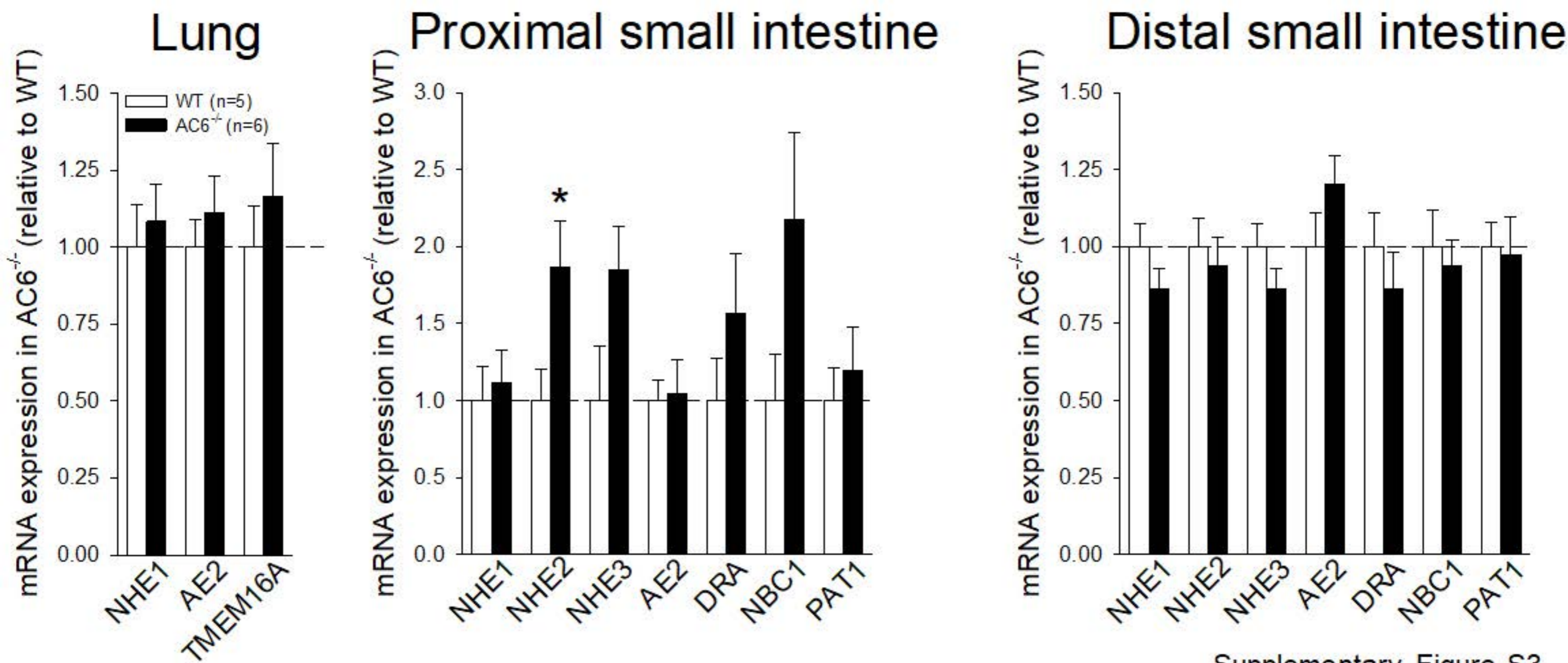


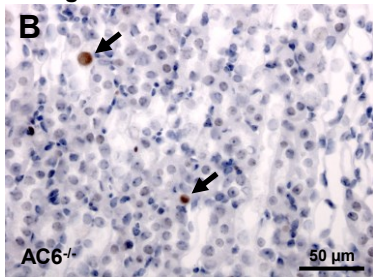
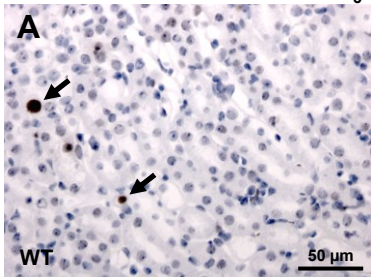
Supplementary Figure S2

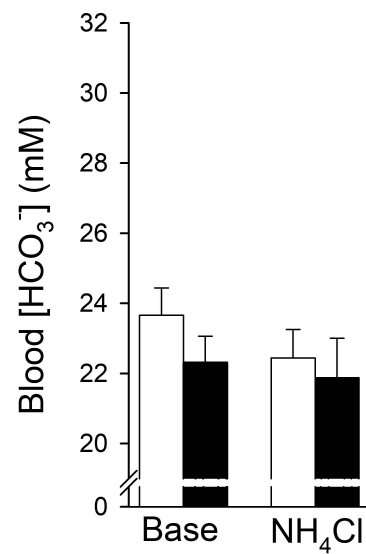
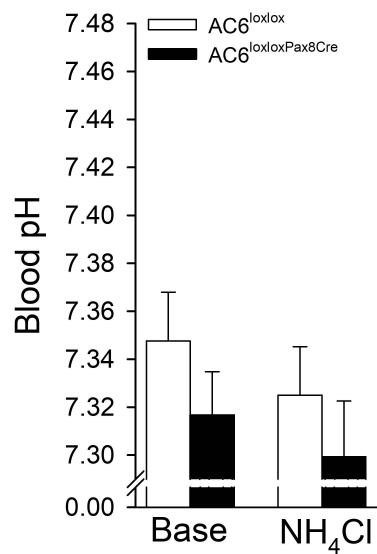
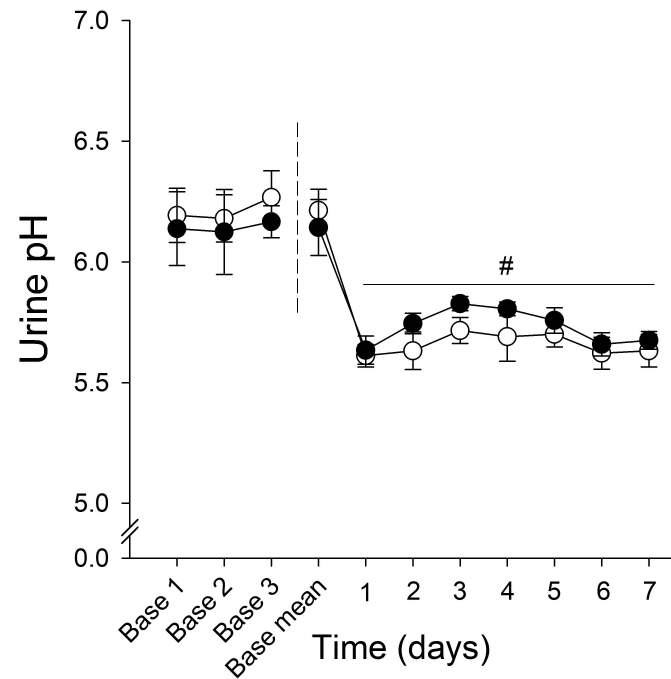
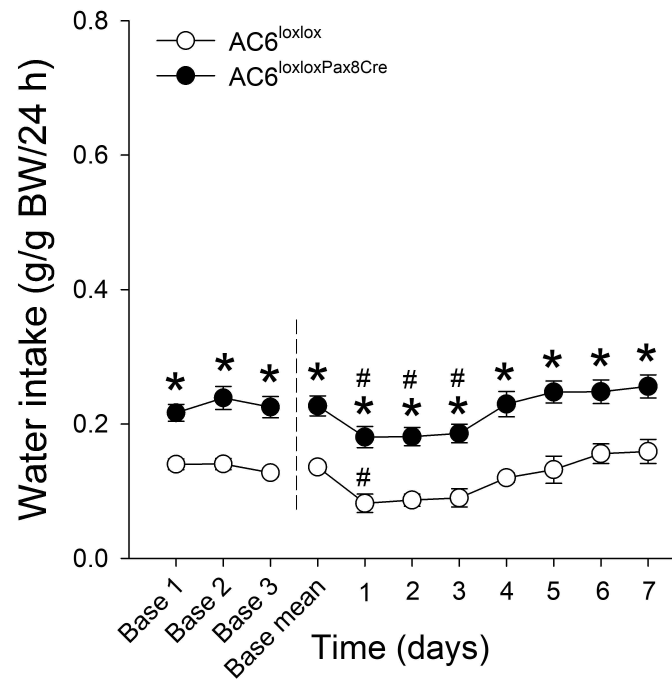


Supplementary Figure S3

HCO₃⁻-loading

PCNA





Supplementary Figure S5

Supplementary Figure Legends

Supplementary Figure S1. AC6^{-/-} mice have normal circadian rhythm but elevated O₂ consumption and CO₂ production. Consistent with our previously published data (1) AC6^{-/-} mice show increased fluid intake. The experiments in PhenoMaster[®] metabolic cages identified a significantly higher water intake during their inactive and active period. No differences in food intake were observed between genotypes. Both, O₂ consumption and CO₂ production were significantly higher during the inactive and active period in AC6^{-/-} versus WT mice. *n*=6/genotype, **P*<0.05 versus WT.

Supplementary Figure S2. AC6^{-/-} mice have increased energy expenditure. Experiments in PhenoMaster[®] metabolic cages identified that activity in the XY-axis was significantly higher in AC6^{-/-} during the inactive period. Activity in the Z-axis is only a fraction compared to the XY activity. Whereas 24-hour Z-axis activity was not different between genotypes, Z-activity in AC6^{-/-} mice was elevated and reduced during the inactive and active period, respectively. AC6^{-/-} mice have, independent of the active/inactive period, increased energy expenditure compared to WT mice. *n*=6/genotype, **P*<0.05 versus WT.

Supplementary Figure S3. mRNA expression of various transporters and channels in lung and proximal/distal small intestine of WT (*n*=5) and AC6^{-/-} (*n*=6) mice. No differences between genotypes were observed in lung tissue mRNA expression. The proximal small intestine of AC6^{-/-} mice had significantly higher expression of NHE2, and tendencies for higher NHE3 and NBC1 expression. No significant differences in transporter profiles were observed between genotypes in the distal small intestine. **P*<0.05 versus WT. Na⁺-H⁺ exchanger 1, NHE1; Na⁺-H⁺ exchanger 2, NHE2; Na⁺-H⁺ exchanger 3, NHE3, anion exchanger 2, AE2; down regulated in adenoma, DRA; Na⁺-HCO₃⁻ cotransporter 1, NBC1; putative anion transporter 1, PAT1; Ca²⁺-activated Cl⁻ channel, TMEM16A.

Supplementary Figure S4. AC6^{-/-} mice do not present more PCNA-positive cells HCO₃⁻ after 8 days of HCO₃⁻ challenge. Representative light microscopy images of 2-μm kidney sections from AC6^{-/-} and WT mice subjected to HCO₃⁻ challenge for 8 days. Immunolabeling was performed using an antibody targeting PCNA. (A and B) No difference was found between the numbers of cells labeled for PCNA in AC6^{-/-} compared to WT mice. *n*=6/genotype. Arrows indicate PCNA labeled cells.

Supplementary Figure S5. No differences were observed between AC6^{loxlox} and AC6^{loxloxPax8Cre} mice in response to 7 days of NH₄Cl challenge. Because daily water intake is double in AC6^{loxloxPax8Cre} mice, NH₄Cl was supplied to this genotype at a 50% lower concentration compared to AC6^{loxlox} mice (0.14 M in AC6^{loxloxPax8Cre} versus 0.28 M in

AC6^{loxlox} mice). Fluid intake was slightly but significantly reduced on day 1 in AC6^{loxlox} mice and on days 1-3 in AC6^{loxlox}Pax8^{Cre} mice. Urinary pH was lowered to a similar extent during NH₄Cl challenge (n=7-8/genotype). Blood pH and HCO₃⁻ concentrations were not significantly different between genotypes and did not significantly change after 7 days NH₄Cl challenge (n=7-8). **P*<0.05 versus AC6^{loxlox}, #*P*<0.05 baseline mean versus NH₄Cl challenge in both genotypes. Statistical comparisons on left and right side of dashed line in panel were performed separately using two-way repeated measurements ANOVAs. Values indicate mean ± S.E.M. Base, Baseline.

1. Rieg T, Tang T, Murray F, Schroth J, Insel PA, Fenton RA, et al. Adenylate cyclase 6 determines cAMP formation and aquaporin-2 phosphorylation and trafficking in inner medulla. *J Am Soc Nephrol.* 2010;21(12):2059-68.