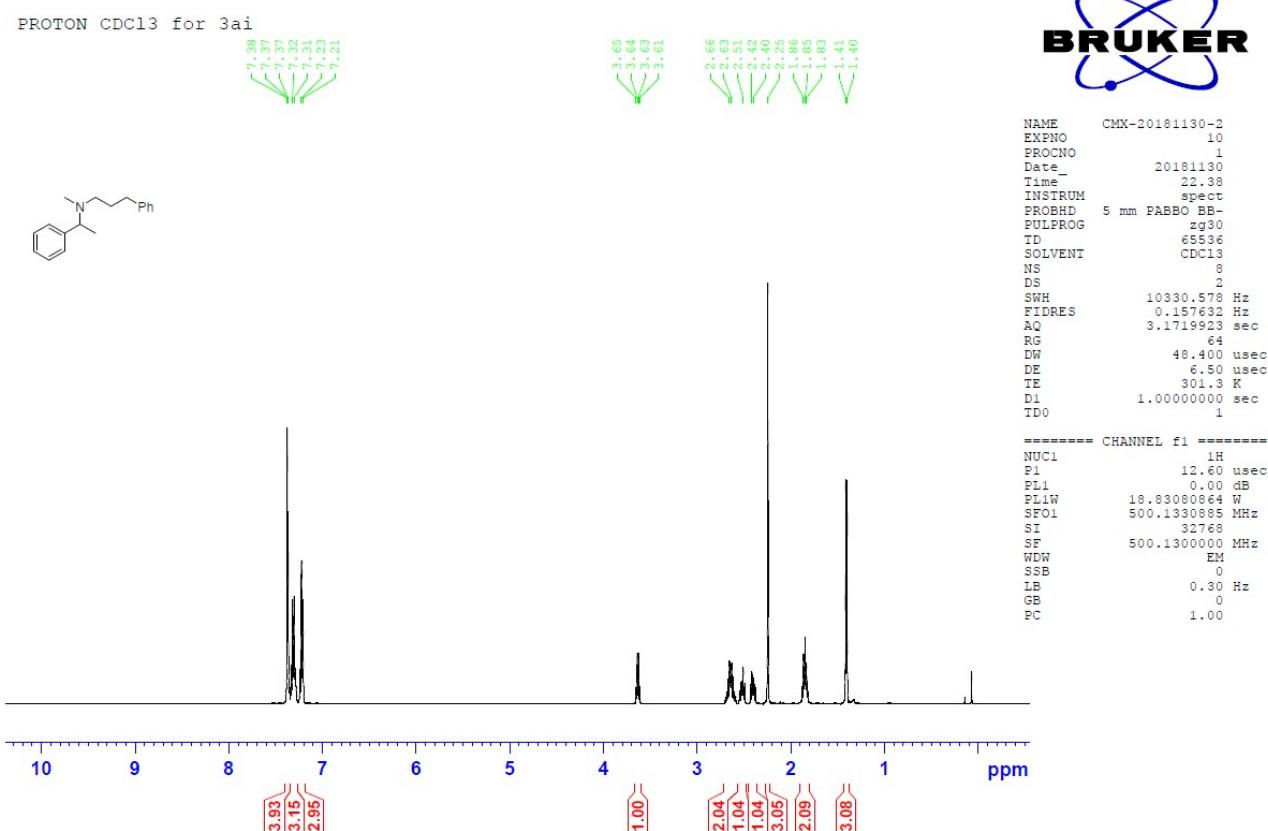
===== CHANNEL f1 =====
NUC1      1H
P1        12.60 usec
PL1      0.00 dB
PL1W    18.8300864 W
SF01    500.1330085 MHz
SI        32768
SF      500.1300000 MHz
WDW      EM
SSB      0
LB      0.30 Hz
GB      0
PC      1.00

```

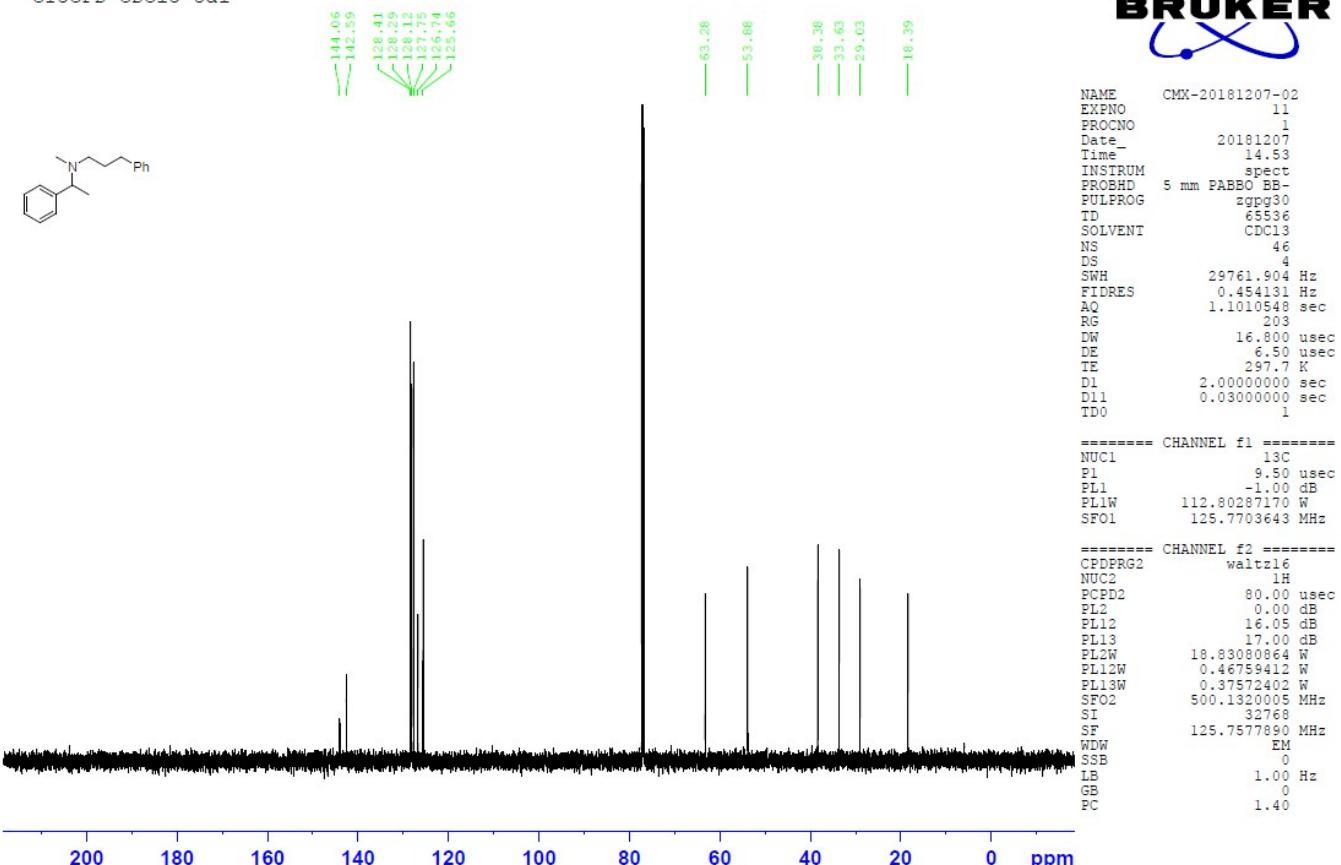
C13CPD CDCl₃ 3ah



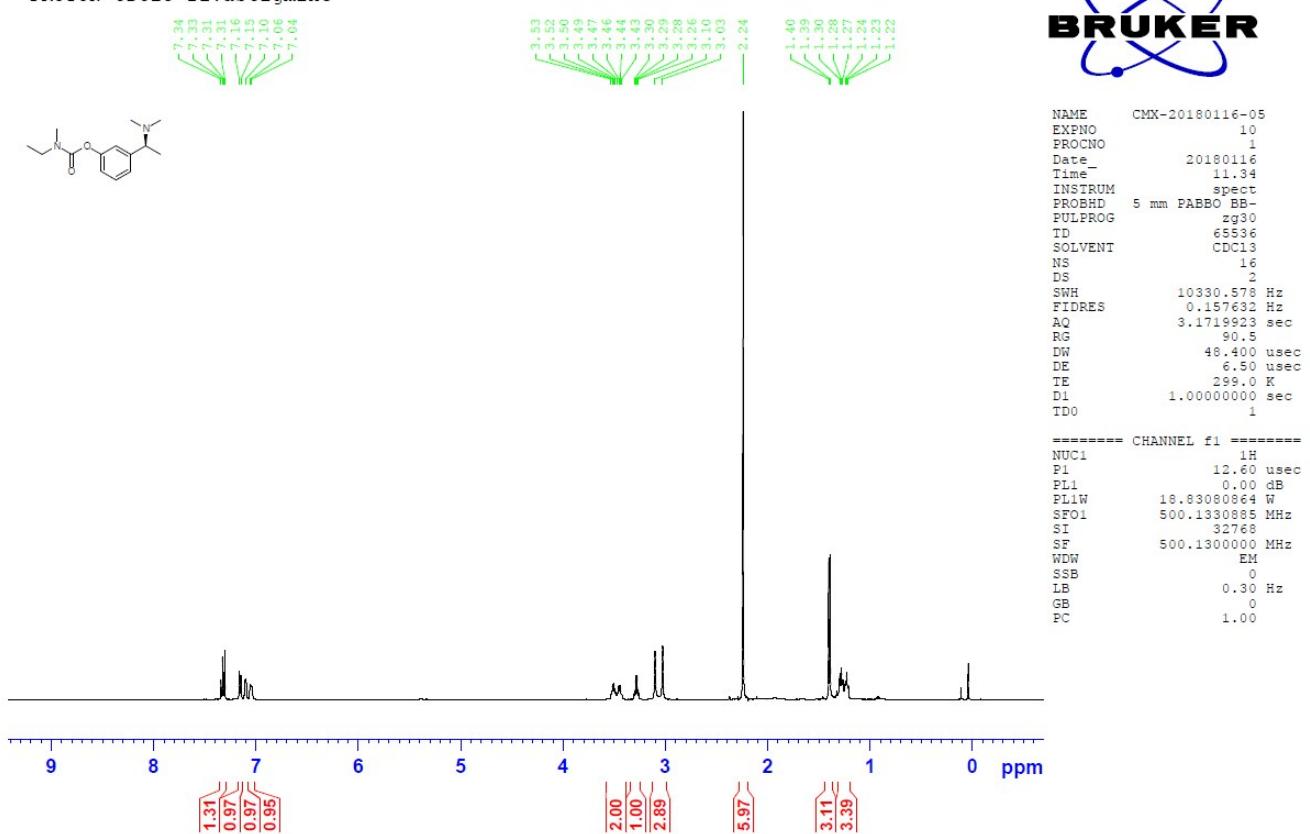
PROTON CDCl₃ for 3ai



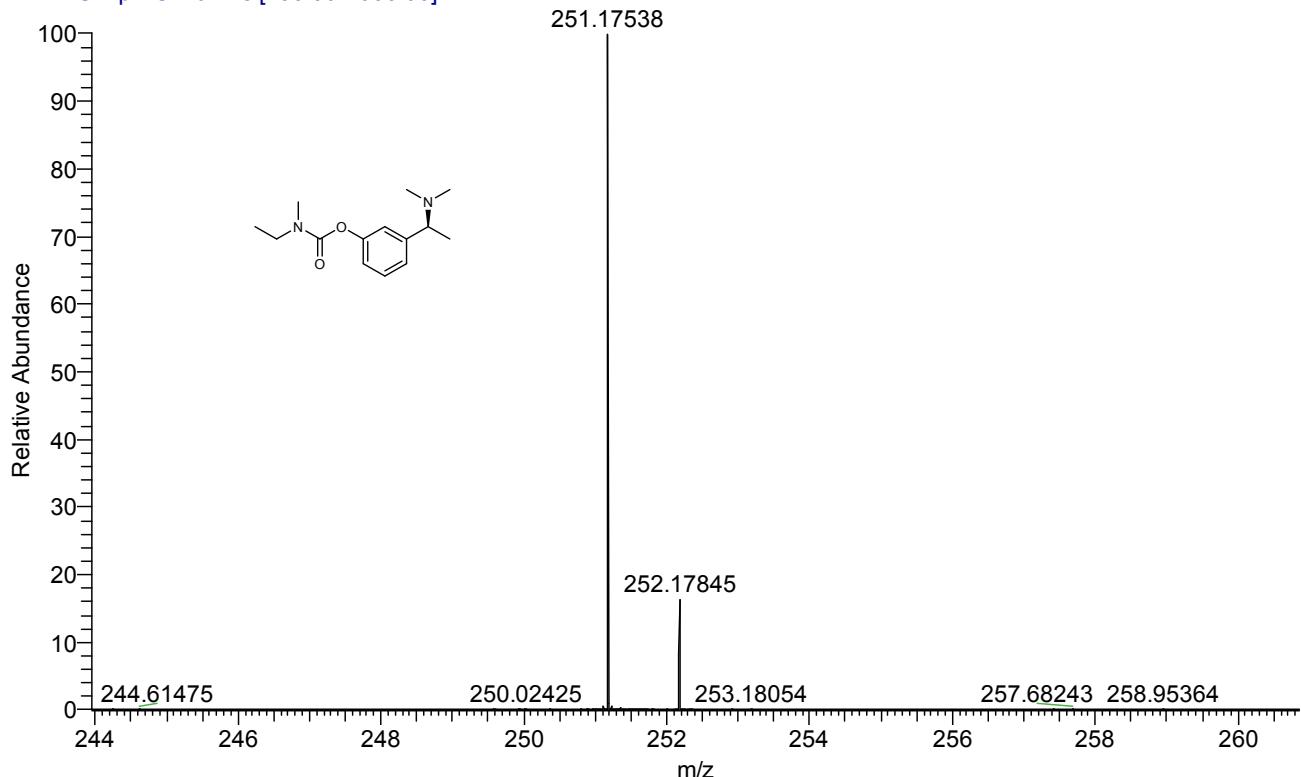
C13CPD CDCl₃ 3ai



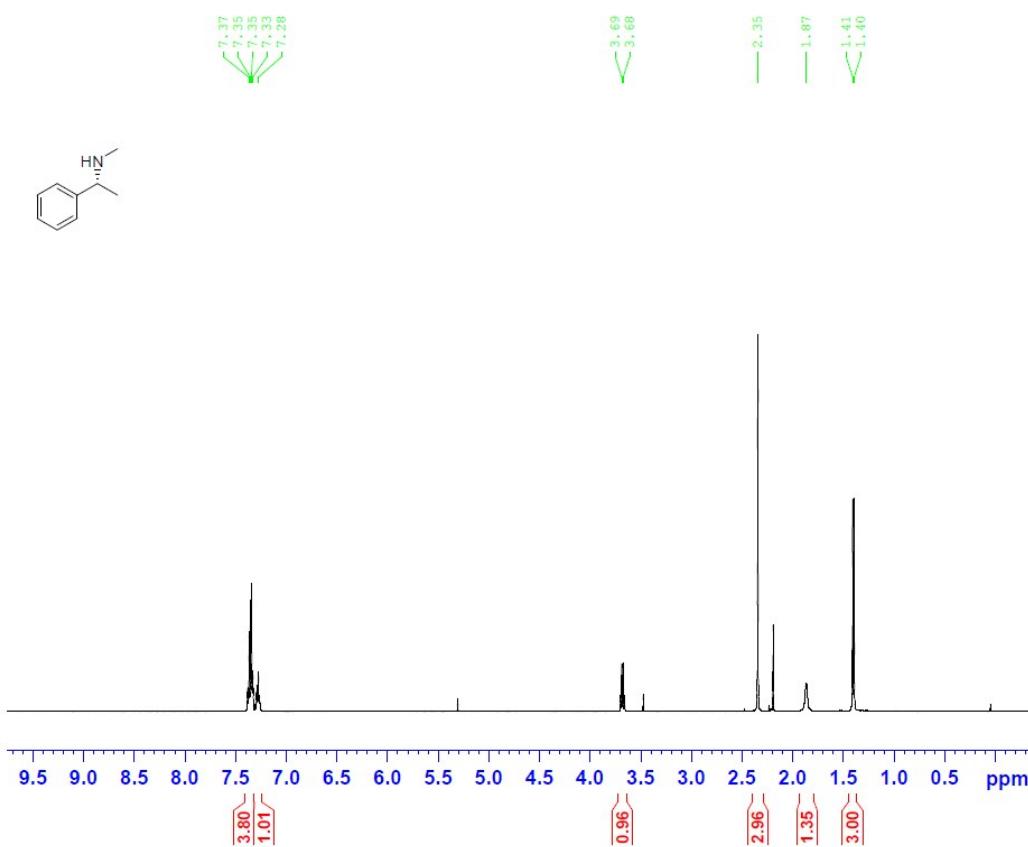
PROTON CDCl₃ rivastigmine



35 #15 RT: 0.21 AV: 1 NL: 9.48E8
T: FTMS + p ESI Full ms [150.00-2000.00]



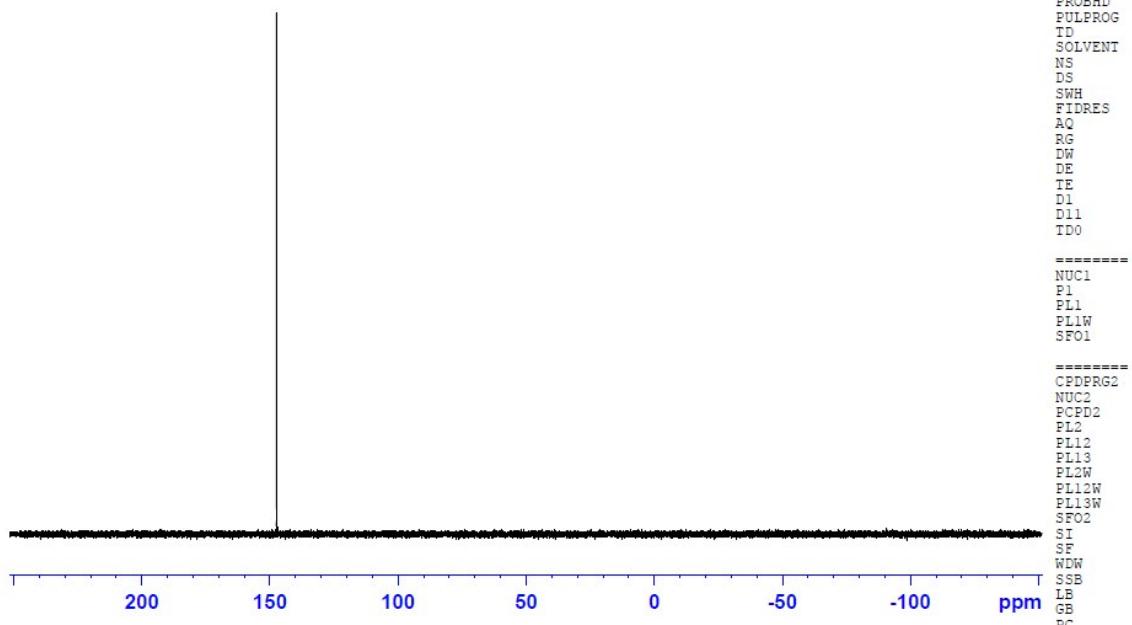
PROTON CDCl₃ 4



P31CPD CDC13 for L9i

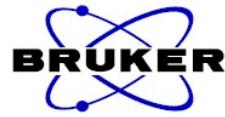


147.79

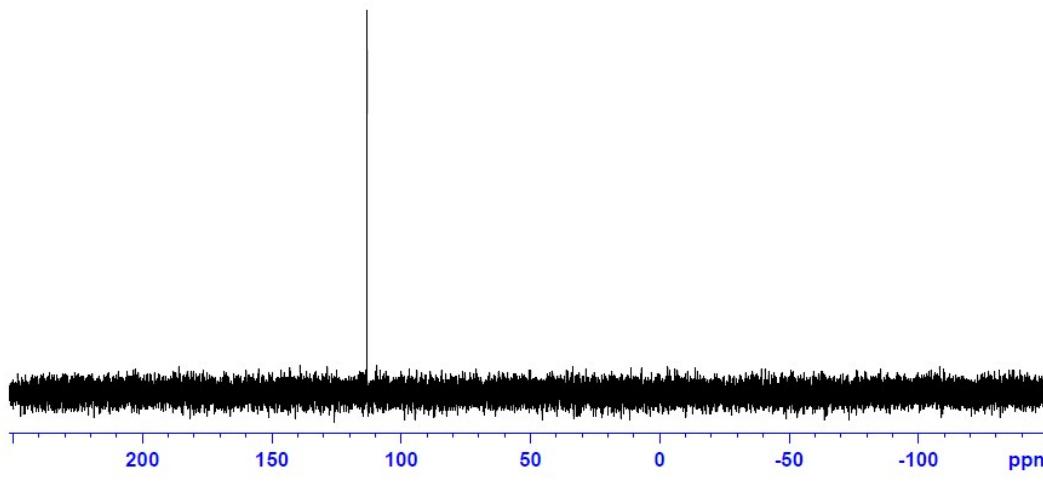


NAME CMX-20190103-09
EXPNO 10
PROCNO 1
Date 20190103
Time 9.30
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 203
DW 6.133 usec
DE 6.50 usec
TE 298.4 K
D1 2.0000000 sec
D11 0.03000000 sec
TDO 1
===== CHANNEL f1 ======
NUC1 31P
P1 11.30 usec
PL1 0.00 dB
PL1W 82.64508057 W
SFO1 202.4664576 MHz
===== CHANNEL f2 ======
CPDPFG2 waitz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 16.05 dB
PL13 17.00 dB
PL2W 18.83080864 W
PL12W 0.46759412 W
PL13W 0.37572402 W
SFO2 500.1320005 MHz
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

P31CPD CDC13 for Ir:L9i 1:1



113.23



NAME CMX-20190103-04
EXPNO 10
PROCNO 1
Date 20190103
Time 9.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 16
DS 4
SWH 81521.742 Hz
FIDRES 1.243923 Hz
AQ 0.4020041 sec
RG 203
DW 6.133 usec
DE 6.50 usec
TE 298.8 K
D1 2.0000000 sec
D11 0.03000000 sec
TDO 1
===== CHANNEL f1 ======
NUC1 31P
P1 11.30 usec
PL1 0.00 dB
PL1W 82.64508057 W
SFO1 202.4664576 MHz
===== CHANNEL f2 ======
CPDPFG2 waitz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 16.05 dB
PL13 17.00 dB
PL2W 18.83080864 W
PL12W 0.46759412 W
PL13W 0.37572402 W
SFO2 500.1320005 MHz
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

P31CPD CDCl₃ for Ir:I9i 1:2

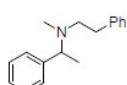


```

NAME      CMX-20190103-05
EXPNO     10
PROCNO    1
Date_     20190103
Time_     9.07
INSTRUM  spect
PROBHD  5 mm PABBO BB-
PULPROG zgpp30
TD       65536
SOLVENT   CDCl3
NS        16
DS         4
SWH      81521.742 Hz
FIDRES   1.243923 Hz
AQ       0.4020041 sec
RG        203
DW       6.133 usec
DE       6.50 usec
TE       298.8 K
D1      2.00000000 sec
D11     0.03000000 sec
TDO      1
===== CHANNEL f1 =====
NUC1      31P
P1        11.30 usec
PL1      0.00 dB
PL1W    82.64508057 MHz
SF01     202.4664576 MHz
===== CHANNEL f2 =====
CPDPG2   waltz16
NUC2      1H
PCPD2    80.00 usec
PL2      0.00 dB
PL12     16.05 dB
PL13     17.00 dB
PL2W    18.83080864 W
PL12W   0.46755412 W
PL13W   0.37572402 W
SF02     500.1320005 MHz
SI        32768
SF      202.4563305 MHz
WDW      EM
SSB      0
LB      1.00 Hz
GB      0
PC      1.40

```

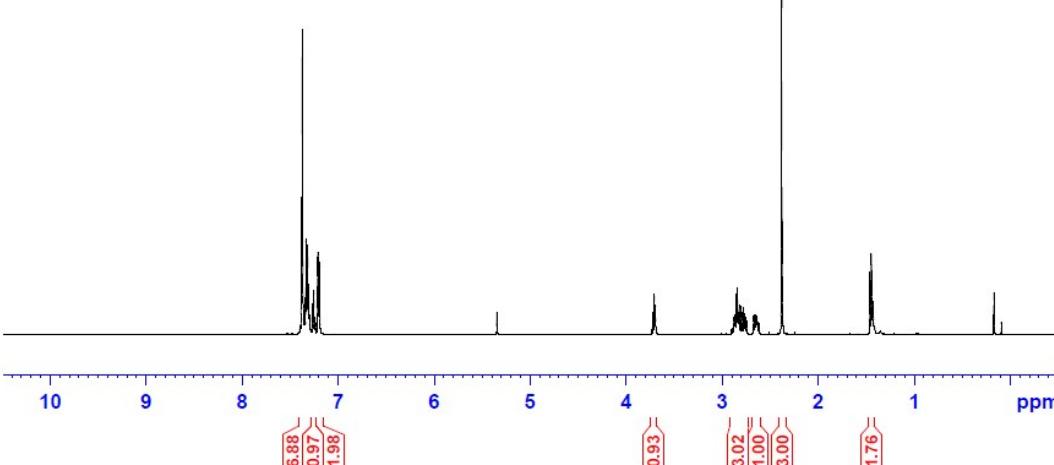
PROTON CDCl₃ for 3ah from 1a-d3



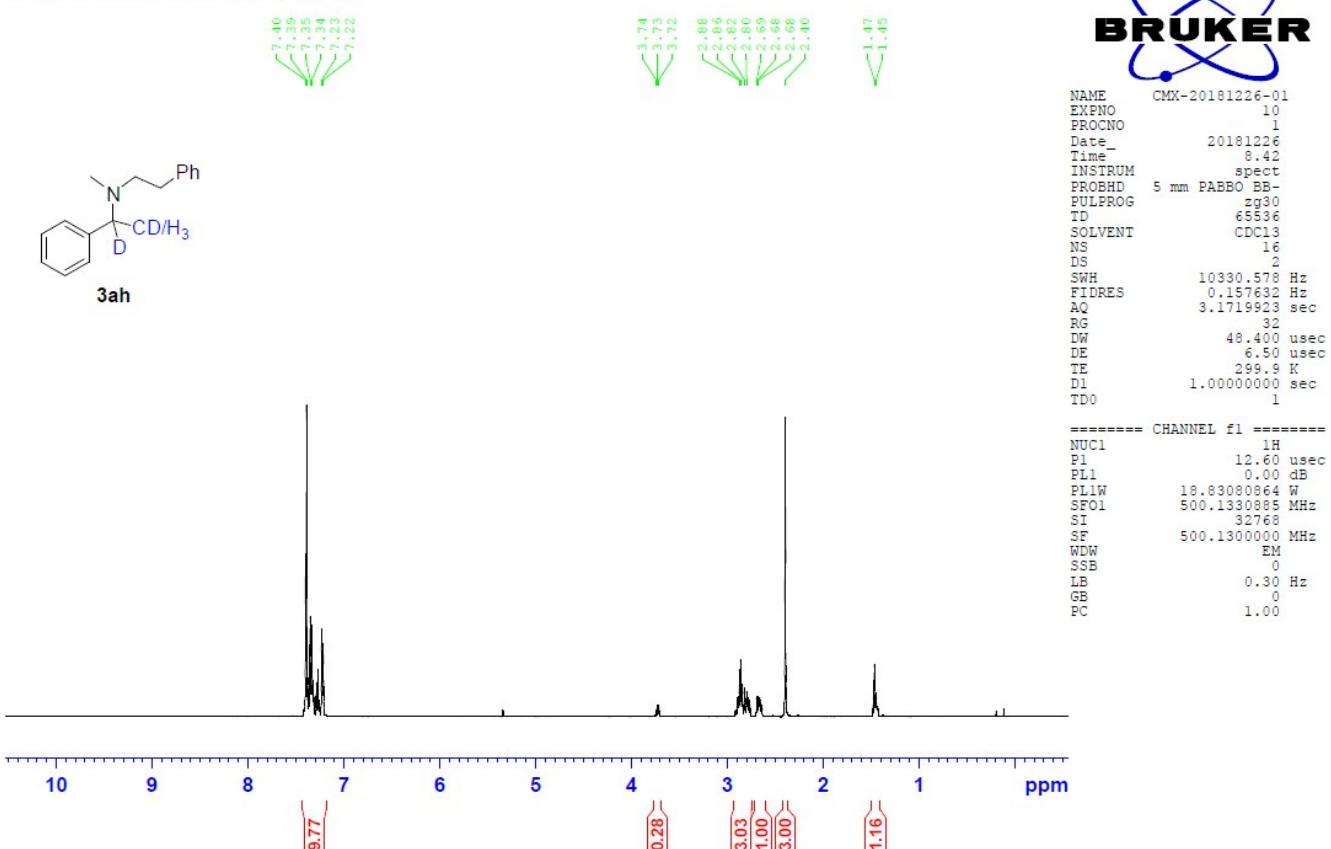
```

NAME      CMX-20190103-01
EXPNO     10
PROCNO    1
Date_     20190103
Time_     8.48
INSTRUM  spect
PROBHD  5 mm PABBO BB-
PULPROG zg30
TD       65536
SOLVENT   CDCl3
NS        16
DS         2
SWH      10330.578 Hz
FIDRES   0.157632 Hz
AQ       3.1719923 sec
RG        40.3
DW       48.400 usec
DE       6.50 usec
TE       298.7 K
D1      1.0000000 sec
TDO      1
===== CHANNEL f1 =====
NUC1      1H
P1        12.60 usec
PL1      0.00 dB
PL1W    18.83080864 W
SF01     500.1330885 MHz
SI        32768
SF      500.1300000 MHz
WDW      EM
SSB      0
LB      0.30 Hz
GB      0
PC      1.00

```



PROTON CDCl₃ for 3ah with D₂



PROTON CDCl₃ for 3ah with MeOD

