# **Supplementary Online Content**

Tosto G, Vardarajan B, Sariya S, et al. Association of variants in *PINX1* and *TREM2* with late-onset Alzheimer disease per a gene-based, transethnic meta-analysis. *JAMA Neurol*. Published online May 6, 2019. doi:10.1001/jamaneurol.2019.1066

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This supplementary material has been provided by the authors to give readers additional information about their work.

#### eMethod 1. Whole-exome sequencing quality control.

Quality control (QC) procedures excluded monomorphic variants, VQSR non-"PASS" variants with call rates <80%, and variants with low average depth of data (DP) and genotype quality (GQ) (8<DP>500 reads and GQ <20, corresponding to a > 1% likelihood of being an incorrect genotype call)<sup>1</sup>. "DP" values represented the number of reads passing QC used to calculate the genotype at a specific site in a specific sample, with higher values for DP generally leading to more accurate genotype calls. "GQ" is a Phred-scaled value representing the confidence that the called genotype is the true genotype. Again, higher values reflected more accurate genotype calls. These thresholds were chosen according to studies of concordance between sequencing experiments and genotyping arrays in order to achieve a 99% genotype likelihood. Because these simulations were focused on SNV only, for INDELS we additionally applied GATK-reccomanded hard filter (QualByDepth (QD) >2.0; FisherStrand (FS) <200.0; ReadPosRankSumTest (ReadPosRankSum) > 20.0) (https://software.broadinstitute.org/gatk/documentation/article?id=2806). We handled multi-allelic site by splitting the alternative alleles in multiple biallelic sites using *bcftools* utility<sup>2</sup>. After this step, we normalized variants by applying parsimonial representation (i.e. coding the variant in as few nucleotides as possible without reducing the length of any allele to 0) and left-alignment (i.e. shifting the start position of that variant to the left till it is no longer possible)<sup>3</sup>. Variants showing strong departure from Hardy Weinberg equilibrium (HWE, p<1-e07) in controls were also filtered out. Principal components (PCs) were estimated using WES data, and filtering out single nucleotide variant (SNV) with minor allele frequency (MAF) <5%, call rate <95%. We used the KING software to detect duplicates and cryptic relatedness, and output the first 10 PCs, separately for each ethnic group. We then excluded outliers that deviated more than six standard deviation from the mean.

## eMethod 2. Replication cohorts' description.

- Alzheimer Disease Sequencing Project (ADSP). Individuals were aged 60 years or older and met NINCDS-ADRDA criteria <sup>4</sup> for possible, probable or definite AD based on clinical assessment, or had presence of LOAD (moderate or high likelihood) upon neuropathology examination. Healthy controls were similar in age and either judged to be cognitively normal or did not meet pathological criteria for LOAD following brain autopsy <sup>5</sup>. We did not include the Caribbean Hispanic part of the ADSP dataset because they overlapped with those already present in our WHICAP WES data.
- 2. The ROS/MAP study. The Religious Orders Study (ROS) is a longitudinal cohort study of aging and Alzheimer's disease from Rush University recruiting Individuals from more than 40 groups of religious orders across the US. Subjects were included at baseline if dementia was not present. The Memory and Aging Project (MAP) is a longitudinal, epidemiologic clinical-pathologic cohort study based at Rush University of aging and risk of LOAD that began in 1997. This study was designed to complement the ROS study by enrolling individuals with a wider range of life experiences and socioeconomic status. The study enrolls older individuals without any signs of dementia, primarily recruiting from continuous care retirement communities throughout northeastern Illinois, USA. LOAD status was determined by an actuarial decision tree that incorporated a computer algorithm and clinical judgment made in series by a neuropsychologist and a clinician who ultimately reviewed all cases. Details of the ROS and MAP study has been described in detail elsewhere <sup>6</sup>. Participants were deemed "non-demented" (healthy or MCI) or LOAD. Other dementia types were excluded from analyses.
- 3. ADSP Family Study. Whole exome data from 67 families of Caribbean Hispanic ancestry (N=358 participants) were selected from the ADSP family dataset (Table 1). No individuals with known early-onset disease mutations (*APP*, *PSEN1*, *PSEN2*, *GRN*, or *MAPT*) were included. All probands were part of families with three or more affected individuals recruited as part of the Estudio Familiar de Influencia Genetica en Alzheimer (EFIGA) study. Detailed description of this cohort has been published elsewhere <sup>7</sup>.
- 4. We used single-marker summary statistics from the International Genomics of Alzheimer's Project (IGAP), a large two-stage genome-wide association study (GWAS) on individuals of European ancestry (<u>http://web.pasteur-lille.fr/en/recherche/u744/igap/igap\_download.php N=54,162</u>)<sup>8</sup>. In stage 1, IGAP employed 7,055,881 genotyped and imputed single nucleotide polymorphisms to meta-analyses four previously-published GWAS datasets (The European Alzheimer's disease Initiative (EADI); the Alzheimer Disease Genetics Consortium (ADGC); The Cohorts for Heart and Aging Research in Genomic Epidemiology consortium (CHARGE) The Genetic and Environmental Risk in AD consortium (GERAD)). In stage 2, top hits from Stage 1 were genotyped and tested for association in an independent set of 8,572 LOAD cases and 11,312 controls and then again meta-analyzed. We used stage 1 summary results in order to include all SNPs available within each gene.

## eMethod 3. Expression data description and analyses methods.

- <u>Description</u>. The Myers and colleagues <sup>9</sup> neocortical transcriptome data. Briefly, a set of 188 individuals without dementia and 176 autopsy confirmed LOAD. Data were downloaded from the NCBI GEO archive (ID: GSE15222). All data were generated using the Illumina HumanRef-8 expression BeadChip (GPL2700) v2 Rev0. <u>Methods</u>. Data were analyzed in R using residual corrected profiles for each individual and for each transcript. Expression measures were corrected for sex, *APOE* status, age at death, cortical region, day of expression hybridization, study cohort, post-mortem interval and transcript detection rate. One-way ANOVA compared expression profile between affected brain sand normal control brains for genes prioritized in genetic analyses.
- 2. <u>Description</u>. ROS/MAP Next-generation RNA sequencing (RNA-seq) data. Briefly, RNA was purified from frozen dorsolateral prefrontal cortex tissue of ROS-MAP participants with the miRNeasy Mini Kit and RNase-Free DNase Set (Qiagen, Germantown, MD). RNA concentration and quality were measured with a Nanodrop (Thermo Fisher Scientific, Wilmington, DE) and Bioanalyzer (Agilent Technologies, Santa Clara, CA). A RIN score of >5 was required for library construction, which was assembled using the strand-specific dUTP method. Sequencing was performed with Illumina HiSeq with 101 base pair paired-end reads and a goal coverage of >85 million paired-end reads as previously described <sup>10</sup>. After QC, 508 subjects were available to analyze. Fragments per kb of transcript per million fragments mapped (FPKM) were quantile-normalized, correcting for batch effect with Combat <sup>11</sup>. <u>Methods</u>. Linear regressions were applied with neuropathological measure as the dependent variables, Combat-adjusted FPKM values as the independent variable, and technical factors as covariates (RNA integrity score, log2[total aligned reads], postmortem interval, and number of ribosomal bases) as detailed elsewhere <sup>12</sup>.
- 3. <u>Description</u>. The Narayanan and colleagues dataset<sup>12</sup> comprised DLPFC (BA9) brain tissues of 624 individual (AD patients, HD patients and non-demented controls samples), obtained from Harvard Brain tissue resource center (HBTRC). The HBTRC samples were primarily of Caucasian ancestry, as only eight non-Caucasian outliers were identified, and therefore excluded for further analysis. Post-mortem interval (PMI) was 17.8+8.3 hours (mean ± standard deviation), sample pH was 6.4±0.3 and RNA integrity number (RIN) was 6.8±0.8 for the average sample in the overall cohort. Tissues were profiled on a custom-made Agilent 44K array (GPL4372). 310 LOAD cases and 157 controls were included in the analyses.

## eFigure 1. Flowchart of the study for the MODERATE-HIGH VEP annotation model.



**eFigure 2**. Quantile-quantile plot for the moderate-high SKAT-O model in the WHICAP metaanalysis.



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**eFigure 3**. Quantile-quantile plot for the moderate-high SKAT-O model in the ROS/MAP (unadjusted and adjusted by minimum achievable p-values – MAP).



**eFigure 4.** Quantile-quantile plot for the moderate-high SKAT-O model in the ADSP (unadjusted and adjusted by minimum achievable p-values – MAP).



Expected Quantiles (-log10 P-values)

**eFigure 5**. Correlation between WHICAP meta-analysis SKAT-O p-values according to annotation models, i.e. VEP Moderate-high and CADD20 (Spearman coefficient was =0.49, p-values <0.001).



MODERATE-HIGH CADD20 p-values correlation

**eFigure 6**. Principal components (PC) scatterplot matrix for each ethnic group of the WHICAP dataset. We included the PCs #1,#2,#3 in each statistical model.



WHICAP Caribbean Hispanics PCs Scatterplot Matrix



**eFigure 7**. Diffierential expression boxplot between LOAD cases and normal controls in Myers et al. dataset for *PINX1*. For LOAD brains, mean standardized pinx1 expression= 0.087; for control brains, mean standardized pinx1 expression= -0.085.



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**eTable 1.** *PINX1* variants included in the VEP MODERATE-HIGH analyses. Table shows base pair (BP, minor allele frequency (MAF), missing rate, major and minor allele, p-value for single-marker analysis (in bold significant p-values) along with effect size (beta) and standard error (s.e.), CADD score. LoF variants are highlighted in red.

SNP	BP	MAF	<b>Missing Rate</b>	MajorAllele	MinorAllele	#SNP in controls	#SNP in cases	beta	s.e.	single-marker pvalue	CADD score	snp138
8:10622923:TTTC:T	10622923	0.006896552	0.000255363	TTTC	T	35	12	-0.788816595	0.306351562	0.010027668		rs201660183
8:10622930:T:C	10622930	0.004473415	0.00102145	Т	С	17	12	0.2291413	0.401678461	0.568366073	17.38	rs61746595
8:10623000:T:TC	10623000	0.000255428	0.000255363	Т	TC	0	2	1.918055022	1.43784195	0.182209815		
8:10623027:G:C	10623027	0.000127845	0.001276813	G	С	0	1	2.100124255	2.005695283	0.29506249	0.852	
8:10623094:C:G	10623094	0.000511117	0.000766088	С	G	2	2	0.204283648	1.01167179	0.83997395	18.77	rs369418829
8:10623104:C:T	10623104	0.000127779	0.000766088	С	Т	1	0	-1.678203154	2.045461306	0.411958389	0.047	rs150489215
8:10623134:G:C	10623134	0.000383436	0.00102145	G	С	3	0	-1.721853739	1.183360734	0.14565432	0.002	rs377260866
8:10623135:C:T	10623135	0.000255493	0.000510725	С	Т	0	1	1.964770358	2.006240402	0.327417212	5.588	rs202202690
8:10623137:CTCT:C	10623137	0.000127714	0.000255363	СТСТ	С	0	1	7.080972646	2.872535284	0.01369902		
8:10623141:T:A	10623141	0.000127714	0.000255363	Т	А	0	1	7.080972646	2.872535284	0.01369902	37	rs201016513
8:10623203:GC:G	10623203	0.000127714	0.000255363	GC	G	1	0	-1.879564594	2.020813157	0.352317703		
8:10623240:T:C	10623240	0.038825032	0.000255363	Т	С	192	96	-0.315669347	0.131495673	0.016367934	0.001	rs17711777
8:10623254:C:A	10623254	0.01073071	0.000510725	С	А	52	27	-0.380986489	0.235443248	0.105626617	24.2	rs17855458
8:10623280:C:G	10623280	0.009200102	0.000766088	С	G	33	34	0.294594541	0.257680527	0.252932749	0.007	rs35530857
8:10623301:C:A	10623301	0	0.000510725	С	А	0	0	NA	NA	NA	10.82	rs202176931
8:10623309:C:T	10623309	0.000127747	0.000510725	С	Т	1	0	-1.373771675	2.262806759	0.543778189	5.326	
8:10623344:C:T	10623344	0.000127714	0.000255363	С	Т	0	1	1.860406497	2.0077784	0.354134506	23.8	rs199675528
8:10623361:C:G	10623361	0.00038324	0.000510725	С	G	2	0	-1.743443751	1.513589574	0.249378482	21.8	rs375581032
8:10623379:G:C	10623379	0.000127714	0.000255363	G	С	0	1	2.218315869	2.014700316	0.270868398	4.485	
8:10623383:G:T	10623383	0.00038324	0.000510725	G	T	3	0	-1.519737169	1.229890354	0.21658169	13.62	rs202049952
8:10623385:C:T	10623385	0.000638733	0.000510725	С	T	2	2	0.359249756	1.034425968	0.728370585	12.68	rs374083715
8:10623386:G:A	10623386	0.000127714	0.000255363	G	А	1	0	-1.826653777	2.013811482	0.364373511	14.1	rs377311809
8:10623393:C:T	10623393	0.000127714	0.000255363	С	Т	1	0	-1.373771675	2.262806759	0.543778189	4.312	rs369880397
8:10623396:TCTC:T	10623396	0.000510856	0.000255363	TCTC	Т	3	1	0.143017387	1.219164593	0.906616228		
8:10623407:G:C	10623407	0.000127714	0.000255363	G	С	0	1	3.858862262	2.2865729	0.091484525	0.119	
8:10623423:C:T	10623423	0.001021711	0.000255363	С	Т	6	2	-0.508481321	0.809915074	0.530121508	6.407	rs200616748
8:10677710:G:A	10677710	0.000127812	0.00102145	G	A	0	1	2.160866408	2.016170191	0.283824313	13.07	rs189167078
8:10677806:G:C	10677806	0.000255624	0.00102145	G	С	1	1	0.279738036	1.493422653	0.851414938		rs368049075
8:10683689:G:T	10683689	0.000127812	0.00102145	G	Т	1	0	-1.711168477	2.038053008	0.401127423	32	
8:10683721:C:G	10683721	0.000127812	0.00102145	С	G	1	0	-1.82932458	2.013645853	0.363633174	26.9	
8:10683752:A:G	10683752	0.000127812	0.00102145	Α	G	0	1	2.370250715	2.030440971	0.243065983	15.88	
8:10689208:C:T	10689208	0.000127747	0.000510725	С	Т	0	1	1.863226453	2.01129488	0.354247733	24.8	rs201784803
8:10689232:C:A	10689232	0	0.000766088	С	А	0	0	NA	NA	NA	32	
8:10690414:G:GGTA	10690414	0.000127714	0.000255363	G	GGTA	0	1	2.450764209	2.036470665	0.228807245		
8:10690422:C:T	10690422	0.000127714	0.000255363	С	Т	1	0	-1.361704771	2.277752881	0.549954612	33	rs377715499
8:10690474:T:G	10690474	0.001405571	0.000766088	Т	G	5	5	0.909309109	0.712155695	0.201658626	25.4	rs189959562
8:10690476:G:T	10690476	0.000127779	0.000766088	G	Т	1	0	-2.377636724	2.027578635	0.240936855	29	
8:10692193:C:T	10692193	0.000127747	0.000510725	С	Т	1	0	-1.963378374	2.005330672	0.327540645	34	
8:10692229:C:T	10692229	0.000127714	0.000255363	С	T	1	0	-1.740323036	2.025811988	0.39029952	32	rs142521930
8:10692232:CA:C	10692232	0	0.000255363	CA	С	0	0	NA	NA	NA		
8:10692283:G:A	10692283	0.000127747	0.000510725	G	A	1	0	-1.789379059	2.018317319	0.375310592	26.4	
8:10697250:T:A	10697250	0.000127747	0.000510725	Т	А	0	1	3.19360382	2.1627751	0.139776678	32	
8:10697260:T:C	10697260	0.000255493	0.000510725	Т	С	1	1	-0.016001327	1.4200209	0.991009329	23	rs200886591
8:10697266:T:A	10697266	0.000255493	0.000510725	Т	А	0	2	1.991418163	1.418068329	0.160224327	24.4	

**Table 2.** *TREM2* variants included in the CADD15/CADD20 analyses. Table shows base pair (BP, minor allele frequency (MAF), missing rate, major and minor allele, p-value for single-marker analysis along with effect size (beta) and standard error (s.e.), CADD

- score

SNP	MAF	<b>Missing Rate</b>	MajorAllele	MinorAllele	beta	se	single-marker pvalue	CADD score	ExAC_ALL	snp138
6:41127543:G:A	0.004984663	0.00102145	G	А	0.829210707	0.343249645	0.016	23.1	0.0036	rs2234255
6:41127561:C:T	0.000638733	0.000510725	С	Т	0.271610307	0.994602461	0.78	23.2	0.0001	rs79011726
6:41129105:G:A	0.000894912	0.001276813	G	А	-0.070395503	0.884520054	0.94	24.3	0.0003	
6:41130779:A:G	0.000255754	0.001532176	А	G	-2.145244892	1.430444422	0.13	23.6		

**eTable 3**. Minor allele frequencies for *PINX1* variants in 1000G and ExAC databases ("\_ALL"=cumulative, 1000g2015aug\_afr= Africans from 1000 Genomes Project; 1000g2015aug\_eur = Europeans from 1000 Genomes Project; ExAC\_AFR= Africans/African Americans from Exome Aggregation Consortium; ExAC\_NFE= Non-Finnish Europeans from Exome Aggregation Consortium).

VARIANT	1000g2015aug_all	1000g2015aug_eur	1000g2015aug_afr	ExAC_ALL	ExAC_AFR	ExAC_NFE
8:10622930:T:C	0.000998403		0.0038	0.0012	0.0133	1.71E-05
8:10623027:G:C						
8:10623094:C:G	0.000199681		0.0008	7.50E-05	0.0009	0
8:10623104:C:T	0.00259585	0.001		0.0024	0.0001	0.0011
8:10623134:G:C				3.33E-05	0.0004	0
8:10623135:C:T				5.83E-05	0.0006	0
8:10623141:T:A				0.0001	0.0001	0.0002
8:10623240:T:C	0.0319489	0.0487	0.003	0.0653	0.014	0.0624
8:10623254:C:A	0.0161741	0.0119	0.0015	0.0158	0.0034	0.0123
8:10623280:C:G	0.00738818		0.0272	0.002	0.023	1.50E-05
8:10623301:C:A	0.000199681	0.001		0.0001	0	0.0002
8:10623309:C:T						
8:10623344:C:T				5.80E-05	0.0001	4.50E-05
8:10623361:C:G				4.97E-05	0.0003	0
8:10623379:G:C				4.14E-05	0	7.50E-05
8:10623383:G:T	0.000199681		0.0008	8.29E-05	0.001	0
8:10623385:C:T				0.0002	0.0021	1.50E-05
8:10623386:G:A				4.14E-05	0.0001	6.00E-05
8:10623393:C:T				1.66E-05	0.0002	0
8:10623407:G:C						
8:10623423:C:T	0.000199681			0.0009	0.0001	0.0015
8:10677710:G:A	0.000399361		0.0015	6.96E-05	0.0007	0
8:10677806:G:C				3.84E-05	0.0004	0
8:10683689:G:T				2.10E-05	0	3.89E-05
8:10683721:C:G				1.05E-05	0	1.93E-05
8:10683752:A:G				1.25E-05	0.0001	0
8:10689208:C:T	0.000199681			8.82E-05	0	0.0001
8:10689232:C:A				3.30E-05	0.0002	3.11E-05
8:10690422:C:T				2.79E-05	0	3.29E-05
8:10690474:T:G	0.00159744	0.006		0.0054	0.0006	0.0084
8:10690476:G:T						
8:10692193:C:T						
8:10692229:C:T	0.000199681	0.001		1.66E-05	0	1.50E-05
8:10692283:G:A				3.40E-05	0	4.61E-05
8:10697250:T:A				1.64E-05	0	2.99E-05
8:10697260:T:C	0.000199681			6.27E-05	0.0004	2.85E-05
8:10697266:T:A	0.000199681			1.56E-05	0	0
8:10622923:TTTC:T	0.00778754		0.028	0.0026	0.0229	0.0007
8:10623000:T:TC	•	•	•	2.50E-05	0.0003	0
8:10623137:CTCT:C	•	•	•	0.0001	0.0001	0.0002
8:10623203:GC:G						
8:10623206:T:TAA						
8:10623396:TCTC:T				0.0001	0	7.50E-05
8:10690414:G:GGTA						
8:10692232:CA:C				•		
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**eTable 4.** LOAD known genes VEP MODERATE-HIGH SKAT-O results (Meta-analysis of WHICAP, ADSP, ROS/MAP). In bold genes that show a p-value<=0.05.

GENE	P.value (WHICAP-ADSP-ROS/MAP MODERATE-HIGH SKATO metanalysis)	symbol
ENSG0000095970	6.10E-05	TREM2
ENSG00000140090	0.00254	SLC24A4
ENSG0000166961	0.008017	MS4A15
ENSG0000137642	0.01175	SORL1
ENSG0000086288	0.01686	NME8
ENSG0000166928	0.01906	MS4A14
ENSG0000166927	0.02066	MS4A7
ENSG00000100599	0.0216	RIN3
ENSG0000108798	0.02523	ABI3
ENSG00000166926	0.06901	MS4A6E
ENSG0000130203	0.08161	APOE
ENSG00000110077	0.08168	MS4A6A
ENSG0000156738	0.09308	MS4A1
ENSG0000073921	0.09368	PICALM
ENSG00000116032	0.1076	GRIN3B
ENSG00000110079	0.1186	MS4A4A
ENSG0000105383	0.1314	CD33
ENSG0000203710	0.1514	CR1
ENSG0000064687	0.1788	ABCA7
ENSG00000172689	0.1795	MS4A10
ENSG00000183580	0.1854	FBXL7
ENSG00000149534	0.1936	MS4A2
ENSG0000046604	0.207	DSG2
ENSG00000120885	0.2243	CLU
ENSG0000073712	0.2399	FERMT2
ENSG0000198087	0.2796	CD2AP
ENSG0000081189	0.2809	MEF2C
ENSG0000197943	0.2825	PLCG2
ENSG0000182168	0.3067	UNC5C
ENSG0000168918	0.3277	INPP5D
ENSG0000166959	0.3386	MS4A8
ENSG0000204979	0.3583	MS4A13
ENSG00000149516	0.4002	MS4A3
ENSG0000149187	0.4011	CELF1
ENSG00000166930	0.4132	MS4A5
ENSG0000087589	0.4216	CASS4
ENSG0000071203	0.4274	MS4A12
ENSG0000120899	0.4524	РТК2В
ENSG0000078487	0.4723	ZCWPW1
ENSG0000136717	0.4788	BIN1
ENSG0000198502	0.526	HLA-DRB5
ENSG00000146904	0.6476	EPHA1
ENSG0000196126	0.7991	HLA-DRB1

## eAppendix 4. Acknowledgment.

## WHICAP and EFIGA

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The ADGC cohorts include: Adult Changes in Thought (ACT supported by NIA grant U01AG006781 to Drs. Eiic Larson and Palu Crane), the Alzheimer's Disease Centers (ADC), the Chicago Health and Aging Project (CHAP), the Memory and Aging Project (MAP), Mayo Clinic (MAYO), Mayo Parkinson's Disease controls, University of Miami, the Multi-Institutional Research in Alzheimer's Genetic Epidemiology Study (MIRAGE), the National Cell Repository for Alzheimer's Disease (NCRAD), the National Institute on Aging Late Onset Alzheimer's Disease Family Study (NIA-LOAD), the Religious Orders Study (ROS), the Texas Alzheimer's Research and Care Consortium (TARC), Vanderbilt University/Case Western Reserve University (VAN/CWRU), the Washington Heights-Inwood Columbia Aging Project (WHICAP supported by NIA grant RF1AG054023 to Dr. Mayeux) and the Washington University Sequencing Project (WUSP), the Columbia University Hispanic- Estudio Familiar de Influencia Genetica de Alzheimer (EFIGA supported by NIA grant RF1AG015473 to Dr. Mayeux), the University of Toronto (UT), and Genetic Differences (GD). Analysis of ADGC cohorts us supported by NIA grants R01AG048927 and RF1AG057519 to Dr. Farrer. Efforts of ADGC investigators were also supported by grants from the NIA (R03AG054936) and National Library of Medicine (R01LM012535).

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#### eReferences. REFERENCES.

- 1. Carson AR, Smith EN, Matsui H, et al. Effective filtering strategies to improve data quality from population-based whole exome sequencing studies. *BMC bioinformatics*. 2014;15(1):125.
- 2. Narasimhan V, Danecek P, Scally A, Xue Y, Tyler-Smith C, Durbin R. BCFtools/RoH: a hidden Markov model approach for detecting autozygosity from next-generation sequencing data. *Bioinformatics*. 2016;32(11):1749-1751.
- 3. Tan A, Abecasis GR, Kang HM. Unified representation of genetic variants. *Bioinformatics*. 2015;31(13):2202-2204.
- McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease Report of the NINCDS-ADRDA Work Group\* under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology*. 1984;34(7):939-939.
- 5. Beecham GW, Bis J, Martin E, et al. The Alzheimer's Disease Sequencing Project: Study design and sample selection. *Neurology Genetics*. 2017;3(5):e194.
- 6. Bennett DA, Buchman AS, Boyle PA, Barnes LL, Wilson RS, Schneider JA. Religious Orders Study and Rush Memory and Aging Project. *Journal of Alzheimer's Disease*. 2018;(Preprint):1-28.
- 7. Vardarajan BN, Barral S, Jaworski J, et al. Whole genome sequencing of Caribbean Hispanic families with lateonset Alzheimer's disease. *Annals of clinical and translational neurology*. 2018;5(4):406-417.
- 8. Naj AC, Jun G, Beecham GW, et al. Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nature genetics*. 2011;43(5):436-441.
- 9. Myers AJ, Gibbs JR, Webster JA, et al. A survey of genetic human cortical gene expression. *Nature genetics*. 2007;39(12):1494.
- 10. Mostafavi S, Gaiteri C, Sullivan SE, et al. A molecular network of the aging human brain provides insights into the pathology and cognitive decline of Alzheimer's disease. *Nature neuroscience*. 2018;21(6):811.
- 11. Johnson WE, Li C, Rabinovic A. Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics*. 2007;8(1):118-127.
- 12. White CC, Yang H-S, Yu L, et al. Identification of genes associated with dissociation of cognitive performance and neuropathological burden: Multistep analysis of genetic, epigenetic, and transcriptional data. *PLoS medicine*. 2017;14(4):e1002287.