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Supplemental Data

**Disease-Associated Genetic Variation
in Human Mitochondrial Protein Import**

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Supplemental Data

Supplemental Table 1: Human genes with reported variants associated with defective mitochondrial import and phenotype of knockout mice. Data for human diseases are reported with genes and phenotype entries in OMIM <https://www.omim.org/> downloaded on 12/09/2018. When the OMIM annotations are not up to date with literature, relevant publications cited in the current study are added. The data for the phenotype of the knockout mice are from <http://www.informatics.jax.org/> and presented with record IDs.

Supplemental Table 2: Genes with variants associated with 3-MGA-uria. A. as reported in OMIM phenotypic series [PS250950], <https://www.omim.org/phenotypicSeries/PS250950> downloaded on 12/11/2018, **B.** additional genes reported in literature.

Supplemental References

1: to the main text

2: to Supplemental Table 1

3: to Supplemental Table 2

Supplemental Table 1

Import pathway component	Gene	Gene (MIM #)	Contribution last update	Last entry edit history	Map location	Disease phenotype	Phenotype (MIM #)	Inheritance	Knockout mouse phenotype (MGI #)
Transporter machinery	<i>AGK</i>	610345	20/06/2012	05/10/2016	7q34	Cataract 38	614691	AR	
						Sengers syndrome	212350	AR	
	<i>DNAJC19</i>	608977	18/03/2014	02/08/2017	3q26.33	3-methylglutaconic aciduria, type V	610198	AR	
	<i>OXA1L</i>	601066	02/06/1999	06/04/2010	14q11.2	Not in OMIM. Supp. Ref 72 Thompson <i>et al.</i>			
	<i>PAM16</i>	614336	23/03/2015	14/04/2017	16p13.3	Spondylometaphyseal dysplasia, Megarbane-Dagher-Melike type	613320	AR	Embryonic or preweaning lethality (MGI:1913699)
	<i>TIMM8A</i>	300356	20/04/2016	20/04/2016	Xq22.1	Mohr-Tranebjaerg syndrome	304700	XLR	
	<i>TIMM22</i>	607251	10/30/2017	10/30/2017	17p13.3	Not in OMIM. Supp. Ref 73 Pacheu-Grau <i>et al.</i>			Embryonic or preweaning lethality (MGI:1929742)
	<i>TIMM50</i>	607381	03/10/2017	03/10/2017	19q13.2	3-methylglutaconic aciduria, type IX	617698	AR	Preweaning lethality (MGI:1913775)
MIA pathway						Myopathy, isolated mitochondrial	616209	AD	
	<i>CHCHD10</i>	615903	29/01/2015	25/10/2016	22q11.23	Frontotemporal dementia and/or amyotrophic lateral sclerosis 2	615911	AD	Mild mitochondrial respiration, skeletal-muscle anomalies (MGI:2143558)
	<i>GFER</i>	600924	07/10/2009	09/10/2009	16p13.3	Myopathy, mitochondrial progressive, congenital cataract, hearing loss and developmental delay	613076		Embryonic lethality (MGI:107757)
Chaperone	<i>HSPD1</i>	118190	13/09/2018	19/09/2018	2q33.1	Leukodystrophy, hypomyelinating, 4 Spastic paraplegia 13	612233 605280	AR AD	Embryonic lethality (MGI:96242)
Client Processing	<i>AFG3L2</i>	604581	20/12/2016	05/01/2018	18p11.21	Spastic ataxia 5 Spinocerebellar ataxia 28	614487 610246	AR AD	Perinatal lethality (MGI:1916847)
	<i>CLPB</i>	616254	16/06/2015	02/08/2017	11q13.4	3-methylglutaconic aciduria, type VII, with cataracts, neurologic involvement and neutropenia	616271	AR	
	<i>CLPP</i>	601119	12/06/2018	13/06/2018	19p13.3	Perrault syndrome 3	614129	AR	Infertility and premature death (MGI:1858213)
	<i>HTRA2</i>	606441	03/01/2017	20/07/2017	2p13.1	3-methylglutaconic aciduria, type VIII	617248	AR	Parkinsonian symptoms, striatal neurons loss, spleen and thymus atrophy, failure to thrive, premature death (MGI:1928676)
	<i>LONP1</i>	605490	06/02/2015	09/02/2015	19p13.3	CODAS syndrome	600373	AR	Embryonic lethality (MGI:1921392)
	<i>MIPEP</i>	602241	30/11/2016	01/12/2016	13q12.12	Combined oxidative phosphorylation deficiency 31	617228	AR	
	<i>PITRM1</i>	618211	30/11/2018	30/11/2018	10p15.2	Not in OMIM. Supp. Ref 74 Brunetti <i>et al.</i>			Preweaning lethality (MGI:1916867)
	<i>PMPCA</i>	613036	03/02/2016	12/10/2016	9q34.3	Spinocerebellar ataxia, 2	213200	AR	
	<i>PMPCB</i>	603131	03/05/2018	29/08/2018	7q22.1	Multiple mitochondrial dysfunctions syndrome 6	617954	AR	Embryonic lethality (MGI:1920328)
	<i>SPG7</i>	602783	10/04/2013	22/07/2015	16q24.3	Spastic paraplegia 7	607259	AD, AR	Impaired motor skills (MGI:2385906)
	<i>XPNPEP3</i>	613553	08/09/2010	06/09/2013	22q13.2	Nephronophthisis-like nephropathy 1	613159	AR	
	<i>YME1L1</i>	607472	21/02/2018	21/02/2018	10p12.1	Optic atrophy 11	617302	AR	Embryonic lethality (MGI:1351651)
Lipid environment	<i>SERAC1</i>	614725	19/02/2018	23/02/2018	6q25.3	3-methylglutaconic aciduria with deafness, encephalopathy, and Leigh-like syndrome	614739	AR	Mild neurological phenotype (MGI: 2447813)
	<i>TMEM70</i>	612418	01/07/2014	21/05/2015	8q21.11	Mitochondrial complex V (ATP synthase) deficiency, nuclear type 2	614052	AR	Embryonic lethality (MGI:1915068)
	<i>TAZ</i>	300394	17/07/2012	13/07/2018	Xq28	Barth syndrome	302060	XLR	Male infertility (MGI:109626)

Supplemental Table 2

A

OMIM type	Gene/locus	Location	Gene/locus MIM number	Phenotype MIM number	Syndrome	Inheritance
I or MGA1	<i>AUH</i>	9q22.31	600529	250950		AR
II or MGCA2	<i>TAZ</i>	Xq28	300394	302060	Barth	XLR
III or MGCA3	<i>OPA3</i>	19q13.32	606580	258501	Costeff	AR
IV or MGCA4	<i>MGCA1</i>	not mapped				
V or MGCA5	<i>DNAJC19</i>	3q26.33	608977	610198	DCMA	AR
VI or MGCA6	<i>SERAC1</i>	6q25.3	614725	614739	MEGDEL	AR
VII or MGCA7	<i>CLPB</i>	11q13.4	616254	616271		AR
VIII or MGCA8	<i>HTRA2</i>	2p13.1	606441	617248		AR
IX or MGCA9	<i>TIMM50</i>	19q13.2	607381	617698		AR

B

	Gene/locus	Location	Gene/locus MIM number	Phenotype MIM number	Syndrome	Evidence for 3-MGA-uria
TIM22 component						
	<i>AGK</i>	7q34	610345	212350	Sengers	Supp. Ref 30 Mayr <i>et al.</i>
Complex V						
	<i>ATPAF2</i>	17p11.2	608918	604273		Supp. Ref 76 De Meirleir <i>et al.</i>
	<i>ATP5F1D</i>	19p13.3	603150	618120		Supp. Ref 77 Olahova <i>et al.</i>
	<i>ATP5F1E</i>	20q13.32	606153	614053		Supp. Ref 75 Mayr <i>et al.</i>
Complex V assembly factor gene						
	<i>TMEM70</i>	8q21.11	612418	614052		Supp. Ref 78 Spiegel <i>et al.</i>
Cristae junction organizing system						
	<i>MICOS13</i>	19p13.3	616658			Supp. Ref 79 Guarani <i>et al.</i>

Supplemental References

Additional references supporting the main text

The references were not included in the main text due to limit on number of allowed citations.

Introduction¹⁻⁵

1. Anderson, S., Bankier, A.T., Barrell, B.G., de Bruijn, M.H., Coulson, A.R., Drouin, J., Eperon, I.C., Nierlich, D.P., Roe, B.A., Sanger, F., et al. (1981). Sequence and organization of the human mitochondrial genome. *Nature* 290, 457-465.
2. Kim, S.J., Xiao, J., Wan, J., Cohen, P., and Yen, K. (2017). Mitochondrially derived peptides as novel regulators of metabolism. *The Journal of physiology* 595, 6613-6621.
3. McCormick, E.M., Zolkipli-Cunningham, Z., and Falk, M.J. (2018). Mitochondrial disease genetics update: recent insights into the molecular diagnosis and expanding phenotype of primary mitochondrial disease. *Current opinion in pediatrics*.
4. Stenton, S.L., and Prokisch, H. (2018). Advancing genomic approaches to the molecular diagnosis of mitochondrial disease. *Essays in biochemistry* 62, 399-408.
5. Rub, C., Wilkening, A., and Voos, W. (2017). Mitochondrial quality control by the Pink1/Parkin system. *Cell and tissue research* 367, 111-123.

The basic machinery for mitochondrial import.^{6; 7}

6. Harbauer, A.B., Zahedi, R.P., Sickmann, A., Pfanner, N., and Meisinger, C. (2014). The protein import machinery of mitochondria-a regulatory hub in metabolism, stress, and disease. *Cell metabolism* 19, 357-372.
7. Wasilewski, M., Chojnacka, K., and Chacinska, A. (2017). Protein trafficking at the crossroads to mitochondria. *Biochimica et biophysica acta* 1864, 125-137.

1. Complexes mediating import of proteins to mitochondria⁸⁻¹¹

8. Stiller, S.B., Hopker, J., Oeljeklaus, S., Schutze, C., Schrempp, S.G., Vent-Schmidt, J., Horvath, S.E., Frazier, A.E., Gebert, N., van der Laan, M., et al. (2016). Mitochondrial OXA Translocase Plays a Major Role in Biogenesis of Inner-Membrane Proteins. *Cell metabolism* 23, 901-908.
9. Tokatlidis, K. (2005). A disulfide relay system in mitochondria. *Cell* 121, 965-967.
10. Craig, E.A. (2018). Hsp70 at the membrane: driving protein translocation. *BMC biology* 16, 11.
11. Mai, N., Chrzanowska-Lightowlers, Z.M., and Lightowlers, R.N. (2017). The process of mammalian mitochondrial protein synthesis. *Cell and tissue research* 367, 5-20.

2. Mitochondrial targeting sequence (MTS)¹²⁻¹⁸

12. Edman, P., and Begg, G. (1967). A protein sequenator. *European journal of biochemistry* 1, 80-91.

13. von Heijne, G. (1986). Towards a comparative anatomy of N-terminal topogenic protein sequences. *Journal of molecular biology* 189, 239-242.
14. von Heijne, G. (1986). Mitochondrial targeting sequences may form amphiphilic helices. *The EMBO journal* 5, 1335-1342.
15. Yeom, J., Ju, S., Choi, Y., Paek, E., and Lee, C. (2017). Comprehensive analysis of human protein N-termini enables assessment of various protein forms. *Scientific reports* 7, 6599.
16. Yamamoto, H., Itoh, N., Kawano, S., Yatsukawa, Y., Momose, T., Makio, T., Matsunaga, M., Yokota, M., Esaki, M., Shodai, T., et al. (2011). Dual role of the receptor Tom20 in specificity and efficiency of protein import into mitochondria. *Proceedings of the National Academy of Sciences of the United States of America* 108, 91-96.
17. Vaca Jacome, A.S., Rabilloud, T., Schaeffer-Reiss, C., Rompais, M., Ayoub, D., Lane, L., Bairoch, A., Van Dorsselaer, A., and Carapito, C. (2015). N-terminome analysis of the human mitochondrial proteome. *Proteomics* 15, 2519-2524.
18. Kalef-Ezra, E., Kotzamani, D., Zaganas, I., Ktrakili, N., Plaitakis, A., and Tokatlidis, K. (2016). Import of a major mitochondrial enzyme depends on synergy between two distinct helices of its presequence. *The Biochemical journal* 473, 2813-2829.

3. Transport-associated protein processing¹⁹⁻²²

19. Falkevall, A., Alikhani, N., Bhushan, S., Pavlov, P.F., Busch, K., Johnson, K.A., Eneqvist, T., Tjernberg, L., Ankarcrona, M., and Glaser, E. (2006). Degradation of the amyloid beta-protein by the novel mitochondrial peptidosome, PreP. *The Journal of biological chemistry* 281, 29096-29104.
20. Dudek, J. (2017). Role of Cardiolipin in Mitochondrial Signaling Pathways. *Frontiers in cell and developmental biology* 5, 90.
21. Osman, C., Merkwirth, C., and Langer, T. (2009). Prohibitins and the functional compartmentalization of mitochondrial membranes. *Journal of cell science* 122, 3823-3830.
22. Sengers, R.C., Trijbels, J.M., Willems, J.L., Daniels, O., and Stadhouders, A.M. (1975). Congenital cataract and mitochondrial myopathy of skeletal and heart muscle associated with lactic acidosis after exercise. *The Journal of pediatrics* 86, 873-880.

Conditions linked to rare genetic defects in mitochondrial import machinery or its clients.

1. Pathological genetic variation affecting the TIM and OXA complexes
 2. The special case of 3-MGA-uria²³⁻³⁰
23. Su, B., and Ryan, R.O. (2014). Metabolic biology of 3-methylglutaconic acid-uria: a new perspective. *Journal of inherited metabolic disease* 37, 359-368.
 24. Claypool, S.M., and Koehler, C.M. (2012). The complexity of cardiolipin in health and disease. *Trends in biochemical sciences* 37, 32-41.
 25. Wortmann, S.B., Vaz, F.M., Gardeitchik, T., Vissers, L.E., Renkema, G.H., Schuurs-Hoeijmakers, J.H., Kulik, W., Lammens, M., Christin, C., Kluijtmans, L.A., et al. (2012). Mutations in the phospholipid remodeling gene SERAC1 impair mitochondrial function and intracellular cholesterol trafficking and cause dystonia and deafness. *Nature genetics* 44, 797-802.
 26. Richter-Dennerlein, R., Korwitz, A., Haag, M., Tatsuta, T., Dargazanli, S., Baker, M., Decker, T., Lamkemeyer, T., Rugarli, E.I., and Langer, T. (2014). DNAJC19, a mitochondrial cochaperone associated with cardiomyopathy, forms a complex with prohibitins to regulate cardiolipin remodeling. *Cell metabolism* 20, 158-171.

27. Malhotra, K., Modak, A., Nangia, S., Daman, T.H., Günsel, U., Robinson, V.L., Mokranjac, D., May, E.R., and Alder, N.N. (2017). Cardiolipin mediates membrane and channel interactions of the mitochondrial TIM23 protein import complex receptor Tim50. *Science advances* 3, e1700532.
28. Kovacs-Nagy, R., Morin, G., Nouri, M.A., Brandau, O., Saadi, N.W., Nouri, M.A., van den Broek, F., Prokisch, H., Mayr, J.A., and Wortmann, S.B. (2018). HTRA2 Defect: A Recognizable Inborn Error of Metabolism with 3-Methylglutaconic Aciduria as Discriminating Feature Characterized by Neonatal Movement Disorder and Epilepsy-Report of 11 Patients. *Neuropediatrics*.
29. Liu, R., and Chan, D.C. (2017). OPA1 and cardiolipin team up for mitochondrial fusion. *Nature cell biology* 19, 760-762.
30. Mayr, J.A., Haack, T.B., Graf, E., Zimmermann, F.A., Wieland, T., Haberberger, B., Superti-Furga, A., Kirschner, J., Steinmann, B., Baumgartner, M.R., et al. (2012). Lack of the mitochondrial protein acylglycerol kinase causes Sengers syndrome. *American journal of human genetics* 90, 314-320.

3. Pathological genetic variation in the MIA pathway ³¹⁻³⁴

31. Chia, R., Chio, A., and Traynor, B.J. (2018). Novel genes associated with amyotrophic lateral sclerosis: diagnostic and clinical implications. *The Lancet Neurology* 17, 94-102.
32. Modjtahedi, N., Tokatlidis, K., Dessen, P., and Kroemer, G. (2016). Mitochondrial Proteins Containing Coiled-Coil-Helix-Coiled-Coil-Helix (CHCH) Domains in Health and Disease. *Trends in biochemical sciences* 41, 245-260.
33. Schorr, S., and van der Laan, M. (2018). Integrative functions of the mitochondrial contact site and cristae organizing system. *Seminars in cell & developmental biology* 76, 191-200.
34. Daithankar, V.N., Schaefer, S.A., Dong, M., Bahnsen, B.J., and Thorpe, C. (2010). Structure of the human sulfhydryl oxidase augments liver regeneration and characterization of a human mutation causing an autosomal recessive myopathy. *Biochemistry* 49, 6737-6745.

4. Pathological genetic variation affecting the MTS ³⁵⁻³⁷

35. Patel, K.P., O'Brien, T.W., Subramony, S.H., Shuster, J., and Stacpoole, P.W. (2012). The spectrum of pyruvate dehydrogenase complex deficiency: clinical, biochemical and genetic features in 371 patients. *Molecular genetics and metabolism* 105, 34-43.
36. Rosenblum, J.S., Gilula, N.B., and Lerner, R.A. (1996). On signal sequence polymorphisms and diseases of distribution. *Proceedings of the National Academy of Sciences of the United States of America* 93, 4471-4473.
37. Lek, M., Karczewski, K.J., Minikel, E.V., Samocha, K.E., Banks, E., Fennell, T., O'Donnell-Luria, A.H., Ware, J.S., Hill, A.J., Cummings, B.B., et al. (2016). Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 536, 285-291.

5. Pathological genetic variation affecting mitochondrial proteases ³⁸⁻⁴³

38. Bagli, E., Zikou, A.K., Agnantis, N., and Kitsos, G. (2017). Mitochondrial Membrane Dynamics and Inherited Optic Neuropathies. *In vivo (Athens, Greece)* 31, 511-525.
39. Gaier, E.D., Boudreault, K., Nakata, I., Janessian, M., Skidd, P., DelBono, E., Allen, K.F., Pasquale, L.R., Place, E., Cestari, D.M., et al. (2017). Diagnostic genetic testing for patients with bilateral optic neuropathy and comparison of clinical features according to OPA1 mutation status. *Molecular vision* 23, 548-560.
40. Strauss, K.A., Jinks, R.N., Puffenberger, E.G., Venkatesh, S., Singh, K., Cheng, I., Mikita, N., Thilagavathi, J., Lee, J., Sarafianos, S., et al. (2015). CODAS syndrome is associated with

- mutations of LONP1, encoding mitochondrial AAA+ Lon protease. *American journal of human genetics* 96, 121-135.
41. Rampello, A.J., and Glynn, S.E. (2017). Identification of a Degradation Signal Sequence within Substrates of the Mitochondrial i-AAA Protease. *Journal of molecular biology* 429, 873-885.
 42. Ding, B., Martin, D.W., Rampello, A.J., and Glynn, S.E. (2018). Dissecting Substrate Specificities of the Mitochondrial AFG3L2 Protease. *Biochemistry* 57, 4225-4235.
 43. Gersch, M., Stahl, M., Poreba, M., Dahmen, M., Dziedzic, A., Drag, M., and Sieber, S.A. (2016). Barrel-shaped ClpP Proteases Display Attenuated Cleavage Specificities. *ACS chemical biology* 11, 389-399.

Rare pathologic genetic variations leading to protein mistargeting to the mitochondria ⁴⁴⁻

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44. Kunze, M., and Berger, J. (2015). The similarity between N-terminal targeting signals for protein import into different organelles and its evolutionary relevance. *Frontiers in physiology* 6, 259.
45. Montioli, R., Fargue, S., Lewin, J., Zamparelli, C., Danpure, C.J., Borri Voltattorni, C., and Cellini, B. (2012). The N-terminal extension is essential for the formation of the active dimeric structure of liver peroxisomal alanine:glyoxylate aminotransferase. *The international journal of biochemistry & cell biology* 44, 536-546.
46. Nakai, M., Endo, T., Hase, T., and Matsubara, H. (1993). Intramitochondrial protein sorting. Isolation and characterization of the yeast MSP1 gene which belongs to a novel family of putative ATPases. *The Journal of biological chemistry* 268, 24262-24269.
47. Wolf, N.I., Zschocke, J., Jakobs, C., Rating, D., and Hoffmann, G.F. (2018). ATAD1 encephalopathy and stiff baby syndrome: a recognizable clinical presentation. *Brain : a journal of neurology* 141, e49.
48. Ren, H., Fu, K., Mu, C., Zhen, X., and Wang, G. (2012). L166P mutant DJ-1 promotes cell death by dissociating Bax from mitochondrial Bcl-XL. *Molecular neurodegeneration* 7, 40.
49. Chen, Y.C., Umanah, G.K., Dephoure, N., Andrabi, S.A., Gygi, S.P., Dawson, T.M., Dawson, V.L., and Rutter, J. (2014). Msp1/ATAD1 maintains mitochondrial function by facilitating the degradation of mislocalized tail-anchored proteins. *The EMBO journal* 33, 1548-1564.
50. Purdue, P.E., Allsop, J., Isaya, G., Rosenberg, L.E., and Danpure, C.J. (1991). Mistargeting of peroxisomal L-alanine:glyoxylate aminotransferase to mitochondria in primary hyperoxaluria patients depends upon activation of a cryptic mitochondrial targeting sequence by a point mutation. *Proceedings of the National Academy of Sciences of the United States of America* 88, 10900-10904.

Deregulated mitochondrial import in cancer ⁵¹⁻⁵⁸

51. Vander Heiden, M.G., Cantley, L.C., and Thompson, C.B. (2009). Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science (New York, NY)* 324, 1029-1033.
52. Madamba, S.M., Damri, K.N., Dejean, L.M., and Peixoto, P.M. (2015). Mitochondrial Ion Channels in Cancer Transformation. *Frontiers in oncology* 5, 120.
53. Sotgia, F., Whitaker-Menezes, D., Martinez-Outschoorn, U.E., Salem, A.F., Tsirigos, A., Lamb, R., Sneddon, S., Hult, J., Howell, A., and Lisanti, M.P. (2012). Mitochondria "fuel" breast cancer metabolism: fifteen markers of mitochondrial biogenesis label epithelial cancer cells, but are excluded from adjacent stromal cells. *Cell cycle (Georgetown, Tex)* 11, 4390-4401.

54. Yang, J., Staples, O., Thomas, L.W., Briston, T., Robson, M., Poon, E., Simoes, M.L., El-Emir, E., Buffa, F.M., Ahmed, A., et al. (2012). Human CHCHD4 mitochondrial proteins regulate cellular oxygen consumption rate and metabolism and provide a critical role in hypoxia signaling and tumor progression. *The Journal of clinical investigation* 122, 600-611.
55. Chatre, L., Fernandes, J., Michel, V., Fiette, L., Ave, P., Arena, G., Jain, U., Haas, R., Wang, T.C., Ricchetti, M., et al. (2017). Helicobacter pylori targets mitochondrial import and components of mitochondrial DNA replication machinery through an alternative VacA-dependent and a VacA-independent mechanisms. *Scientific reports* 7, 15901.
56. Arena, G., Cisse, M.Y., Pyrdziak, S., Chatre, L., Riscal, R., Fuentes, M., Arnold, J.J., Kastner, M., Gayte, L., Bertrand-Gaday, C., et al. (2018). Mitochondrial MDM2 Regulates Respiratory Complex I Activity Independently of p53. *Molecular cell* 69, 594-609.e598.
57. Quiros, P.M., Espanol, Y., Acin-Perez, R., Rodriguez, F., Barcena, C., Watanabe, K., Calvo, E., Loureiro, M., Fernandez-Garcia, M.S., Fueyo, A., et al. (2014). ATP-dependent Lon protease controls tumor bioenergetics by reprogramming mitochondrial activity. *Cell reports* 8, 542-556.
58. Luo, B., Wang, M., Hou, N., Hu, X., Jia, G., Qin, X., Zuo, X., Liu, Y., Luo, K., Song, W., et al. (2016). ATP-Dependent Lon Protease Contributes to Helicobacter pylori-Induced Gastric Carcinogenesis. *Neoplasia (New York, NY)* 18, 242-252.

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37. Lek, M., Karczewski, K.J., Minikel, E.V., Samocha, K.E., Banks, E., Fennell, T., O'Donnell-Luria, A.H., Ware, J.S., Hill, A.J., Cummings, B.B., et al. (2016). Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 536, 285-291.
59. Allot, A., Peng, Y., Wei, C.H., Lee, K., Phan, L., and Lu, Z. (2018). LitVar: a semantic search engine for linking genomic variant data in PubMed and PMC. *Nucleic acids research* 46, W530-w536.
60. Bentley, A.R., Callier, S., and Rotimi, C.N. (2017). Diversity and inclusion in genomic research: why the uneven progress? *Journal of community genetics* 8, 255-266.
61. Falk, M.J., Shen, L., Gonzalez, M., Leipzig, J., Lott, M.T., Stassen, A.P., Diroma, M.A., Navarro-Gomez, D., Yeske, P., Bai, R., et al. (2015). Mitochondrial Disease Sequence Data Resource (MSeqDR): a global grass-roots consortium to facilitate deposition, curation, annotation, and integrated analysis of genomic data for the mitochondrial disease clinical and research communities. *Molecular genetics and metabolism* 114, 388-396.
62. Hirano, M., Emmanuele, V., and Quinzii, C.M. (2018). Emerging therapies for mitochondrial diseases. *Essays in biochemistry* 62, 467-481.
63. Kenney, M.C., Chwa, M., Atilano, S.R., Falatoonzadeh, P., Ramirez, C., Malik, D., Tarek, M., Del Carpio, J.C., Nesburn, A.B., Boyer, D.S., et al. (2014). Molecular and bioenergetic differences between cells with African versus European inherited mitochondrial DNA haplogroups: implications for population susceptibility to diseases. *Biochimica et biophysica acta* 1842, 208-219.
64. Landrum, M.J., Lee, J.M., Riley, G.R., Jang, W., Rubinstein, W.S., Church, D.M., and Maglott, D.R. (2014). ClinVar: public archive of relationships among sequence variation and human phenotype. *Nucleic acids research* 42, D980-985.
65. Nambot, S., Gavrillov, D., Thevenon, J., Bruel, A.L., Bainbridge, M., Rio, M., Goizet, C., Rotig, A., Jaeken, J., Niu, N., et al. (2017). Further delineation of a rare recessive encephalomyopathy linked to mutations in GFER thanks to data sharing of whole exome sequencing data. *Clinical genetics* 92, 188-198.

66. Nurminen, A., Farnum, G.A., and Kaguni, L.S. (2017). Pathogenicity in POLG syndromes: DNA polymerase gamma pathogenicity prediction server and database. *BBA clinical* 7, 147-156.
67. Petrovski, S., and Goldstein, D.B. (2016). Unequal representation of genetic variation across ancestry groups creates healthcare inequality in the application of precision medicine. *Genome biology* 17, 157.
68. Vrbacky, M., Kovalcikova, J., Chawengsaksophak, K., Beck, I.M., Mracek, T., Nuskova, H., Sedmera, D., Papousek, F., Kolar, F., Sobol, M., et al. (2016). Knockout of Tmem70 alters biogenesis of ATP synthase and leads to embryonal lethality in mice. *Human molecular genetics* 25, 4674-4685.
69. Carapito, C., Kuhn, L., Karim, L., Rompais, M., Rabilloud, T., Schwenzer, H., and Sissler, M. (2017). Two proteomic methodologies for defining N-termini of mature human mitochondrial aminoacyl-tRNA synthetases. *Methods (San Diego, Calif)* 113, 111-119.
70. Zhang, C., Montooth, K.L., and Calvi, B.R. (2017). Incompatibility between mitochondrial and nuclear genomes during oogenesis results in ovarian failure and embryonic lethality. *Development (Cambridge, England)* 144, 2490-2503.
71. Crawford, N., Prendergast, D., Oehlert, J.W., Shaw, G.M., Stevenson, D.K., Rappaport, N., Sirota, M., Tishkoff, S.A., and Sondheimer, N. (2018). Divergent Patterns of Mitochondrial and Nuclear Ancestry Are Associated with the Risk for Preterm Birth. *The Journal of pediatrics* 194, 40-46.e44.

References for Supplemental Table 1 ⁷²⁻⁷⁴

72. Thompson, K., Mai, N., Olahova, M., Scialo, F., Formosa, L.E., Stroud, D.A., Garrett, M., Lax, N.Z., Robertson, F.M., Jou, C., et al. (2018). OXA1L mutations cause mitochondrial encephalopathy and a combined oxidative phosphorylation defect. *EMBO molecular medicine*.
73. Pacheu-Grau, D., Callegari, S., Emperador, S., Thompson, K., Aich, A., Topol, S.E., Spencer, E.G., McFarland, R., Ruiz-Pesini, E., Torkamani, A., et al. (2018). Mutations of the mitochondrial carrier translocase channel subunit TIM22 cause early-onset mitochondrial myopathy. *Human molecular genetics* 27, 4135-4144.
74. Brunetti, D., Torsvik, J., Dallabona, C., Teixeira, P., Sztromwasser, P., Fernandez-Vizarra, E., Cerutti, R., Reyes, A., Preziuso, C., D'Amati, G., et al. (2016). Defective PITRM1 mitochondrial peptidase is associated with Abeta amyloidotic neurodegeneration. *EMBO molecular medicine* 8, 176-190.

References for Supplemental Table 2 ^{30 75-79}

30. Mayr, J.A., Haack, T.B., Graf, E., Zimmermann, F.A., Wieland, T., Haberberger, B., Superti-Furga, A., Kirschner, J., Steinmann, B., Baumgartner, M.R., et al. (2012). Lack of the mitochondrial protein acylglycerol kinase causes Sengers syndrome. *American journal of human genetics* 90, 314-320.
75. Mayr, J.A., Havlickova, V., Zimmermann, F., Magler, I., Kaplanova, V., Jesina, P., Pecinova, A., Nuskova, H., Koch, J., Sperl, W., et al. (2010). Mitochondrial ATP synthase deficiency due to a mutation in the ATP5E gene for the F1 epsilon subunit. *Human molecular genetics* 19, 3430-3439.

76. De Meirleir, L., Seneca, S., Lissens, W., De Clercq, I., Eyskens, F., Gerlo, E., Smet, J., and Van Coster, R. (2004). Respiratory chain complex V deficiency due to a mutation in the assembly gene ATP12. *Journal of medical genetics* 41, 120-124.
77. Olahova, M., Yoon, W.H., Thompson, K., Jangam, S., Fernandez, L., Davidson, J.M., Kyle, J.E., Grove, M.E., Fisk, D.G., Kohler, J.N., et al. (2018). Biallelic Mutations in ATP5F1D, which Encodes a Subunit of ATP Synthase, Cause a Metabolic Disorder. *American journal of human genetics* 102, 494-504.
78. Spiegel, R., Khayat, M., Shalev, S.A., Horovitz, Y., Mandel, H., Hershkovitz, E., Barghuti, F., Shaag, A., Saada, A., Korman, S.H., et al. (2011). TMEM70 mutations are a common cause of nuclear encoded ATP synthase assembly defect: further delineation of a new syndrome. *Journal of medical genetics* 48, 177-182.
79. Guarani, V., Jardel, C., Chretien, D., Lombes, A., Benit, P., Labasse, C., Lacene, E., Bourillon, A., Imbard, A., Benoist, J.F., et al. (2016). QIL1 mutation causes MICOS disassembly and early onset fatal mitochondrial encephalopathy with liver disease. *eLife* 5.