DNA polymerase η modulates replication fork progression and DNA damage responses in platinum-treated human cells.

Anna M. Sokol^{1**}, Séverine Cruet-Hennequart^{1***}, Philippe Pasero² and Michael P. Carty^{1*}

- * To whom correspondence should be addressed. Tel: (353) 91-493695; Fax: (353) 91-495504; Email: michael.carty@nuigalway.ie
- ** Present Address: Laboratory of Mitochondrial Biogenesis, International Institute of Molecular and Cell Biology, 4 Ks. Trojdena Street, 02-109 Warsaw, Poland
- *** Present Address: Microenvironment and Pathology Laboratory (MILPAT, EA 4652), Niveau 3, Faculté de Médecine, Avenue de la Côte de Nacre, 14032 Caen cedex. France

¹ DNA Damage Response Laboratory, Centre for Chromosome Biology, Biochemistry, School of Natural Sciences, National University of Ireland, Galway, Ireland

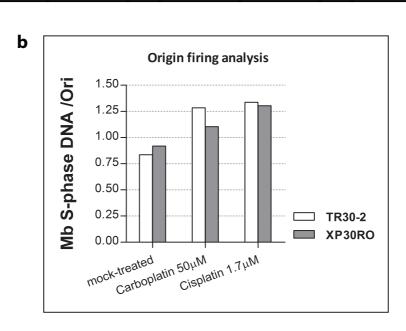
² Department of Molecular Basis of Human Diseases, Institute of Human Genetics, CNRS UPR 1142, Montpellier, France

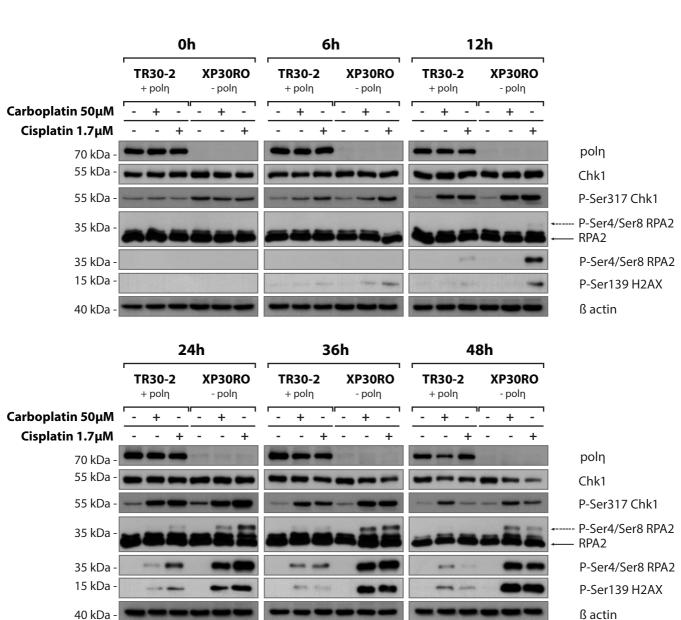
Supplementary Figure 1. DNA combing parameters. 1a Mean length and synthesis rate values from two independent DNA combing experiments ± SEM, are presented in Table format. N stands for the number of measured objects. **1b** Graph presents the frequency of origin firing based on data from DNA combing experiments carried out under the indicated conditions. BrdU incorporation data from FACS analysis was used to calculate the S-phase DNA fraction.

Supplementary Figure 2. Kinetics of carboplatin- and cisplatin-induced DNA damage responses in cells lacking and expressing DNA polymerase η. Polη-deficient XP30RO cells and polη expressing TR30-2 cells were treated with indicated doses of carboplatin and cisplatin and harvested at indicated times post-treatment. Expression of DNA damage response proteins and protein phosphorylation were analysed by western blotting using specific antibodies as described in Materials and Methods. For direct comparison, samples obtained from both cell lines at the same time-point were run on the same gels.

Supplementary Table 1. EdU intensity analysis by quantitative immunofluorescence. Cells expressing (TR30-2) or lacking (XP30RO) pol η were grown on glass coverslips and treated with indicated doses of cisplatin and carboplatin. 75 minutes before fixing, cells were incubated with 10 μ M EdU. Click chemistry was used to visualise EdU incorporation. 1a Mean relative EdU intensities values \pm SEM from three independent experiments under the indicated conditions are presented. The number of individual values is indicated as n. 1b Fold-decrease in EdU intensity \pm SEM relative to mock-treated samples is presented. 1c The EdU intensity threshold, obtained by subtracting double the SEM value from the mean relative EdU intensity value.

	Mock - treated			Carboplatin 50μM			Cisplatin 1.7μM		
а	Mean length [kb]	Mean synthesis rate [kb/min]	n	Mean length [kb]	Mean synthesis rate [kb/min]	n	Mean length [kb]	Mean synthesis rate [kb/min]	n
TR30-2	82.0 ± 2.8	1.37 ± 0.05	183	82.8 ± 2.8	1.38 ± 0.05	185	78.9 ± 2.8	1.31 ± 0.05	163
XP30RO	82.3 ± 2.1	1.37 ± 0.03	268	69.1 ± 2.6	1.15 ± 0.04	180	48.2 ± 1.6	0.80 ± 0.03	360





a	Mock - treated		Carboplatin 50μM		Cisplatin 1.7μM		
"	Mean relative EdU intensity	n	Mean relative EdU intensity	n	Mean relative EdU intensity	n	
TR30-2	1175.0 ± 31.5	394	447.9 ± 12.4	487	385.7 ± 14.1	402	
XP30RO	1220.0 ± 26.7	362	255.1 ± 9.4	523	177.0 ± 9.5	489	

	Carboplatin 50μM	Cisplatin 1.7μM			
b	Fold decrease in relative EdU intensity	Fold decrease in relative EdU intensity			
TR30-2	2.7 ± 0.14	3.2 ± 0.22			
XP30RO	5.7 ± 1.75	7.3 ± 1.19			

	Carboplatin 50µM	Cisplatin 1.7μM		
C	Threshold value for EdU intensity Mean relative EdU intensity - 2xSEM	Threshold value for EdU intensity Mean relative EdU intensity - 2xSEM		
XP30RO	236.7	157.9		