

AJP-17-04-0469R2
Supplemental Material

Altered Brain Developmental Trajectories in Adolescents after Initiating Drinking

Adolf Pfefferbaum, M.D.,^{1,2} Dongjin Kwon, Ph.D.,^{1,2} Ty Brumback, Ph.D.,³
Wesley K. Thomson, Ph.D.,^{3,4} Kevin Cummins, M.A.,³ Susan F. Tapert, Ph.D.,³
Sandra A. Brown, Ph.D.,³ Ian M. Colrain, Ph.D.,¹ Fiona C. Baker, Ph.D.,¹ Devin Prouty, Ph.D.,¹
Michael D. De Bellis, M.D., M.P.H.,⁵ Duncan B. Clark, M.D., Ph.D.,⁶ Bonnie J. Nagel, Ph.D.,⁷
Weiwei Chu, M.A.,¹ Sang Hyun Park, Ph.D.,^{1,8} Kilian M. Pohl, Ph.D.,¹ Edith V. Sullivan, Ph.D.^{2*}

¹Center for Health Sciences, SRI International, Menlo Park, CA;

²Department of Psychiatry & Behavioral Sciences,
Stanford University School of Medicine, Stanford, CA;

³Department of Psychiatry, University of California, San Diego, La Jolla, CA;

⁴Institute of Biological Psychiatry, Roskilde, Denmark;

⁵Healthy Childhood Brain Development Research Program, Department of
Psychiatry & Behavioral Sciences, Duke University School of Medicine, Durham, NC;

⁶Department of Psychiatry, University of Pittsburgh, Pittsburgh, PA;

⁷Departments of Psychiatry and Behavioral Neuroscience,
Oregon Health & Sciences University, Portland, OR

⁸Department of Robotics Engineering,
Daegu Gyeongbuk Institute of Science and Technology, South Korea

INTRODUCTION

Here we present initial evidence of altered adolescent brain growth trajectory associated with moderate and heavy alcohol use from a national, multi-site, prospective study of hundreds of participants studied before and after they initiated harmful levels of alcohol use.

Cortical shrinkage in normal development is typically interpreted as normal pruning of neuropil constituents in response to absence of environmental or interoceptive input (e.g., 1, 2) (for review, 3). Support for this hypothesis derives from studies based on sleep physiology and on positron emission tomography (PET). Specifically, pubertal maturational changes in sleep electrophysiology measured longitudinally in adolescents revealed a steep decline in delta power density, which paralleled thinning in cortical layers known to evidence synaptic pruning (4, 5). Glucose metabolism measured cross-sectionally with PET characterized a rise in regional cortical metabolism from 5 weeks of age, peaking at about 9 years (5), and declining thereafter (6, 7), paralleling the sleep markers of maturation, the temporal course of the rise and fall of cortical volumes (8), and synaptic remodeling through pruning (9). In contrast to cortical thinning, growth of white matter determines ultimate intracranial volume (8) and is thought to underlie maturing connectivity with experience (10).

METHODS

Informed consent. The Institutional Review Boards (IRB) of each site approved this study. All participants underwent IRB-approved informed consent processes at each visit. Adult participants or the parents of minor participants provided written informed consent before participation at each annual visit; minor participants provided assent.

Alcohol history determination. Participants completed the Customary Drinking and Drug use Record (CDDR) (11) to characterize past and current alcohol and substance use. Historical variables regarding substance use obtained with the CDDR included 9 temporally-linked measures: cumulative days consuming alcohol over a lifetime; maximum drinks per session, total number of drinks, binges, and hangovers in the past year; and days drinking, total drinks, maximum drinks per occasion, and hangover symptoms in the last month quantified with the Hangover Symptoms Scale (12). Lifetime and past month marijuana use data were also collected.

Demographics

In light of the substantial differences in salaries, incomes, and occupational categories across the five geographically-distributed data collection sites, we expressed SES with reference to parental education level, which is less subject to geographical differences in the U.S. Most subjects reported a single self-identified ethnicity (Caucasian, African-American, Asian, Pacific Islander, and Native American) with some reporting mixed heritage. There were adequate numbers of the first three types to assign categorical ethnicity, with dual-heritage identifications assigned to the minority ethnicity group (e.g., Asian-Caucasian was categorized as Asian). Male youth were disproportionately represented in the heavy relative to the moderate drinking group ($\chi^2=6.5821$, $p=.0103$) (reviewed in 13).

Internalizing and externalizing symptoms, considered high risk for alcohol use and problems (14), were quantified using the Achenbach System of Empirically Based Assessment (15, 16). Participants under age 18 years completed the Youth Self-Report; participants over age 18 completed the Adult Self-Report. Each scale yielded age- and sex-normed continuous measures, where T-scores >60 from the externalizing and internalizing scales were in the subclinical psychopathology range.

Cahalan et al. alcohol consumption criteria

Heavy drinkers ranged from moderate frequency (e.g., 2x/month) with high quantity consumption (e.g., with 3-4 drinks on average and > 4 drinks maximum) to higher frequency (e.g., 1x/week or more) with moderate quantity consumption (e.g., with 2-3 drinks on average and >4 drinks maximum). Moderate drinkers ranged from low drinking frequency (e.g., <1x/month) with moderate quantity consumption (e.g., with 2-3 drinks on average and 4-5 drinks maximum) to moderate frequency (e.g., 1x/week) and low quantity consumption (e.g., with 2 drinks on average and <4 drinks maximum). No/low drinkers reported no or low quantity and frequency consumption (e.g., <1x/month, <2 drinks on average, and <4 drinks maximum).

Drug use

All participants also submitted samples to a 14-panel urine toxicology screen for tetrahydrocannabinol, amphetamine, methamphetamine, methylenedioxy-methamphetamine, cocaine, phencyclidine, benzodiazepines, barbiturates, morphine, oxycodone, methadone, buprenorphine, propoxyphene, and tricyclic antidepressants, and a breathalyzer for alcohol to

confirm absence of evidence for recent use of drugs of abuse. Participants were asked to abstain from substance use for 72 hours prior to testing and scanning. Participants with clinical or biological evidence of recent use were excluded from that day, rescheduled, and tested again for recent use of substances on the return visit.

MRI Acquisition

The longitudinal data for the current analysis comprised MR images collected on 483 of the initial 674 no-to-low baseline participants (17) who had 2-year (and in most cases, 1 year) follow-up MRI and CDDR data, met the double alcohol inclusion criteria, met FreeSurfer SNR criteria, and had adequate quality imaging data.

MRIs were acquired in the sagittal plane on systems from two manufacturers: 3T General Electric (GE) Discovery MR750 at three sites (UCSD, SRI, and Duke) and 3T Siemens TIM TRIO scanners at two sites (University of Pittsburgh and OHSU). MRI acquisition and analysis details appear in supplemental material. The GE sites used an Array Spatial Sensitivity Encoding Technique (ASSET) for parallel and accelerated imaging with an 8-channel head coil and acquired an Inversion Recovery-Spoiled Gradient Recalled (IR-SPGR) echo sequence (TR=5.904ms, TI=400ms, TE=1.932ms, flip angle=11°, NEX=1, matrix=256x256, FOV=24cm, slice dimensions=1.2 x 0.9375 x 0.9375mm, 146 slices). The Siemens sites used a 12-channel head coil and parallel imaging and temporal acceleration with iPAT and acquired an MPRAGE sequence (TR=1900ms, TI=900ms, TE=2.92 ms, flip angle=9°, NEX=1, matrix=256x256, FOV=24cm, slice dimensions=1.2 x 0.9375 x 0.9375mm, 160 slices). All sites also collected sagittal T2-weighted images with the same geometric prescription as the T1-weighted acquisitions for use in skull stripping.

Scalable Informatics for Biomedical Imaging Studies (SIBIS)

The data were based on a formal, locked data release (NCANDA_PUBLIC_2Y_STRUCTURAL_MEASUREMENTS_V01) provided by the software platform Scalable Informatics for Biomedical Imaging Studies (SIBIS; <https://github.com/sibis-platform>). SIBIS consists of IT infrastructure for collecting behavioral and imaging data at the NCANDA sites, Internet, and application programming interfaces for uploading the acquired data to a central biomedical data repository, a validated workflow to perform quality control, a multi-modal image processing pipeline for structural scores, and a release mechanism for disseminating the data to be used for publications. Below is a brief review the structural MR imaging pipeline; the non-imaging component of SIBIS is described elsewhere (18-20).

MRI Analysis

Preprocessing of the T1-weighted (T1w) and T2-weighted (T2w) MRI data involved noise removal (21), correcting field inhomogeneity via N4ITK (22), aligning T2w to T1w MRIs using CMTK (23), repeating image inhomogeneity correction of both modalities confined to the brain mask defined by aligning SRI24 atlas (24) to T1w MRI using the symmetric, diffeomorphic non-rigid registration called ANTS (25). The brain mask was refined by majority voting (26) accomplished across maps extracted by FSL BET (27), AFNI 3dSkullStrip (28), FreeSurfer `mri_gcut` (29), and the Robust Brain Extraction (ROBEX) method (30), which were applied on combinations of bias and non-bias corrected T1w and T2w images. Using the refined masked, we repeated the image inhomogeneity correction.

For baseline visits only, the skull-stripped T1w image was registered to the SRI24 atlas (24) via ANTS (25). To ensure the longitudinal consistency of the structural measures, ANTS also

registered the baseline to the follow-up T1w MRIs (with skull) to align the brain mask of the baseline to each visit. Using the aligned brain mask, the skull-stripping and image inhomogeneity correction of the follow-up scans was repeated. Afterwards, the inter-visit alignment was refined by ANTS registering the processed T1w MRIs (without skull) of baseline to the ones of the follow-up visits. The corresponding transformation was then used to align the SRI24 atlas to each visit. Furthermore, the intensity profile of the registered follow-up scan was matched to the baseline by smoothing both images (Gaussian Filter with 3mm kernel), computing the ratio between the smoothed intensities of the two time points at each image location, and then applying that ratio to the follow-up scan.

We extracted longitudinal image scores using two different atlases. To produce volume scores based on the SRI24 atlas, longitudinal brain tissue segmentation (gray matter, white matter, and cerebrospinal fluid) was performed via Atropos (31). The resulting label maps of each time point was parcellated by the SRI24 atlas, which identified supratentorial volume (svol), pons, corpus callosum, and a large central white matter sample including the centrum semiovale and internal capsule. To compute structural scores based on the FreeSurfer software (32) [<http://www.sciencedirect.com/science/article/pii/S1053811912000389>], we applied its cross-sectional approach to the skull-stripped MRI of each time point, which, in part, refined the brain masks removing voxels having low T2-weighted intensities near the brain surface. Based on the refined brain masks, longitudinal FreeSurfer (32, 33) applied to the aligned baseline and follow-up T1-weighted MRIs resulted in bilateral surface area, volume, and thickness measures. Initial testing collapsed the Desikan-Killiany regions-of-interest (ROI) (34) [surfer.nmr.mgh.harvard.edu/fswiki/CorticalParcellation] into bilateral frontal, temporal, parietal, occipital, cingulate, and insular cortices. Secondary analyses used all 34 individual Desikan-Killiany bilateral cortical ROIs. For this report, only volume (a function of surface area and thickness) was considered.

Statistical Analysis

For display (Figure 1, left panel), Linear Mixed-Effects Models (lme4, R Version 3.2.4 [<http://www.r-project.org/>]) of native values, before and after svol was removed by regression, demonstrate svol as a major contribution to apparent sex differences.

The group matching procedure used "MatchIt" in R with exact sex and ethnicity and nearest age (35).

DISCUSSION

As would be expected from epidemiological (for review,13) and laboratory studies (e.g.,36, 37, 38), the heavy drinking group had proportionately greater male (60%) than female (40%) representation than the even sex distribution in the full baseline group and a higher percentage of family history positive youth (14.5%) than the moderate (7.7%) or no/low (7.6%) drinking groups. In addition, the female moderate drinkers tended to have the highest internalizing scores (mean $T=46.2$, $F(1,123)=2.654$, $p=.106$), whereas the male heavy drinkers had the highest externalizing scores (mean $T=46.8$, $F(1,123)=4.318$, $p=.04$). Other studies have identified these variables as predictors of alcohol use in youth (e.g.,39).

Examination of marijuana co-use with alcohol on trajectories revealed no additional effect of marijuana, thus providing further support for the conclusion that regionally accelerated tissue decline in the alcohol transitioners could be attributable to alcohol

consumption itself. Of note, the marijuana co-use group comprised 67% male youth, whereas the non-marijuana drinkers comprised only 43% male youth ($\chi^2=4.585$, $p=.0323$). Another characteristic difference between these groups was the higher level of externalizing symptomatology in marijuana users regardless of drinking level ($F(1,122)=6.102$, $p=.0149$).

While two earlier longitudinal studies showed attenuated white matter volume growth in pons and corpus callosum in heavy-drinking youth, the current study found decreased growth only for central white matter volume ($p=.0344$, uncorrected). The tissue-based trajectory differences suggest that initiating heavy drinking during the growth years of adolescence has potentially differential effects on gray matter and white matter volume development, possibly with respect to mechanisms of gray matter pruning and white matter expansion and myelination. Alternatively, measures of gray matter volume may be more sensitive than those of white matter. Tissue shrinkage in normal development is typically interpreted as normal pruning of neuropil constituents in response to absence of environmental or interoceptive input (e.g., 15, 16). One speculative interpretation of the apparent acceleration of the pruning trend notable in young adolescent drinkers is an over-exuberance of the typical synaptic refinements, suggesting an alteration of progression into the later stages of neurodevelopment.

References for Supplemental Material

1. Feinberg I, Thode HC, Chugani HT, March JD. Gamma distribution model describes maturational curves for delta wave amplitude, cortical metabolic rate and synaptic density. *J Theor Biol.* 1990;142:149-161.
2. Feinberg I, Campbell IG. Longitudinal sleep EEG trajectories indicate complex patterns of adolescent brain maturation. *Am J Physiol Regul Integr Comp Physiol.* 2013;304:R296-303.
3. Selemon LD. A role for synaptic plasticity in the adolescent development of executive function. *Transl Psychiatry.* 2013;3:e238.
4. Campbell IG, Feinberg I. Longitudinal trajectories of non-rapid eye movement delta and theta EEG as indicators of adolescent brain maturation. *Proceedings of the National Academy of Sciences of the United States of America.* 2009;106:5177-5180.
5. Campbell IG, Grimm KJ, de Bie E, Feinberg I. Sex, puberty, and the timing of sleep EEG measured adolescent brain maturation. *Proceedings of the National Academy of Sciences of the United States of America.* 2012;109:5740-5743.
6. Chugani HT, Phelps ME, Mazziotta JC. Positron emission tomography study of human brain functional development. *Ann Neurol.* 1987;22:487-497.
7. Chugani HT. A critical period of brain development: studies of cerebral glucose utilization with PET. *Prev Med.* 1998;27:184-188.
8. Pfefferbaum A, MATHALON DH, Sullivan EV, Rawles JM, Zipursky RB, Lim KO. A quantitative magnetic resonance imaging study of changes in brain morphology from infancy to late adulthood. *Arch Neurol.* 1994;51:874-887.
9. Huttenlocher PR. Synaptic density in human frontal cortex: Developmental changes and effects of aging. *Brain Res.* 1979;163:195-205.
10. Giedd JN, Raznahan A, Alexander-Bloch A, Schmitt E, Gogtay N, Rapoport JL. Child psychiatry branch of the National Institute of Mental Health longitudinal structural

- magnetic resonance imaging study of human brain development. *Neuropsychopharmacology*. 2015;40:43-49.
11. Brown SA, Myers MG, Lippke L, Tapert SF, Stewart DG, Vik PW. Psychometric evaluation of the Customary Drinking and Drug Use Record (CDDR): a measure of adolescent alcohol and drug involvement. *Journal of Studies on Alcohol*. 1998;59:427-438.
 12. Slutske WS, Piasecki TM, Hunt-Carter EE. Development and initial validation of the Hangover Symptoms Scale: prevalence and correlates of Hangover Symptoms in college students. *Alcohol Clin Exp Res*. 2003;27:1442-1450.
 13. Patrick ME, Schulenberg JE. Prevalence and predictors of adolescent alcohol use and binge drinking in the United States. *Alcohol Research: Current Reviews*. 2013;35:193-200.
 14. Brown SA, Brumback T, Tomlinson K, Cummins K, Thompson WK, Nagel BJ, De Bellis MD, Hooper SR, Clark DB, Chung T, Hasler BP, Colrain IM, Baker FC, Prouty D, Pfefferbaum A, Sullivan EV, Pohl KM, Rohlfing T, Nichols BN, Chu W, Tapert SF. The National Consortium on Alcohol and NeuroDevelopment in Adolescence (NCANDA): A multi-site study of adolescent development and substance use. *Journal of Studies on Alcohol and Drugs*. 2015;76:895-908.
 15. Achenbach T, Rescorla L: Manual for the ASEBA school-age forms & profiles. Burlington, VT, University of Vermont, Research Center for Children, Youth, and Families; 2001.
 16. Achenbach TM, Rescorla LA: Manual for the ASEBA School-Age Forms & Profiles. Burlington, VT, University of Vermont, Research Center for Children, Youth, & Families; 2001.
 17. Pfefferbaum A, Rohlfing T, Pohl KM, Lane B, Chu W, Kwon D, Nichols BN, Brown SA, Tapert SF, Cummins K, Thompson WK, Brumback T, Meloy MJ, Jernigan TL, Dale A, Colrain IM, Baker FC, Prouty D, De Bellis MD, Voyvodic JT, Clark DB, Luna B, Chung T, Nagel BJ, Sullivan EV. Adolescent Development of Cortical and White Matter Structure in the NCANDA Sample: Role of Sex, Ethnicity, Puberty, and Alcohol Drinking. *Cereb Cortex*. 2016;26:4101-4121.
 18. Rohlfing T, Cummins K, Henthorn T, Chu W, Nichols BN. N-CANDA data integration: anatomy of an asynchronous infrastructure for multi-site, multi-instrument longitudinal data capture. *J Am Med Inform Assoc*. 2014;21:758-762.
 19. Nichols BN, Pohl KM. Neuroinformatics Software Applications Supporting Electronic Data Capture, Management, and Sharing for the Neuroimaging Community. *Neuropsychol Rev*. 2015;25:356-368.
 20. Pohl KM, Sullivan EV, Rohlfing T, Chu W, Kwon D, Nichols BN, Zhang Y, Brown SA, Tapert SF, Cummins K, Thompson WK, Brumback T, Colrain IM, Baker FC, Prouty D, De Bellis MD, Voyvodic JT, Clark DB, Schirda C, Nagel BJ, Pfefferbaum A. Harmonizing DTI measurements across scanners to examine the development of white matter microstructure in 803 adolescents of the NCANDA study. *Neuroimage*. 2016;130:194-213.
 21. Coupe P, Yger P, Prima S, Hellier P, Kervrann C, Barillot C. An optimized blockwise nonlocal means denoising filter for 3-D magnetic resonance images. *IEEE Trans Med Imaging*. 2008;27:425-441.
 22. Tustison NJ, Avants BB, Cook PA, Zheng Y, Egan A, Yushkevich PA, Gee JC. N4ITK: improved N3 bias correction. *IEEE Trans Med Imaging*. 2010;29:1310-1320.

23. Rohlfing T, Maurer CR. Nonrigid image registration in shared-memory multiprocessor environments with application to brains, breasts, and bees. *IEEE Trans Inf Technol Biomed.* 2003;7:16-25.
24. Rohlfing T, Zahr NM, Sullivan EV, Pfefferbaum A. The SRI24 multi-channel atlas of normal adult human brain structure. *Hum Brain Mapp.* 2010;31:798-819.
25. Avants BB, Epstein CL, Grossman M, Gee JC. Symmetric diffeomorphic image registration with cross-correlation: evaluating automated labeling of elderly and neurodegenerative brain. *Med Image Anal.* 2008;12:26-41.
26. Rohlfing T, Russakoff DB, Maurer Jr CR. Performance-Based Classifier Combination in Atlas-Based Image Segmentation Using Expectation-Maximization Parameter Estimation. *IEEE Trans Med Imaging.* 2004;23:983-994.
27. Smith S. Fast robust automated brain extraction. *Hum Brain Mapp.* 2002;17:143-155.
28. Cox RW. AFNI: software for analysis and visualization of functional magnetic resonance neuroimages. *Comput Biomed Res.* 1996;29:162-173.
29. Sadananthan SA, Zheng W, Chee MW, Zagorodnov V. Skull stripping using graph cuts. *Neuroimage.* 2010;49:225-239.
30. Iglesias JE, Liu CY, Thompson PM, Tu Z. Robust brain extraction across datasets and comparison with publicly available methods. *IEEE Trans Med Imaging.* 2011;30:1617-1634.
31. Avants BB, Tustison NJ, Wu J, Cook PA, Gee JC. An open source multivariate framework for n-tissue segmentation with evaluation on public data. *Neuroinformatics.* 2011;9:381-400.
32. Dale AM, Fischl B, Sereno MI. Cortical surface-based analysis. I. Segmentation and surface reconstruction. *Neuroimage.* 1999;9:179-194.
33. Reuter M, Schmansky NJ, Rosas HD, Fischl B. Within-subject template estimation for unbiased longitudinal image analysis. *Neuroimage.* 2012;61:1402-1418.
34. Desikan RS, Segonne F, Fischl B, Quinn BT, Dickerson BC, Blacker D, Buckner RL, Dale AM, Maguire RP, Hyman BT, Albert MS, Killiany RJ. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *Neuroimage.* 2006;31:968-980.
35. Ho DE, Imai K, King G, Stuart EA. Matching as nonparametric preprocessing for reducing model dependence in parametric causal inference. *Political Analysis.* 2007;15:199-236.
36. Puttler LI, Fitzgerald HE, Heitzeg MM, Zucker RA. Boys, Early Risk Factors for Alcohol Problems, and the Development of the Self: An Interconnected Matrix. *Infant Ment Health J.* 2017;38:83-96.
37. Weiland BJ, Korycinski ST, Soules M, Zubieta JK, Zucker RA, Heitzeg MM. Substance abuse risk in emerging adults associated with smaller frontal gray matter volumes and higher externalizing behaviors. *Drug Alcohol Depend.* 2014;137:68-75.
38. Squeglia LM, Ball TM, Jacobus J, Brumback T, McKenna BS, Nguyen-Louie TT, Sorg SF, Paulus MP, Tapert SF. Neural Predictors of Initiating Alcohol Use During Adolescence. *The American journal of psychiatry.* 2017;174:172-185.
39. Brumback TY, Worley M, Nguyen-Louie TT, Squeglia LM, Jacobus J, Tapert SF. Neural predictors of alcohol use and psychopathology symptoms in adolescents. *Dev Psychopathol.* 2016;28:1209-1216.

Supplemental Table. Mean residualized volumes and t-test differences between the no/low and heavy drinking groups and p-values for the 34 FreeSurfer ROIs

Region of interest (ROI)	Mean	t	uncorrected p	adjusted p
frontalpole	-1.8634838	0.28267049	0.777573493	0.85282254
caudalmiddlefrontal	-2.2474363	-3.20350528	0.001464748	0.02377815
lateralorbitofrontal	-1.7027995	-1.40706427	0.16017338	0.32034676
medialorbitofrontal	-1.6641957	-1.1039058	0.270287835	0.43489767
<i>paracentral</i>	<i>-1.6607567</i>	<i>-2.04893725</i>	<i>0.041110569</i>	<i>0.1164799</i>
<i>parsopercularis</i>	<i>-1.4757981</i>	<i>-2.328787</i>	<i>0.020360429</i>	<i>0.0865318</i>
parsorbitalis	-2.1688183	-1.5904716	0.112506864	0.23907709
<i>parstriangularis</i>	<i>-2.1404769</i>	<i>-2.43113188</i>	<i>0.01548368</i>	<i>0.0865318</i>
<i>precentral</i>	<i>-1.2290262</i>	<i>-2.47463877</i>	<i>0.013744708</i>	<i>0.0865318</i>
<i>rostralmiddlefrontal</i>	<i>-2.6154937</i>	<i>-2.14102999</i>	<i>0.032866508</i>	<i>0.1110648</i>
superiorfrontal	-1.5783155	-3.0959195	0.002098072	0.02377815
inferiorparietal	-2.6762046	-1.28074383	0.201014796	0.36169955
<i>postcentral</i>	<i>-1.7962486</i>	<i>-2.10468575</i>	<i>0.035932744</i>	<i>0.1110648</i>
<i>precuneus</i>	<i>-2.007058</i>	<i>-2.2048668</i>	<i>0.028023508</i>	<i>0.1058666</i>
superiorparietal	-2.4252064	-1.62125987	0.105738236	0.23907709
supramarginal	-2.2596892	-1.66747494	0.0961911	0.23500586
<i>cuneus</i>	<i>-1.7377284</i>	<i>-2.35963883</i>	<i>0.018765222</i>	<i>0.0865318</i>
lateraloccipital	-1.9101856	-0.59640048	0.551240099	0.65666749
lingual	-1.1967916	-1.27758391	0.20212622	0.36169955
pericalcarine	0.3092154	-0.92059083	0.357810756	0.52893764
bankssts	-2.7540927	-0.68335118	0.494774738	0.64701312
temporalpole	-0.8573634	-0.29603932	0.767351413	0.85282254
entorhinal	-0.666269	-0.06781642	0.945965154	0.94596515
fusiform	-1.5436273	-0.68575904	0.493256456	0.64701312
inferiortemporal	-2.4838566	-0.19402511	0.846253259	0.87796252
middletemporal	-2.170363	-0.58317581	0.560098741	0.65666749
parahippocampal	-1.1170502	-0.18650732	0.852140091	0.87796252
superiortemporal	-1.3828514	-1.6645848	0.09676712	0.23500586
transversetemporal	-1.0428155	-0.60183123	0.547622408	0.65666749
caudalanteriorcingulate	-0.9365606	-1.0786047	0.281404377	0.43489767
<i>isthmuscingulate</i>	<i>-1.8069184</i>	<i>-2.38640357</i>	<i>0.017471324</i>	<i>0.0865318</i>
posteriorcingulate	-1.6443726	-3.12310425	0.001917771	0.02377815
rostralanteriorcingulate	-0.8838348	-1.2118194	0.226286304	0.38468672
insula	-0.6734757	-0.78103711	0.435235837	0.6165841

Italic font: unadjusted $p \leq 0.05$ and displayed on Figure 5

bold font: FDR-adjusted p-value and displayed in orange in Figure 5