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**Supplemental Information** 

Dorsal-Ventral Differences in Neural Stem Cell Quiescence Are Induced by p57<sup>KIP2</sup>/Dacapo Leo Otsuki and Andrea H. Brand



## Figure S1, Related to Figure 1. *dap* knockdown in NSCs leads to a switch from G<sub>0</sub> to G<sub>2</sub> <u>quiescence and acceleration in stem cell activation timing</u>

(A) Proportions of G<sub>0</sub> and G<sub>2</sub> quiescent NSCs in control (*wor*-GAL4>*mCherry* RNAi) *versus dap* knockdown (*wor*-GAL4>*dap* RNAi) brains at 0ALH. Control:  $28\pm1.0\%$  G<sub>0</sub>. *dap* knockdown:  $10\pm0.5\%$  G<sub>0</sub>. *n*=10 tVNCs (control) or 7 tVNCs (knockdown), ~140 NSCs each. \*\*\*: *p*=5.04x10<sup>-10</sup>, Welch's test.

(**B**) Activated NSCs in control (*wor*-GAL4>*mCherry* RNAi) *versus dap* knockdown (*wor*-GAL4>*dap* RNAi) tVNCs at 20 hours ALH. Wor expression was used to label activated stem cells (Otsuki and Brand, 2018). *n*=10 tVNCs (control) or 9 tVNCs (RNAi), ~130 NSCs each. \*\*\*: *p*=7.65x10<sup>-4</sup>, Student's *t*-test.



## Figure S2, Related to Figure 2. Quiescent NSCs do not transcribe or translate dap

(**A-B**) RNA polymerase II occupancy at the *dap* and *dpn* loci in quiescent NSCs, as determined by Targeted DamID. RNA polymerase II is poised at the 5' end of the *dap* locus, but *dap* is not transcribed (A), in contrast to *dpn* which is actively transcribed in quiescent NSCs (B). Axis depicts log<sub>2</sub> ratio change between test and reference samples. Data from Otsuki and Brand, 2018.

(C) Quiescent NSCs do not express Dap protein at 0ALH. Single hemi-segment of a tVNC co-stained to visualise NSCs (red) and Dap (green) at 0ALH. Dotted line indicates the ventral midline. Anterior is up and dorsal is to the right. Maximum intensity projection.



## Figure S3, Related to Figure 3. Dorsal-ventral patterning of NSCs in the tVNC

(A) Map overlaying the distribution of  $G_0$  (red) and  $G_2$  (cyan) quiescent NSCs with the Msh<sup>+</sup> (pale green), Ind<sup>+</sup> (pale red) and Vnd<sup>+</sup> (pale blue) regions of the tVNC. Map assembled using data from (Chu et al., 1998; D'Alessio and Frasch, 1996; Isshiki et al., 1997; McDonald et al., 1998; Otsuki and Brand, 2018; Weiss et al., 1998). Dotted line indicates ventral midline. A: anterior. P: posterior.

(**B**) Proportions of G<sub>0</sub> and G<sub>2</sub> quiescent NSCs in control ( $msh^{\Delta 68}$  heterozygous) versus  $msh^{\Delta 68}$  mutant brains at stage 17 embryogenesis. n=10 tVNCs, ~140 NSCs each. \*\*\*:  $p=3.70 \times 10^{-6}$ , Welch's test. Error bars indicate S.E.M.

(C) The ventral  $G_0$  NSC NB2-2 (arrowed) still arrests in  $G_0$  (CycA<sup>-</sup>; red) in *msh* mutants. Single section confocal image.

(**D-D'**) In control embryos, the ventral  $G_0$  NSC NB2-2 (arrowed) can be identified by its proximity to the midline (dotted line) and expression of the transcription factor Runt (Run, green). NB2-2 does not express CycA (cyan), identifying it as a  $G_0$  NSC. Stage 17 embryo, single section confocal image.

(E-E') NB2-2 is not always formed in  $vnd^6$  mutant embryos (Chu et al., 1998). However, in 33% of hemi-segments, a G<sub>0</sub> (CycA<sup>-</sup>), Run<sup>+</sup> NSC is present close to the midline (dotted line), which we interpret to be the ventral G<sub>0</sub> NSC NB2-2. n=10 embryos, 6 hemi-segments each. Asterisk indicates expected position of NB2-2 in a hemi-segment lacking NB2-2. Stage 17 embryo, single section confocal image.

Anterior is up in all images.



## Figure S4, Related to Figure 4. Msh regulates Dap expression in NSCs

(A-B) Single hemi-segments from control (A) and *msh* mutant (B) tVNCs, co-stained to visualise NSCs (red) and Dap (green). Dap<sup>+</sup> NSCs (yellow) are circled. Anterior is up; dorsal is right. Dorsal NSCs lose Dap expression in *msh* mutants. Dotted line indicates midline. Maximum intensity projections.

(C) Msh binding intensity across ~175kb of chromosome 2R, encompassing the *dap* locus (green box). Msh binds to the *dap* locus. Msh binding was assessed specifically in embryonic NSCs *in vivo* using TaDa (Southall et al., 2013). Unlogged data assembled from three biological replicates.