

1 **Electronic Supplemental Material for “Common Caribbean corals exhibit highly variable**
2 *responses to future acidification and warming”*

3
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7
8 **Supplemental Methods:**

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10 **(a) Coral collection**

11 In June 2015, 6 colonies each of 4 reef-building coral species (*Siderastrea siderea*,
12 *Pseudodiploria strigosa*, *Porites astreoides*, and *Undaria tenuifolia*; figure S1) were collected
13 from an inshore reef (Port Honduras Marine Reserve; 16°11'23.5314"N, 88°34'21.9360"W) and
14 6 colonies of each of the 4 coral species were collected from an offshore reef (Sapodilla Cayes
15 Marine Reserve; 16°07'00.0114"N, 88°15'41.1834"W) along the Belize Mesoamerican Barrier
16 Reef System (MBRS) at a depth of 3 to 5 m. A total of 48 coral colonies were collected from
17 both reef environments (2 reef environments x 4 species x 6 colonies). The inshore reef is 9 km
18 from the mainland of Belize, while the offshore reef is approximately 37 km from the mainland.

19
20 **(b) Experimental design and setup**

21 Corals were transported to Northeastern University's natural flow-through seawater
22 system located at the Marine Science Centre, where corals were sectioned with a seawater-
23 cooled tile-cutting saw. Each sectioned coral fragment (approximate surface area: 5 cm x 3 cm =
24 15 cm²; approximate thickness: 2 cm) was mounted on to the outer surface of a 47 mm
25 polystyrene petri dish (EMD Millipore; Billerica, Massachusetts, USA) using Loctite®
26 cyanoacrylate adhesive (Düsseldorf, Germany). All 384 coral fragments (i.e., 48 colonies x 8
27 fragments) were placed into 1 of 8 treatments (4 fragments per species per tank; 16 fragments per
28 tanks; 384 fragments in total; figure S2) filled with 5 µm-filtered seawater obtained from
29 Massachusetts Bay off the coast of Boston, Massachusetts (see table S1 for *in situ* water
30 chemistry data from Belize) [1, 2]. Corals were maintained in natural seawater at a salinity (±SD)
31 of 30.7 (±0.8) and temperature (±SD) of 28.2°C (±0.5) for a recovery period of 23 days. After
32 recovery, temperature and *p*CO₂ were adjusted every other day over a 20-day interval until target
33 experimental conditions were approximately achieved for each treatment (temperature: 28 and
34 31°C; *p*CO₂: 280, 400, 700, 2800 µatm). Seawater temperatures in experimental tanks were
35 incrementally increased by 0.4°C every 3 days and experimental *p*CO₂ was adjusted by -12 µatm
36 (pre-industrial), 0 µatm (current-day), +30 µatm (end-of-century), and +240 µatm (extreme)
37 during the 20-day adjustment interval before starting the 30-day acclimation period. Four *p*CO₂
38 treatments corresponding to pre-industrial (311/288 µatm), current-day (*p*CO₂ control; 405/447
39 µatm), end-of-century (701/673 µatm), and an extreme (3309/3285 µatm) *p*CO₂ were maintained
40 at two temperatures corresponding to the corals' approximate present day mean annual
41 temperature (28°C; determined by over 10 years of *in situ* records) [3-5] and projected end-of-
42 century annual mean temperature (31°C) [6].

43 Experimental 42 L acrylic tanks were illuminated by full spectrum LED lights
44 (Euphotica; 120W, 20000K) on a 10:14 h light:dark cycle with photosynthetically active
45 radiation (PAR) of ca. 300 µmol photons m⁻² s⁻¹ to simulate natural light cycles occurring within
46 the corals' native habitat [7]. PAR was regularly measured within each tank using a LI-COR LI-

47 1500 data logger affixed with a LI-COR LI-192 2π underwater quantum sensor (LI-COR;
48 Lincoln, Nebraska, USA; figure S3). Experimental tanks were covered with an acrylic lid and
49 wrapped in cellophane plastic to facilitate equilibrium between the gas mixtures and the
50 experimental seawaters and to minimize evaporative water loss. Circulation and turbulence in the
51 experimental tanks were maintained with a Maxi-Jet[®] 400 L h⁻¹ powerhead (Marineland;
52 Blacksburg, Virginia, USA), which have been used in previous common garden experiments on
53 corals from Belize [7, 8]. Freshly filtered natural seawater was added via the flow-through
54 system so that the water in each tank was replenished *ca.* 1.3 times per day.

55 Experimental $p\text{CO}_2$ gas mixtures were measured using Qubit S151 (range 0-2000 μatm ;
56 accuracy $\pm 1 \mu\text{atm}$) and S153 (range 0-10%; accuracy $\pm 0.3\%$) infrared $p\text{CO}_2$ analyzers (Qubit
57 Systems; Kingston, Ontario, Canada) calibrated with certified air- CO_2 gas standards. High-
58 precision digital solenoid-valve mass flow controllers (*Aalborg* Instruments and Controls;
59 Orangeburg, NY, USA) were used to bubble air alone (401; 447 μatm), or in combination with
60 CO_2 -free air (311; 288 μatm) or CO_2 gas (701; 673; 3309; 3285 μatm) with compressed air to
61 achieve gas mixtures of the desired $p\text{CO}_2$, and bubbled into each tank and sump via flexible air
62 bubblers (table 2; figure S4). Because temperature affects the solubility of CO_2 in seawater, the
63 two temperature treatments averaged different carbonate parameters for each of the $p\text{CO}_2$
64 treatments, despite being sparged with the same gas mixture ratios (figure S4). These eight
65 $p\text{CO}_2$ -temperature combinations were replicated three-fold (24 tanks total) and yielded the
66 following treatment conditions ($\pm\text{SD}$): 311 (± 96), 405 (± 91), 701 (± 94), 3309 (± 414) $\mu\text{atm } p\text{CO}_2$
67 at 28°C (± 0.4); and 288 (± 65), 447 (± 152), 673 (± 104), 3285 (± 484) $\mu\text{atm } p\text{CO}_2$ at 31.0°C
68 (± 0.4). The temperature of both the 28 and 31°C treatments were maintained using 50W glass
69 aquarium heaters within each tank and 75W glass aquarium heaters (EHEIM; Deizisau,
70 Germany) in each sump. Temperature, salinity, and pH were measured every other day and water
71 samples were taken using 250 mL ground-glass-stoppered borosilicate glass bottles around 13:00
72 Eastern Time every 10 days throughout the 93-day experimental period (9 September – 17
73 December 2015). Total alkalinity was determined by closed-cell potentiometric Gran titration
74 and DIC was determined by coulometry (UIC 5400), with both methods calibrated with certified
75 Dickson Laboratory standards for seawater CO_2 measurements (Scripps Institution of
76 Oceanography; San Diego, California, USA). Measured temperature, salinity, TA, and DIC were
77 used to calculate carbonate parameters using CO_2SYS [9] with Roy et al. (1993) carbonic acid
78 constants K_1 and K_2 [10], the Mucci (1983) value for the stoichiometric aragonite solubility
79 product [11], and an atmospheric pressure of 1.015 atm (electronic supplementary material;
80 figure S4; tables S2, S3). Moderate deviations between calculated and targeted parameters
81 throughout the duration of the experiment resulted largely from biological activity within the
82 aquaria and from minor seasonal changes in source water chemistry. Temperature was measured
83 using a high precision partial-immersion glass thermometer (precision $\pm 0.3\%$; accuracy $\pm 0.4\%$).
84 Salinity ($\pm\text{SD}$) was measured using a YSI 3200 (Yellow Springs, Ohio, USA) conductivity meter
85 with a 10.0 cm⁻¹ cell and maintained at 31.7 (± 0.2), with slight natural seasonal variation as
86 expected in Massachusetts Bay waters. An AccuFet[™] Solid-State pH probe (Fisher
87 Scientific[™]; Waltham, Massachusetts, USA) calibrated with 7.00 and 10.01 NBS buffers
88 maintained at experimental temperatures was used to measure pH in each tank (table S2; figure
89 S4). Coral fragments within each tank were fed every other day with a mixture of *ca.* 6 g frozen
90 adult *Artemia* sp. and 250 mL concentrated newly hatched live *Artemia* sp. (500 mL⁻¹) to satisfy
91 any heterotrophic feeding by each species [12, 13].

92

93 (c) Buoyant weight quantification

94 Coral fragments were suspended in a 38 L aquarium 4 cm below the surface in seawater
95 (temperature, 28.2°C; salinity, 32.4) using an aluminum wire hanging from a Nimbus NBL 423e
96 Precision Balance (± 0.0002 precision, ± 0.002 accuracy; AE Adam[®]; Oxford, Connecticut, USA).
97 A standard of a known mass was weighed three times before weighing corals in each tank to
98 monitor any deviations in the balance over the course of the experiment. Each coral fragment
99 was weighed three times, averaged, and normalized to surface area. Surface area was quantified
100 in triplicate from photos of each nubbin taken at corresponding intervals using imaging software
101 (IMAGE J).

102 A subsample of fragments from each coral species was selected for constructing the
103 linear regression that relates the coral species' buoyant weight to their dry weight. Buoyant
104 weight ('BW') and dry weight of the fragments are highly correlated for each species (R^2 *S. siderea*
105 = 0.970, $p < 0.001$; R^2 *P. strigosa* = 0.900, $p < 0.001$; R^2 *P. astreoides* = 0.980, $p < 0.001$; R^2 *U. tenuifolia* =
106 0.983, $p < 0.001$), therefore the change in buoyant weight should be proportional to the
107 corresponding change in dry weight (figure S5).

108
109 *S. siderea*: Dry weight (mg) = 1.9 * BW + 3.47, $R^2 = 0.970$

110 *P. strigosa*: Dry weight (mg) = 1.78 * BW + 5.47, $R^2 = 0.900$

111 *P. astreoides*: Dry weight (mg) = 1.93 * BW + 4.51, $R^2 = 0.980$

112 *U. tenuifolia*: Dry weight (mg) = 1.66 * BW + 5.04, $R^2 = 0.983$

114 (d) Linear Extension

115 A calcein horizon was emplaced into coral skeletons at the beginning of the experiment
116 to establish a marker from which linear extension throughout the experiment could be measured
117 [14]. Each experimental tank was dosed with 213.4 g of a 1% calcein solution for 5 days. During
118 this period, the light cycle was increased to 14 h light in all tanks to ensure sufficient uptake of
119 fluorescent marker into skeletons. At the completion of the experiment, tissue was removed from
120 all coral fragments using a precision seawater sprayer (PointZero; Sunrise, Florida, USA).
121 Sections 5mm thick were cut from the middle of each fragment using a DB-100 ReefKeeperTM
122 diamond band saw (Inland; Madison Heights, Michigan, USA). The full thin sections were
123 imaged under a stereo microscope outfitted with a blue fluorescent adapter with excitation 440–
124 460nm (NIGHTSEATM; Lexington, Massachusetts, USA). Linear extension was measured as the
125 total area of new growth above the calcein line (figure S7) measured using imaging software
126 (IMAGE J) divided by the measured length of the coral's lateral growth surface. Extension was
127 then divided by the number of months in the experimental treatments resulting in linear
128 extension per month (mm month^{-1}).

130 (e) Estimation of gross calcification rates

131 Gross calcification rates were estimated by subtracting the corals' calculated gross
132 dissolution rates from their net calcification rates at the aragonite saturation states of each
133 treatment. Gross dissolution was calculated using gross dissolution regression equations derived
134 in Ries et al. [15] for two coral species. The gross dissolution equation ('y') for the massive coral
135 *S. siderea* was used to estimate gross dissolution of the massive corals *S. siderea*, *P. strigosa*,
136 and *P. astreoides* from the current experiment, while the gross dissolution equation for the
137 branching coral *O. arbuscula* was used to estimate gross dissolution of the branching coral *U.*
138 *tenuifolia* [15] (figure S9).

139

140 *S. siderea*: y (%-wt/day) = $0.055 - 0.638 * e^{(-6.187 * \Omega_A + 2.039 * \Omega_A)}$

141 *O. arbuscula*: y (%-wt/day) = $0.073 - 0.638 * e^{(-5.632 * \Omega_A + 2.039 * \Omega_A)}$

142

143 (f) Survival quantification and analysis

144 Coral fragments were assessed for mortality every 30 days and considered dead when no
145 living tissue remained. Impacts of $p\text{CO}_2$ and temperature on survival rates were assessed using a
146 Kaplan-Meier estimate of survival (*survfit*, *survival*, 2.39-5) [16]. Cox proportional hazard
147 models, with colony nested within tank as a random effect, were performed using *coxme* (2.2-5)
148 [17].

149

150 (g) Further explanation of statistical analyses

151 Linear mixed effects models were fit to the calcification and linear extension data.
152 Models were run to include species, $p\text{CO}_2$ (factor), and temperature (factor) as fixed effects with
153 colony (genotype) as a random effect:

154

$$\text{lmer}(\text{rate} \sim \text{species} * (p\text{CO}_2 + \text{temperature}) + (1 | \text{colony}))$$

155

156 This model was selected using AIC and log likelihood tests to determine the best fit for the data.
157 A parametric bootstrap of the data was run 1500 times for each model, resulting in the modelled
158 mean and 95% confidence intervals. Colonies were pooled by natal reef environment in all
159 analyses because this was not a significant predictor of any measured parameter. All statistical
160 analyses were performed using R 3.3.2 for OS X [18].

161

162 A Bayesian hierarchical regression model was fit to calculate credible intervals of the
163 corresponding extracted correlation coefficients using Hamiltonian MCMC, using default
164 uninformative priors. Four chains were run for 1000 iterations after a 1000-iteration warmup.
165 Chains mixed well and all Rhats were less than 1.0. The model was fit with species, $p\text{CO}_2$
166 (factor), and temperature (factor) as fixed effects with colony (genotype) as a random intercept
167 and temperature and $p\text{CO}_2$ as random slopes:

168

$$\text{brms}(\text{rate} \sim \text{species} * (p\text{CO}_2 + \text{temperature}) + (1 + p\text{CO}_2 + \text{temperature} | \text{colony}), \text{family} = \text{gaussian}())$$

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170

171 Supplemental Results:

172

173 (a) Coral survivorship

174 *Siderastrea siderea* maintained nearly 100% survival across treatments, resulting in no
175 significant effect of temperature ($p = 0.23$), $p\text{CO}_2$ ($p = 0.60$), or their interaction ($p = 1.0$) on
176 survival (figure S6a). Survival of *P. strigosa*, *P. astreoides*, and *U. tenuifolia* reared at 31°C was
177 significantly reduced compared to conspecifics reared at 28°C ($p < 0.01$, $p < 0.01$, $p < 0.01$,
178 respectively; figure 3b-d). No *U. tenuifolia* fragments under extreme $p\text{CO}_2$ conditions at 31°C
179 survived the acclimation period, indicating that this species is extremely sensitive to these
180 conditions. Increasing $p\text{CO}_2$ had no effect on survival of *P. astreoides* or *U. tenuifolia* ($p = 0.09$
181 and $p = 0.22$, respectively), while increasing $p\text{CO}_2$ significantly increased survivorship of *P.*
182 *strigosa* ($p < 0.01$), a trend driven by relatively low survival at present-day $p\text{CO}_2$. Finally, the

183 interaction between $p\text{CO}_2$ and temperature had no significant effect on survivorship of *P.*
184 *strigosa*, *P. astreoides*, or *U. tenuifolia* ($p < 0.08$, $p < 0.25$, $p < 0.21$, respectively; figure S6b-d;
185 tables S9, S10, S11).
186

187 **(b) Effects of exposure duration on calcification rate**

188 Differences in calcification rates for the four species were also examined across three 30-
189 day observation intervals (T0-T30, T31-60, and T61-T90) to assess the impact of duration of
190 exposure to treatment conditions on coral calcification rates. Although responses are complex,
191 some general patterns emerged.

192 Specimens of *S. siderea* exhibited a slight increase in calcification rates from the first
193 (T0-T30) to second (T31-T60) intervals in most treatments, followed by a decline from the
194 second to third (T61-T90) interval (figure S13a). In addition, calcification rates for coral reared
195 at 28°C and 31°C under extreme $p\text{CO}_2$ are lower at each interval when compared with the lower
196 $p\text{CO}_2$ treatments.

197 Calcification rates of *P. strigosa* were generally higher at 28°C than at 31°C at every 30-
198 day interval, regardless of $p\text{CO}_2$ treatment. Excluding specimens reared under current-day $p\text{CO}_2$
199 at 28°C, calcification rates progressively declined across the three 30-day observational intervals
200 of the experiment (figure S13b).

201 *Porites astreoides* calcification rates demonstrated a declining trend across observational
202 intervals within most temperature- $p\text{CO}_2$ treatment combinations, and exhibited net dissolution
203 during the final interval (figure S13C). However, some specimens failed to exhibit net
204 calcification during any of the three intervals at either temperature.

205 Calcification rates of *U. tenuifolia* exhibited a decreasing trend across the three
206 observational intervals for all $p\text{CO}_2$ and temperature treatment combinations (figure S13d).
207 Missing data from the 31°C treatment in both the current-day and extreme $p\text{CO}_2$ treatments
208 reflects the low survival rates in these treatments.
209
210

211 **Supplemental Discussion:**

213 **(a) Corals' natal reef environment does not influence resilience to $p\text{CO}_2$ or thermal stress**

214 Rates of calcification, linear extension, and survival were not significantly impacted by
215 natal reef environment (i.e., inshore vs. offshore) of the four coral species investigated here
216 (figures S11, S12; tables S11, S12, S13). This result is consistent with previous laboratory
217 experiments on some of the same and other species of zooxanthellate corals, which found no
218 difference in responses to thermal and $p\text{CO}_2$ stress due to natal reef environment [7, 8], but
219 inconsistent with historical growth records of *S. siderea* obtained from century-scale coral cores
220 that showed that the extension rate of forereef colonies has declined much faster than that of
221 backreef and nearshore colonies [19]. However, it is possible that natal-reef-environment
222 differences in resilience to thermal stress may emerge with more prolonged exposure to
223 acidification and warming stress, as well as with larger sample sizes.
224
225

226 **Supplemental tables and figures:**

227

Reef environment	T (°C)	pCO₂ (µatm)	pH	TA (µM)	DIC (µM)	Ω_A	Salinity
Inshore	26.7	346.7	8.05	2495.9	2112	4.56	32.8
Inshore	26.7	326.0	8.04	2485.9	2090	4.68	32.7
Offshore	27.5	302.5	8.06	2572.8	2124	5.2	34.8
Offshore	27.5	298.1	8.06	2579.3	2126	5.25	34.8
Offshore	27.5	287.5	8.06	2583.8	2120	5.37	34.8

228

229 **Table S1.** Carbonate system parameters of seawater samples obtained in December 2016 from
 230 inshore and offshore locations in southern Belize near coral sampling sites demonstrating
 231 similarity to experimental seawater treatments (see table 1 in the main text).

MEASURED PARAMETERS										
pCO₂ (μatm-v)	311	405	701	3309	288	447	673	3285		
Sal	31.72	31.77	31.69	31.77	31.74	31.72	31.69	31.74		
SD	0.21	0.22	0.22	0.23	0.25	0.25	0.24	0.21		
Range	31.26 - 32.06	31.26 - 32.13	31.23 - 32.03	31.26 - 32.06	31.19 - 32.12	31.03 - 32.16	31.16 - 32.12	31.23 - 32.06		
n	120	120	120	120	120	120	120	120		
Temp (°C)	27.9	28.0	28.1	28.1	31.0	31.1	30.9	31.0		
SD	0.4	0.4	0.5	0.2	0.4	0.5	0.3	0.5		
Range	27.2 - 29.6	27.0 - 29.0	27.1 - 30.2	27.7 - 28.7	30.0 - 32.2	30.4 - 32.5	30.1 - 31.7	30.0 - 33.0		
n	120	120	120	120	120	120	120	120		
pH_{M-NBS}	8.30	8.20	8.01	7.31	8.34	8.21	8.00	7.29		
SD	0.11	0.09	0.34	0.07	0.12	0.11	0.12	0.10		
Range	8.03 - 8.46	7.93 - 8.33	7.62 - 11.62	7.13 - 7.45	7.97 - 8.55	7.94 - 8.51	7.61 - 8.20	7.12 - 7.53		
n	120	120	120	120	120	120	120	120		
TA (μM)	2052	2081	2092	2131	2101	2077	2082	2123		
SD	43	17	37	25	32	32	35	22		
Range	1947 - 2104	2053 - 2121	2012 - 2128	2076 - 2160	2048 - 2152	2010 - 2125	2021 - 2134	2071 - 2148		
n	29	30	30	30	29	30	30	30		
DIC (μM)	1708	1788	1901	2156	1710	1773	1865	2135		
SD	78	52	46	34	57	80	42	28		
Range	1551 - 1829	1702 - 1859	1830 - 1981	2082 - 2217	1611 - 1795	1625 - 1905	1757 - 1917	2084 - 2194		
n	29	30	30	30	29	30	30	30		

Table S2. Average measured parameters for all treatments: salinity (Sal), temperature (Temp), pH, total alkalinity (TA), and dissolved inorganic carbon (DIC). 'SD' represents standard deviation and 'n' is the sample size.

CALCULATED PARAMETERS												
$p\text{CO}_2$ (gas-e)	($\mu\text{atm-v}$)	311	405	701	3309	288	447	673	3285			
SD		96	91	94	414	65	152	104	484			
Range		165 - 520	252 - 553	555 - 981	2442 - 4299	214 - 416	236 - 792	462 - 879	2681 - 4438			
n		29	30	30	30	29	30	30	30			
$\text{pH}_{\text{c-NBS}}$		8.27	8.18	7.97	7.37	8.29	8.15	7.99	7.38			
SD		0.10	0.08	0.05	0.05	0.07	0.11	0.06	0.06			
Range		8.07 - 8.45	8.06 - 8.33	7.85 - 8.05	7.25 - 7.48	8.16 - 8.38	7.93 - 8.34	7.89 - 8.11	7.25 - 7.46			
n		29	30	30	30	29	30	30	30			
$[\text{CO}_3^{2-}]$	(μM)	241	209	145	42	274	217	162	47			
SD		39	28	12	5	31	40	18	6			
Range		173 - 312	170 - 260	115 - 164	32 - 54	217 - 315	144 - 288	129 - 195	34 - 57			
n		29	30	30	30	29	30	30	30			
$[\text{HCO}_3^-]$	(μM)	1459	1568	1737	2029	1429	1545	1687	2009			
SD		109	77	51	29	82	114	51	23			
Range		1235 - 1643	1435 - 1666	1652 - 1841	1967 - 2076	1301 - 1553	1332 - 1742	1551 - 1748	1965 - 2052			
n		29	30	30	30	29	30	30	30			
$[\text{CO}_2]$ (sw)	(μM)	8	10	18	85	7	11	16	79			
SD		2	2	2	11	2	4	2	12			
Range		4 - 13	7 - 14	14 - 25	63 - 111	5 - 10	6 - 19	11 - 21	64 - 109			
n		29	30	30	30	29	30	30	30			
Ω_A		4.0	3.4	2.4	0.7	4.6	3.6	2.7	0.8			
SD		0.6	0.5	0.2	0.1	0.5	0.7	0.3	0.1			
Range		2.8 - 5.1	2.8 - 4.3	1.9 - 2.7	0.5 - 0.9	3.6 - 5.2	2.4 - 4.8	2.2 - 3.3	0.6 - 0.9			
n		29	30	30	30	29	30	30	30			

Table S3. Average measured parameters for all treatments: $p\text{CO}_2$ of the mixed gases in equilibrium with seawaters ($p\text{CO}_2$ (gas-e)); calculated pH (pH_{c}); carbonate ion concentration ($[\text{CO}_3^{2-}]$); bicarbonate ion concentration ($[\text{HCO}_3^-]$); dissolved carbon dioxide ($[\text{CO}_2]_{\text{sw}}$); and aragonite saturation state (Ω_A). ‘SD’ represents standard deviation and ‘n’ is the sample size.

Model	AIC	df
Temperature * Reef	508.5704	6
Reef	506.5132	4
Temperature	505.4378	4
Species * Reef	481.4342	10
Species	477.2899	6
Reef * $p\text{CO}_2$ * Temperature	476.3029	18
$p\text{CO}_2$ * Reef	475.0965	10
$p\text{CO}_2$	473.5468	6
Temperature * $p\text{CO}_2$	471.7448	10
Species * Temperature * Reef	470.4927	17
Species * Temperature	459.8295	10
Species * $p\text{CO}_2$ * Reef	458.286	34
Species * $p\text{CO}_2$ * Reef + Temperature	457.005	35
Species * $p\text{CO}_2$ + Temperature + Reef	451.5823	20
Species * $p\text{CO}_2$	451.1012	18
Species * $p\text{CO}_2$ + Temperature	449.6505	19
Species * $p\text{CO}_2$ * Temperature * Reef	449.1111	59
$p\text{CO}_2$ * Temperature * Reef + Species	448.8439	21
Species + $p\text{CO}_2$ * Temperature + Reef	446.5169	14
Species * Reef + $p\text{CO}_2$ + Temperature	446.3451	14
Species + $p\text{CO}_2$ + Temperature * Reef	445.5166	12
$p\text{CO}_2$ * Temperature + Species	444.6838	13
Species + $p\text{CO}_2$ + Temperature + Reef	444.5401	11
$p\text{CO}_2$ * Reef + Species + Temperature	443.6495	14
Species + $p\text{CO}_2$ + Temperature	442.6241	10
Species * $p\text{CO}_2$ * Temperature + Reef	440.0991	33
Species * $p\text{CO}_2$ * Temperature	438.1393	32
Species * Temperature * Reef + $p\text{CO}_2$	438.0031	20
Species * ($p\text{CO}_2$ + Temperature)	432.9082	22
Species * Temperature + Reef + $p\text{CO}_2$	430.5345	14
Species * Temperature + $p\text{CO}_2$	428.5378	13

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Table S4. Summary of AIC and degrees of freedom (df) for all model combinations. The model combination in bold is the final model used in this analysis.

Species	Treatment	N	Mean Calcification (mg cm ² day ⁻¹)	Lower 95% CI	Upper 95% CI	
<i>S. siderea</i>	28°C	311 µatm	10	1.106	0.872	1.342
		405 µatm	12	1.256	1.038	1.468
		701 µatm	11	1.084	0.875	1.302
		3309 µatm	12	0.280	0.070	0.492
	31°C	288 µatm	8	1.093	0.854	1.335
		447 µatm	11	1.243	1.026	1.448
		673 µatm	11	1.071	0.856	1.286
		3285 µatm	12	0.267	0.047	0.468
<i>P. strigosa</i>	28°C	311 µatm	15	1.198	0.989	1.408
		405 µatm	5	0.504	0.209	0.828
		701 µatm	14	0.665	0.443	0.871
		3309 µatm	16	0.181	-0.015	0.374
	31°C	288 µatm	9	0.202	-0.023	0.450
		447 µatm	6	-0.493	-0.801	-0.184
		673 µatm	7	-0.332	-0.606	-0.088
		3285 µatm	8	-0.815	-1.058	-0.564
<i>P. astreoides</i>	28°C	311 µatm	11	0.072	-0.159	0.304
		405 µatm	12	0.010	-0.233	0.231
		701 µatm	10	-0.196	-0.438	0.050
		3309 µatm	12	-0.680	-0.903	-0.456
	31°C	288 µatm	6	0.229	-0.039	0.497
		447 µatm	8	0.166	-0.073	0.419
		673 µatm	9	-0.039	-0.280	0.219
		3285 µatm	4	-0.523	-0.803	-0.246
<i>U. tenuifolia</i>	28°C	311 µatm	11	0.147	-0.138	0.432
		405 µatm	7	0.237	-0.125	0.611
		701 µatm	4	0.029	-0.398	0.465
		3309 µatm	5	-0.241	-0.650	0.177
	31°C	288 µatm	4	0.129	-0.304	0.583
		447 µatm	0	NA	NA	NA
		673 µatm	1	0.011	-0.565	0.601
		3285 µatm	0	NA	NA	NA

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240 **Table S5.** Bootstrapped modelled mean calcification rate for each species in all $p\text{CO}_2$ and
241 temperature treatments reported in $\text{mg cm}^2 \text{ day}^{-1}$. Sample sizes (N) and 95% confidence intervals
242 (CI) are reporter for each modelled mean calcification rate (figure 1).

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Fixed effect	Value	SE	<i>t</i> -value
(Intercept)	1.089	0.163	6.664
Species (PSTR)	0.106	0.224	0.471
Species (PAST)	-1.020	0.231	-4.412
Species (UTEN)	-0.947	0.251	-3.769
<i>p</i> CO ₂ - current	0.163	0.148	1.102
<i>p</i> CO ₂ - end-of-century	-0.002	0.150	-0.013
<i>p</i> CO ₂ - extreme	-0.809	0.146	-5.522
Temperature (31°C)	-0.011	0.100	-0.113
Species (PSTR) * <i>p</i> CO ₂ - current	-0.887	0.228	-3.886
Species (PAST) * <i>p</i> CO ₂ - current	-0.224	0.215	-1.039
Species (UTEN) * <i>p</i> CO ₂ - current	-0.074	0.280	-0.263
Species (PSTR) * <i>p</i> CO ₂ - end-of-century	-0.523	0.205	-2.558
Species (PAST) * <i>p</i> CO ₂ - end-of-century	-0.267	0.220	-1.216
Species (UTEN) * <i>p</i> CO ₂ - end-of-century	-0.121	0.295	-0.410
Species (PSTR) * <i>p</i> CO ₂ - extreme	-0.189	0.199	-0.950
Species (PAST) * <i>p</i> CO ₂ - extreme	0.063	0.221	0.284
Species (UTEN) * <i>p</i> CO ₂ - extreme	0.420	0.298	1.409
Species (PSTR) * Temperature (31°C)	-1.066	0.154	-6.923
Species (PAST) * Temperature (31°C)	0.166	0.153	1.080
Species (UTEN) * Temperature (31°C)	-0.013	0.273	-0.048
Colony (intercept)	0.147		
Residual	0.215		

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Table S6. Summary output of the linear mixed effects model used to determine the relationship between calcification rates, *p*CO₂, and temperature for all four coral species (PSTR = *P. strigosa*; PAST = *P. astreoides*; UTEN = *U. tenuifolia*). Temperature and *p*CO₂ were treated as factors.

Species	Treatment	N	Mean LE (mm day ⁻¹)	Lower 95% CI	Upper 95% CI	
<i>S. sidera</i>	28°C	311 µatm	11	0.0080	0.0070	0.0090
		405 µatm	9	0.0082	0.0074	0.0091
		701 µatm	11	0.0086	0.0076	0.0095
		3309 µatm	12	0.0075	0.0066	0.0083
	31°C	288 µatm	10	0.0069	0.0059	0.0079
		447 µatm	8	0.0071	0.0062	0.0081
		673 µatm	11	0.0075	0.0066	0.0083
		3285 µatm	12	0.0063	0.0055	0.0072
<i>P. astreoides</i>	28°C	311 µatm	9	0.0059	0.0048	0.0069
		405 µatm	9	0.0047	0.0037	0.0058
		701 µatm	9	0.0046	0.0036	0.0056
		3309 µatm	12	0.0033	0.0023	0.0043
	31°C	288 µatm	7	0.0054	0.0042	0.0066
		447 µatm	5	0.0042	0.0031	0.0053
		673 µatm	6	0.0041	0.0029	0.0051
		3285 µatm	1	0.0028	0.0014	0.0042

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Table S7. Bootstrapped modelled mean linear extension for each species in all $p\text{CO}_2$ and temperature treatments reported in mm day^{-1} . Sample sizes (N) and 95% confidence intervals (CI) are reported for each mean extension rate (figure 2).

Fixed effect	Estimate	SE	<i>t</i> -value
Intercept	7.86E-03	6.31E-04	12.5
Species (PAST)	-1.95E-03	9.14E-04	-2.14
<i>p</i> CO ₂ - current	3.62E-04	6.24E-04	0.058
<i>p</i> CO ₂ - end-of-century	7.32E-04	6.11E-04	1.20
<i>p</i> CO ₂ - extreme	-4.50E-04	6.01E-04	-0.075
Temperature (31°C)	-1.08E-03	4.12E-04	-2.62
Species (PAST) * <i>p</i> CO ₂ - current	-1.51E-03	9.35E-04	-1.62
Species (PAST) * <i>p</i> CO ₂ - end-of-century	-2.01E-03	9.38E-04	-2.15
Species (PAST) * <i>p</i> CO ₂ - extreme	-2.15E-03	9.60E-04	-2.24
Species (PAST) * Temperature (31°C)	5.01E-04	6.94E-04	0.072
Colony	1.68E-06		
Residual	3.46E-06		

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Table S8. Summary output of the linear mixed effects model used to determine the relationship between linear extension, *p*CO₂ and temperature for *S. siderea* and *P. astreoides* (PAST). Temperature and *p*CO₂ were treated as factors.

Species	Treatment	T0	T30	T60	T90	
<i>S. siderea</i>	28°C	311 µatm	10	10	10	10
		405 µatm	12	12	12	12
		701 µatm	11	11	11	11
		3309 µatm	12	12	12	12
	31°C	288 µatm	8	8	8	8
		447 µatm	11	11	11	11
		673 µatm	12	11	11	11
		3285 µatm	12	12	12	12
<i>P. strigosa</i>	28°C	311 µatm	16	16	15	15
		405 µatm	8	6	5	5
		701 µatm	14	14	14	14
		3309 µatm	16	16	16	16
	31°C	288 µatm	14	11	9	9
		447 µatm	13	11	6	6
		673 µatm	15	13	7	7
		3285 µatm	13	11	8	8
<i>P. astreoides</i>	28°C	311 µatm	11	11	11	11
		405 µatm	12	12	12	12
		701 µatm	12	11	10	10
		3309 µatm	12	12	12	12
	31°C	288 µatm	11	8	6	6
		447 µatm	9	8	8	8
		673 µatm	12	12	9	9
		3285 µatm	10	6	4	4
<i>U. tenuifolia</i>	28°C	311 µatm	12	11	11	11
		405 µatm	7	7	7	7
		701 µatm	8	5	4	4
		3309 µatm	8	6	5	5
	31°C	288 µatm	8	8	4	4
		447 µatm	1	0	0	0
		673 µatm	4	2	1	1
		3285 µatm	0	0	0	0

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Table S9. Sample size surviving for each species at each time point per treatment that was used for constructing survival curves (figure S6).

Species	Fixed Effect	Hazard rate	Hazard ratio	Hazard ratio SE	<i>z</i>	<i>P</i>
<i>S. siderea</i>	<i>p</i> CO ₂	-5.39E-06	1.00	0.00	0	1.00
	Temperature (31°C)	22.09	3.92E+09	0.00	Inf	0.00
	<i>p</i> CO ₂ * Temperature (31°C)	-5.87E-04	1.00	0.00	-Inf	0.00
<i>P. strigosa</i>	<i>p</i> CO ₂	-3.72E-03	1.00	0.00	-1.02	0.31
	Temperature (31°C)	0.58	1.79	1.51	0.39	0.70
	<i>p</i> CO ₂ * Temperature (31°C)	3.54E-03	1.00	0.00	0.97	0.33
<i>P. astreoides</i>	<i>p</i> CO ₂	3.12E-04	1.00	0.00	1.20	0.23
	Temperature (31°C)	0.47	1.60	1.17	0.40	0.69
	<i>p</i> CO ₂ * Temperature (31°C)	3.28E-03	1.00	0.00	1.52	0.13
<i>U. tenuifolia</i>	<i>p</i> CO ₂	3.41E-04	1.00	2.66E-04	1.28	0.20
	Temperature (31°C)	0.52	1.68	1.17	0.44	0.66
	<i>p</i> CO ₂ * Temperature (31°C)	3.26E-03	1.00	2.17E-03	1.51	0.13

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Table S10. Cox mixed effects proportional hazards analysis for survival of all four species. The ‘hazard rate’ represents the modelled risk of death, so that positive values represent increased risk. The ‘hazard ratio’ indicates the hazard in the treatment compared to the control.

Species	Fixed Effect	loglik	χ^2	DF	P
<i>S. siderea</i>	NULL	-4.48			
	pCO ₂	-4.34	0.27	1	0.6
	Temperature (31°C)	-3.61	1.47	1	0.23
	Reef environment	-2.94	1.35	1	0.225
	pCO ₂ * Temperature (31°C)	-3.61	0	1	1
<i>P. strigosa</i>	NULL	-131.95			
	pCO ₂	-121.63	20.64	1	5.53E-06 ***
	Temperature (31°C)	-113.32	16.61	1	4.60E-05 ***
	Reef environment	-113.29	0.07	1	0.79
	pCO ₂ * Temperature (31°C)	-111.80	3.06	1	0.08
<i>P. astreoides</i>	NULL	-74.67			
	pCO ₂	-73.25	2.84	1	0.09
	Temperature (31°C)	-66.06	14.38	1	1.49E-04 ***
	Reef environment	-64.55	3.02	1	0.08
	pCO ₂ * Temperature (31°C)	-65.41	1.3	1	0.25
<i>U. tenuifolia</i>	NULL	-59.12			
	pCO ₂	-58.36	1.5	1	0.22
	Temperature (31°C)	-54.28	8.18	1	4.24E-03 **
	Reef environment	-54.16	0.24	1	0.63
	pCO ₂ * Temperature (31°C)	-53.49	1.56	1	0.21

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Table S11. Statistical outcomes for coral survival analyses of all four species, using Cox mixed effects proportional hazards models.

Species	Reef Environment	Treatment	N	Mean Calcification (mg cm ² day ⁻¹)	Lower 95% CI	Upper 95% CI	
<i>S. siderea</i>	Offshore	28°C	311 µatm	6	1.045	0.803	1.284
			405 µatm	6	1.192	0.974	1.411
			701 µatm	6	1.023	0.808	1.252
			3309 µatm	6	0.217	-0.002	0.441
		31°C	288 µatm	4	1.031	0.789	1.275
			447 µatm	5	1.177	0.956	1.398
			673 µatm	6	1.008	0.789	1.228
			3285 µatm	6	0.202	-0.022	0.405
	Inshore	28°C	311 µatm	4	1.173	0.926	1.421
			405 µatm	6	1.320	1.094	1.539
			701 µatm	5	1.151	0.926	1.374
			3309 µatm	6	0.345	0.129	0.564
		31°C	288 µatm	4	1.159	0.905	1.407
			447 µatm	6	1.305	1.073	1.522
<i>P. strigosa</i>	Offshore	28°C	311 µatm	10	1.141	0.935	1.354
			405 µatm	3	0.444	0.146	0.778
			701 µatm	8	0.605	0.387	0.822
			3309 µatm	10	0.124	-0.078	0.322
		31°C	288 µatm	5	0.144	-0.088	0.386
			447 µatm	3	-0.553	-0.859	-0.233
			673 µatm	4	-0.392	-0.672	-0.141
			3285 µatm	5	-0.874	-1.136	-0.621
	Inshore	28°C	311 µatm	5	1.269	1.042	1.488
			405 µatm	2	0.572	0.265	0.904
			701 µatm	6	0.733	0.495	0.952
			3309 µatm	6	0.252	0.044	0.466
		31°C	288 µatm	4	0.272	0.036	0.527
			447 µatm	3	-0.425	-0.744	-0.107
		673 µatm	3	-0.264	-0.544	-0.006	
		3285 µatm	3	-0.746	-0.997	-0.482	

Species	Reef Environment	Treatment	N	Mean Calcification (mg cm ² day ⁻¹)	Lower 95% CI	Upper 95% CI
<i>P. astreoides</i>	Offshore	311 µatm	6	0.012	-0.226	0.255
		405 µatm	6	-0.053	-0.296	0.171
		701 µatm	5	-0.259	-0.496	-0.010
		3309 µatm	6	-0.749	-0.991	-0.508
		288 µatm	3	0.163	-0.119	0.435
		447 µatm	4	0.098	-0.146	0.353
		673 µatm	4	-0.108	-0.356	0.155
		3285 µatm	0	NA	NA	NA
	Inshore	311 µatm	4	0.140	-0.102	0.385
		405 µatm	6	0.075	-0.180	0.301
		701 µatm	5	-0.131	-0.379	0.121
		3309 µatm	6	-0.621	-0.853	-0.399
		288 µatm	4	0.291	0.015	0.574
		447 µatm	6	0.226	-0.020	0.485
<i>U. tenuifolia</i>	Offshore	311 µatm	3	0.060	-0.233	0.361
		405 µatm	2	0.152	-0.233	0.539
		701 µatm	1	-0.062	-0.513	0.380
		3309 µatm	1	-0.337	-0.773	0.099
		288 µatm	0	NA	NA	NA
		447 µatm	0	NA	NA	NA
		673 µatm	0	NA	NA	NA
		3285 µatm	0	NA	NA	NA
	Inshore	311 µatm	8	0.188	-0.099	0.479
		405 µatm	5	0.280	-0.071	0.650
		701 µatm	3	0.066	-0.369	0.515
		3309 µatm	4	-0.209	-0.621	0.210
		288 µatm	4	0.150	-0.284	0.597
		447 µatm	0	NA	NA	NA
31°C	673 µatm	1	0.028	-0.536	0.622	
	3285 µatm	0	NA	NA	NA	

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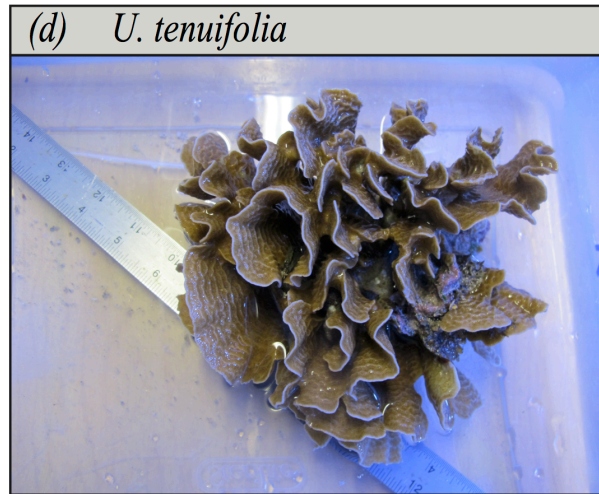
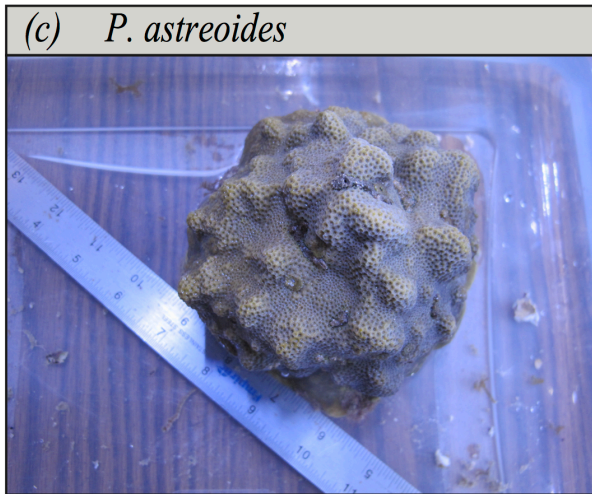
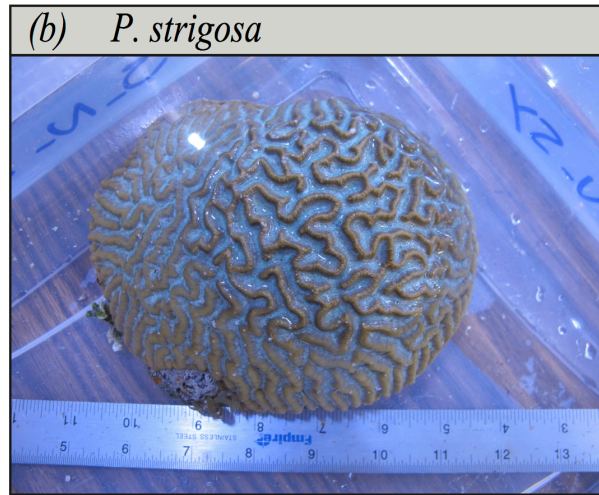
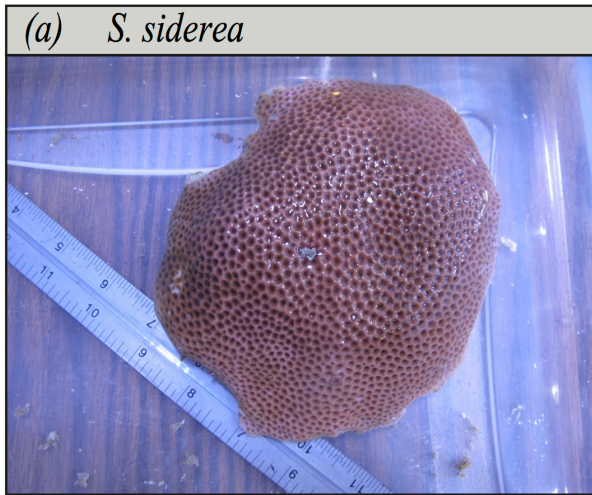
277 **Table S12.** Bootstrapped modelled mean calcification rate for each species by reef environment
278 in all $p\text{CO}_2$ and temperature treatments reported in $\text{mg cm}^{-2} \text{day}^{-1}$. Sample sizes (N) and 95%
279 confidence intervals (CI) are reported for each mean calcification rate (figure S11).

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Species	Reef Environment	Treatment	N	Mean LE (mm day ⁻¹)	Lower 95% CI	Upper 95% CI	
<i>S. siderea</i>	Offshore	28°C	311 µatm	6	0.0076	0.0066	0.0087
			405 µatm	6	0.0078	0.0069	0.0088
			701 µatm	6	0.0082	0.0072	0.0091
			3309 µatm	6	0.0071	0.0062	0.0080
		31°C	288 µatm	4	0.0065	0.0054	0.0076
			447 µatm	4	0.0067	0.0057	0.0077
			673 µatm	6	0.0071	0.0061	0.0080
			3285 µatm	6	0.0059	0.0050	0.0069
	Inshore	28°C	311 µatm	3	0.0084	0.0073	0.0096
			405 µatm	5	0.0086	0.0077	0.0096
			701 µatm	5	0.0090	0.0080	0.0100
			3309 µatm	6	0.0079	0.0069	0.0088
		31°C	288 µatm	4	0.0073	0.0063	0.0084
			447 µatm	6	0.0075	0.0065	0.0085
<i>P. astreoides</i>	Offshore	28°C	311 µatm	5	0.0055	0.0043	0.0066
			405 µatm	3	0.0043	0.0031	0.0055
			701 µatm	5	0.0042	0.0031	0.0053
			3309 µatm	6	0.0029	0.0018	0.0040
		31°C	288 µatm	2	0.0049	0.0037	0.0062
			447 µatm	3	0.0038	0.0026	0.0050
			673 µatm	3	0.0037	0.0025	0.0048
			3285 µatm	0	NA	NA	NA
	Inshore	28°C	311 µatm	4	0.0063	0.0052	0.0074
			405 µatm	6	0.0051	0.0040	0.0062
			701 µatm	4	0.0050	0.0039	0.0061
			3309 µatm	6	0.0037	0.0027	0.0048
		31°C	288 µatm	3	0.0057	0.0046	0.0070
			447 µatm	4	0.0045	0.0034	0.0057
		673 µatm	3	0.0045	0.0033	0.0056	
		3285 µatm	1	0.0032	0.0017	0.0046	

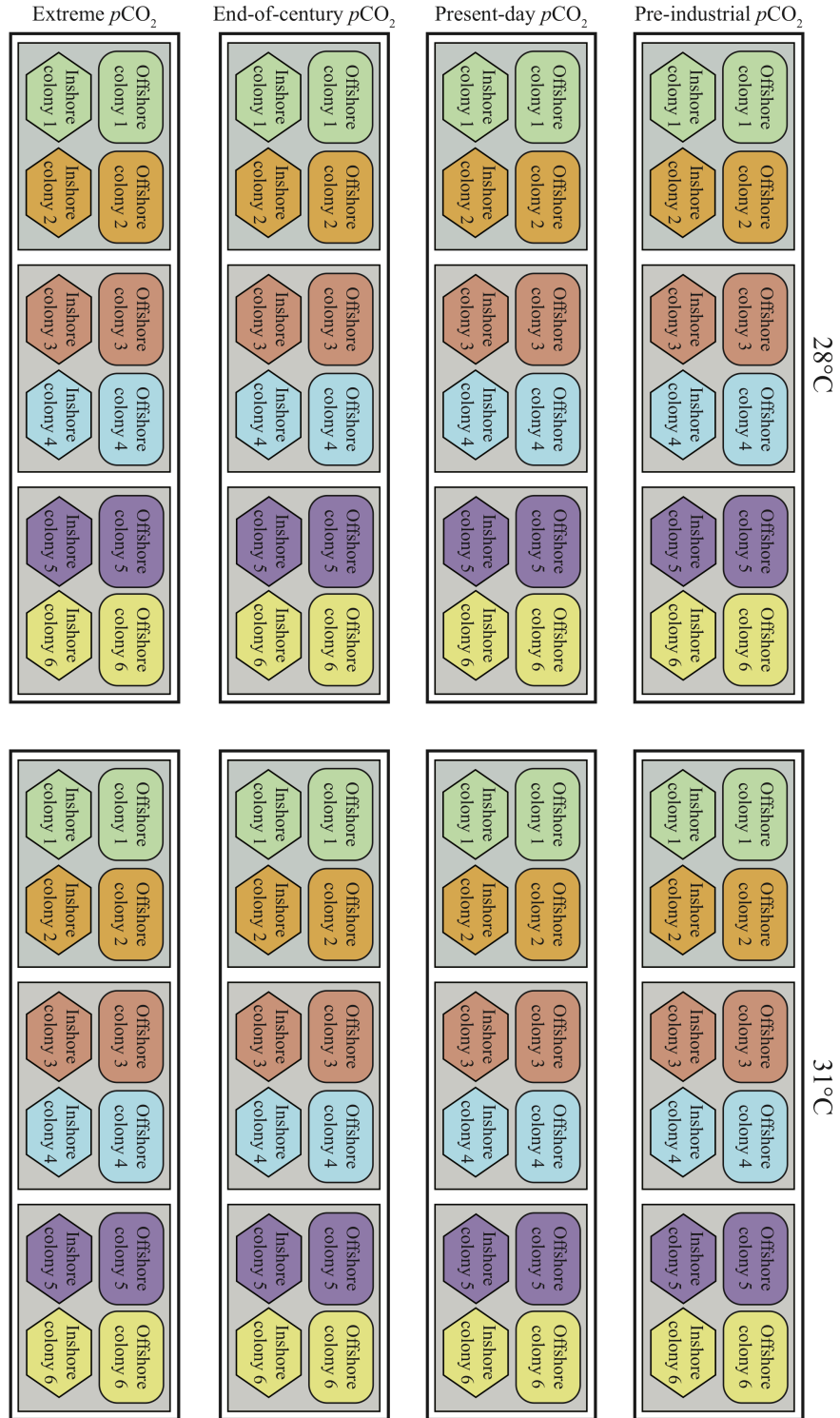
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Table S13. Bootstrapped modelled mean linear extension for each species by reef environment in all $p\text{CO}_2$ and temperature treatments reported in mm day^{-1} . Sample sizes (N) and 95% confidence intervals (CI) are reporter for each mean extension rate (figure S12).

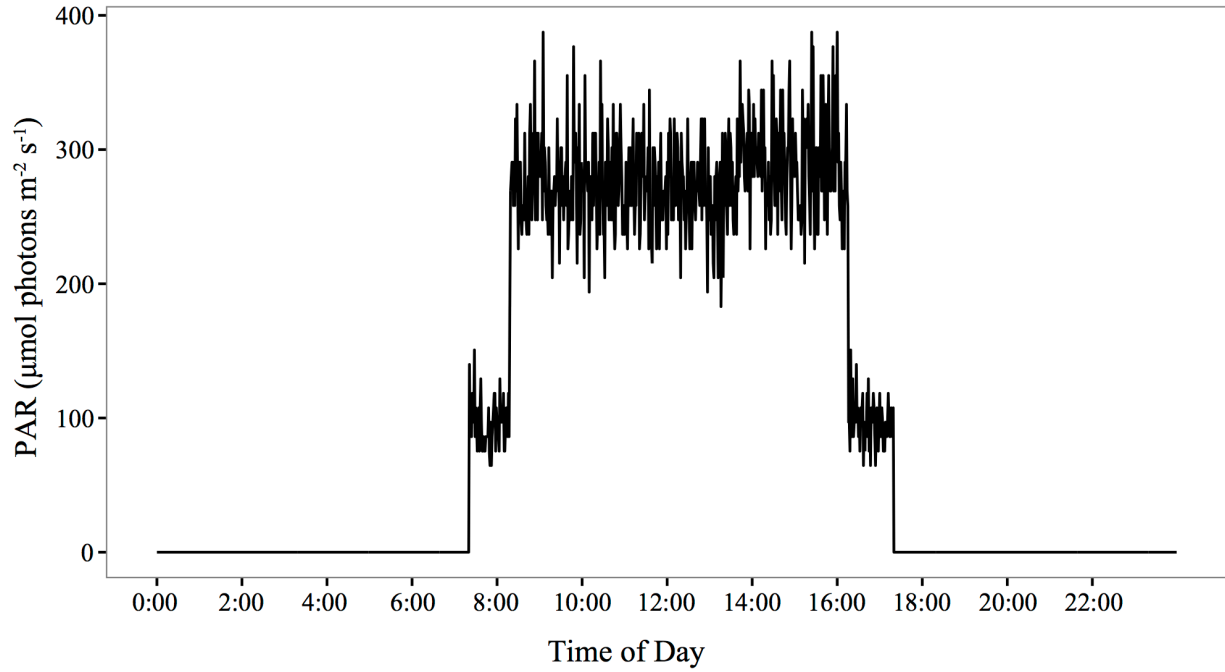


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Figure S1. Representative specimens of the collected colonies of (a) *S. siderea*, (b) *P. strigosa*, (c) *P. astreoides*, and (d) *U. tenuifolia* from the Belize Barrier Reef System prior to sectioning.

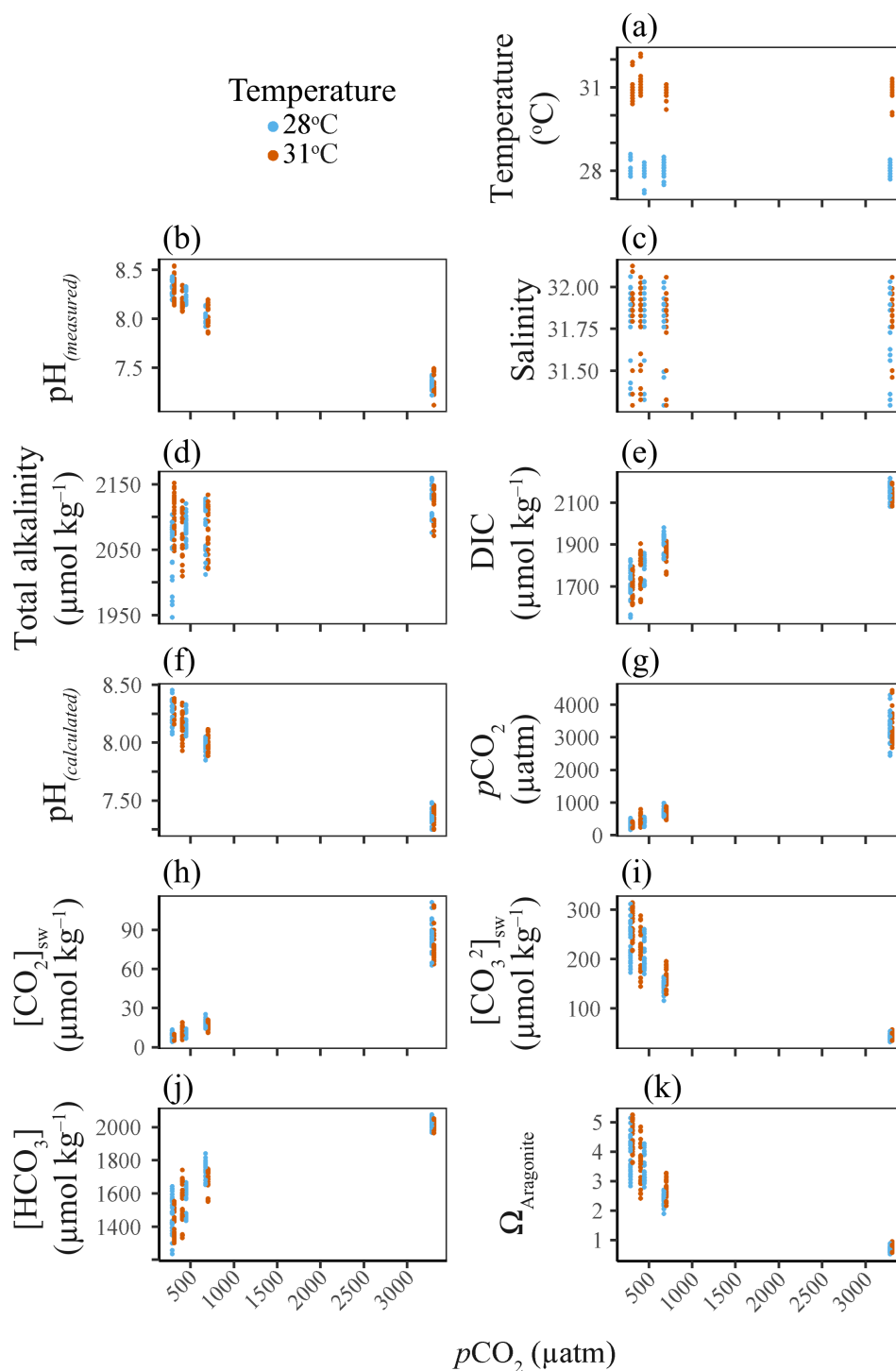


290 **Figure S2.** Diagram showing allocation of coral fragments for a single species throughout
 291 experimental tank array. Colour represent a different colony and shape represents reef
 292 environment. Four colonies (two from each reef environment) are reared within each tank (grey
 293 box), with three tanks comprising a treatment (white box). This is repeated for each $p\text{CO}_2$
 294 treatment at both temperatures. This same experimental design was used for all four species.

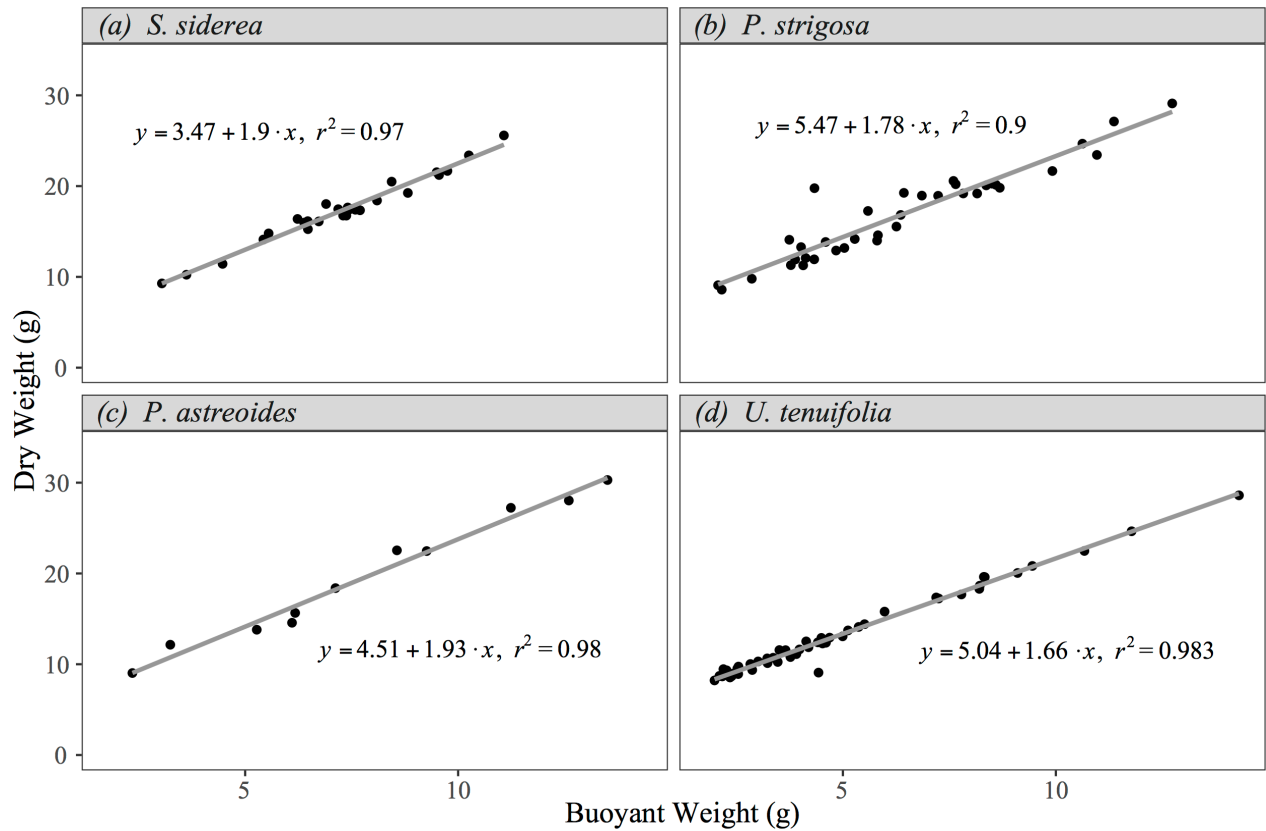


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Figure S3. Ten hour light cycle for all 24 experimental treatment tanks reported in PAR (photosynthetically active radiation; $\mu\text{mol photons m}^{-2} \text{s}^{-1}$).

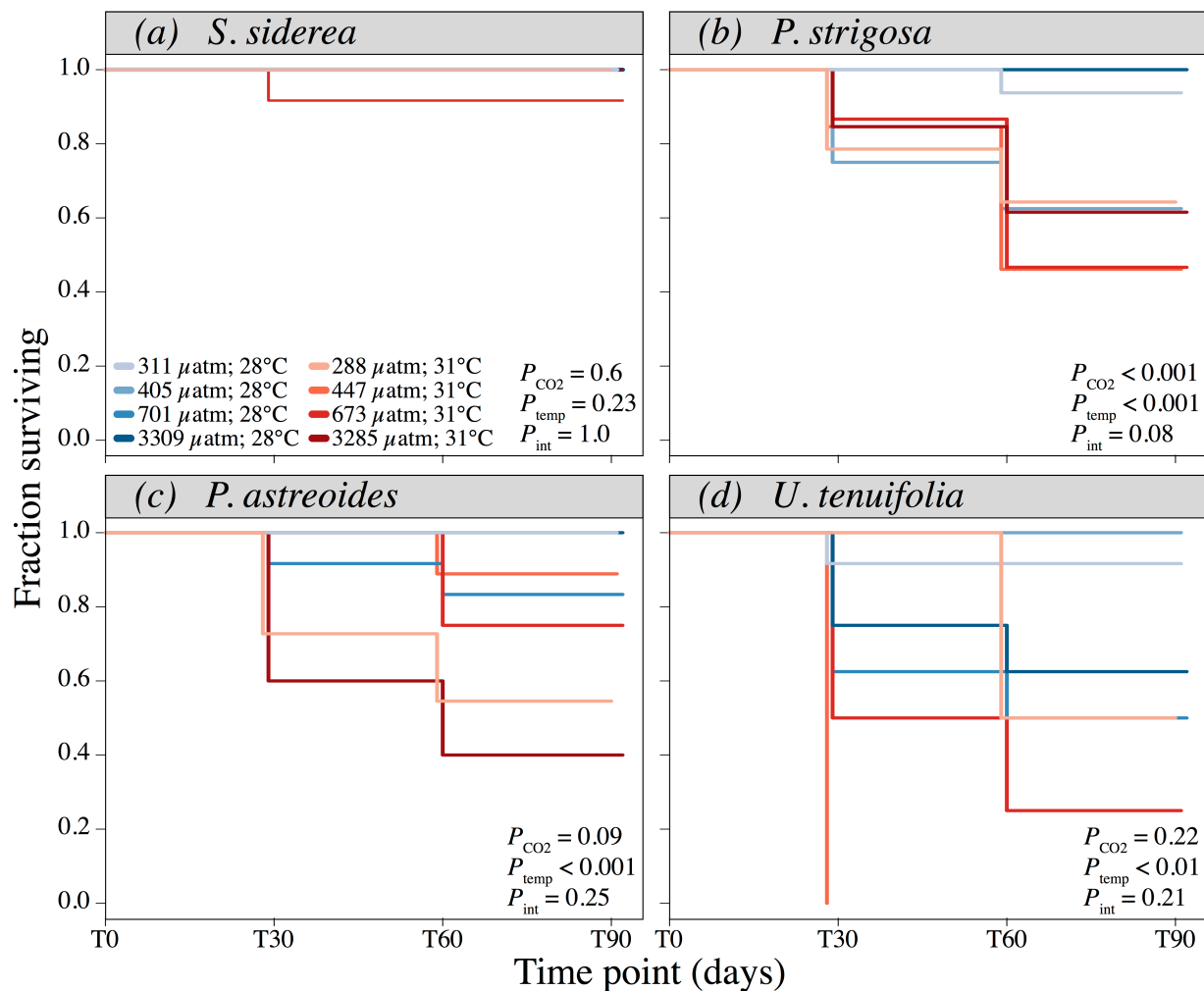


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 300 **Figure S4.** Calculated and measured parameters for all 24 experimental tanks over the 93-day
 301 experimental interval: (a) measured temperature; (b) measured pH; (c) measured salinity; (d)
 302 measured total alkalinity; (e) measured dissolved inorganic carbon; (f) calculated pH; (g)
 303 calculated $p\text{CO}_2$ of the mixed gases in equilibrium with the experimental seawaters; (h)
 304 calculated dissolved carbon dioxide; (i) calculated carbonate ion concentration; (j) calculated
 305 bicarbonate ion concentration; and (k) calculated aragonite saturation state.

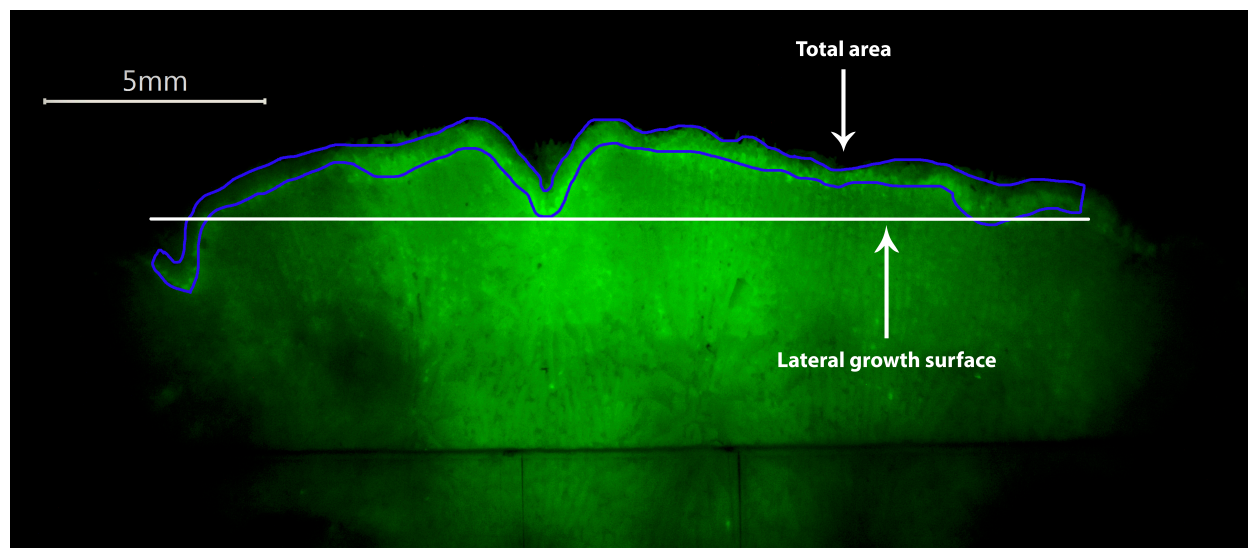


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Figure S5. Linear relationship between buoyant weight (mg) and dry weight (mg) for (a) *S. siderea*, (b) *P. strigosa*, (c) *P. astreoides*, and (d) *U. tenuifolia*.

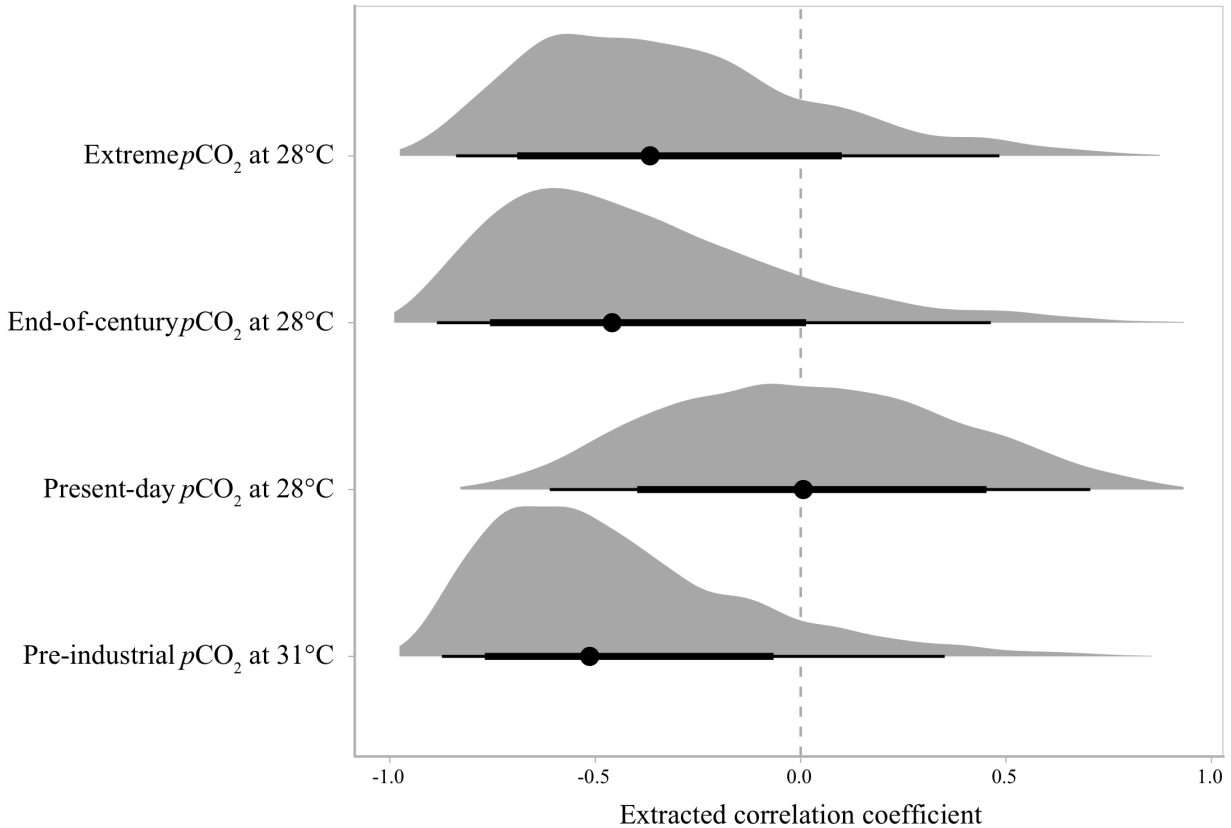


311
 312 **Figure S6.** Fraction of fragments surviving from the start of the experiment for *S. siderea* (a), *P.*
 313 *strigosa* (b), *P. astreoides* (c), and *U. tenuifolia* (d). Blue represents 28°C treatments and red
 314 represents 31°C treatments. Colour intensity corresponds to $p\text{CO}_2$ level, with the lowest intensity
 315 representing pre-industrial $p\text{CO}_2$ and the highest intensity representing an extreme $p\text{CO}_2$
 316 condition.
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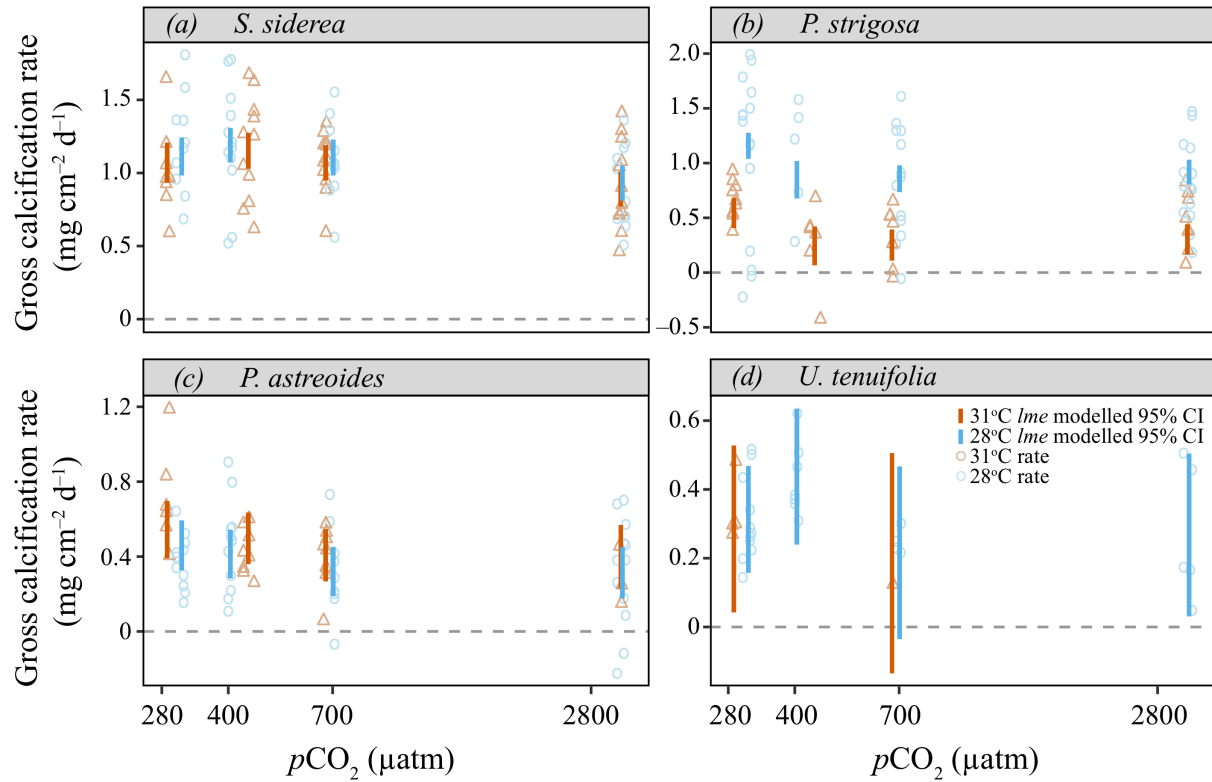


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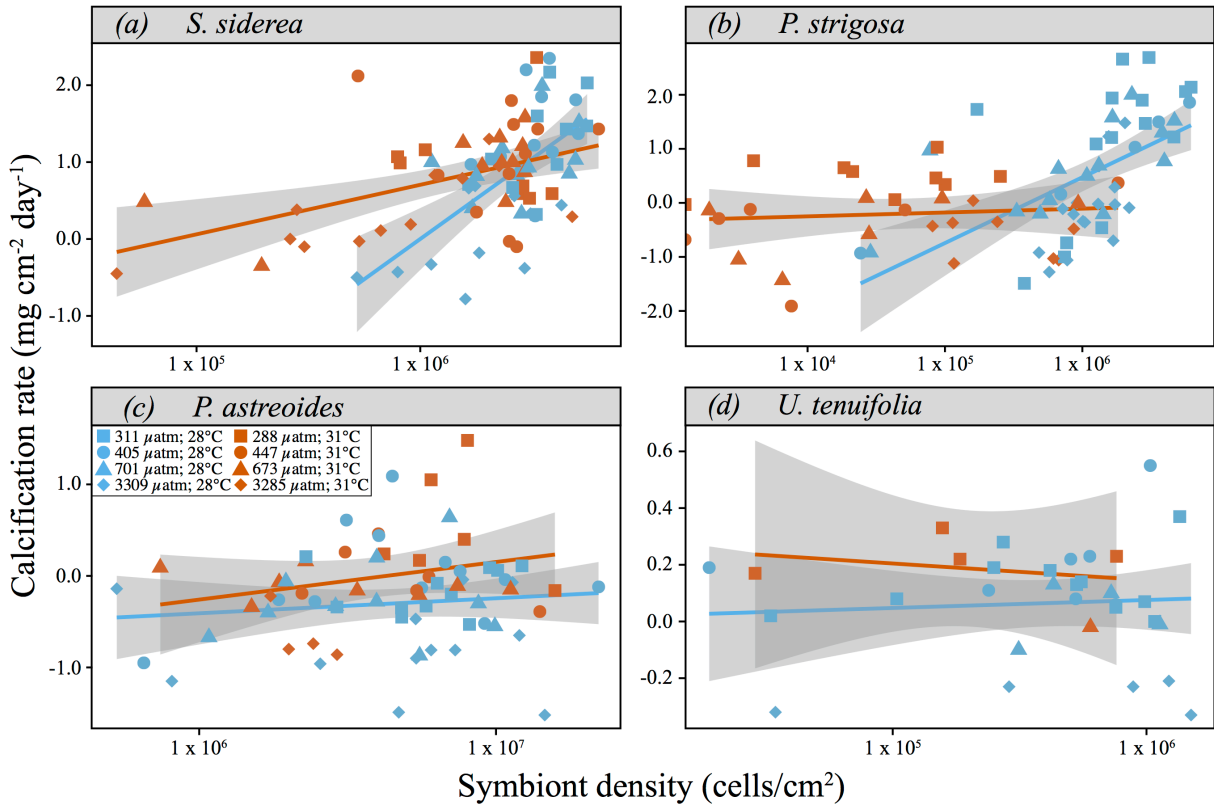
Figure S7. Example of linear extension measurement for *S. siderea* sample, indicating total growth area and lateral growth surface determination using image analysis software (IMAGE J). Linear extension was calculated by dividing total growth area by lateral growth surface



324
 325 **Figure S8.** Density plot of the extracted correlation coefficients describing the correlation
 326 between the Bayesian random effects of colony on calcification rate under the control treatment
 327 (pre-industrial $p\text{CO}_2$ at 28°C) versus each stress treatment. The black circle represents the
 328 estimated mean, the thick black bar is the 75% credible interval, the thin black bar is the 95%
 329 credible interval, and the grey area represents the range of the Bayesian model output of the
 330 extracted correlation coefficients. Intervals that do not overlap zero denote significant effects of
 331 colony basal calcification rate on colony-level calcification response to $p\text{CO}_2$ or thermal stress.
 332

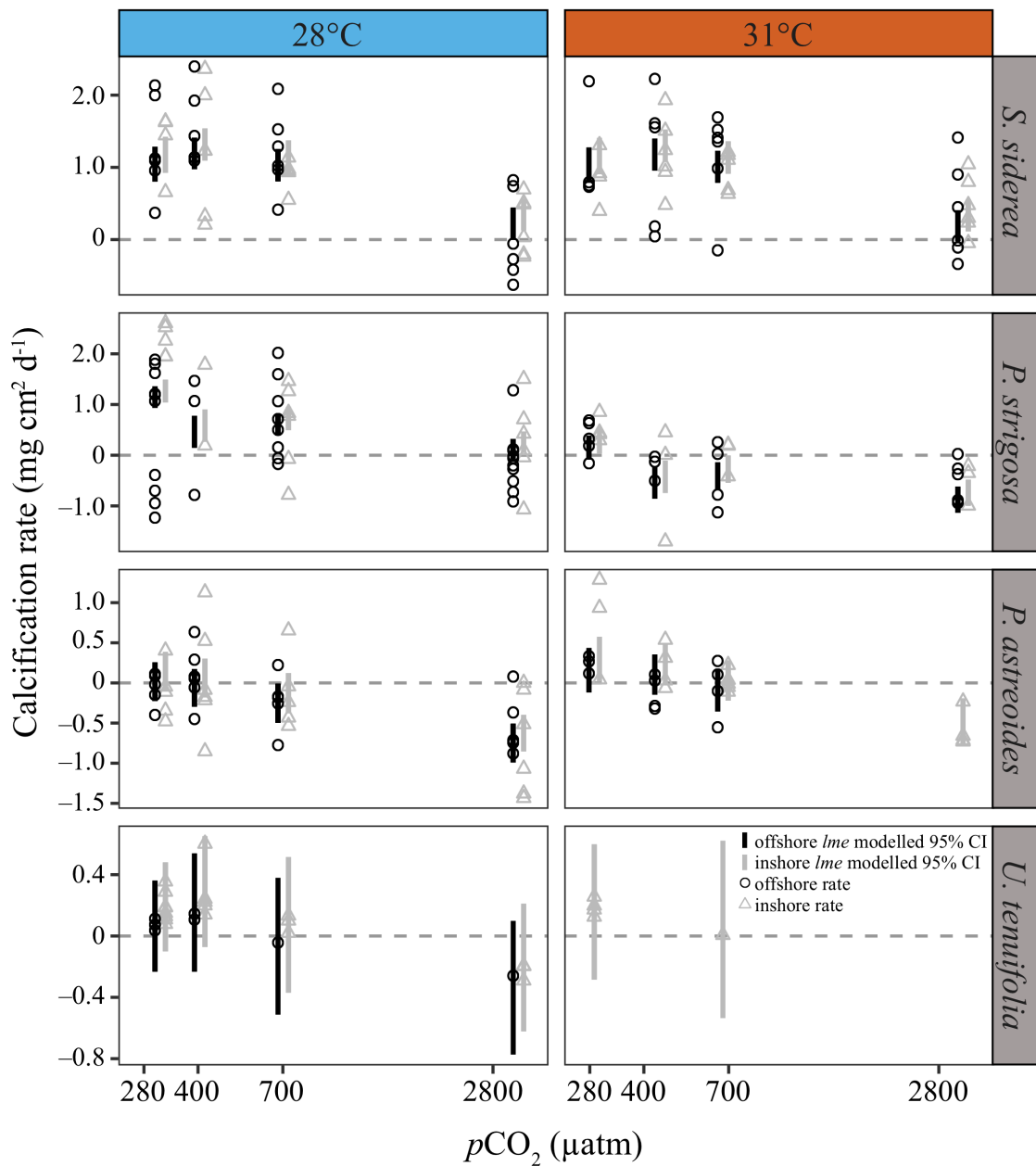


333
 334 **Figure S9.** Modelled 95% confidence intervals of gross calcification rate for the 90-day
 335 experimental period in $\text{mg cm}^{-2} \text{day}^{-1}$ for (a) *S. siderea*, (b) *P. strigosa*, (c) *P. astreoides*, and (d)
 336 *U. tenuifolia*. Blue bars represent 28°C treatment 95% confidence intervals and orange bars
 337 represent 31°C treatment 95% confidence intervals, with $p\text{CO}_2$ along the x-axis (μatm). Blue
 338 open circles represent gross calcification rates for individual fragments in the 28°C treatment,
 339 and orange open circles represent gross calcification rates for individual fragments in the 31°C
 340 treatment.
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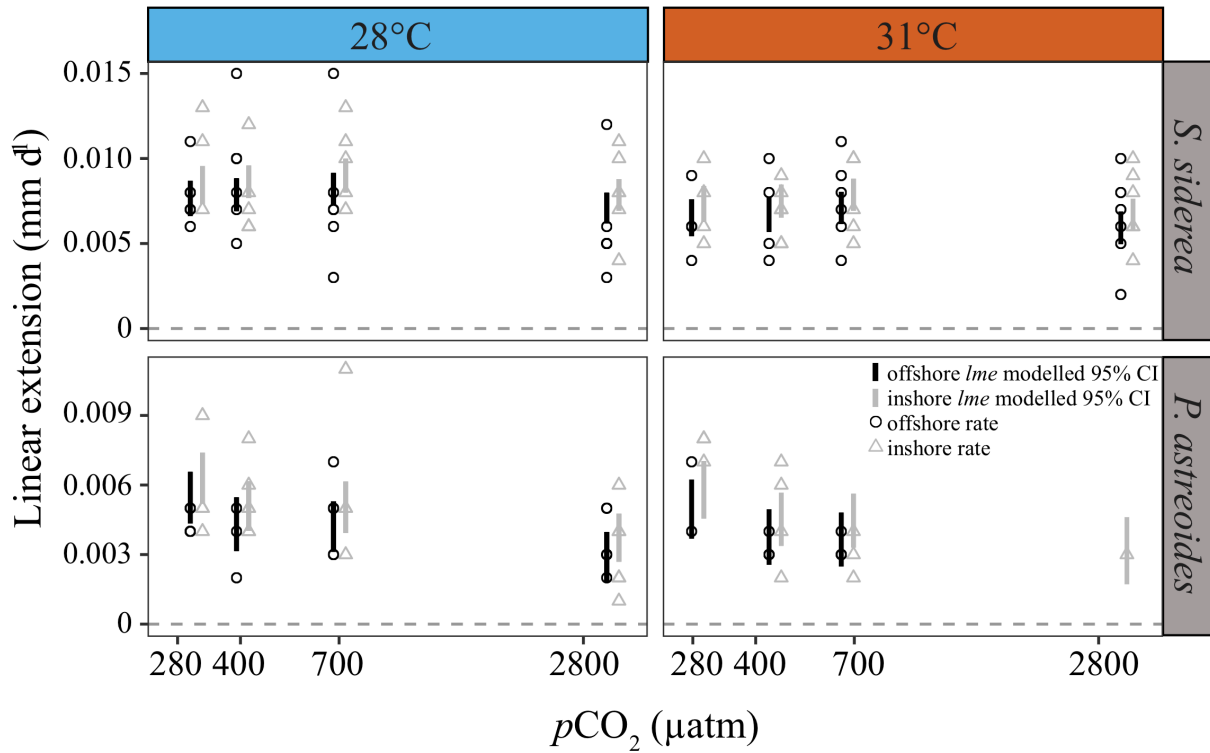


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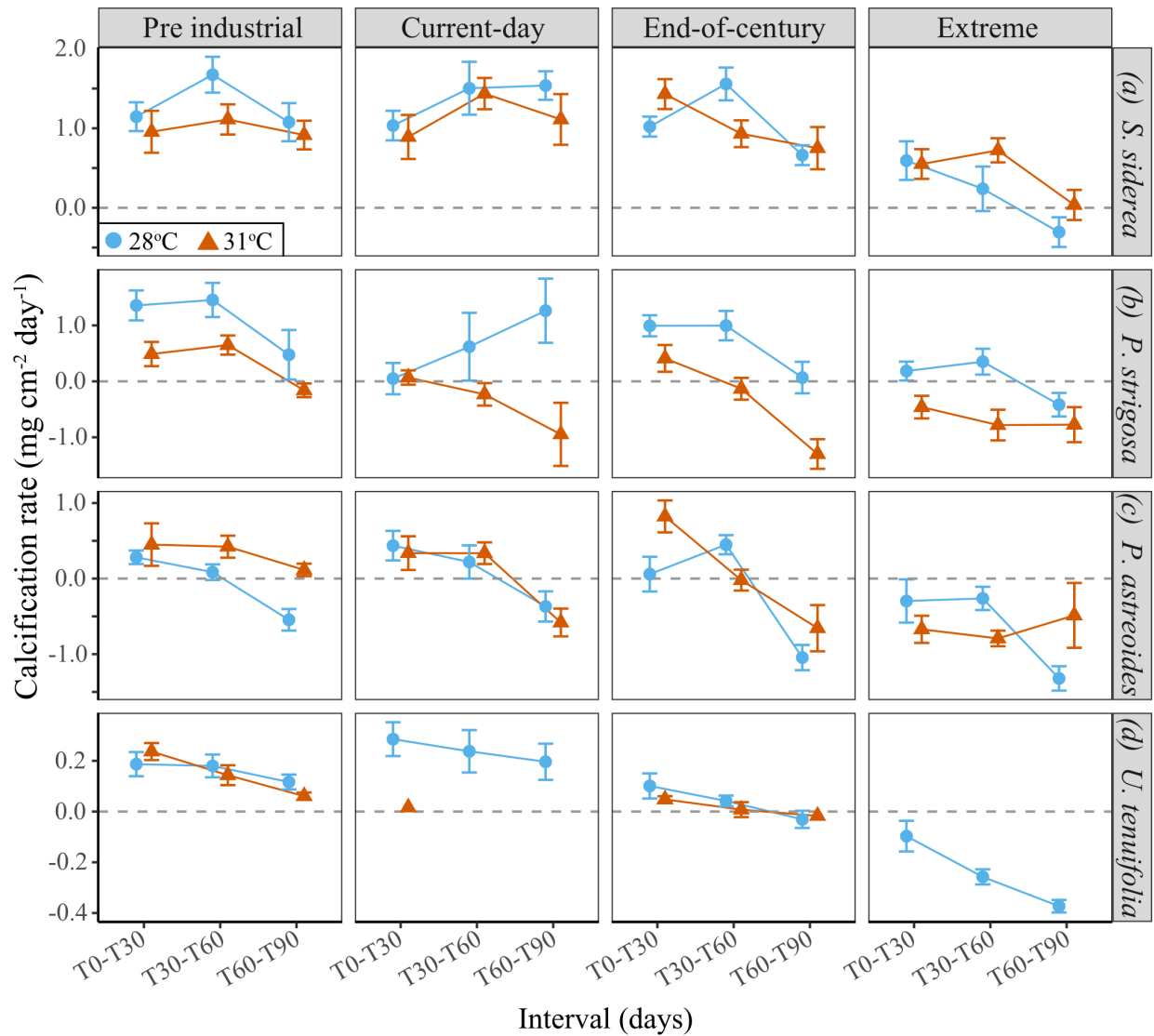
Figure S10. Relationship between calcification rate and symbiont density (cell counts cm⁻²) for (a) *S. siderea*, (b) *P. strigosa*, (c) *P. astreoides*, and (d) *U. tenuifolia*. Shape represents pCO₂ treatments and colour represents temperature treatments. The line denotes a simple linear regression with standard error denoted by grey shading.



348
 349 **Figure S11.** Modelled mean calcification rate for the 93-day experimental period in mg cm⁻²
 350 day⁻¹ separated by reef environment for (a) *S. siderea*, (b) *P. strigosa*, (c) *P. astreoides*, and (d) *U.*
 351 *tenuifolia*. Grey triangles denote inshore corals and black circles denote offshore corals. Left
 352 panel demonstrates mean calcification rate at 28°C and the right panel shows calcification at
 353 31°C, with pCO₂ along the x-axis (µatm) on a log scale. Error bars denote 95% confidence
 354 intervals of each estimated mean.
 355



356
 357 **Figure S12.** Modelled mean linear extension rate for the 93-day experimental period in mm cm⁻²
 358 day⁻¹ separated by reef environment for (a) *S. sideraea* and (b) *P. astreoides*. Grey triangles denote
 359 inshore corals and black circles denote offshore corals. Left panel demonstrates mean
 360 calcification rate at 28°C and the right panel shows calcification at 31°C, with pCO₂ along the x-
 361 axis (μatm) on a log scale. Error bars denote 95% confidence intervals of each estimated mean.
 362



363
 364 **Figure S13.** Mean calcification rate (mg cm⁻² day⁻¹) at each 30-day experimental interval at all
 365 pCO₂ treatments for (a) *S. siderea*, (b) *P. strigosa*, (c) *P. astreoides*, and (d) *U. tenuifolia*. Blue
 366 circles represent 28°C treatments and orange triangles represent 31°C treatments, with time
 367 interval along the x-axis. Error bars denote standard error of each mean.

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