

Supplementary Materials for

Inhibition of somatosensory mechanotransduction by annexin A6

Ramin Raouf, Stéphane Lolignier, Jane E. Sexton, Queensta Millet,
Sonia Santana-Varela, Anna Biller, Alice M. Fuller, Vanessa Pereira, Jyoti S. Choudhary,
Mark O. Collins, Stephen E. Moss, Richard Lewis, Julie Tordo, Els Henckaerts,
Michael Linden, John N. Wood*

*Corresponding author. Email: j.wood@ucl.ac.uk

Published 19 June 2018, *Sci. Signal.* **11**, eaao2060 (2018)

DOI: 10.1126/scisignal.aao2060

The PDF file includes:

Fig. S1. Isolation of NMB-1 peptide-binding partners.
Fig. S2. Interaction between NMB-1 and annexin A6.
Fig. S3. Representative traces of MA currents in vitro.
Fig. S4. ATP6V1B2 does not confer mechanosensitivity to HEK or ND-C cells but inhibits Piezo2 current.
Fig. S5. Transduction of DRG neurons using AAV-TT serotype.
Table S1. Top NMB-1-binding candidates in ND-C cells.
Legends for data files S1 and S2

Other Supplementary Material for this manuscript includes the following:

(available at www.sciencesignaling.org/cgi/content/full/11/535/eaao2060/DC1)

Data file S1 (Microsoft Excel format). NMB-1-binding candidates in DRG neurons.

Data file S2 (Microsoft Excel format). NMB-1-binding candidates from differentiated ND-C cells.

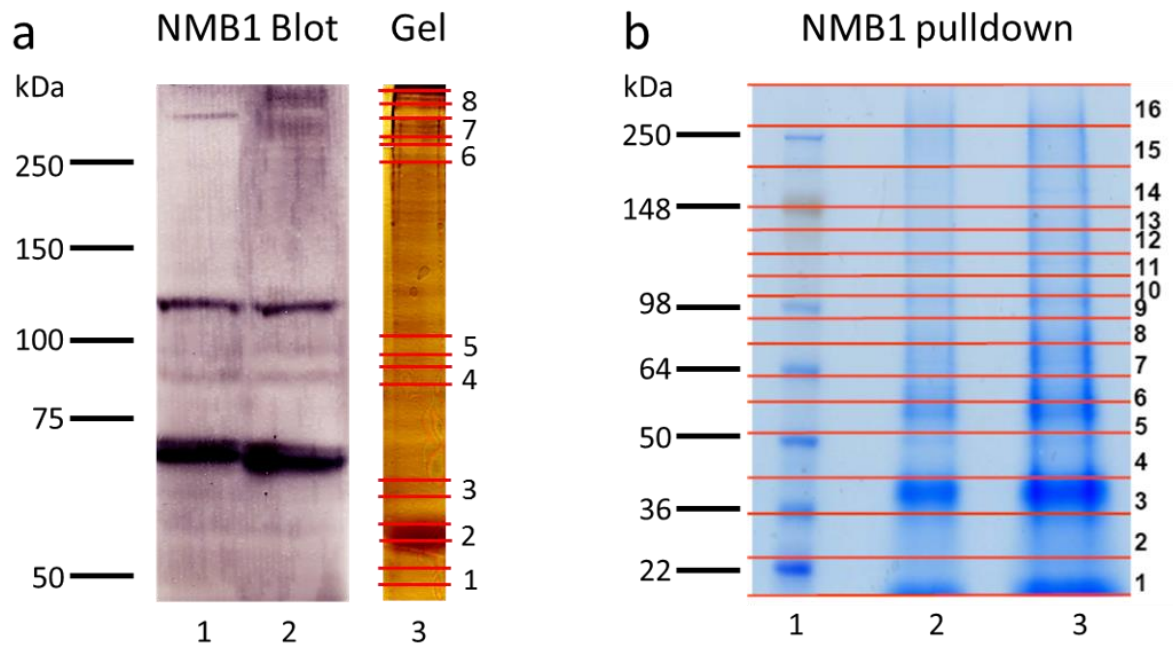


Figure S1. Isolation of NMB-1 peptide-binding partners. (a) Biotinylated NMB-1 was used to visualize binding silver stained SDS gel loaded with DRG protein extract processed under the same conditions is juxtaposed alongside the NMB-1 stained blot (3). Silver stained gel with reactive bands on the NMB-1 blot that were excised (location of the bands shown with red lines) and processed for LC MS/MS analysis. (b) NMB-1 pulled from differentiated ND-C cell protein extracts was treated with 2% (lane 2) or 4% (lane 3) beta-mercaptoethanol and resolved on 10% SDS gel. The gel was cut into 16 strips and processed for LC MS/MS analysis.

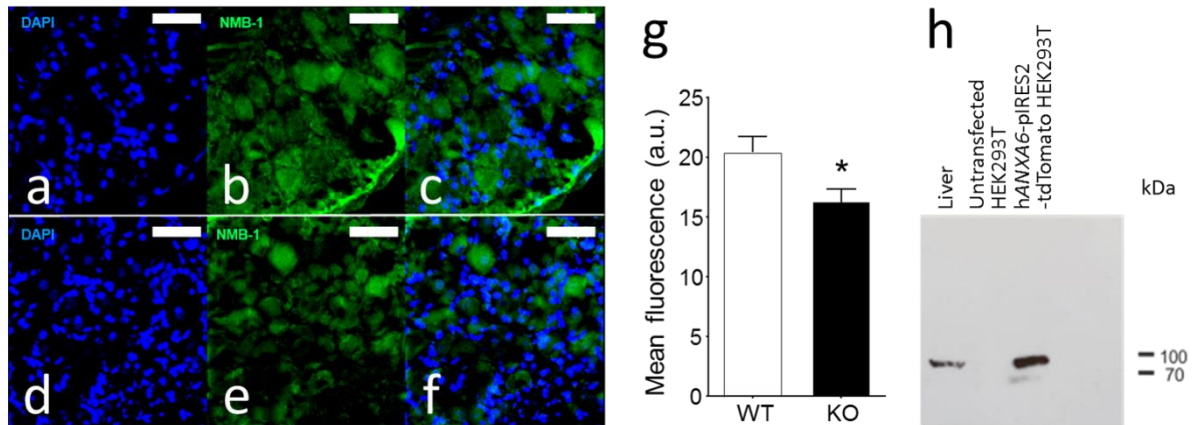


Figure S2. Interaction between NMB-1 and annexin A6. (a-g) WT (a-c) and *Anxa6* KO (d-f) DRG sections were incubated with bNMB-1 followed by FITC-Avidin (scale bars = 50 μ m) and the fluorescence intensity was quantified (g) ($p=0.0298$, $n=10$ and represent sections obtained from 2 mice per group). (h) Direct interaction of annexin A6 and NMB-1 was probed using Western blot. The panel shows expression of annexin A6 in mouse liver, *hANXA6*-pIRES2-tdTomato expressing HEK293T cells, and untransfected HEK293T cells (negative control).

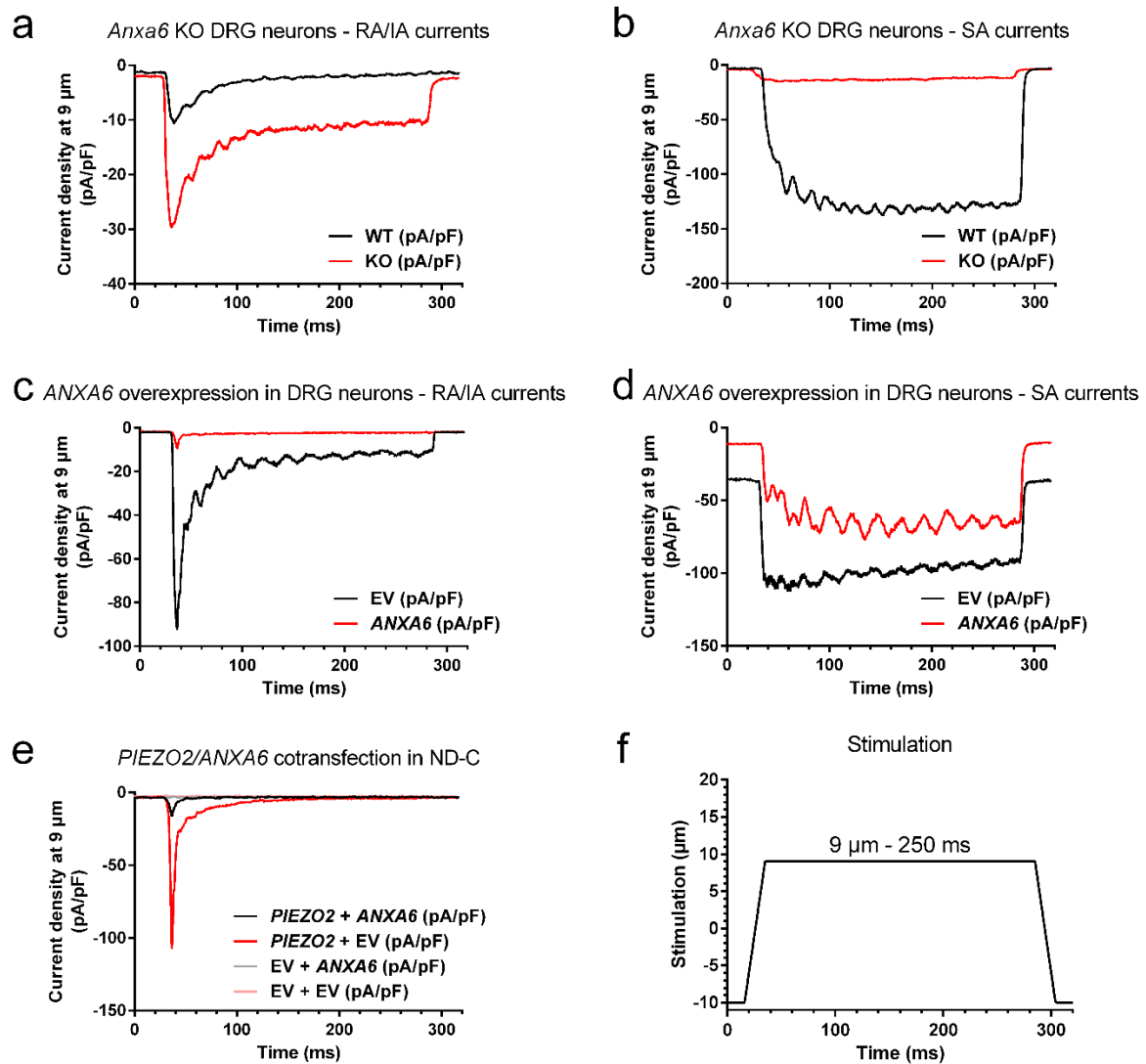


Figure S3. Representative traces of MA currents in vitro. (a-b) RA/IA and SA currents recorded in *Anxa6* KO DRG neurons, representative of the data given in Fig. 2. (c-d) RA/IA and SA currents recorded in WT DRG neurons overexpressing *hANXA6*, representative of the data given in Fig. 3. (e) RA currents recorded in ND-C cells transfected with *hPIEZO2* ± *hANXA6* or their respective empty vectors, representative of the data given in Fig. 4. (f) All current traces shown were recorded in response to a 9 μm deformation of the plasma membrane by a firepolished glass pipette, lasting for 250 ms.

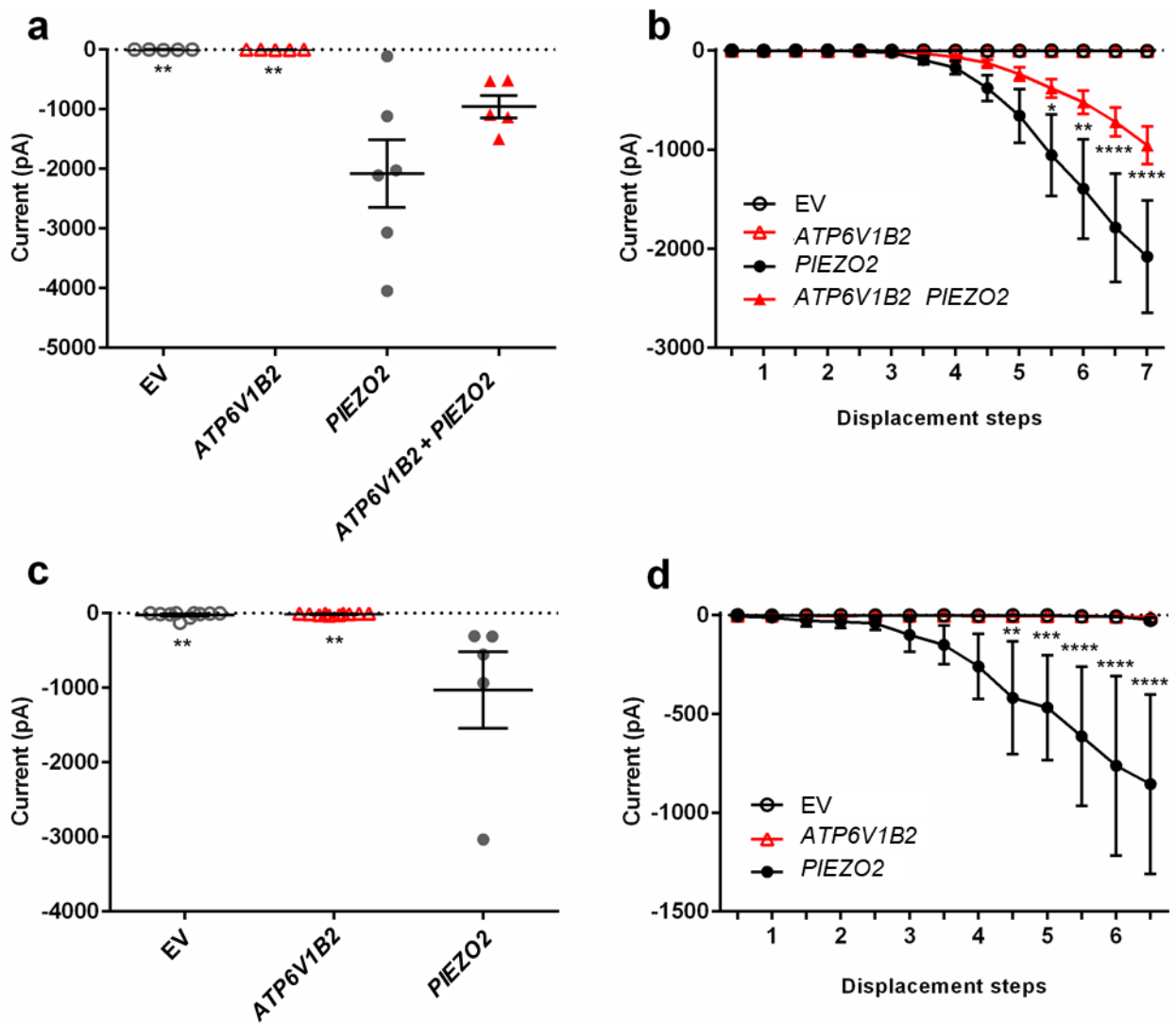


Figure S4. ATP6V1B2 does not confer mechanosensitivity to HEK or ND-C cells but inhibits Piezo2 current. Peak current (a) and current-displacement curve (b) for HEK cells transfected with hATP6V1B2. Peak current (c) and current-displacement curve (b) for ND-C cells transfected with hATP6V1B2. (HEK cells: ATP6V1B2 n=10; EV n=10; PIEZO2 n=5; ATP6V1B2 + PIEZO2 n=5; ND-C cells: ATP6V1B2 n=10; EV n=10; PIEZO2 n=5). Data are shown as Mean±SEM. One way ANOVA with Dunnett's post-hoc test (statistics shown on graph is each group compared to PIEZO2) (a,c); Two-way repeated measures ANOVA with Tukey post-hoc test (statistics shown on graph is ATP6V1B2+PIEZO2 compared to PIEZO2) (b); Two-way repeated measure ANOVA with Tukey post-hoc test (statistics shown on graph is ATP6V1B2 compared to PIEZO2) (d). *p<0.05, **p<0.01, ***p<0.001; ****p<0.0001.

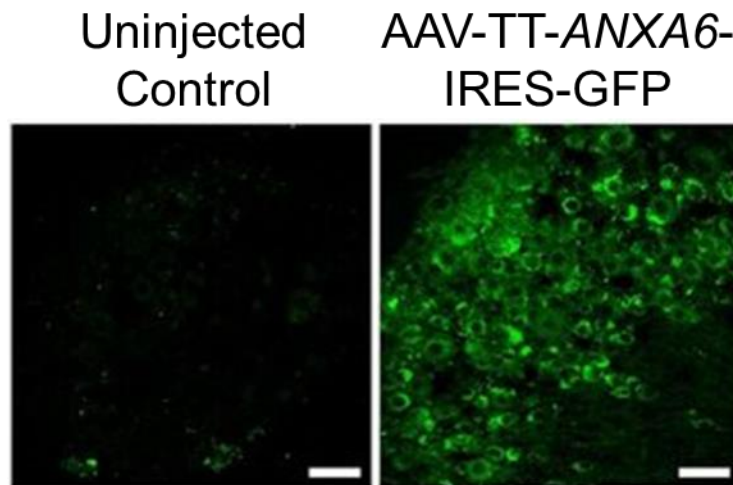


Figure S5. Transduction of DRG neurons using AAV-TT serotype. 15 weeks after intrathecal injection, robust GFP expression was observed in lumbar DRG of animals treated with AAV-TT-*hANXA6*-IRES-GFP compared to untreated animals. Similar results were obtained from 3 different animals per condition. Scale bar, 50 μ m.

Table S1. Top NMB-1-binding candidates in ND-C cells.

Protein		UniprotKB	MW	NMB-1 Peptide hits
Anxa6	Annexin A6	P14824	75,838	24
Atic	5-aminoimidazole-4-carboxamide ribonucleotide formyltransferase/IMP cyclohydrolase	B0LAC5	64,117	6
Atp6v1b2	Vacuolar ATP synthase subunit B, brain isoform	P62814	56,515	4
Abcb10	ATP-binding cassette sub-family B member 10, mitochondrial	Q9JI39	73,972	4
Anxa5	Annexin A5	P48036	35,730	4
G6pdx	Glucose-6-phosphate 1-dehydrogenase X	Q00612	59,225	4
Hspa9	Stress-70 protein, mitochondrial precursor	P38647	44,145	3
Rps3	40S ribosomal protein S3	P62908	26,674	3
Msn	Moesin	P26041	67,725	3
Stxbp1	Isoform 1 of Syntaxin-binding protein 1	O08599	67,526	3
Hspa11	Heat shock 70 kDa protein 1L	P16627	70,593	3
Ppp2r1a	Serine/threonine-protein phosphatase 2A 65 kDa regulatory subunit A alpha isoform	Q76MZ3	65,281	3
Vim	Vimentin	P20152	53,655	2

Data File S1. NMB-1-binding candidates in DRG neurons.

Data File S2. NMB-1-binding candidates from differentiated ND-C cells.