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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see <u>Authors & Referees</u> and the <u>Editorial Policy Checklist</u>.

Statistics

For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Cor	firmed
		The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
		A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
		The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\boxtimes		A description of all covariates tested
		A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
		A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	\boxtimes	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	\square	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
	1	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about <u>availability of computer code</u>				
Data collection	The data processing and normalization is noted in manuscript. Data is uploaded on SRA (PRJNA503740).			
Data analysis	No commercial software was used.			

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Life sciences

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable: - Accession codes, unique identifiers, or web links for publicly available datasets

- A list of figures that have associated raw data

- A description of any restrictions on data availability

Data can be found on SRA (PRJNA503740).

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>

Life sciences study design

Human research participants

Clinical data

 \boxtimes

 \boxtimes

All studies must disclose on these points even when the disclosure is negative.

Sample size	As bulk biochemical assays are involved, each experiment reports the behavior of a large number of molecules in ensemble, with molecule numbers calculable from concentrations provided. Individual variants are represented in most cases by multiple barcoded species in each library, with numbers of such species noted in the bar graphs. Each species in turn is reported only in cases where a minimum number of reads (50) is observed for the tag. Minimum reference read counts were chosen to avoid substantial stochastic variation in signal.
Data exclusions	N/A
Replication	To ensure reproducibility of the high throughput assays, parallel assessments for each primary input library were done at least twice and amplified independently, with consistent results in each case. Additional verification comes from the parallel use of two library types (RVL and TypeII), which gave equivalent results (for the in vitro assessments using Cas9). Validating the high throughput assays, top hits from were further verified with native agarose gel migration assays.
Randomization	Randomization was not part of the experimental design for this study.
Blinding	Blinding was not part of the experimental design for this study.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems Methods n/a Involved in the study n/a Involved in the study \boxtimes \boxtimes Antibodies ChIP-seq \boxtimes Eukaryotic cell lines \boxtimes Flow cytometry Palaeontology \boxtimes MRI-based neuroimaging \boxtimes Animals and other organisms

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