

Exome sequencing in patients with chronic central serous chorioretinopathy

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Supplementary files

- Figure S1: Principal component analysis (PCA) of the cCSC patients and controls.
- Figure S2: Kinship analysis of cCSC patients and controls
- Figure S3: Single variant analysis of the exome sequencing results in cCSC patients and controls.
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- Table S5: Gene-based results of genes in the known cCSC loci
- Script File S1: R commands performed to run the Burden, SKAT, SKAT-O and single variant analysis.

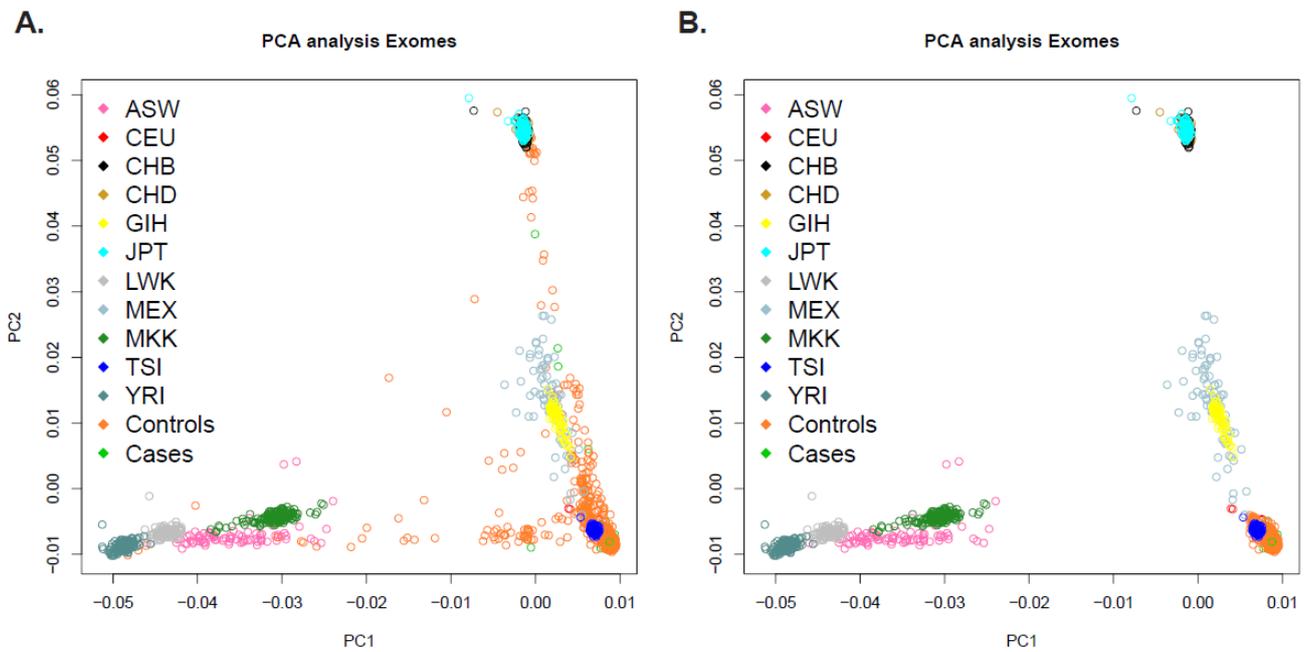


Figure S1. Principal component analysis (PCA) of the cCSC patients and controls. Graphical representation of the first 2 principal components of the PCA analysis, before (A) and after (B) filtering based on European ancestry. Various colours indicate the Hapmap3 samples according to the legend, orange depicts the controls and light green depicts the cCSC patients.

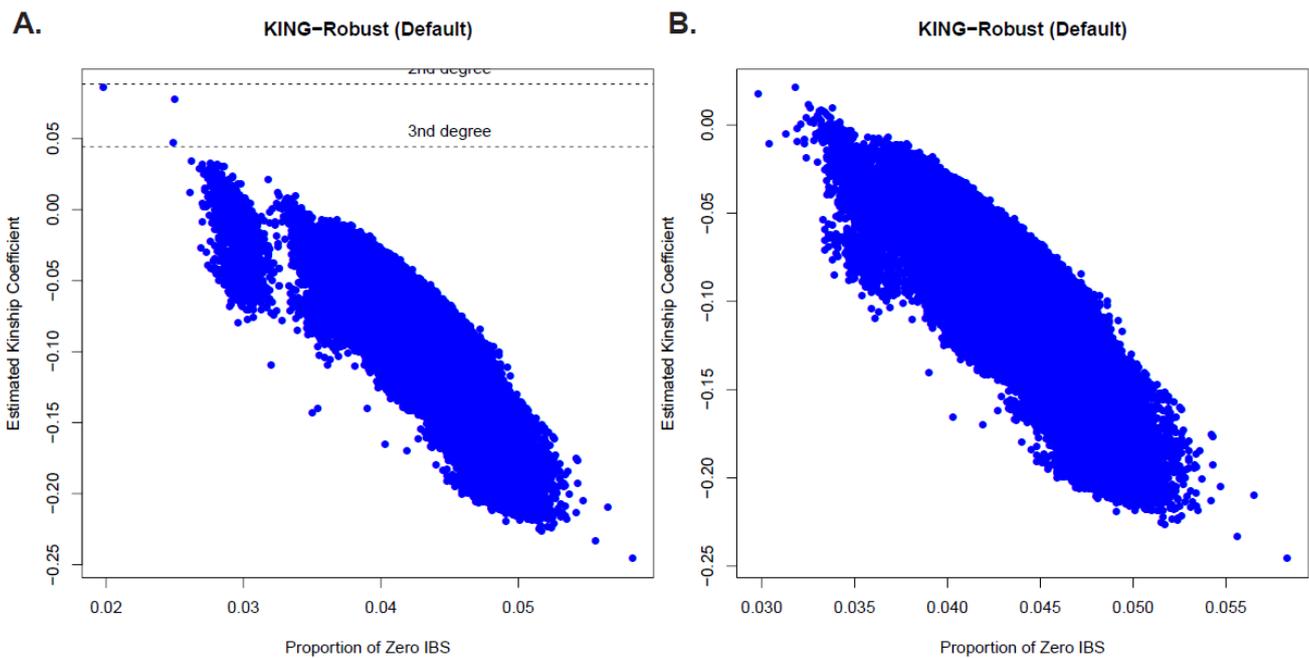


Figure S2. Kinship analysis of cCSC patients and controls. Kinship analysis was performed with KING, and estimated kinship coefficients of individuals in the study were calculated (A). Individuals with a kinship > 0.0442 were removed from the analysis (B).

Single variant analysis

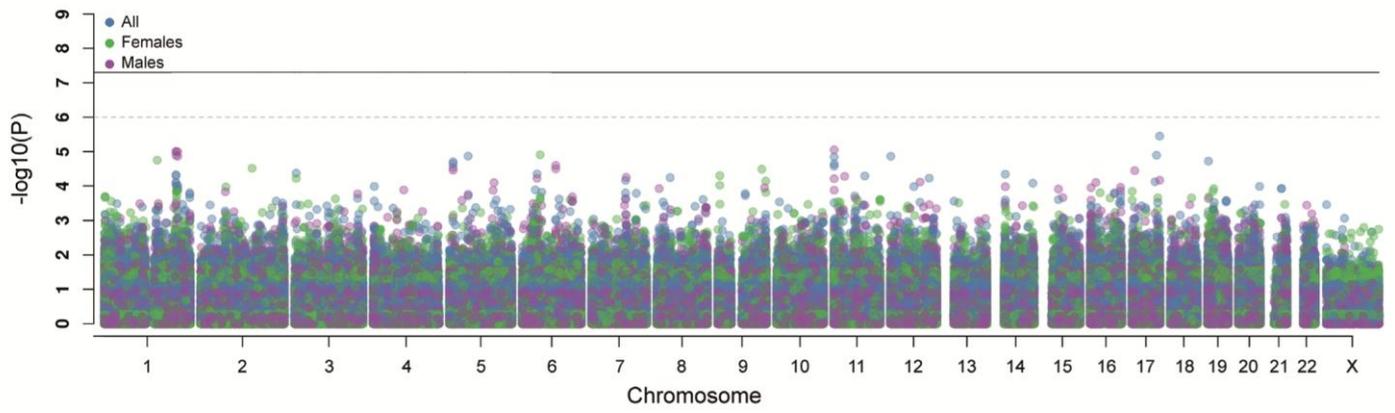


Figure S3. Single variant analysis of the exome sequencing results in cCSC patients and controls. Single variant analysis was performed with correction for 2 principal components and gender in the complete cohort (263 cCSC patients vs. 1352 controls [blue]), while for the females (55 cCSC patients vs. 681 controls [green]) and males (208 cCSC patients vs. 671 controls [purple]) only correction with the 2 principal components was applied. The dark horizontal line indicates the significance threshold after correction for multiple testing of 5×10^{-8} , while the dotted line indicates the suggestive threshold of $P=1 \times 10^{-5}$.

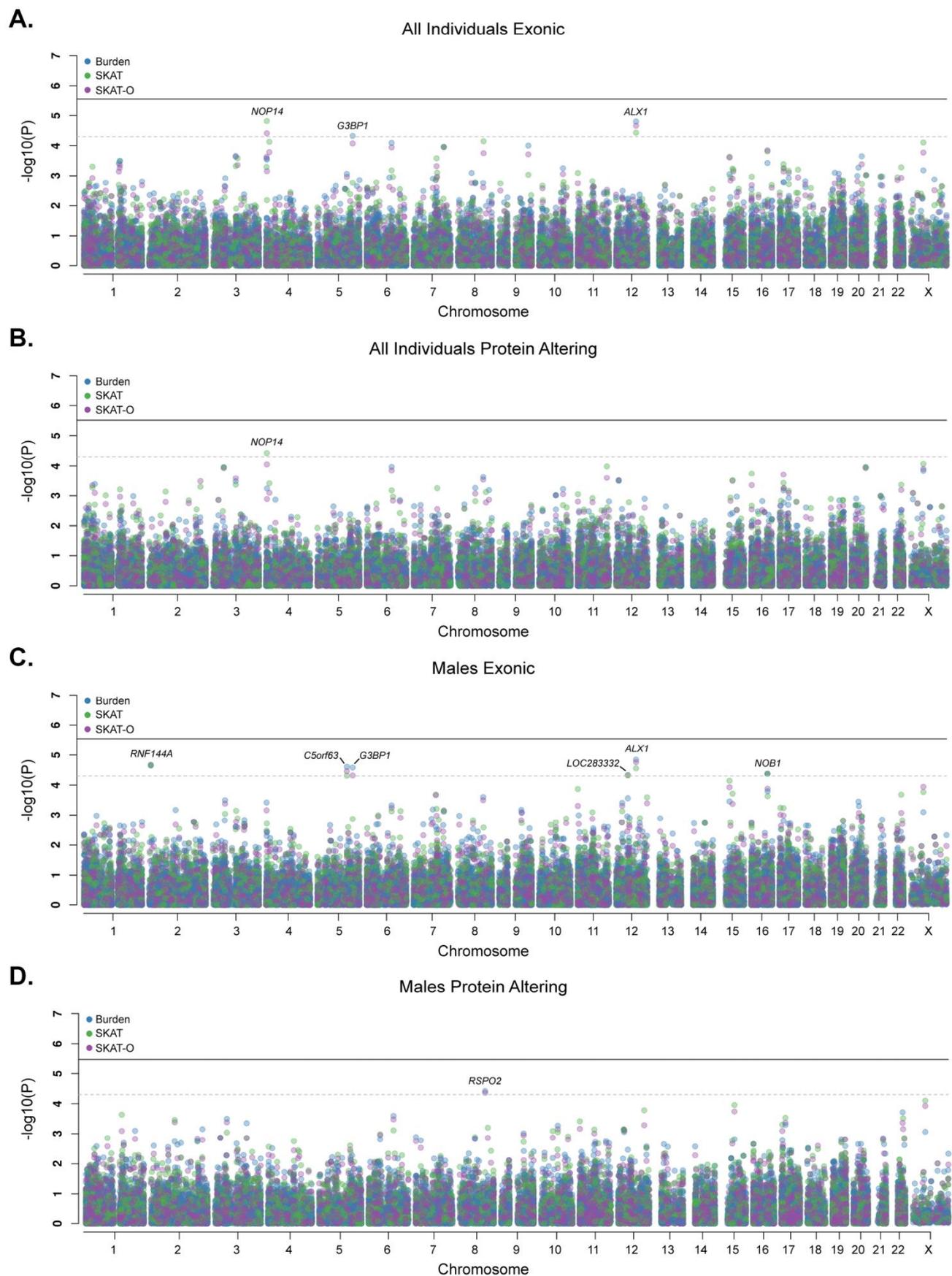


Figure S4. Gene-burden analysis all cCSC individuals and the male cCSC cohort.

Burden (blue), SKAT (green) and SKAT-O (purple) association results are depicted for the complete cohort of 263 cCSC patients and 1352 controls corrected for gender and 2 principal components, using the Exonic (A) and Protein Altering (B) group files. The results the gene-burden test in the 208 male cCSC patients and 671 male controls, for both group files are also depicted (C+D). The dark horizontal line indicates the significance threshold after correction for multiple testing: $0.05/18,173 \text{ genes}=2.75 \times 10^{-6}$ (A), $0.05/16,370 \text{ genes}=3.05 \times 10^{-6}$ (B), $0.05/17,348 \text{ genes}=2.88 \times 10^{-6}$ (C) and $0.05/15,083 \text{ genes}=3.31 \times 10^{-6}$ (D), while the dotted line indicates the suggestive threshold of $P=5 \times 10^{-5}$.

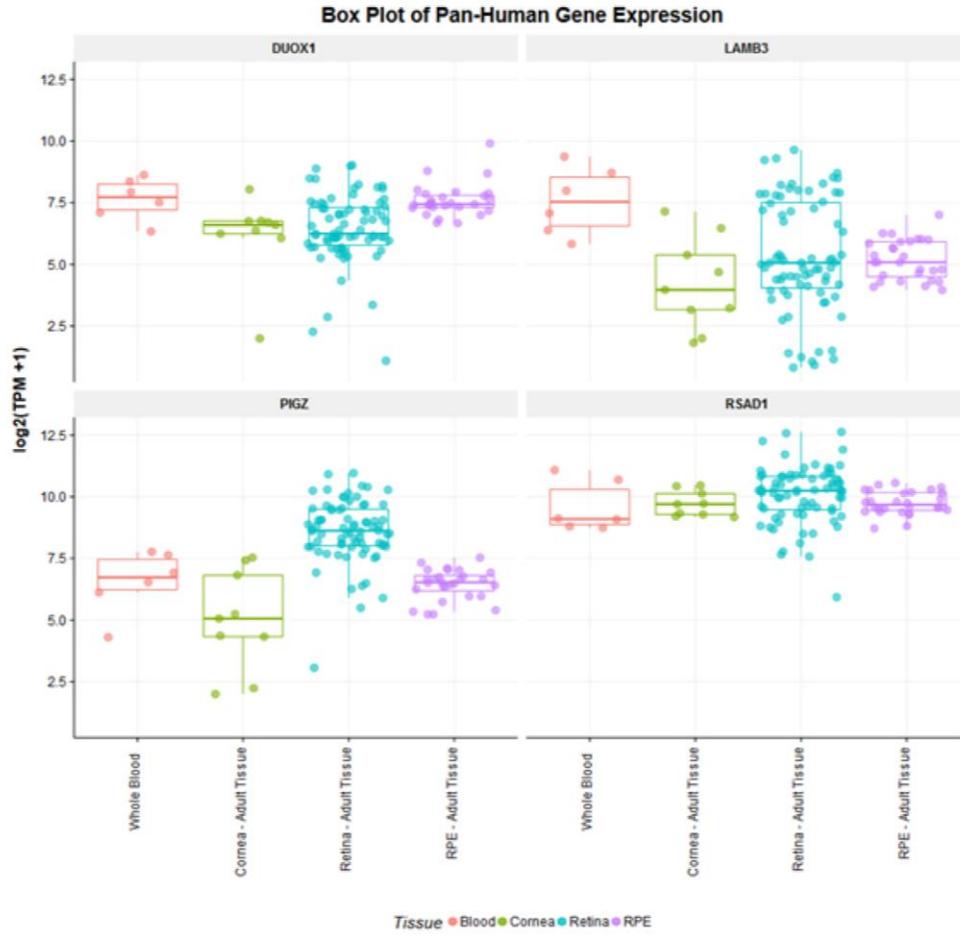


Figure S5. Gene expression of *DUOX1*, *LAMB3*, *PIGZ* and *RSAD1* in different tissues.

Pan-Human Gene Expression levels of the four susceptibility genes in females according to the eyeintegration database v0.63 (27). Whole blood (red), Cornea (green), retina (blue) and retinal pigment epithelium (RPE, purple) expression levels are depicted.

Table S1. Suggestive gene-based associations using the Burden, SKAT and SKAT-O tests				
<i>Gene</i>	<i>Variants</i>	<i>Burden p-value</i>	<i>SKAT p-value</i>	<i>SKAT-O p-value</i>
All individuals				
<i>ALX1</i>	Exonic	1.57x10 ⁻⁵	3.71x10 ⁻⁵	2.17x10 ⁻⁵
<i>NOP14</i>	Protein-altering/ Exonic	3.02x10 ⁻² / 2.49x10 ⁻²	3.76x10 ⁻⁵ / 1.52x10 ⁻⁵	8.88x10 ⁻⁵ / 3.88x10 ⁻⁵
<i>G3BP1</i>	Exonic	4.72x10 ⁻⁵	1.71x10 ⁻³	8.45 x10 ⁻⁵
Males				
<i>RSPO2</i>	Protein-altering	3.81x10 ⁻⁵	1.95x10 ⁻²	4.37x10 ⁻⁵
<i>ALX1</i>	Exonic	1.43x10 ⁻⁵	2.80x10 ⁻⁵	1.74x10 ⁻⁵
<i>RNF144A</i>	Exonic	2.25x10 ⁻⁵	2.25x10 ⁻⁵	2.12x10 ⁻⁵
<i>C5orf63</i>	Exonic	2.48x10 ⁻⁵	4.92x10 ⁻⁵	3.49x10 ⁻⁵
<i>NOB1</i>	Exonic	4.24x10 ⁻⁵	4.24x10 ⁻⁵	4.24x10 ⁻⁵
<i>LOC283332</i>	Exonic	2.77x10 ⁻⁴	4.81x10 ⁻⁵	4.64x10 ⁻⁵
<i>G3BP1</i>	Exonic	2.63x10 ⁻⁵	1.36x10 ⁻³	4.81x10 ⁻⁵
Females				
<i>ZNF713</i>	Exonic	1.68x10 ⁻⁴	3.40x10 ⁻⁶	4.86x10 ⁻⁶
<i>SPATA7</i>	Exonic	7.11x10 ⁻⁶	2.30x10 ⁻⁵	5.35x10 ⁻⁶
<i>C1QL3</i>	Exonic	1.68x10 ⁻⁴	1.03x10 ⁻⁵	1.10x10 ⁻⁵
<i>GAPVD1</i>	Exonic	1.45x10 ⁻²	7.43x10 ⁻⁶	1.99x10 ⁻⁵
<i>ZNF300P1</i>	Exonic	3.33x10 ⁻⁵	3.33x10 ⁻⁵	3.16x10 ⁻⁵
<i>LIPH</i>	Exonic	4.73x10 ⁻⁵	4.73x10 ⁻⁵	4.64x10 ⁻⁵
<i>TDGF1</i>	Protein-altering	4.89x10 ⁻⁵	4.89x10 ⁻⁵	4.64x10 ⁻⁵
<i>OR5H14</i>	Exonic	4.67 x10 ⁻⁵	9.33x10 ⁻⁴	5.42x10 ⁻⁵
<i>RSL1D1</i>	Protein-altering	3.52x10 ⁻⁵	4.28x10 ⁻⁴	9.66x10 ⁻⁵

Table S4. Male rare variant (MAF <0.05) carriers in <i>PIGZ</i> , <i>DUOX1</i> , <i>LAMB3</i> and <i>RSAD1</i>					
	No. rare variants in gene	Controls (n=671)	cCSC patients (n=208)	P-value	OR (95% CI)
<i>LAMB3</i>	32	81 (12.1%)	20 (9.6%)	0.38	0.78 (0.46-1.30)
<i>PIGZ</i>	19	35 (5.2%)	7 (3.4%)	0.35	0.63 (0.28-1.45)
<i>DUOX1</i>	23	24 (3.6%)	8 (3.8%)	1.00	1.08 (0.48-2.44)
<i>RSAD1</i>	4	7 (1.0%)	1 (0.5%)	0.69	0.46 (0.06-3.73)
Variant in 1 or more genes		137 (20.4%)	33 (15.9%)	0.15	0.74 (0.49-1.12)

Table S2. Variants observed in the *DUOX1*, *PIGZ*, *LAMB3* and *RSAD1* genes in the female cohort

Gene	Variant	Exon	Effect	Nucleotide change	Protein change	SKAT P-value	SAS P-value	OR (95% CI)	Weight	Counts	Counts	MAF cCSC	MAF controls	Exac_NFE	PhyloP	Grantham	Mutation Taster				
										female cCSC (n=55)	female controls (n=681)						(score)	CADD	Sift (score)	Polyphen (score)	Trap(score)
<i>DUOX1</i> (NM_175940)	rs764911806	5	nonsyn	c.C411G	p.D137E	0.50	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	1.00E-04	0.25	45	N(1)	15.8	T (0.361)	B (0.036)	0.147
	rs200421952	14	nonsyn	c.G1627A	p.A543T	0.55	0.64	2.46 (0.06-102.04)	1.00E+00	0	2	0	1.47E-03	7.00E-04	1.61	58	N(0.953)	13.6	T (0.159)	B (0.124)	0.001
	rs201198610	15	nonsyn	c.T1778C	p.F593S	0.54	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	6.00E-04	1.29	155	D(0.558)	20.9	T (0.45)	B (0.138)	0.012
	rs370633829	15	nonsyn	c.G1786A	p.G596R	0.044	0.12	37.16 (0.40->999.99)	1.00E+00	1	0	9.09E-03	0	7.50E-05	1.53	125	N(0.643)	24.4	T (0.068)	D (0.999)	0.234
	rs143304688	17	stopgain	c.C1948T	p.Q650*	0.55	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	1.70E-03	.	.	A(1)	44.0	.	.	0.773
	rs368978619	18	nonsyn	c.T2152A	p.L718M	0.57	0.64	2.46 (0.06-102.04)	1.00E+00	0	2	0	1.47E-03	2.00E-04	0.06	15	N(1)	8.1	T (0.266)	B (0.003)	0.004
	rs373898472	21	nonsyn	c.G2750T	p.W917L	0.54	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	3.00E-05	3.56	61	D(1)	17.0	T (0.831)	D (0.957)	0.021
	rs145668427	21	nonsyn	c.C2773T	p.R925W	0.0024	0.03	62.69 (1.52->999.99)	1.00E+00	2	0	0.018	0	2.40E-03	0.99	101	D(1)	34.0	D (0.004)	D (1)	0.048
	rs148902727	21	nonsyn	c.C2794T	p.R932C	0.57	0.64	2.46 (0.06-102.04)	1.00E+00	0	2	0	1.47E-03	5.00E-04	3.23	180	D(1)	26.6	D (0.014)	D (1)	0.038
	rs16939752	24	nonsyn	c.T3076C	p.C1026R	0.33	0.30	1.35 (0.77-2.37)	1.09E-03	15	146	0.136	0.107	0.127	0.98	180	P(0.899)	15.4	T (0.364)	B (0)	0.026
	rs777590867	24	nonsyn	c.G3170C	p.G1057A	0.051	0.08	12.41 (0.77-199.80)	1.00E+00	1	1	0.01	7.34E-04	1.52E-05	1.46	60	D(1)	10.9	T (0.38)	Prob. D (0.661)	0.014
	rs2458236	27	nonsyn	c.C3532T	p.L1178F	0.99	0.91	0.98 (0.65-1.47)	2.24E-17	37	467	0.336	0.343	0.674	0.85	22	P(0.999)	20.5	T (0.439)	B (0.002)	0.09
	rs142004175	28	nonsyn	c.G3644A	p.R1215H	0.56	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	1.60E-03	3.38	29	D(1)	33.0	D (0.002)	D (1)	0.059
	rs138894830	28	nonsyn	c.G3656A	p.R1219Q	0.51	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	1.30E-03	3.38	43	D(1)	23.8	T (0.277)	Prob. D (0.706)	0.332
rs773444369	33	nonsyn	c.C4462T	p.R1488C	0.55	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	4.50E-05	2.20	180	D(1)	33.0	D (0)	D (1)	0.116	
<i>PIGZ</i> (NM_025163)	rs74554271	2	nonsyn	c.C65T	p.P22L	0.54	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	2.00E-04	0.26	98	N(1)	16.23	D (0.015)	B (0.002)	0.021
	rs780890452	3	nonsyn	c.C308T	p.T103I	0.53	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	9.98E-05	2.40	89	D(1)	15.80	D (0.018)	B (0.036)	0.214
	rs200901195	3	nonsyn	c.G323A	p.R108K	0.53	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	2.00E-04	0.29	26	D(1)	9.08	T (1)	B (0)	0.475
	rs199852771	3	nonsyn	c.C344T	p.P115L	0.60	0.75	1.76 (0.06-54.12)	1.00E+00	0	3	0	2.20E-03	4.00E-04	-0.93	98	N(1)	0.00	T (0.407)	B (0)	0.123
	rs144088027	3	nonsyn	c.C371T	p.A124V	0.00046	0.00030	16.40 (3.64-73.94)	1.00E+00	4	3	0.036	2.20E-03	5.70E-03	-1.40	64	N(1)	0.00	T (1)	B (0.002)	0.144
	rs150001262	3	nonsyn	c.G464A	p.R155H	0.53	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	1.30E-03	0.66	29	D(1)	23.20	T (0.129)	D (1)	0.128
	rs148186466	3	nonsyn	c.G725T	p.G242V	0.63	0.85	1.364 (0.052-36.04)	1.00E+00	0	4	0	2.94E-03	2.00E-04	0.78	109	D(0.821)	16.08	T (0.164)	Prob. D (0.934)	0.327
	rs574365	3	nonsyn	c.G796A	p.A266T	0.58	0.56	0.87 (0.55-1.39)	2.11E-12	24	334	0.218	0.245	0.744	0.20	58	P(1)	0.27	T (0.323)	B (0.001)	0.063
	rs140946995	3	nonsyn	c.C821T	p.A274V	0.57	0.64	2.46 (0.06-102.04)	1.00E+00	0	2	0	1.47E-03	1.50E-03	1.61	64	N(1)	0.00	T (1)	B (0.136)	0.064
	rs148546943	3	nonsyn	c.C864A	p.N288K	0.54	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	1.57E-04	0.52	94	.	0.03	T (0.49)	B (0.002)	0.011
	rs149631795	3	nonsyn	c.C1010T	p.A337V	0.55	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	4.00E-04	0.76	64	N(1)	7.99	T (0.164)	Prob. D (0.919)	0.078
	rs4916589	3	nonsyn	c.G1019A	p.R340Q	0.75	0.72	0.93 (0.63-1.38)	9.96E-23	43	557	0.391	0.409	0.596	0.46	43	P(1)	8.52	T (0.505)	B (0)	0.166
	rs150517093	3	nonsyn	c.C1078T	p.R360W	0.54	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	4.00E-04	-2.76	101	N(1)	0.00	T (0.188)	B (0.001)	0.009
	rs1147238	3	nonsyn	c.T1250C	p.V417A	0.47	0.42	1.24 (0.73-2.09)	3.51E-05	18	189	0.164	0.139	0.141	0.98	64	P(0.947)	7.25	T (0.252)	B (0.007)	0.081
rs150414171	3	nonsyn	c.G1401C	p.M467I	0.11	0.09	7.45 (0.71-77.77)	1.00E+00	1	2	9.09E-03	1.47E-03	1.90E-03	3.82	10	D(0.937)	25.20	D (0.036)	D (1)	0.043	
rs17855662	3	nonsyn	c.G1461A	p.M487I	0.14	0.19	0.55 (0.23-1.33)	8.97E-02	5	117	0.045	0.086	0.084	2.20	10	N(0.712)	22.10	T (0.143)	Prob. D (0.524)	0.431	
rs202156268	3	nonsyn	c.G1466A	p.G489E	0.50	0.54	4.09 (0.04-381.40)	1.00E+00	0	1	0	7.34E-04	3.00E-05	3.82	98	D(0.931)	24.90	D (0.005)	D (0.998)	0.297	
rs146219795	3	nonsyn	c.G1591A	p.V531M	0.042	0.12	37.16 (0.40->999.99)	1.00E+00	1	0	9.09E-03	0	5.00E-04	-0.60	21	N(1)	2.73	T (0.18)	Prob. D (0.539)	0.01	

Table S5. Gene-burden results of genes in the known cCSC loci

Gene	Locus	All individuals																Males								Females											
		exonic				protein altering				SKAT-O				exonic				protein altering				SKAT-O				exonic				protein altering				SKAT-O			
		Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O	Vars	Burden	SKAT	SKAT-O				
ARMS2	ARMS2-HTRA1	6	0.358	0.681	0.498	5	0.593	0.865	0.725	5	0.377	0.602	0.500	4	0.635	0.633	0.636	4	0.784	0.781	1.000	4	0.784	0.781	1.000	4	0.784	0.781	1.000	4	0.784	0.781	1.000				
HTRA1	ARMS2-HTRA1	10	0.763	0.910	0.918	6	0.844	0.632	0.866	9	0.716	0.859	0.887	5	0.885	0.470	0.637	4	0.807	0.571	0.913	2	0.634	0.634	0.634	9	0.204	0.854	0.349								
PLEKHA1	ARMS2-HTRA1	18	0.572	0.267	0.432	12	0.118	0.248	0.201	15	0.209	0.149	0.254	10	0.020	0.096	0.036	11	0.157	1.000	0.274	9	0.204	0.854	0.349												
CR1L	CD46	8	0.542	0.844	0.645	7	0.546	0.877	0.663	7	0.673	1.000	0.779	6	0.671	1.000	0.775	7	0.689	0.801	0.797	6	0.673	0.879	0.773												
CD46	CD46	10	0.405	0.689	0.504	7	0.347	0.666	0.431	7	0.163	0.151	0.179	5	0.131	0.147	0.165	7	0.373	0.050	0.072	6	0.389	0.049	0.071												
CD34	CD46	16	0.219	0.033	0.047	9	0.113	0.033	0.047	12	1.000	0.575	0.712	7	0.795	0.539	0.667	10	0.0091	0.0016	0.0026	6	0.0067	0.0017	0.0027												
BEAN1	CDH5	24	0.758	1.000	0.892	16	0.583	0.668	0.694	16	0.589	0.633	0.756	9	0.525	0.594	0.635	17	1.000	0.390	0.554	10	1.000	1.000	1.000												
CDH5	CDH5	39	0.787	0.892	1.000	23	0.867	0.577	0.798	25	0.549	0.596	0.721	15	0.321	0.264	0.434	29	0.122	1.000	0.247	17	0.126	0.982	0.228												
CFHR2	CFH	6	0.090	0.389	0.136	4	0.193	0.494	0.270	6	0.249	0.685	0.342	4	0.326	0.729	0.432	4	0.128	0.507	0.193	3	0.424	0.678	0.590												
CFHR4	CFH	7	0.294	0.146	0.242	4	0.262	0.709	0.400	6	1.000	0.410	0.607	4	0.255	0.709	0.391	2	0.069	0.069	0.071	NA	NA	NA	NA												
KCNT2	CFH	26	0.733	0.188	0.271	11	0.140	0.057	0.119	14	0.552	0.278	0.384	4	0.033	0.130	0.100	17	0.086	0.204	0.135	8	0.957	0.539	0.957												
CFHR5	CFH	24	0.374	0.358	0.524	19	0.350	0.338	0.497	17	0.943	0.641	0.813	15	0.952	0.613	0.787	17	0.085	0.050	0.091	13	0.064	0.047	0.080												
CFH	CFH	33	0.717	0.333	0.515	19	0.276	0.464	0.423	24	0.184	0.073	0.126	14	0.059	0.128	0.099	23	0.182	0.924	0.315	13	0.254	0.562	0.373												
NR3C2	NR3C2	15	0.227	0.737	0.348	8	0.109	0.782	0.185	12	0.332	0.881	0.485	5	0.159	0.783	0.233	14	0.465	0.593	0.641	7	0.379	0.677	0.581												
ARHGAP10	NR3C2	31	0.570	0.630	0.742	16	0.202	0.932	0.276	24	0.866	0.363	0.523	12	0.282	1.000	0.378	18	0.419	1.000	0.590	7	0.422	1.000	0.548												
PTF2	RCCX	4	0.170	0.109	0.151	3	0.789	0.793	1.000	4	0.128	0.143	0.190	3	0.454	0.455	0.455	2	0.210	0.210	0.210	2	0.210	0.210	0.210												
STK19	RCCX	10	0.590	0.600	0.754	4	0.187	0.519	0.380	8	0.846	0.965	0.966	3	0.638	0.763	0.741	5	0.058	0.210	0.149	3	0.105	0.294	0.294												
ZBTB12	RCCX	8	0.482	0.617	0.566	5	0.372	0.776	0.481	5	0.516	0.599	0.608	2	0.343	0.721	0.721	5	1.000	1.000	1.000	3	0.603	0.603	0.603												
FKBP1	RCCX	7	0.699	0.729	0.852	6	0.889	0.746	0.885	7	0.740	0.264	0.394	6	1.000	0.283	0.416	6	1.000	0.289	0.433	5	1.000	0.270	0.406												
RDBP	RCCX	10	0.040	0.018	0.040	8	0.046	0.021	0.028	7	0.035	0.011	0.020	5	0.012	0.010	0.017	7	0.541	0.490	0.710	5	0.718	0.906	0.906												
DOM3Z	RCCX	19	1.000	0.777	1.000	9	1.000	0.783	1.000	15	1.000	0.502	0.673	8	0.600	0.478	0.586	13	1.000	0.443	0.607	5	0.526	0.299	0.371												
EGFL8	RCCX	14	0.655	0.223	0.332	10	0.064	0.162	0.107	10	1.000	0.374	0.495	7	0.119	0.201	0.194	10	0.320	0.323	0.426	7	0.184	1.000	0.298												
CFB	RCCX	17	0.350	0.703	0.505	11	0.204	0.646	0.310	15	0.836	1.000	1.000	9	0.891	0.895	1.000	13	0.117	0.101	0.149	9	0.0060	0.056	0.012												
ATF6B	RCCX	21	0.173	0.682	0.268	11	0.248	1.000	0.373	16	0.522	1.000	0.668	8	0.402	0.808	0.578	13	0.070	0.124	0.116	9	0.569	0.804	0.835												
SLC44A4	RCCX	25	0.483	0.641	0.656	15	0.404	0.706	0.597	14	0.257	1.000	0.431	10	0.348	0.837	0.527	21	1.000	0.611	0.774	12	0.870	0.863	1.000												
EHHMT2	RCCX	36	0.208	0.183	0.284	16	0.153	0.073	0.113	22	0.656	0.538	0.745	11	0.548	0.317	0.471	25	0.129	0.108	0.159	12	0.086	0.070	0.101												
C2	RCCX	24	0.811	0.615	0.786	20	0.660	0.591	0.728	18	0.682	0.676	0.811	14	0.679	0.654	0.787	14	1.000	0.316	0.430	13	1.000	0.393	0.512												
AGER	RCCX	31	0.209	0.745	0.324	20	0.275	0.368	0.385	25	0.320	0.818	0.475	15	0.397	0.521	0.535	19	0.384	0.582	0.545	11	0.396	0.713	0.509												
SKIVL2	RCCX	45	0.922	0.283	0.429	24	0.798	0.215	0.327	33	0.596	0.050	0.080	19	0.343	0.077	0.125	30	0.447	0.420	0.601	14	0.167	0.273	0.262												
NOTCH4	RCCX	72	0.685	1.000	0.856	47	0.544	0.939	0.740	55	0.591	1.000	0.769	32	0.756	0.695	0.863	49	0.093	0.220	0.150	29	0.074	0.448	0.128												
TNXX	RCCX	116	0.519	0.404	0.576	84	0.221	0.346	0.351	87	0.228	0.048	0.085	63	0.048	0.050	0.066	71	0.394	0.159	0.251	52	0.284	0.141	0.238												
GPSM3	RCCX	2	0.221	0.221	0.221	NA	NA	NA	NA	2	0.094	0.094	0.094	NA	NA	NA	NA	2	1.000	1.000	1.000	NA	NA	NA	NA												
MIR1236	RCCX	3	0.104	0.379	0.196	NA	NA	NA	NA	2	0.076	0.279	0.123	NA	NA	NA	NA	1	NA																		
RNF5	RCCX	3	0.530	0.482	0.575	NA	NA	NA	NA	3	0.591	1.000	0.716	NA	NA	NA	NA	2	0.037	0.037	0.037	NA	NA	NA	NA												
RNF5P1	RCCX	3	0.544	0.544	0.544	NA	NA	NA	NA	3	0.372	0.372	0.372	NA	NA	NA	NA	3	0.729	0.729	0.729	NA	NA	NA	NA												
AGPAT1	RCCX	4	0.793	0.357	0.723	NA	NA	NA	NA	3	0.347	0.747	0.541	NA	NA	NA	NA	3	0.115	0.115	0.115	NA	NA	NA	NA												
TNFRSF10C	TNFRSF10A	7	0.755	0.554	0.699	3	0.970	0.267	0.369	3	0.786	0.809	0.926	NA	NA	NA	NA	7	0.176	0.335	0.338	3	0.079	0.079	0.079												
CHMP7	TNFRSF10A	6	0.544	0.791	0.815	6	0.544	0.791	0.815	4	0.527	0.753	0.787	4	0.527	0.753	0.787	2	0.585	0.585	0.585	2	0.585	0.585	0.585												
R3HC1	TNFRSF10A	18	0.936	0.829	1.000	8	0.674	0.980	0.779	15	0.836	0.687	0.813	6	0.632	1.000	0.736	13	0.785	1.000	1.000	6	0.934	1.000	1.000												
TNFRSF10D	TNFRSF10A	20	0.483	0.030	0.051	10	0.330	0.505	0.454	12	0.786	0.114	0.181	8	0.317	0.535	0.439	18	0.337	0.179	0.275	8	0.823	0.782	1.000												
TNFRSF10A	TNFRSF10A	18	0.893	0.049	0.089	15	0.271	0.017	0.030	14	0.575	0.119	0.203	12	0.890	0.057	0.100	12	0.250	0.193	0.317	10	0.097	0.118	0.143												
LXLL2	TNFRSF10A	36	0.881	0.518	0.707	15	0.470	0.061	0.112	29	0.360	0.182	0.290	11	1.000	0.603	0.806	23	0.131	0.569	0.218	10	0.079	0.102	0.148												
VIPR2	VIPR2	19	0.608	0.659	0.809	7	0.818	0.649	0.815	11	0.603	0.754	0.798	4	0.764	0.683	0.781	15	0.867	0.590	0.800	5	0.954	0.307	0.307												
WDR60	VIPR2	41	0.499	0.282	0.415	27	0.547	0.843	0.733	29	0.417	0.355	0.512	20	0.396	0.817	0.569	22	0.979	0.596	0.754	13	1.000	0.495	0.674												

Yellow color indicated P-values < 0.05. Correction for multiple testing was applied for 42 for the exonic groupfile (P<0.0119) and for 37 genes for the protein altering groupfile (P<0.001351). No genes were significantly associated after correction for multiple testing.

Script file S1

```
#Convert data to plink files and make a dummy gene list based on the
variants in the bim files
#In the following format:
#dummygene      SNP1
#dummygene      SNP2
awk '{print("dummygene"\t"$2)}' input.bim | sort -k2 -u >
SNPsets_dummy.txt

#Generate weight file based on minor allele frequency
R
library(SKAT)
#Generate the File.SSD and File.Info
Generate_SSD_SetID("input.bed", "input.bim", "input.fam",
"SNPsets_dummy.txt", "File.SSDdummy", "File.Infodummy")
#read the covariate file and fam file if applicable
FAM_Cov<-Read_Plink_FAM_Cov("input.fam", "input_cov.txt",
Is.binary=TRUE, flag1=0, cov_header=TRUE)
SSD.INFOdummy<-Open_SSD("File.SSDdummy", "File.Infodummy")
#Get genotype matrix of the dummy geneset
dummy_geneset<-Get_Genotypes_SSD(SSD.INFOdummy, 1, is_ID = TRUE)
weight<-Get_Logistic_Weights(dummy_geneset, par=0.07, par2=150)
weights<-data.frame(weight)
weights[,2] <- colnames(dummy_geneset)
weights = weights[, c(2,1)]
write.table(weights, file="snp_weights.txt", col.names=FALSE,
row.names=FALSE, sep="\t", quote = FALSE)

#perform single variant association with dummy geneset dataset
#Define the covariates if applicable
Phenotype = FAM_Cov$Phenotype
PC1 = FAM_Cov$PC1
PC2 = FAM_Cov$PC2
Sex = FAM_Cov$Sex
#Run the null model
obj<-SKAT_Null_Model_MomentAdjust(Phenotype ~ PC1 + PC2 + Sex,
type.Resampling="bootstrap.fast")
#Run the loop through the genotype matrix and output the p-values
single_pvalue <- NULL
for(snp in colnames(dummy_geneset)){out <-
SKATBinary_Single(dummy_geneset[,snp], obj); single_pvalue <-
rbind(single_pvalue, data.frame(snp, out$p.value, out$method.bin,
out$m))}
#print the output file
write.table(single_pvalue, file="Single_pvalue_all.txt",
col.names=TRUE, row.names=FALSE, sep="\t", quote = FALSE)

#Run the burden, SKAT and SKAT-O test using the snp_weights_file.txt
Generate_SSD_SetID("input.bed", "input.bim", "input.fam",
"groupfile.txt", "File.SSD_groupfile", "File.Info_groupfile")
#read the covariate file and fam file if applicable
FAM_Cov<-Read_Plink_FAM_Cov("input.fam", "input_cov.txt",
Is.binary=TRUE, flag1=0, cov_header=TRUE)
SSD.INFO_groupfile<-Open_SSD("File.SSD_groupfile",
"File.Info_groupfile")
Phenotype = FAM_Cov$Phenotype
PC1 = FAM_Cov$PC1
PC2 = FAM_Cov$PC2
Sex = FAM_Cov$Sex.x
obj<-SKAT_Null_Model_MomentAdjust(Phenotype ~ Sex + PC1 + PC2,
type.Resampling="bootstrap.fast")
custom_weights<-Read_SNP_WeightFile("snp_weights.txt")
```

```

#Burden test
out.burden<-SKATBinary.SSD.All(loaded_SSD, obj, method="Burden",
obj.SNPWeight=custom_weights, kernel="linear.weighted")
#SKAT test
out.skate<-SKATBinary.SSD.All(loaded_SSD, obj, method="SKAT",
obj.SNPWeight=custom_weights, kernel="linear.weighted")
#SKAT-O test
out.skato<-SKATBinary.SSD.All(loaded_SSD, obj, method="SKATO",
obj.SNPWeight=custom_weights, kernel="linear.weighted")

#keep only genes with more than 2 variants and perform correction
for multiple testing
out.burden_select <- out.burden[out.burden$N.Marker.Test>1,]
out.burden_select$FDR <- p.adjust(out.burden_select$P.value,
method="fdr")
out.burden_select$Bonf <- p.adjust(out.burden_select$P.value,
method="bonferroni")
write.table(out.burden_select, file="burden_output_groupfile.txt",
col.names=TRUE, row.names=FALSE, sep="\t", quote = FALSE)

out.skate_select <- out.skate[out.skate$N.Marker.Test>1,]
out.skate_select$FDR <- p.adjust(out.skate_select$P.value,
method="fdr")
out.skate_select$Bonf <- p.adjust(out.skate_select$P.value,
method="bonferroni")
write.table(out.skate_select, file="skate_output_groupfile.txt",
col.names=TRUE, row.names=FALSE, sep="\t", quote = FALSE)

out.skato_select <- out.skato[out.skato$N.Marker.Test>1,]
out.skato_select$FDR <- p.adjust(out.skato_select$P.value,
method="fdr")
out.skato_select$Bonf <- p.adjust(out.skato_select$P.value,
method="bonferroni")
write.table(out.skato_select, file="skato_output_groupfile.txt",
col.names=TRUE, row.names=FALSE, sep="\t", quote = FALSE)

#plot the data in CMplot
library(CMplot)
ref_genes=read.table("genes_location.txt", header=T)
data_combined1 <- merge(out.burden_select[,c(1,2)],
out.skate_select[,c(1,2)], by="SetID", all=TRUE)
data_combined2 <- merge(data_combined1, out.skato_select[,c(1,2)],
by="SetID", all=TRUE)
data_combined3 <- merge(ref_genes, data_combined2 , by=1,
no.dups=TRUE)
colnames(data_combined3) =
c("GENE", "CHR", "BP", "burden", "skate", "skato")
nrow(data_combined3)
#genome-wide significant line --> 0.05/nrow=pvalue_cutoff
data_combined3[is.na(data_combined3)] <- 1

CMplot(data_combined3, plot.type="m", multitracks=TRUE,
threshold=c(pvalue_cutoff,suggestive_line),threshold.lty=c(1,2),
threshold.lwd=c(1,1), threshold.col=c("black","grey"),
amplify=TRUE,bin.size=1e6,
chr.den.col=NULL,
signal.col=c("red","green"),signal.cex=c(1,1),
file="pdf",memo="results")

```