

Supplemental material

Goel et al., <https://doi.org/10.1083/jcb.201807165>

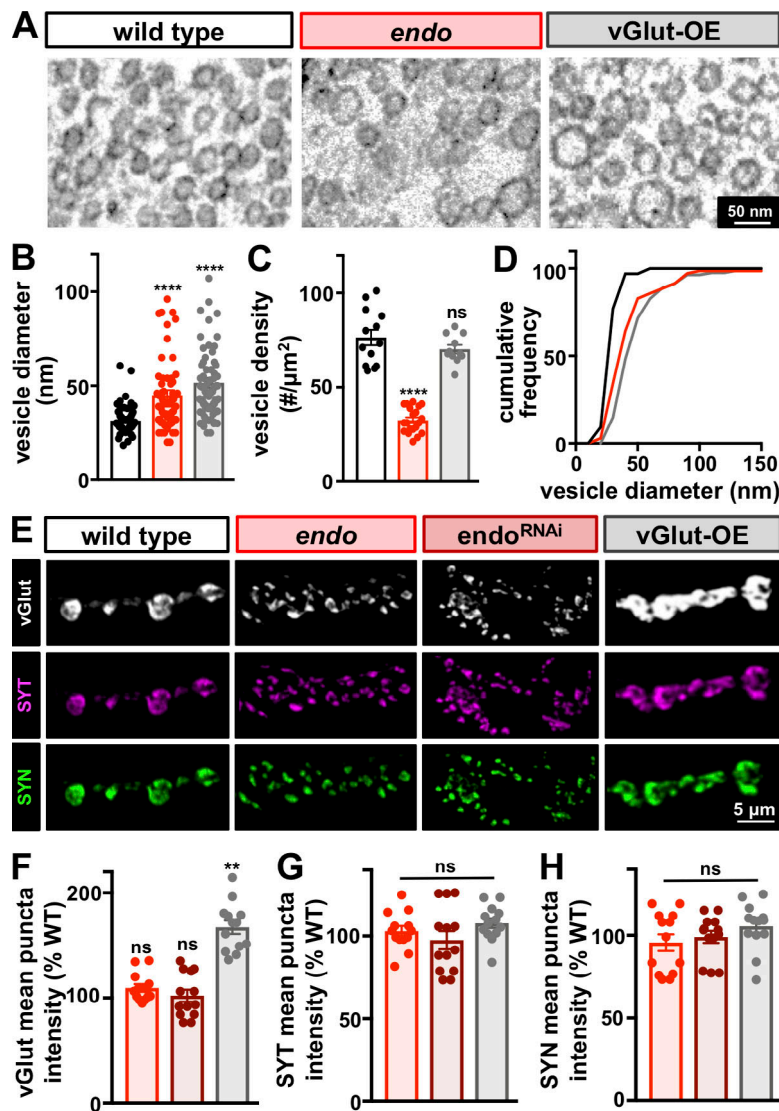


Figure S1. **Distinct mechanisms enlarge SVs and quantal size at *endo* and vGlut-OE synapses (related to Fig. 1).** (A) Representative electron micrographs of SVs at NMJ terminals from wild type, *endo*, and vGlut-OE. (B) Quantification of increased SV diameter in *endo* and vGlut-OE, consistent with increased glutamate emitted from these vesicles. (C) Quantification of SV density reveals a significant reduction in *endo* but no change in vGlut-OE compared with wild type ( $n \geq 65$ ; one-way ANOVA, K-S test in D; Table S1). (D) Cumulative frequency distribution of SV diameter shows a rightward shift of the entire population in *endo* and vGlut-OE. (E) Representative images of individual boutons in the indicated genotypes immunostained with antibodies that recognize the SV markers vGlut (vesicular glutamate transporter), SYN (Synapsin), and SYT (Synaptotagmin). (F-H) Quantification of mean intensities of individual vGlut, SYN, and SYT puncta demonstrates a specific increase in vGlut intensity in vGlut-OE, as expected, with no change in puncta intensities of SYN or SYT. In contrast, intensity is unchanged in all SV markers in *endo* or *endo*<sup>RNAi</sup>. Error bars indicate  $\pm$  SEM ( $n \geq 7$ ; one-way ANOVA; Table S1). \*\*,  $P < 0.01$ ; \*\*\*\*,  $P < 0.0001$ ; ns, not significant.

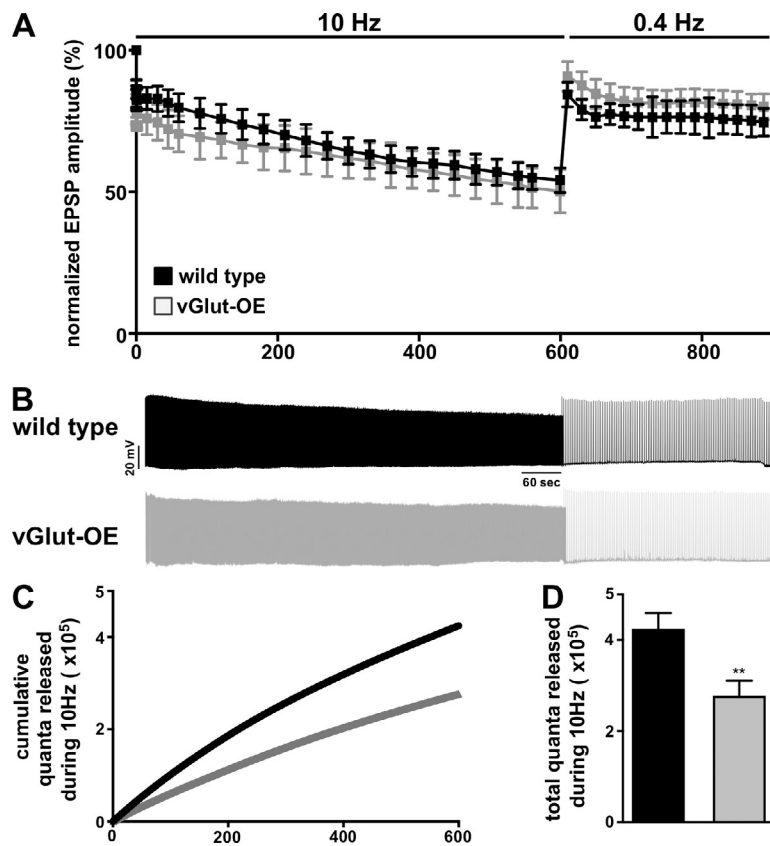


Figure S2. **SV recycling is unperturbed in vGlut-OE (related to Fig. 1).** (A) Wild-type and vGlut-OE NMJs were stimulated at 10 Hz for 10 min in 2 mM extracellular Ca<sup>2+</sup> and subsequently allowed to recover with stimulation at 0.4 Hz. EPSP amplitudes were averaged (binning 2 s of responses for each time point), normalized to prestimulus amplitude, and plotted as a function of time. (B) Representative traces of experiments in A for the indicated genotypes. (C) Quantification of cumulative quanta released during the 10-min stimulation protocol in each genotype shows the expected reduction in vGlut-OE. (D) Quantification of the total quanta released in each genotype during the 10-Hz rundown period reveals a reduction in vGlut-OE. All error bars indicate  $\pm$  SEM ( $n \geq 7$ ;  $t$  test; Table S1). \*\*,  $P < 0.01$ .

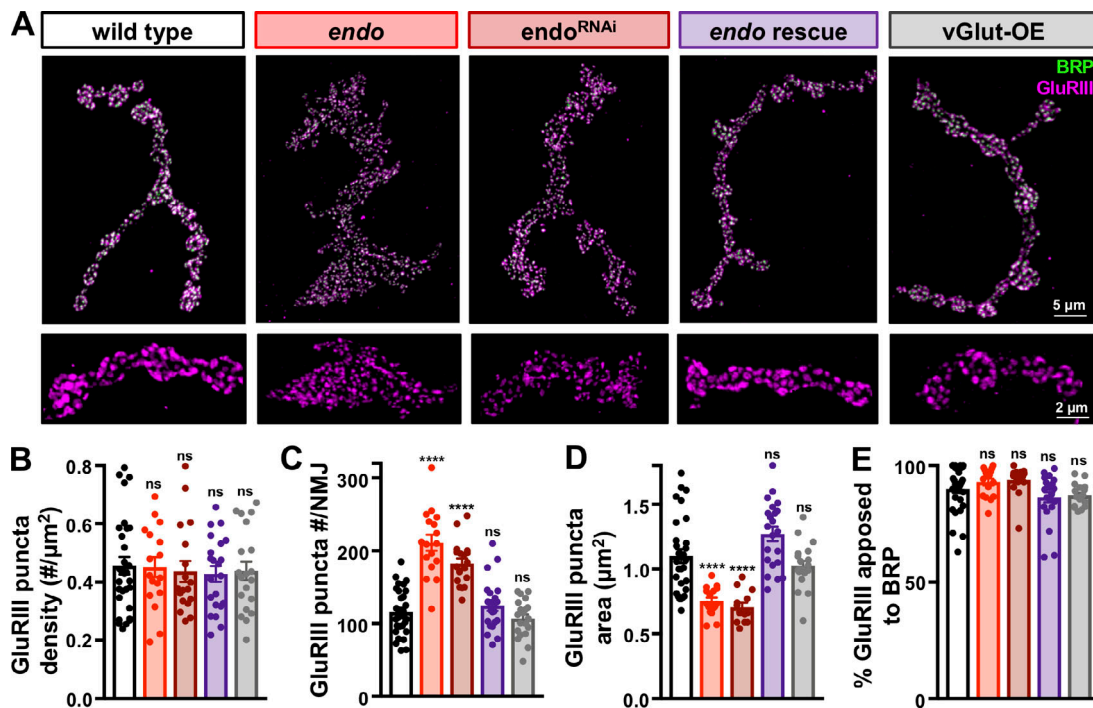


Figure S3. **Postsynaptic glutamate receptor fields mirror presynaptic AZ structure in *endo* mutants (related to Fig. 2).** (A) Representative NMJs in the indicated genotypes immunostained with antibodies against the essential postsynaptic glutamate receptor subunit GluRIII, with insets below representing individual boutons from the same NMJ. (B) No change in GluRIII density is observed in any genotype. (C) GluRIII puncta numbers per NMJ are significantly increased in *endo* and *endo<sup>RNAi</sup>* and rescued in *endo rescue*, and no change is found in *vGlut-OE*. (D) A significant reduction in the area of individual GluRIII puncta is observed in *endo* and *endo<sup>RNAi</sup>*, with no change in *vGlut-OE*. (E) No significant difference in the apposition of GluRIII puncta to BRP puncta is observed in any genotype. Error bars indicate  $\pm$  SEM ( $n \geq 12$ ; one-way ANOVA; Table S1). \*\*\*\*,  $P < 0.0001$ ; ns, not significant.

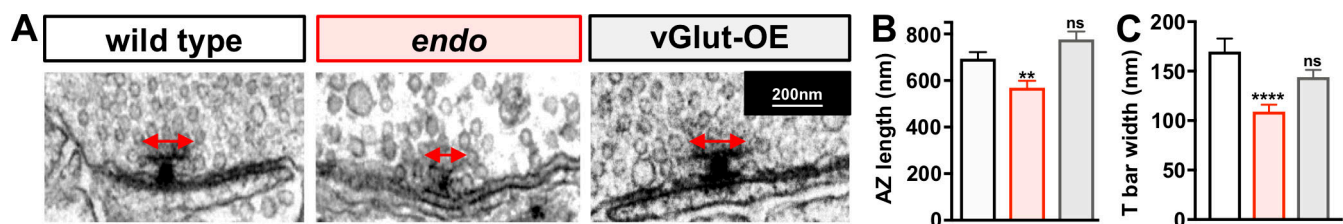


Figure S4. **Reduced AZ length and T-bar width in *endo* mutants revealed by EM (related to Figs. 2 and 5).** (A) Representative EM images of AZs in wild type, *endo* mutants, and *vGlut-OE*. The width of the T-bar structure at the center of the AZ in shown by the arrows. (B and C) Quantification of AZ length (B) and T bar width (C) in the indicated genotypes. All error bars indicate  $\pm$  SEM ( $n \geq 54$ ; one-way ANOVA; Table S1). \*\*,  $P < 0.01$ ; \*\*\*\*,  $P < 0.0001$ ; ns, not significant.

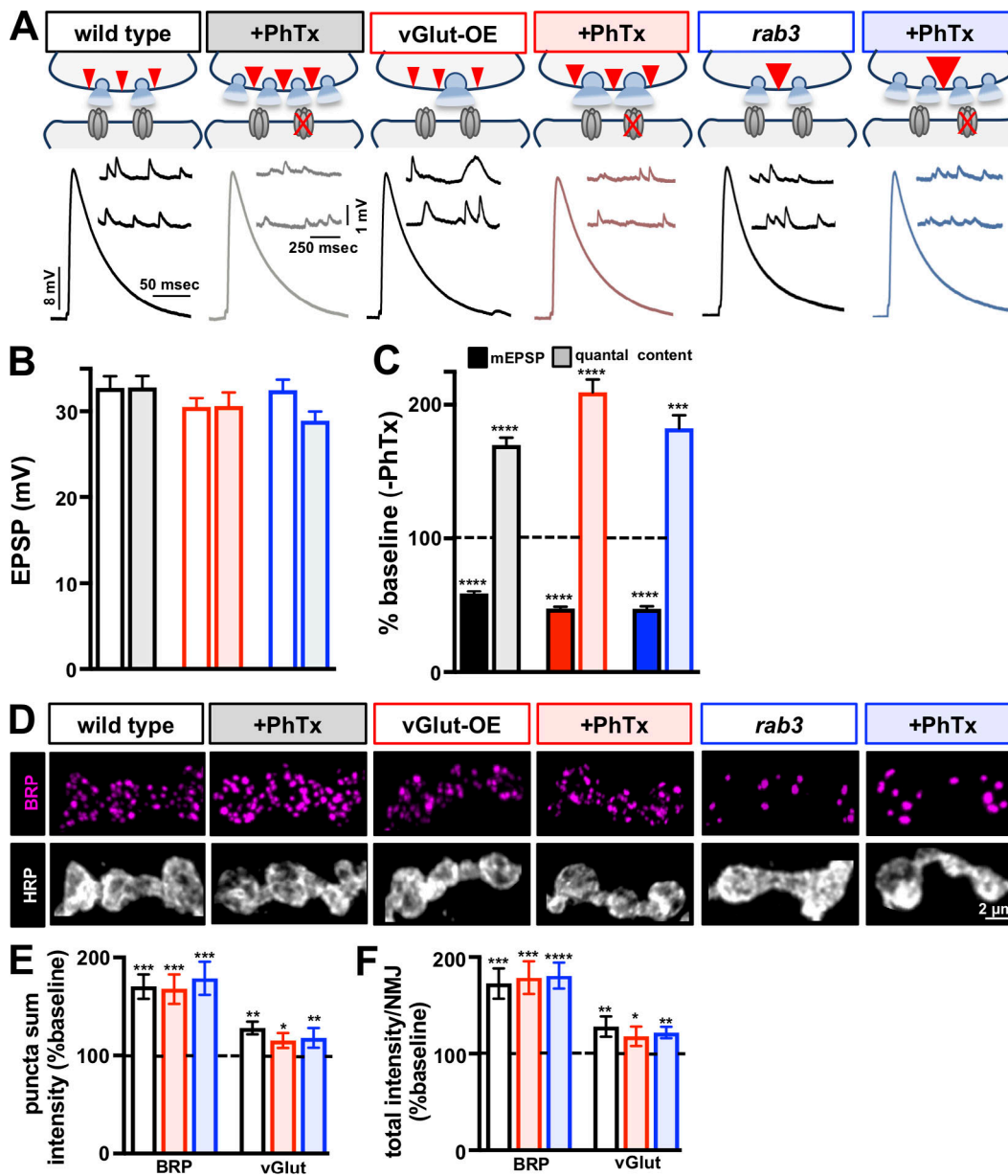


Figure S5. NMJs expressing homeostatic depression or AZ scaling can acutely express homeostatic potentiation (related to Fig. 7). (A) Schematic and representative traces of wild type, vGlut-OE, and *rab3* mutant NMJs before and after PhTx application. Diminished mEPSP amplitudes are observed in all conditions after PhTx application, while EPSP amplitudes are maintained at baseline levels due to a homeostatic increase in quantal content. (B and C) Quantification of EPSP amplitude (B) and normalized mEPSP amplitude and quantal content (C) in the indicated genotypes ( $n \geq 7$ ; t test; Table S1). (D) Representative images of NMJ boutons immunostained with anti-BRP in the indicated genotypes at baseline and after PhTx application. (E) Quantification of mean intensity of BRP and vGlut puncta reveals a significant increase following PhTx application compared with baseline values (no PhTx). (F) An increase in the total fluorescence intensity of BRP and vGlut puncta per NMJ is observed in wild type, vGlut-OE, and *rab3* mutants after PhTx application ( $n \geq 11$ ; t test; Table S1). Error bars indicate  $\pm$  SEM. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ ; \*\*\*\*,  $P < 0.0001$ .

Table S2. **Key resources**

| Reagent/resource                                      | Source                                  | Identifier                            |
|---|---|---------------------------------------|
| <b>Antibodies</b>                                     |   |                                       |
| Mouse anti-Synapsin (3C11)                            | DSHB                                    | AB_2313867                            |
| Guinea pig anti-vGlut                                 | <a href="#">Chen et al., 2017</a>       | N/A                                   |
| Rabbit anti-Synaptotagmin 1                           | <a href="#">Mackler et al., 2002</a>    | N/A                                   |
| Mouse anti-BRP (nc82)                                 | DSHB                                    | AB_2314866                            |
| Guinea pig anti-Unc13A                                | <a href="#">Böhme et al., 2016</a>      | N/A                                   |
| Affinity-purified rabbit anti-GluRIII                 | <a href="#">Kiragasi et al., 2017</a>   | N/A                                   |
| DyLight 405–conjugated secondary antibodies           | Jackson ImmunoResearch                  | 706-475-148, 715-475-150, 711-475-152 |
| Alexa Fluor 488–conjugated secondary antibodies       | Jackson ImmunoResearch                  | 706-545-148, 715-545-150, 711-545-152 |
| Alexa Fluor Cyanine 3–conjugated secondary antibodies | Jackson ImmunoResearch                  | 706-165-148, 715-165-150, 711-165-152 |
| Alexa Fluor 647–conjugated goat anti-HRP              | Jackson ImmunoResearch                  | 123-605-021                           |
| Goat anti-mouse Alexa Fluor 594                       | Thermo Fisher Scientific                | A11032                                |
| Goat anti-guinea pig star635                          | Abberior                                | 1-0101002                             |
| <b>Experimental models: fly lines</b>                 |   |                                       |
| <i>endo</i> <sup>1</sup>                              | <a href="#">Verstreken et al., 2002</a> | N/A                                   |
| <i>endo</i> <sup>Δ4</sup>                             | <a href="#">Verstreken et al., 2002</a> | N/A                                   |
| <i>GluRIIA</i> <sup>sp16</sup>                        | <a href="#">Petersen et al., 1997</a>   | N/A                                   |
| <i>elav</i> <sup>c155</sup> - <i>Gal4</i>             | BDSC                                    | BDSC #458                             |
| <i>OK371-Gal4</i>                                     | <a href="#">Mahr and Aberle, 2006</a>   | N/A                                   |
| <i>OK6-Gal4</i>                                       | <a href="#">Aberle et al., 2002</a>     | N/A                                   |
| <i>UAS-vGlut</i>                                      | <a href="#">Daniels et al., 2004</a>    | N/A                                   |
| <i>rab3</i> <sup>rup</sup>                            | <a href="#">Graf et al., 2009</a>       | N/A                                   |
| <i>Cac</i> <sup>s<sup>GFP-N</sup></sup>               | <a href="#">Gratz et al., 2019</a>      | N/A                                   |
| <i>UAS-arl8-GFP</i>                                   | <a href="#">Vukoja et al., 2018</a>     | N/A                                   |
| <i>arl8</i> <sup>e00336</sup>                         | <a href="#">Vukoja et al., 2018</a>     | N/A                                   |
| <i>UAS-endo RNAi</i> (chromosome 3)                   | BDSC                                    | BDSC #27679                           |
| <i>UAS-Rab3 RNAi</i> (chromosome 3)                   | BDSC                                    | BDSC #31691                           |
| BRP-GFP   | BDSC                                    | BDSC #59292                           |
| vGlut-GFP   | BDSC                                    | BDSC #59411                           |

N/A, not applicable.

**Provided online is Table S1, showing absolute values for normalized data and additional statistics.**

## References

- Aberle, H., A.P. Haghighi, R.D. Fetter, B.D. McCabe, T.R. Magalhães, and C.S. Goodman. 2002. wishful thinking encodes a BMP type II receptor that regulates synaptic growth in *Drosophila*. *Neuron*. 33:545–558. [https://doi.org/10.1016/S0896-6273\(02\)00589-5](https://doi.org/10.1016/S0896-6273(02)00589-5)
- Böhme, M.A., C. Beis, S. Reddy-Alla, E. Reynolds, M.M. Mampell, A.T. Grasskamp, J. Lützkendorf, D.D. Bergeron, J.H. Driller, H. Babikir, et al. 2016. Active zone scaffolds differentially accumulate Unc13 isoforms to tune Ca(2+) channel-vesicle coupling. *Nat. Neurosci.* 19:1311–1320. <https://doi.org/10.1038/nn.4364>
- Chen, X., W. Ma, S. Zhang, J. Paluch, W. Guo, and D.K. Dickman. 2017. The BLOC-1 Subunit Pallidin Facilitates Activity-Dependent Synaptic Vesicle Recycling. *eNeuro*. 4:1–18. <https://doi.org/10.1523/ENEURO.0335-16.2017>
- Daniels, R.W., C.A. Collins, M.V. Gelfand, J. Dant, E.S. Brooks, D.E. Krantz, and A. DiAntonio. 2004. Increased expression of the *Drosophila* vesicular glutamate transporter leads to excess glutamate release and a compensatory decrease in quantal content. *J. Neurosci.* 24:10466–10474. <https://doi.org/10.1523/JNEUROSCI.3001-04.2004>
- Graf, E.R., R.W. Daniels, R.W. Burgess, T.L. Schwarz, and A. DiAntonio. 2009. Rab3 dynamically controls protein composition at active zones. *Neuron*. 64: 663–677. <https://doi.org/10.1016/j.neuron.2009.11.002>

- Gratz, S.J., P. Goel, J.J. Bruckner, R.X. Hernandez, K. Khateeb, G.T. Macleod, D. Dickman, and K.M. O'Connor-Giles. 2019. Endogenous tagging reveals differential regulation of Ca<sup>2+</sup> channels at single AZs during presynaptic homeostatic potentiation and depression. *J. Neurosci.* 28:3068–3018.
- Kiragasi, B., J. Wondolowski, Y. Li, and D.K. Dickman. 2017. A Presynaptic Glutamate Receptor Subunit Confers Robustness to Neurotransmission and Homeostatic Potentiation. *Cell Reports*. 19:2694–2706. <https://doi.org/10.1016/j.celrep.2017.06.003>
- Mackler, J.M., J.A. Drummond, C.A. Loewen, I.M. Robinson, and N.E. Reist. 2002. The C(2)B Ca(2+)-binding motif of synaptotagmin is required for synaptic transmission in vivo. *Nature*. 418:340–344. <https://doi.org/10.1038/nature00846>
- Mahr, A., and H. Aberle. 2006. The expression pattern of the Drosophila vesicular glutamate transporter: a marker protein for motoneurons and glutamatergic centers in the brain. *Gene Expr. Patterns*. 6:299–309. <https://doi.org/10.1016/j.modgep.2005.07.006>
- Petersen, S.A., R.D. Fetter, J.N. Noordermeer, C.S. Goodman, and A. DiAntonio. 1997. Genetic analysis of glutamate receptors in Drosophila reveals a retrograde signal regulating presynaptic transmitter release. *Neuron*. 19:1237–1248. [https://doi.org/10.1016/S0896-6273\(00\)80415-8](https://doi.org/10.1016/S0896-6273(00)80415-8)
- Verstreken, P., O. Kjaerulff, T.E. Lloyd, R. Atkinson, Y. Zhou, I.A. Meinertzhagen, and H.J. Bellen. 2002. Endophilin mutations block clathrin-mediated endocytosis but not neurotransmitter release. *Cell*. 109:101–112. [https://doi.org/10.1016/S0092-8674\(02\)00688-8](https://doi.org/10.1016/S0092-8674(02)00688-8)
- Vukoja, A., U. Rey, A.G. Petzoldt, C. Ott, D. Vollweiler, C. Quentin, D. Puchkov, E. Reynolds, M. Lehmann, S. Hohensee, et al. 2018. Presynaptic Biogenesis Requires Axonal Transport of Lysosome-Related Vesicles. *Neuron*. 99:1216–1232.e7. <https://doi.org/10.1016/j.neuron.2018.08.004>