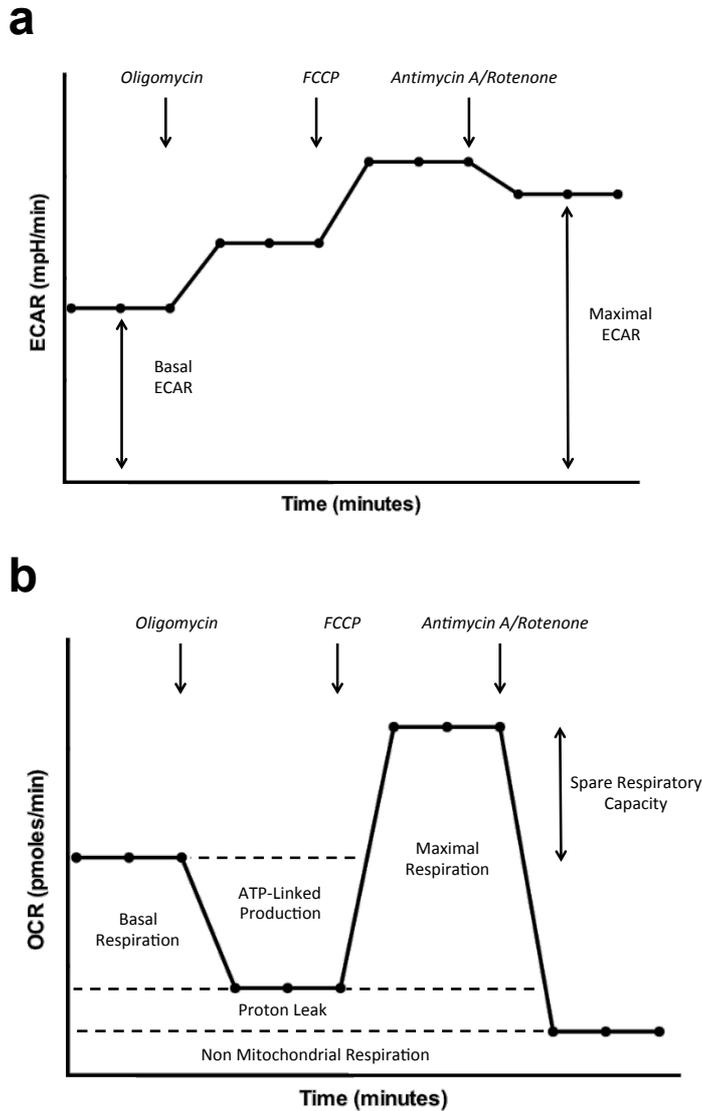


SUPPLEMENTARY INFORMATION

Immediate TCR-induced metabolic response in naïve CD4+ T-cells is mediated by

Akt and STAT5

Jones et al.,



Supplementary Figure 1. Metabolic parameters measured using oxygen consumption rate (OCR) or extracellular acidification rate (ECAR) using the Seahorse XF^e analyser (a) Oxidative parameters of the mitochondria measured using OCR (pmoles/min), specifically basal, ATP-linked, maximal respiration, spare respiratory capacity, proton leak and non-mitochondrial respiration. Calculated with use of injections, oligomycin, FCCP and antimycin A/rotenone. (b) Glycolytic parameters measured using ECAR (mpH/min), basal and maximal ECAR levels with use of injections above.

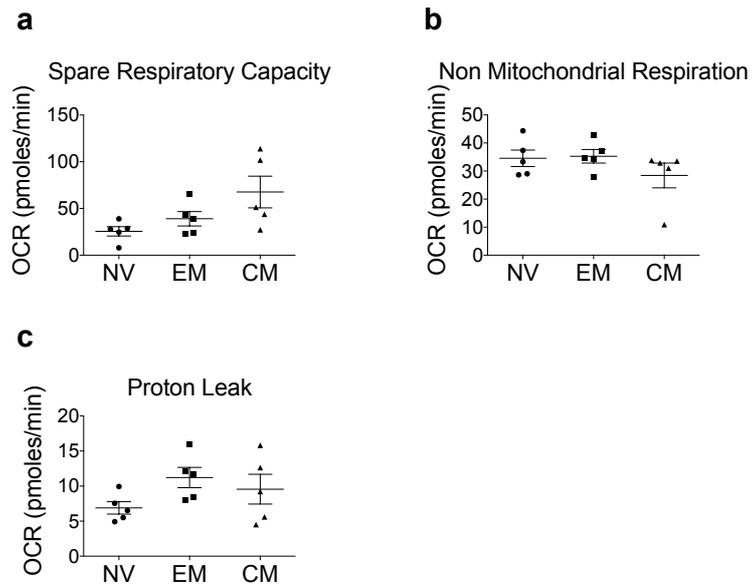
a

Parameter (OCR)	Calculation
Proton leak	Difference between last three and oligomycin injection measurements
Non-mitochondrial respiration	Measurement after antimycin A/rotenone
Basal respiration	Initial three measurements – non-mitochondrial respiration
ATP-linked production	Basal respiration – proton leak
Maximal respiration	Difference between FCCP and antimycin A/rotenone injections
Spare respiratory capacity	Basal respiration – maximal respiration

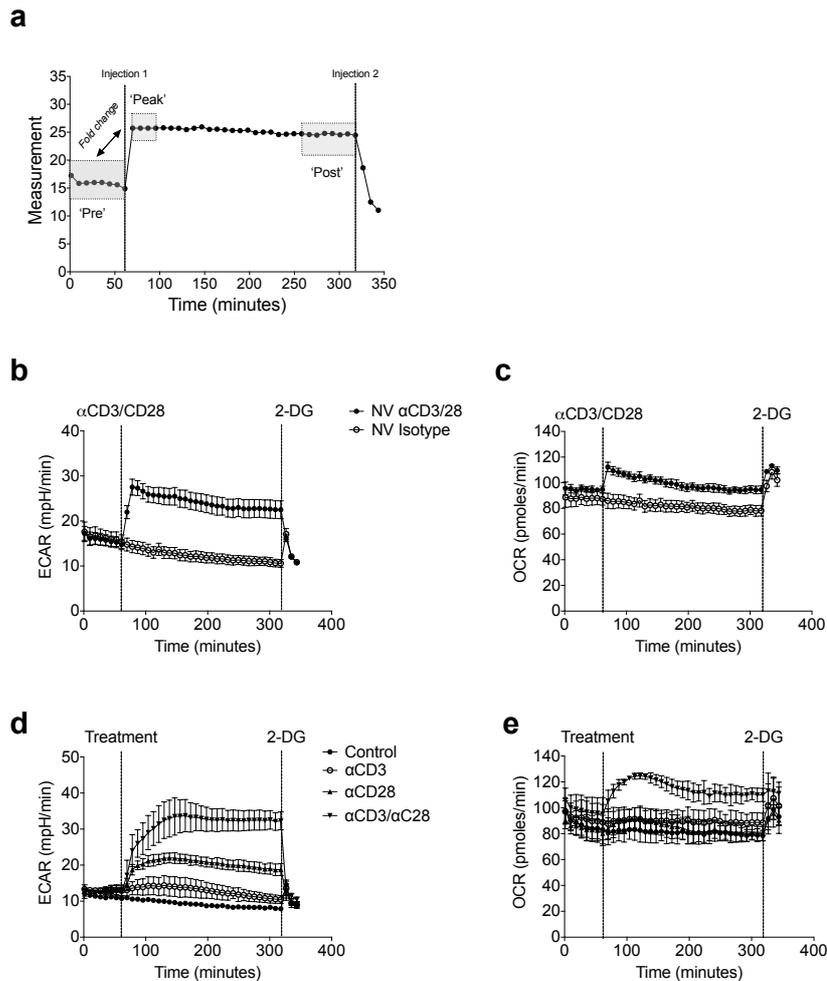
b

Parameter (ECAR)	Calculation
Basal ECAR	Average initial three measurements
Maximal ECAR	Average last three measurements

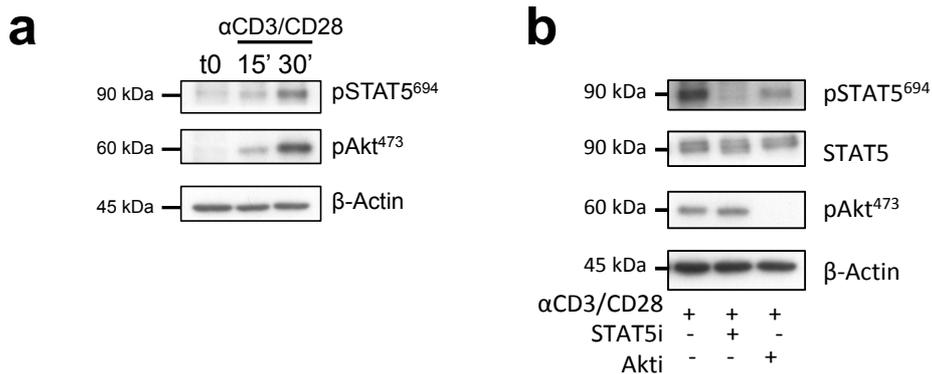
Supplementary Table 1. Various calculations used to measure metabolic parameters. (a) Oxidative phosphorylation parameter calculations and (b) glycolytic parameter calculations. Both measured using averaged triplicate measurements.



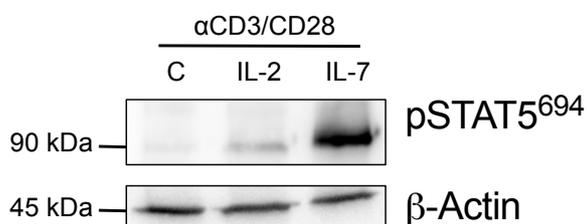
Supplementary Figure 2. Additional oxidative parameters between NV, EM and CM T cells (a) Spare respiratory capacity, (b) Non-mitochondrial respiration and (c) proton leak of NV, EM and CM subset populations. Data are from five independent experiments and are expressed as mean \pm SEM.



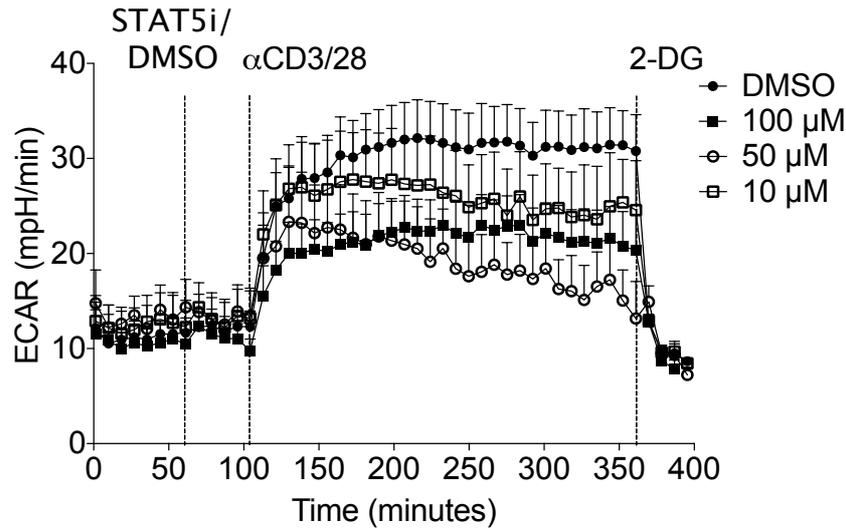
Supplementary Figure 3. NV T cells engage glycolysis and oxygen consumption upon stimulation. (a) Method of calculating fold change and 'pre', 'peak' and 'post' measurements. (b) Extracellular acidification rate (ECAR) and (c) oxygen consumption rate of NV T cells upon stimulation with antibodies, anti-CD3 (0.2 μ g/mL) and anti-CD28 (20 μ g/mL). NV T cells were subjected to anti-CD3 and anti-CD28 injection with 2-deoxy-D-glucose (2-DG; 100 mM) added at the end of the experiment to immediately arrest glycolysis. Simultaneous measurements of ECAR and OCR changes that occurred during the assay. Matched isotype controls for the activating antibodies were used and did not induce any metabolic changes. (d) ECAR and (e) OCR of NV T cells upon stimulation with antibodies, anti-CD3 (0.2 μ g/mL), anti-CD28 (20 μ g/mL) or a combination of anti-CD3 and anti-CD28 (0.2 and 20 μ g/mL respectively). Data are from four (b,c) or three (d,e) independent experiments and are expressed as mean \pm SEM.



Supplementary Figure 4. STAT5 and Akt are immediately phosphorylated upon TCR ligation in NV T cells. (a) NV T cells were activated for 15 or 30 minutes with anti-CD3 (2 μ g/mL) and anti-CD28 (20 μ g/mL). Protein immunoblot for pSTAT5⁶⁹⁴, pAkt⁴⁷³ and housekeeping β -actin. (b) NV T cells were activated for 30 minutes in the presence of either the STAT5 inhibitor (N'-((4-Oxo-4H-chromen-3-yl)methylene)nicotinohydrazide; 100 μ M) or Akt1/2 inhibitor (10 μ M). Protein immunoblot for pSTAT5⁶⁹⁴, STAT5, pAkt⁴⁷³ and housekeeping β -actin. Data from one representative sample of four (a) or two (b) experiments are shown.

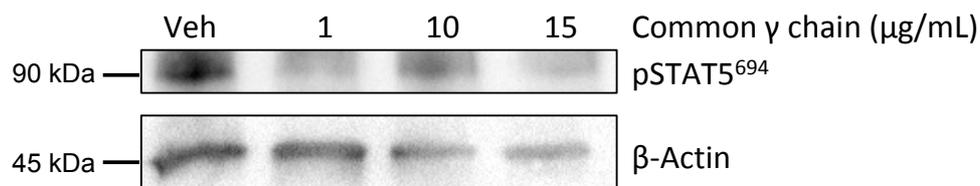


Supplementary Figure 5. The effect of gamma chain cytokines IL-2 and IL-7 on STAT5 phosphorylation. NV T cells were activated for 30 minutes with anti-CD3 (2 μ g/mL) and anti-CD28 (20 μ g/mL) with or without IL-2 or IL-7 (both 10 ng/mL; Miltenyi Biotec). Protein immunoblot for pSTAT5⁶⁹⁴ and housekeeping β -actin; data from one representative sample of two experiments are shown.



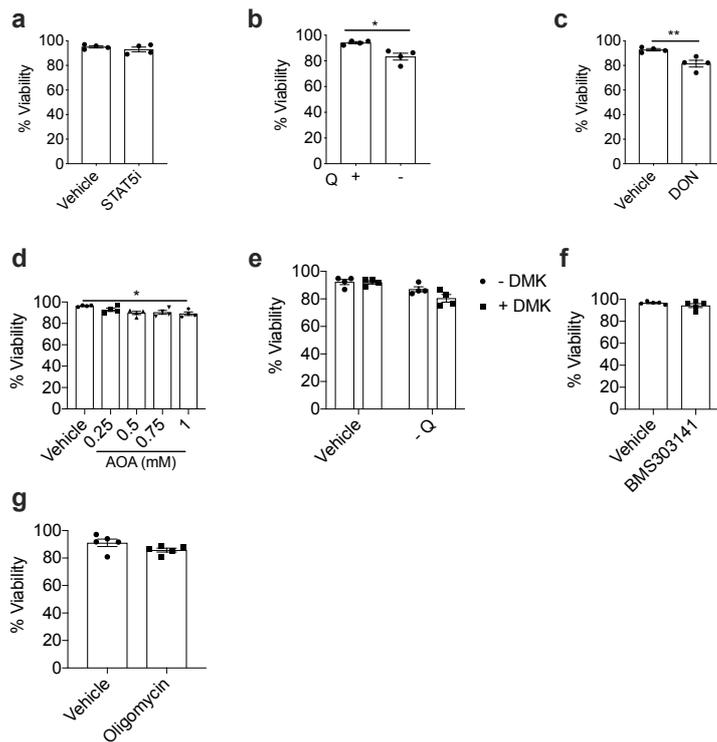
Supplementary Figure 6. Glycolytic response to dose course of STAT5 inhibitor.

NV T cells were exposed to STAT5 inhibitor, N'-((4-Oxo-4H-chromen-3-yl)methylene)nicotinothiazide at varying concentrations (10, 50 and 100 μM) before activation with anti-CD3 (2 μg/mL) and anti-CD28 (20 μg/mL) and the glycolytic response measured via ECAR. A final injection of 2-DG was used to arrest glycolysis. This experiment was repeated three to five times and data are expressed as mean + SEM.

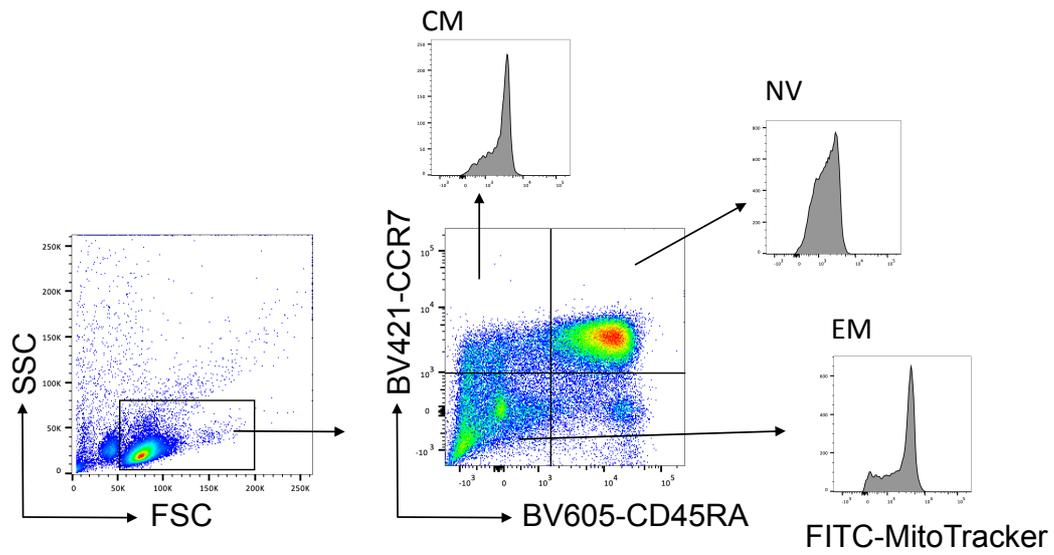


Supplementary Figure 7. The common γ chain antibody reduces IL-2 mediated STAT5 phosphorylation in NV T cells.

Immunoblot of NV T cells stimulated with IL-2 (10 ng/mL; Miltenyi Biotec) and a common gamma chain antibody (1, 10 and 15 μg/mL) for 3 hours. Protein immunoblot for pSTAT5⁶⁹⁴ and housekeeping β-actin; data from one representative sample of two experiments are shown.

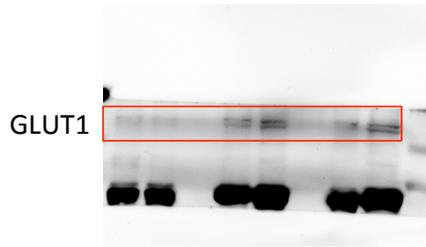


Supplementary Figure 8. The effect of DMK supplementation, oligomycin, AOA and BMS303141 inhibition on NV T cell viability. (a) Viability was determined by DRAQ7 (1 μ M) flow cytometry of CD4+ NV T cells. Stimulated NV T cells with anti-CD3 (2 μ g/mL) and anti-CD28 (20 μ g/mL) were cultured for 24 h with or without (a) STAT5i (100 μ M), (b) glutamine withdrawal, (c) DON treatment, (d) AOA (0.25-1 mM), (e) \pm glutamine withdrawal \pm DMK (0.3 mM), (f) BMS303141 (0.01-1 μ M) or (g) oligomycin (100 nM). Data are from four independent experiments (a-f) or five experiments (g) and are expressed as mean + SEM; * $p \leq 0.05$, ** $p \leq 0.01$.



Supplementary Figure 9. Representative gating strategy. Gating strategy used to analyse MitoTracker Green content of CD4+ NV, EM and CM T cell subsets (Figure 2f).

Figure 1d.



Reprobed for HKII



Figure 4a/5a

NV T cell

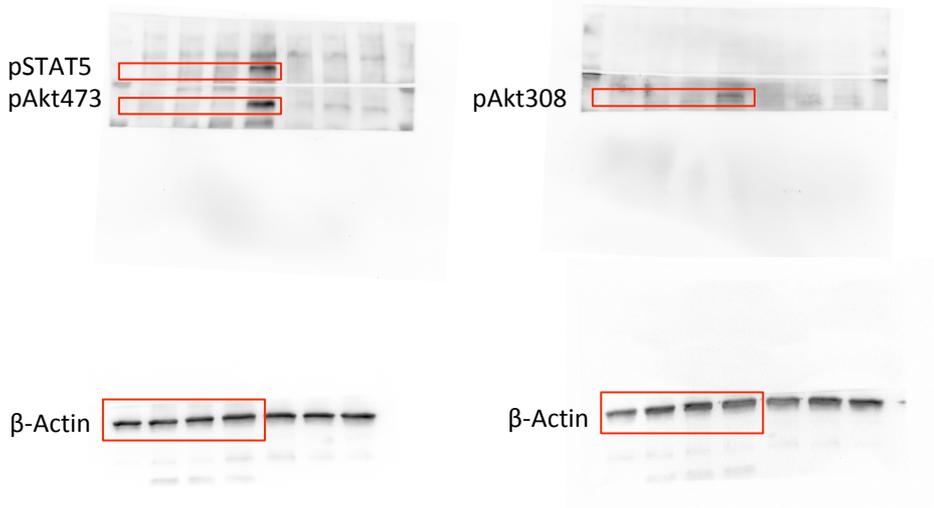


Figure 4a/5a

EM T cell

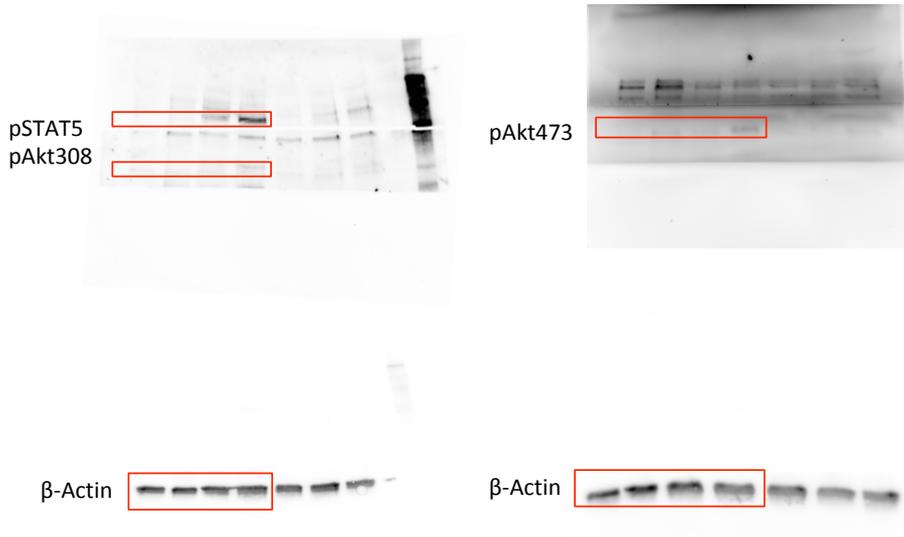


Figure 4a/5a

CM T cell

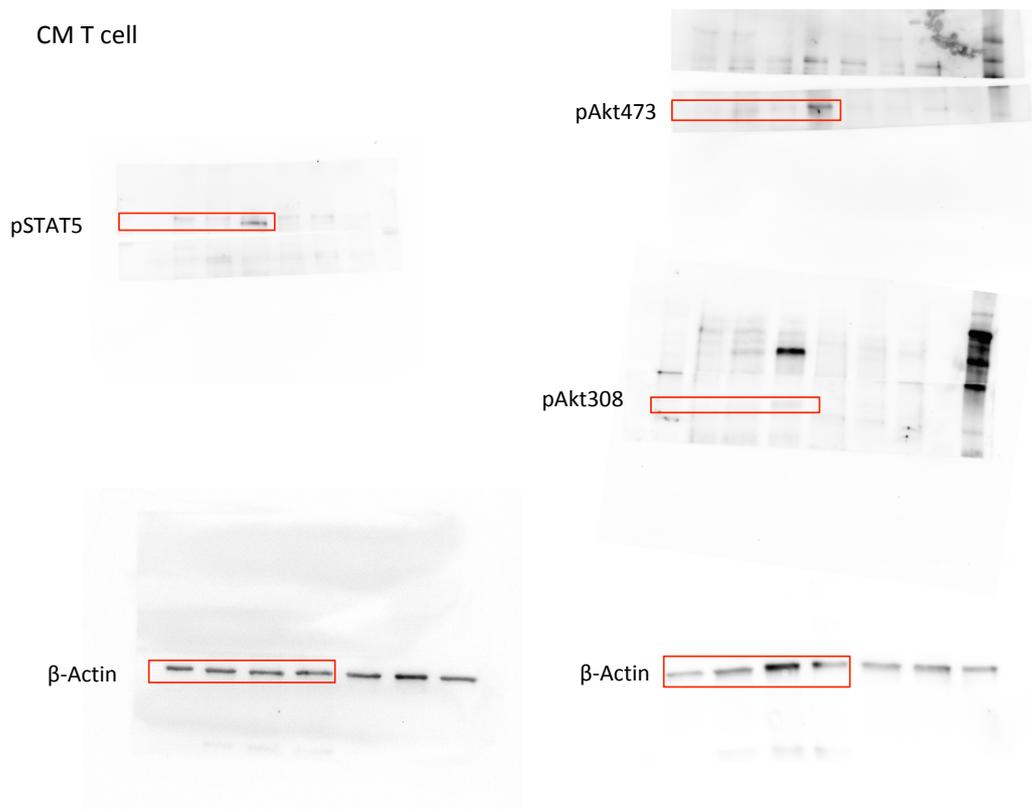


Figure 5b



Figure 5m

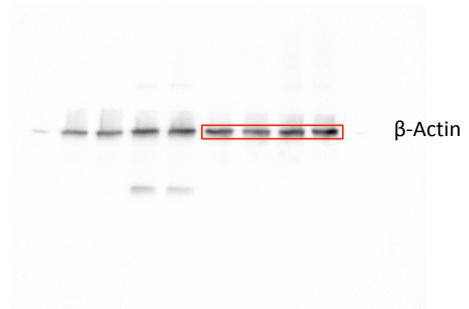
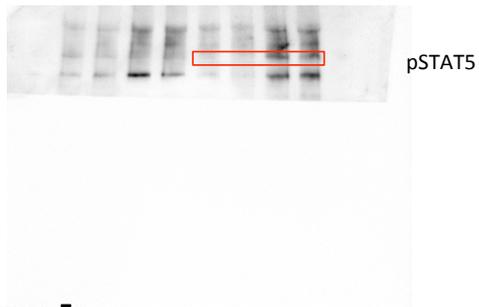


Figure 5n

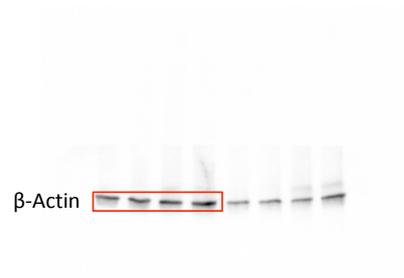
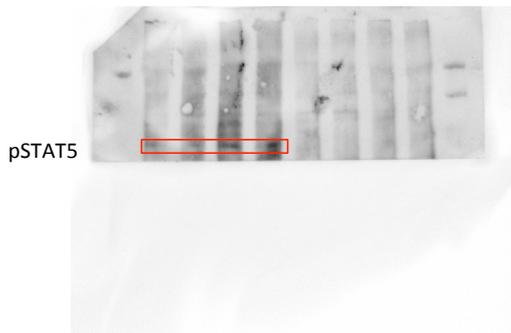
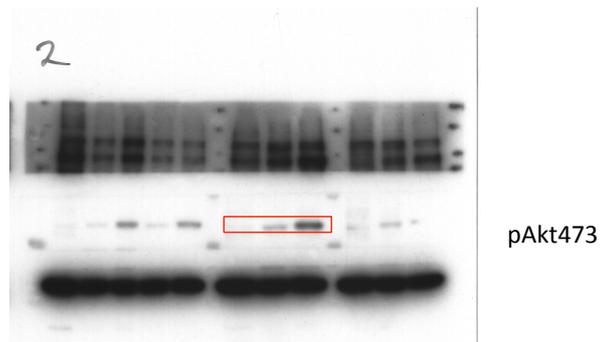
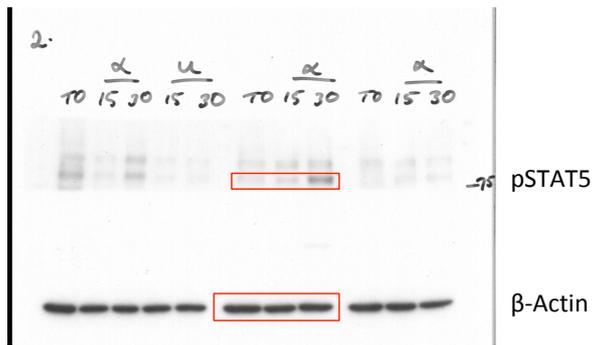


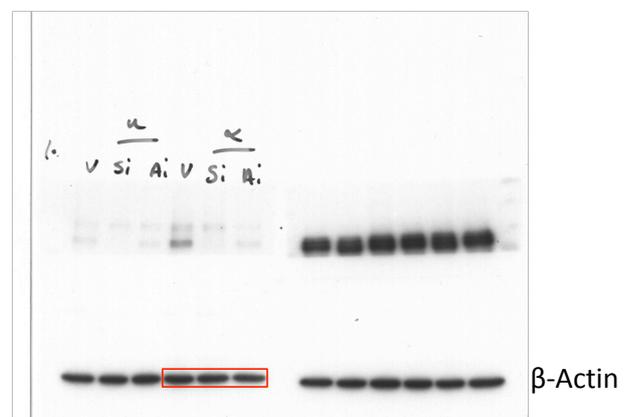
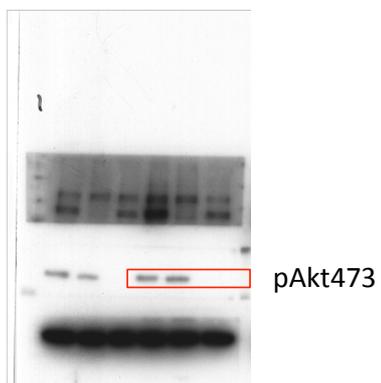
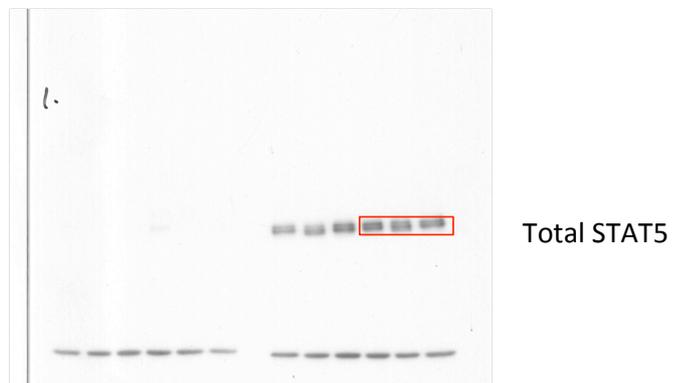
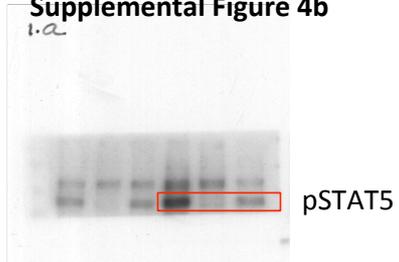
Figure 6f



Supplemental Figure 4a



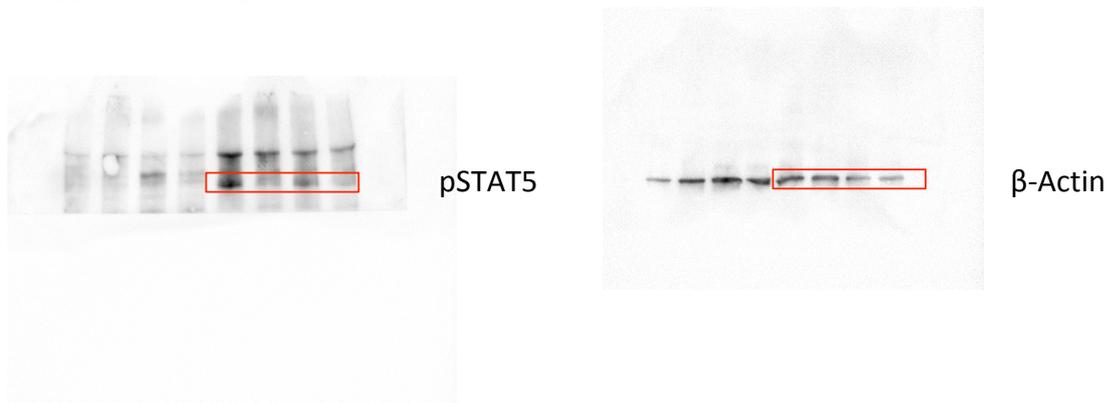
Supplemental Figure 4b



Supplemental Figure 5



Supplemental Figure 7



Supplementary Figure 10. Original uncropped immunoblots for main figures and supplementary figures.