

Figure S1. Summary of structures of PKA R:PKAcα heterodimers and tetrameric PKA holoenzymes determined previously and in this study, Related to Figures 1, 4 and 5.

(A) Previously reported structures of truncated PKA R:PKAcα heterodimers: RIα (91-379):PKAcα (PDB ID 2QCS, left) (Kim et al., 2007), RIIα (90-400):PKAcα (PDB ID 2QVS, middle) (Wu et al., 2007) and RIIβ (108-268):PKAcα (PDB ID 3IDB, right) (Brown et al.,

2009). (B) Previously determined wt R₂:PKAcα₂ holoenzymes: RIβ₂:PKAcα₂ (PDB ID 4DIN, left) (Ilouz et al., 2012) and RIIβ₂:PKAcα₂ (PDB ID 3TNP, midle) (Zhang et al., 2012) holoenzymes structures which were determined by X-ray crystallography, and the RIα₂:PKAcα₂ model based on crystal packing of two truncated RIα(73-244):PKAcα heterodimers (right) (Boettcher et al., 2011). On the top, the right R:PKAcα heterodimers are aligned at the same orientation; on the bottom the bird-eye view is shown for the three holoenzymes. (C) Bird-eye view of RIα₂:J-PKAcα₂ and RIα₂:PKAcα₂ tetrameric holoenzyme structures determined in this study. The twofold axis position for each holoenzyme is shown as a black dot in the middle.



Figure S2. Formation and structural analysis of the chimeric RIα₂:PKAcα₂ holoenzyme, Related to Figure 1.

(A) Analytical gel filtration profile showing formation of RI α_2 :J-PKAc α_2 . (B) The diffracting crystals contain the full-length RI α_2 :J-PKAc α_2 complex that was used for crystallization. The purified proteins RI α (lane 1), J-PKAc α (lane 2) and the dissolved diffracting crystals (lane 3) were run on a 7% tris-acetate SDS-PAGE gel and silver stained. (C) Cross-eyed stereo view of part of the RI α_2 :J-PKAc α_2 holoenzyme structure (left) and the N3A-N3A' interface in the RI α_2 :J-PKAc α_2 holoenzyme structure (right) in the 3.66 Å resolution 2Fo-Fc map at 1 σ . (D) Overlay of J-PKAc α (colored) in the chimeric holoenzyme and PKI-bound PKAc α (gray, PDB ID 1ATP).

(E) Overlay of J-PKAcα in the chimeric holoenzyme (colored) and PKI-bound J-PKAcα (gray,
PDB ID 4WB7). (F) Side-by-side comparison of RIα:J-PKAcα in the chimeric holoenzyme and a canonical R:PKAcα heterodimer in the RIIβ₂:PKAcα₂ holoenzyme (PDB ID 3TNP).



Figure S3. SAXS results from RIa2:J-PKAca2, Related to STAR methods.

(A) SAXS profiles of RI α_2 :J-PKAc α_2 at different concentrations. (B) Calculated scattering curve from crystal structure in continuous green line and SAXS experimental curve extrapolated to infinity dilution in blue dots. (C) Guinier plot, I₀: 0.081, R_g:48.8 ± 2.0 Å and Q_{max}*R_g: 1.26. (D) The P(r) functions from the crystal structure of RI α_2 :J-PKAc α_2 in continuous solid green line and SAXS experimental data for the chimeric holoenzyme in blue dots.



Figure S4. The J-domain configurations during the MD simulations started from the J-in state model (A and B) and the J-out state model (C), Related to Figure 2.

For each simulation of $RI\alpha_2$:J-PKAc α_2 , the top shows the orientation of the J-domain for each copy of the chimera in the holoenzyme, and the bottom shows the Initial (left) and final (right)

configurations of the J-domain in the holoenzyme. Hydrogen bonds that formed between the Jdomain and CNB-B domain during the J-in state model simulation are shown in (B). The angles in both simulations are the same as those defined in Figure 2. The red 'x' indicates the initial conformation of the J-domain.



Figure S5. Comparison among the final conformations from holoenzyme MD simulations, holoenzyme crystal and SAXS experimental curves, Related to STAR Methods.

(A) Calculated scattering curves from holoenzyme crystal and final conformations of MD simulations in solid lines and SAXS experimental curve extrapolated to infinity dilution in blue dots. (B) The P(r) functions from MD final conformations and holoenzyme crystal in solid lines and SAXS experimental data for the chimeric holoenzyme in blue dots. Calculated χ^2 , R_g and D_{max} values are reported in Table S2.





(A) Analytical gel filtration profile showing formation of $RI\alpha_2$:PKAc α_2 . (B) The diffracting crystals contain the full-length $RI\alpha_2$:PKAc α_2 complex that was used for crystallization. The purified proteins RI α (lane 1), PKAc α (lane 2), J-PKAc α (lane 3) and the dissolved diffracting crystals (lane 4) were run on a 7% tris-acetate SDS-PAGE gel and silver stained. (C) Overlay of the RI α_2 :PKAc α_2 holoenzyme 1 (gray) and chimeric RI α_2 :J-PKAc α_2 holoenzyme (colored). (D)

Cross-eyed stereo view of the N3A-N3A' interfaces in the conformations 1 (top) and 2 (bottom) of the RI α_2 :PKAc α_2 holoenzyme structure in the 4.75 Å resolution 2Fo-Fc map at 1 σ . (E) Overlay of the RI α :PKAc α heterodimers in the wt holoenzyme 1 (colored) and 2 (gray). (F) Overlay of RI α :PKAc α in the wt holoenzyme 1 (colored) and the previously reported RI α :PKAc α heterodimer (gray, PDB ID 2QCS). (G) The minimum C α distances between PKAc α and RI α ' in wt holoenzyme 1 (left) and 2 (right). (H) Formation of salt bridges between the C-lobe of PKAc α and the CNB-B' domain of RI α ' in wt holoenzyme 2 during MD simulation.



Figure S7. The RIα-RIα' interfaces and dynamics of the αB/C-helix in the RIα chimeric holoenzyme (Crystal), models of the chimeric holoenzyme with both J-domains in J-in state (J-in) and with both J-domains in J-out state (J-out), and wt holo (WT) during MD simulations, Related to Figure 2.

(A) Orientation between the two α B/C-helices in the RI α dimers as a function of time. (B) Contact area of the RI α -RI α ' interfaces. (C) Linearity of the α B/C-helices as defined by the C α atoms of D225-G235-K250. The α B/C-helices in simulations do not sample the bent conformation that is observed in the cAMP-bound RI α homodimer. Solid lines indicate the α B/C bending angle in one R subunit while dashed lines indicate those in the symmetry-related R subunit in the holoenzyme.

Table S1. Averaged B factors for the J-domain and the rest of J-PKAca bound with RIa or **PKI**, Related to Figure 1.

	$\mathbf{B_{J}}^{\mathrm{a}}$	B_{C}^{b}	Ratio of B _J :B _C
RIa ₂ :J-PKAca ₂	203.45	94.09	2.16:1
J-PKAca:PKI ^c	63.54	34.39	1.85:1

^aB_J: Averaged B factors of the J-domain (residues 1-69) ^bB_C: Averaged B factors of the rest of J-PKAcα (residues 70-405) ^c from PDB ID 4WB7 (Cheung et al., 2015)

Holoenzyme conformation	$R_g(Å)$ in	$R_g(Å)$ in	D _{max} (Å)	$\chi^{2 c}$
	reciprocal space	real space		
SAXS solution scattering	$48.8\pm2.0^{\rm d}$	50.0 ± 1.0	177	N/A
Crystal ^a	44.6	44.7 ± 0.5	142	1.44
Jout-Jout1 ^b	44.6	44.6 ± 0.5	149	1.63
Jout-Jout2 ^b	46.7	46.9 ± 0.7	176	1.30
Jout-Jin ^b	45.3	45.4 ± 0.5	151	1.37

Table S2. SAXS structural parameters, Related to Figure 2.

^a Calculated from the crystal structure
 ^b Calculated from the final conformations of MD simulations
 ^c Compared to experimental SAXS solution data
 ^d Obtained from Guinier plot. The other Rg values were obtained from GNOM.

Table S3. Thermal stability of the RIa2:J-PKAca2 and RIa2:PKAca2 holoenzymes, Related to Figure 4.

Tm ^a (°C)	- MgATP	+ MgATP
RIa ₂ :PKAca ₂	52.93 ± 0.02	59.40 ± 0.03
RIa ₂ :J-PKAca ₂	51.70 ± 0.01	58.90 ± 0.03
ΔTm (wt/mutant)	1.23 ± 0.02	0.50 ± 0.03

^a Tm: temperature at which the protein denatures All data are mean \pm s.d. (n = 2 independent experiments).