

Figure S1. Behavioral preferences of flies to acids and generation of *Ir* mutants. Related to Figure 1.

(A—B) Two-way choice feeding assays showing preferences of control (w^{1118}) and $Ir7a^1$ flies upon presentation of 2 mM sucrose alone, versus 2 mM sucrose with the indicated concentrations of (A) butyric acid, and (B) HCl. n=4-6.

(C—F) Preferences of control (w^{1118}) and $Ir7a^1$ flies to the indicated concentrations of (C) acetic acid, (D) citric acid, (E) glycolic acid, and (F) lactic acid without sucrose. n=4-6. (G) *Ir* mutants generated by ends-out homologous recombination.

The error bars indicate SEMs. The asterisks indicate significant differences from the controls using ANOVA tests with Scheffe's *post hoc* analyses between wild-type and mutants. (**p < 0.01).



Figure S2. Analyses with control flies after starvation, *Ir7a¹/Df*, *Ir25a²*, *Ir76b¹* flies, and behavioral response of bitter GRN ablated and rescue flies in the absence of sucrose. Related to Figures 1 and 2.

(A) Two-way choice feeding assay using flies harboring the $Ir7a^1$ mutation *in trans* with a deficiency that removes Ir7a ($Ir7a^1/Df$). n=4-6.

(B) Two-way choice assay performed separately with males and females. n=4-6.

(C) Frequencies of action potentials elicited by controls, and $Ir7a^{1}/Df$ flies. n=10-12.

(D) Frequencies of 1% acetic acid-induced action potentials elicited from S6 and S10 sensilla from control (w^{1118}), $Ir25a^2$, and $Ir76b^1$ flies. n=20.

(E) Average frequencies of action potentials elicited from S, I, and L type sensilla in response to 1% acetic acid after the control and $Ir7a^{1}$ flies were starved for 24 hrs. n=18-30.

The error bars indicate SEMs. The asterisks indicate significant differences from the controls using ANOVA tests with Scheffe's *post hoc* analyses. **p < 0.01.



Figure S3. Screening of *Ir* mutants to test avoidances to the indicated acids using binary choice assays. Related to Figure 1.

The flies were given a choice between 2 mM sucrose versus 2 mM sucrose and the indicated acids. n=4-6.

Significant differences between the controls and the mutants were tested using ANOVA tests with Scheffe's *post hoc* analyses.



Figure S4. Acid-induced action potentials displayed by *Ir* mutants. Related to Figure 2.

Frequencies of action potentials obtained by performing tip recording on S6 and S10 sensilla from control (w^{1118}) and *Ir* mutant flies in response to the indicated concentrations of organic acids and HCl. n=12-20.

Significant differences between the controls and the mutants were tested using ANOVA tests with Scheffe's *post hoc* analyses.



Figure S5. Testing roles for *Ir7a* for acetic acid inhibition of sugar GRNs, and for sensing molecules with similar carbon backbones as acetic acid. Related to Figure 3.

(A-B) Average frequencies of action potentials elicited by several S-type sensilla from control and *Ir7a*¹ flies in response to ethanol and acetaldehyde, which have similar carbon backbones as acetic acid.

(A) 10% ethanol. n=12-14.

(B) 1% acetaldehyde. n=15-19.

(C-D) Representative tip recording traces obtained from S10 sensilla from control and $Ir7a^{1}$ flies.

(C) 10% ethanol.

(D) 1% acetaldehyde.

(E) Average frequencies of action potentials elicited from L4 and L6 sensilla using 100 mM sucrose and the indicated concentrations of acetic acid. n=10-12. There are no significant differences between the control and the $Ir7a^{1}$ mutant.

(F) Two-way choice assays in response to 1 mM sucrose alone versus 5 mM sucrose plus bitter compounds at the following concentrations: 1 mM quinine, 0.3 mM denatonium, 0.3 mM lobeline, 0.5 mM strychnine, 10 mM caffeine, and 0.1 mM berberine. n=4-6.

(G) Average frequencies of action potentials elicited from S6 and S10 sensilla in response to 1 mM quinine, 1 mM denatonium, 1 mM lobeline, 1 mM strychnine, 10 mM caffeine, and 0.1 mM berberine. n=10-12.

The error bars represent SEMs. The asterisks indicate significant differences (*p < 0.05, **p < 0.01) using unpaired Student *t*-tests for comparing two sets of data or ANOVA with Scheffe's analysis as a *post hoc* test to compare two sets of data.



Figure S6. Testing for rescue of *Ir7a*-dependent acetic acid repulsion and for the effect of killing *Ir7a* GRNs with *hid* using two-way choice assays without sugar. Related to Figure 4.

(A) Two-way choice feeding assays without sucrose showing rescue of the avoidance defect in $Ir7a^1$ in response to 5% acetic acid. n=6.

(B) Two-way food choice assays without sucrose after expressing the cell death gene, *hid* (*UAS-hid*), under control of either the *Ir7a-GAL4* or the *Gr66a-GAL4*. n=6. The error bars indicate SEMs. The asterisks indicate significant differences from the controls using ANOVA tests with Scheffe's *post hoc* analyses. **p < 0.01.



Figure S7. Expression pattern of the *Ir7a-GAL4* reporter in the labellum, legs and pharynx. Related to Figure 5.

UAS-DsRed was expressed under the control of the *Ir7a-GAL4*. The signals were detected by staining with anti-DsRed and viewed by confocal microscopy.

(A) Schematic representation of the 11 S-type sensilla in the labellum that were labeled by the *Ir7a-GAL4* reporter. This is the same image presented in Figure 5A. The scale bars represent 25 μ m.

(B) Expression of the *Ir7a* reporter in the legs of control (w^{1118}) flies. The upper and lower panels show flies harboring the *UAS-GFP* transgene only, and *UAS-GFP* plus the *Ir7a-GAL4* transgenes, respectively. The inset in the image of the prothoracic leg is a magnified region indicated by the box. The scale bars represent 100 μ m.

(C) *Ir7a* reporter expression in the pharynx of control (w^{1118}) flies. The panel on the left is a proboscis from a fly containing *UAS-GFP* transgene only. The panel on the right shows expression of *UAS-GFP* under control of the *Ir7a-GAL4*. The inset is a magnified region indicated by the box. The scale bars represent 100 μ m.