

Supplementary Table S1. Antimicrobial resistance in *A. baumannii* from the 2010-2014 TSAR collection

	TSAR VII (2010)		TSAR VIII (2012)		TSAR IX (2014)		Overall	
	N = 321		N = 332		N = 226		N = 879	
	N	%	N	%	N	%	N	%
AMK	226	70.4	246	74.1	149	65.9	621	70.6
AMS	168	52.3	206	62	106	46.9	480	54.6
CAZ	251	78.2	269	81	169	74.8	689	78.4
CIP	265	82.6	280	84.3	180	79.6	725	82.5
FEP	161	50.2	211	63.6	89	39.4	461	52.4
GEN	255	79.4	267	80.4	164	72.6	686	78
IPM	210	65.4	254	76.5	148	65.5	612	69.6
LEV	174	54.2	240	72.3	147	65	561	63.8
MEM	211	65.7	254	76.5	149	65.9	614	69.9
COL	0	0	0	0	2	0.9	2	0.2
TG > 1	7	2.2	68	20.5	22	9.7	97	11
TZP	225	70.1	268	80.7	156	69	649	73.8

^a AMK, amikacin; AMS, ampicillin/sulbactam; CAZ, ceftazidime; CIP, ciprofloxacin; FEP, cefepime; GEN, gentamicin; IPM,

imipenem; LEV, levofloxacin; MEM, meropenem; COL, colistin; TG > 1, tigecycline MIC > 1 mg/L; TZP, piperacillin/tazobactam

Supplementary Table S2. Antimicrobial resistance in *A. nosocomialis* and *A. pittii* from the 2010-2014 TSAR collection

	TSAR VII (2010)		TSAR VIII (2012)		TSAR IX (2014)		Overall	
	N = 40		N = 58		N = 64		N = 162	
	N	%	N	%	N	%	N	%
AMK	1	2.5	1	1.7	2	3.1	4	2.5
AMS	1	2.5	1	1.7	4	6.3	6	3.7
CAZ	1	2.5	0	0	2	3.1	3	1.9
CIP	6	15	12	20.7	16	25	34	21
FEP	2	5	2	3.4	6	9.4	10	6.2
GEN	6	15	12	20.7	19	29.7	37	22.8
IPM	3	7.5	9	15.5	14	21.9	26	16
LEV	3	7.5	9	15.5	13	20.3	25	15.4
MEM	3	7.5	8	13.8	14	21.9	25	15.4
COL	0	0	0	0	1	1.6	1	0.6
TG > 1	0	0	2	3.4	1	1.6	3	1.9
TZP	2	5	7	12.1	16	25	25	15.4

^a AMK, amikacin; AMS, ampicillin/sulbactam; CAZ, ceftazidime; CIP, ciprofloxacin; FEP, cefepime; GEN, gentamicin; IPM,

imipenem; LEV, levofloxacin; MEM, meropenem; COL, colistin; TG > 1, tigecycline MIC > 1 mg/L; TZP, piperacillin/tazobactam

Supplementary Table S3. Comparison of patient characteristics of *A. baumannii* versus *A.*

nosocomialis and *A. pittii* from the 2010-2014 TSAR collection

	<i>A. baumannii</i>		<i>A. nosocomialis</i> and <i>A. pittii</i>		P-value ^a
	N = 879		N = 162		
	N	%	N	%	
Hospital type					
Medical centers	364	41.4	94	58.0	< 0.001
Region					< 0.001
North	243	27.7	71	43.8	
Central	370	42.1	35	21.6	
South	184	20.9	43	26.5	
East	82	9.3	13	8.0	
Specimen type					< 0.001
Respiratory	476	54.2	61	37.7	
Blood	131	14.9	68	42.0	
Urine	121	13.8	9	5.6	
Pus/discharge	99	11.3	15	9.3	
Others	52	5.9	9	5.6	
Age^b					< 0.001
< 18	24	2.7	9	5.6	
18-65	280	31.9	67	41.4	
> 65	571	65.0	86	53.1	
Patient location^b					< 0.001
OPD/ER	71	8.1	14	8.6	
Non-ICU	469	53.4	118	72.8	
ICU	337	38.3	30	18.5	

^a Chi-squared test

^b Information regarding age and patient location was not available for some patients.

Supplementary Table S4. Comparison of plasmids carrying *bla*_{OXA-58-like} in this study and in the NCBI database

Isolate No. Accession No.	Plasmid size	Upstream insertion sequences	Accession No. of the best matched plasmid, host species	Plasmid size	Carbapenemase gene in the best matched plasmid	Coverage	Similarity
2010S01-197 CP033563	92 K	IS1006- ISAba3	No comparative plasmid	-	-	-	-
2012C01-137 CP033558	91 K	IS15DI- ISAba3	CP026086.1, <i>A. pittii</i>	112 K	<i>bla</i> _{OXA-58-like}	96%	99%
2014N23-120 CP033547	79 K	IS15DI- ISAba3	CP026086.1, <i>A. pittii</i>	112 K	<i>bla</i> _{OXA-58-like}	98%	99%
2012N21-164 CP033537	73 K	IS1008- ISAba3	LN833432.1, <i>A.</i> <i>baumannii</i>	85 K	<i>bla</i> _{NDM-1}	67%	98%
2014S07-126 CP033531	284 K	ISAba3	CP010351.1, <i>A.</i> <i>johnsonii</i>	399 K	Not detected	87%	99%

Supplementary Table S5. Primers used to detect for *bla*_{OXA-58-like} and its upstream regions, pAB-NCGM253, and *bla*_{OXA-213/272-like}

Target	Primers	Sequences	Size (bp)	Reference
<i>bla</i> _{OXA-58-like}	OXA-58-like-F	CGATCAGAATGTTCAAGCGC	523	Poirel et al ^a
	OXA-58-like-R	CCCCTCTGCGCTCTACATAC		Woodford et al ^b
IS <i>Aba3</i> - <i>bla</i> _{OXA-58-like}	IS <i>Aba3</i> -2F	TATACTATCACTGAGGCAGGTT	773	This study
	OXA-58-like-R	CCCCTCTGCGCTCTACATAC		Woodford et al ^b
Hybrid promoters- <i>bla</i> _{OXA-58-like}	Hybrid-P _{C2}	TTGCAACAGTG <u>CC</u> ATTTT	795	This study
	Hybrid-P _{C3}	TTGCAACAGTG <u>CCC</u> ATTTT		This study
	OXA-58-like-R	CCCCTCTGCGCTCTACATAC		Woodford et al ^b
IS15DI- <i>bla</i> _{OXA-58-like}	IS15DI-F	TACAGATACGCCAGCGG	1124	This study
	OXA-58-like-R	CCCCTCTGCGCTCTACATAC		Woodford et al ^b
IS1006- <i>bla</i> _{OXA-58-like}	IS1006-F	GGAAGGTAAATGTCCACCAGACC	1084	This study
	OXA-58-like-R	CCCCTCTGCGCTCTACATAC		Woodford et al ^b
IS1008- <i>bla</i> _{OXA-58-like}	IS1008-F	CCTACAGATTTACATTCATGGC	1330	This study
	OXA-58-like-R	CCCCTCTGCGCTCTACATAC		Woodford et al ^b

Exclusively <i>bla</i> _{OXA-272-like}	AP-OXA272-F	CCATTGGTACGATGT	265	This study
	AP-OXA272-R	TGTTGCTTTATGATGC		This study
<i>bla</i> _{OXA-213-like} not including <i>bla</i> _{OXA-272-like}	AC-OXA213-F	CGCCCTCTAGAATGAGTATTCAAC	150	This study
	AC-OXA213-R	GTGAAGGATCCTTACCAATGCTTAAT		This study
pAB-NCGM253	OXA24-NCGM1-F	TATGTACTCATGATTGTACA	1980 ^c	This study
	OXA24-NCGM1-R	GCTTAATATGACTTTAGCAT		This study
	OXA24-NCGM2-F	CATATTCTGTGTGAGATAGC	1990 ^c	This study
	OXA24-NCGM2-R	ATTAAAGTTTTACAGAATAA		This study
	OXA24-NCGM3-F	CGAATAGAACCAGACATTCC	1373 ^c	This study
	OXA24-NCGM3-R	AACCATGCTCATATTTGTTTCG		This study
	OXA24-NCGM4-F	TCTAAAATTAACATAATACG	2180 ^c	This study
	OXA24-NCGM4-R	GTCACGCCAGTATTAACCAA		This study
	OXA24-NCGM5-F	CGCCATAAATTACACCTCT	1241 ^d	This study
	OXA24-NCGM5-R	AGGCATTGTCCTTTACAAC		This study

	OXA24-NCGM6-F	AAATCATCCGAATCATTAGTGCA	889 ^e	This study
	OXA24-NCGM6-R	GACTTAGCTAAACTTGGTTCAAC		This study

^a Poirel L, Marqué S, Héritier C et al. OXA-58, A Novel Class D β -Lactamase Involved in Resistance to Carbapenems in *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 2005; **49**: 202–208.

^b Woodford N, Ellington MJ, Coelho JM et al. Multiplex PCR for genes encoding prevalent OXA carbapenemases in *Acinetobacter* spp. *Int J Antimicrob Agents* 2006; **27**: 351-3.

^c 30 cycles of denaturation at 94 °C for 30 s, annealing at 53°C for 30s and elongation at 72 °C for 2.5 min.

^d 30 cycles of denaturation at 94 °C for 30 s, annealing at 58°C for 30s and elongation at 72 °C for 1.5 min.

^e 30 cycles of denaturation at 94 °C for 30 s, annealing at 54°C for 30s and elongation at 72 °C for 1 min.

Supplementary Table S6. Detection of *bla*_{OXA-272-like} in *Acinetobacter* spp. by PCR

	<i>A. baumannii</i> (n = 20)		<i>A. nosocomialis</i> (n = 20)		<i>A. pittii</i> (n = 20)
	CR	CS	CR	CS	CS
	(n = 10)	(n = 10)	(n = 10)	(n = 10)	(n = 20)
No. of isolates from each hospital ^a	1/2/5/2	0/7/1/2	3/4/2/1	3/3/1/3	14/0/0/6
Year	2010-2013	2004-2013	2003-2015	2004-2013	2012-2014
Imipenem MIC range	8-128	0.25-2	8-64	0.125-2	0.25-1
Meropenem MIC range	8-64	0.25-2	8-128	0.25-2	0.25-0.5
Carbapenemase genes ^b					
<i>bla</i> _{OXA-23-like}	8	0	1	0	0
<i>bla</i> _{OXA-24-like}	2	0	5	0	0
IS <i>Aba1</i> - <i>bla</i> _{OXA-51-like}	2	0	0	0	0
Hybrid promoter- <i>bla</i> _{OXA-58-like}	1	0	4	0	0
<i>bla</i> _{OXA-272-like}	0	0	0	0	20
<i>bla</i> _{IMP}	0	0	0	0	0
<i>bla</i> _{VIM}	0	0	0	0	0

CR, carbapenem-resistant; CS, carbapenem-susceptible

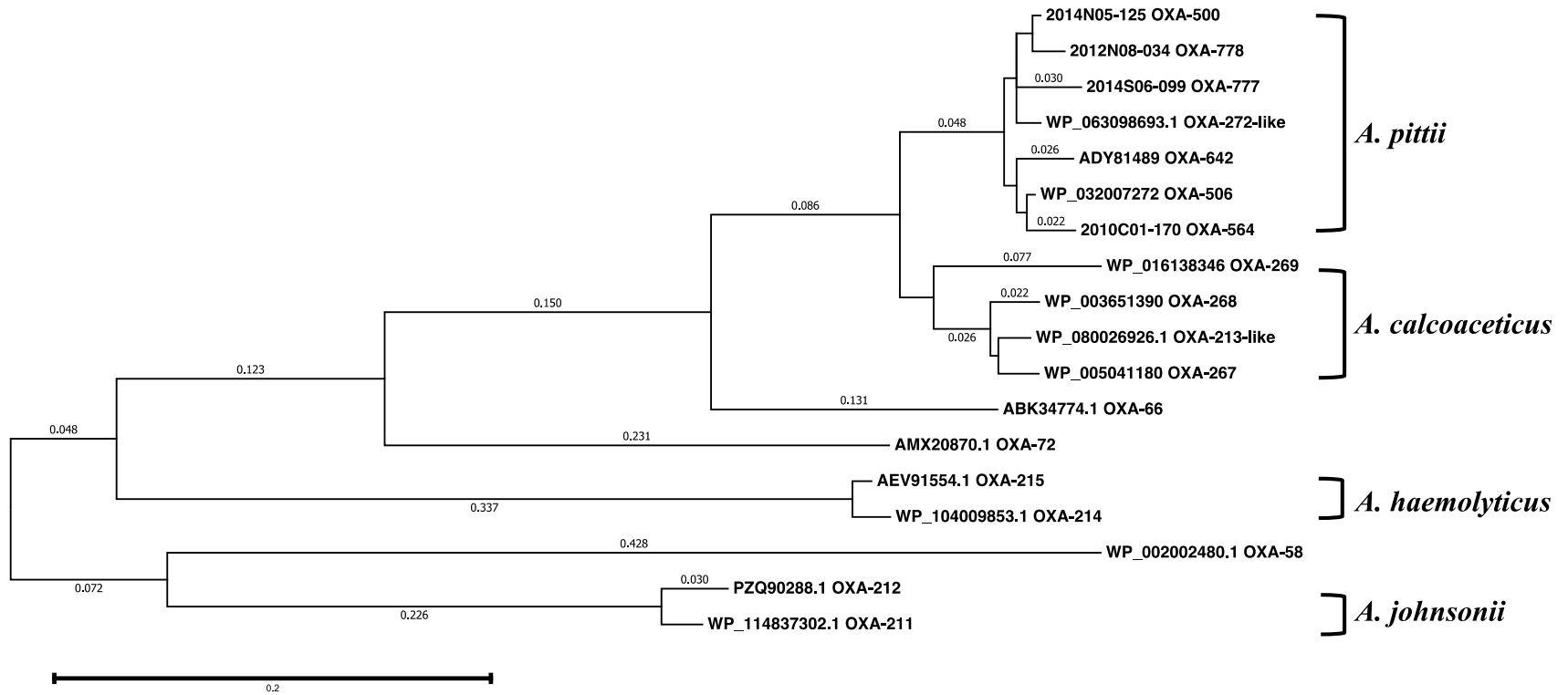
^a Hospitals included Tri-Service General Hospital, Taipei Veterans General Hospital, Mackay Memorial Hospital, and Changhua

Christian Hospital.

^b An isolate may contain multiple carbapenemase genes.

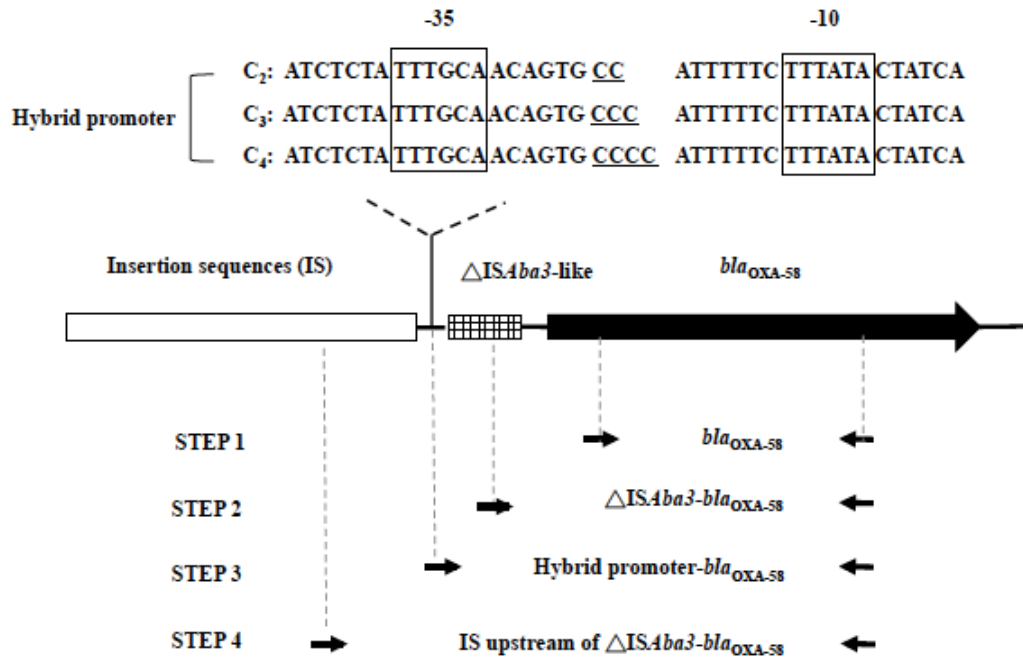
Supplementary Figure S1. Dendrogram of amino acid sequences of carbapenem-hydrolyzing class D β -lactamases (CHDL) in *A. calcoaceticus-baumannii* complex isolates and intrinsic CHDLs in other *Acinetobacter* species (a) and comparison of the amino acid sequences of OXA-272-like and OXA-213-like (b). (a) Amino acid sequences of our isolates and amino acid sequences from NCBI were compared. Accession no. (or isolate no. from this study), and CHDL type are indicated. The CHDL in *A. pittii* belonged to OXA-272-like and those in *A. calcoaceticus* belonged to OXA-213-like. The species was identified based on the sequence of *gyrB*. (b) Multiple amino acid sequence alignment was generated using default settings of CLUSTALW at https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_server.html.

(a)



Supplementary Figure S2. Scheme to identify the genetic background of *bla*_{OXA-58} (a) and validation of the mixture of forward primers in step 3 (b). The number of cytosines in hybrid promoter regions, which are underlined in (a), differed in our isolates and ranged from one to three in isolates C₂ (2012N21-164), C₃ (2014N23-120), and C₄ (2010S01-197), respectively. Sequences deposited in NCBI also showed this variation (accession no. CP026086.1, JQ241789, and GU327621.1). Therefore, three sets of forward primers (Hybrid-P_{C2}, -P_{C3}, and -P_{C4}) were designed. PCR with the mixture of Hybrid-P_{C2} and Hybrid -P_{C3} primers yielded the best results. M, marker; NC negative control.

(a)



(b)

