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## 1. General Information

**Methods and Reagents.** All reactions were performed in oven-dried flasks fitted with septa under a positive pressure of nitrogen atmosphere. Organic solutions were concentrated using a Buchi rotary evaporator below 40 °C at 25 torr. Analytical thin-layer chromatography was routinely utilized to monitor the progress of the reactions and performed using pre-coated glass plates with 230-400 mesh silica gel impregnated with a fluorescent indicator (250 nm). Visualization was then achieved using UV light, iodine, or ceric ammonium molybdate. Flash column chromatography was performed using 40-63  $\mu\text{m}$  silica gel (SiliaFlash F60 from Silicycle). Dry solvents were obtained from a SG Waters solvent system utilizing activated alumina columns under an argon pressure. All other commercial reagents were used as received from Sigma Aldrich, Alfa Aesar, Acros Organics, TCI, and Combi-Blocks, unless otherwise noted.

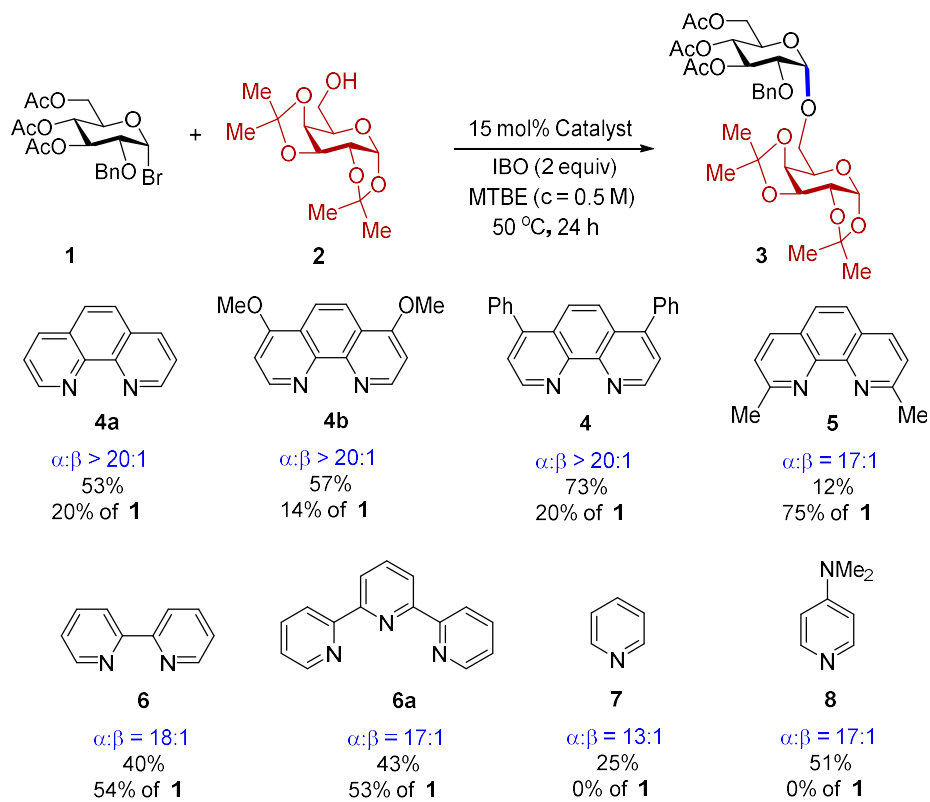
**Instrumentation.** All new compounds were characterized by Nuclear Magnetic Resonance (NMR) spectroscopy and High-Resolution Mass spectrometry (HRMS). All  $^1\text{H}$  NMR spectra were recorded on either Bruker 400 or 500 MHz spectrometers or DRX-400 (400 MHz) spectrometer. All  $^{13}\text{C}$  NMR spectra were recorded on either Bruker 100 or 125 MHz spectrometer or DRX-400 (100 MHz) spectrometer. All  $^{19}\text{F}$  NMR spectra were recorded on DRX-400 (376 MHz) spectrometer. Chemical shifts are expressed in parts per million ( $\delta$  scale) downfield from tetramethylsilane and are referenced to the residual proton in the NMR solvent ( $\text{CDCl}_3$ :  $\delta$  7.26 ppm,  $\delta$  77.00 ppm). Data are presented as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and bs = broad singlet), integration, and coupling constant in hertz (Hz).

High resolution (ESI-TOF) mass spectrometry was performed at the University of Iowa and at Wayne State University.

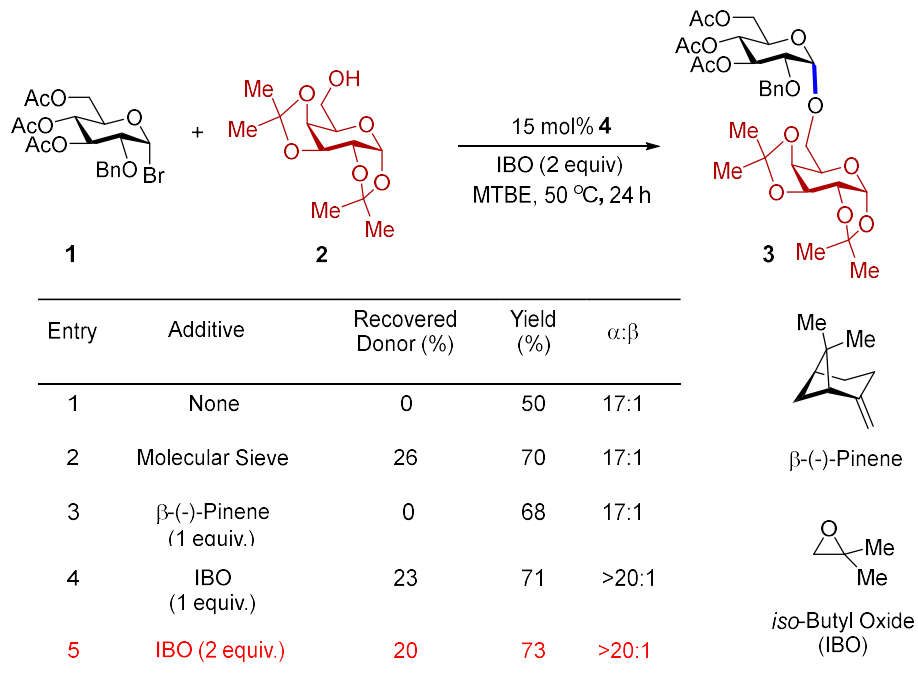
Abbreviations were used as follows: MTBE (*tert*-butyl methyl ether), IBO (isobutylene oxide) and DFT (density functional theory).

## 2. Optimization Studies

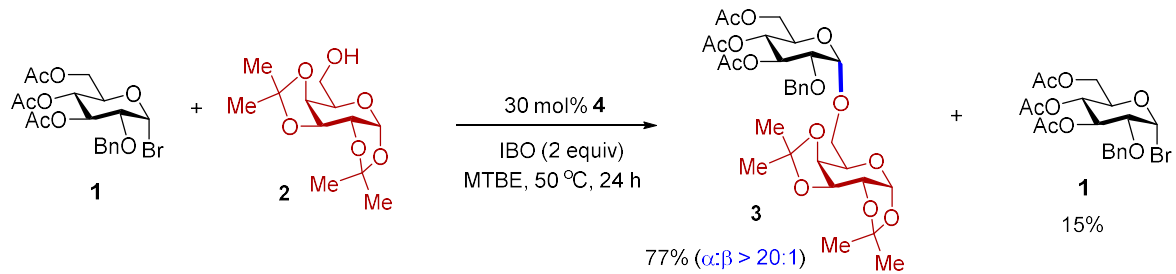
Figure S1. Screening of Small-Molecule Catalysts



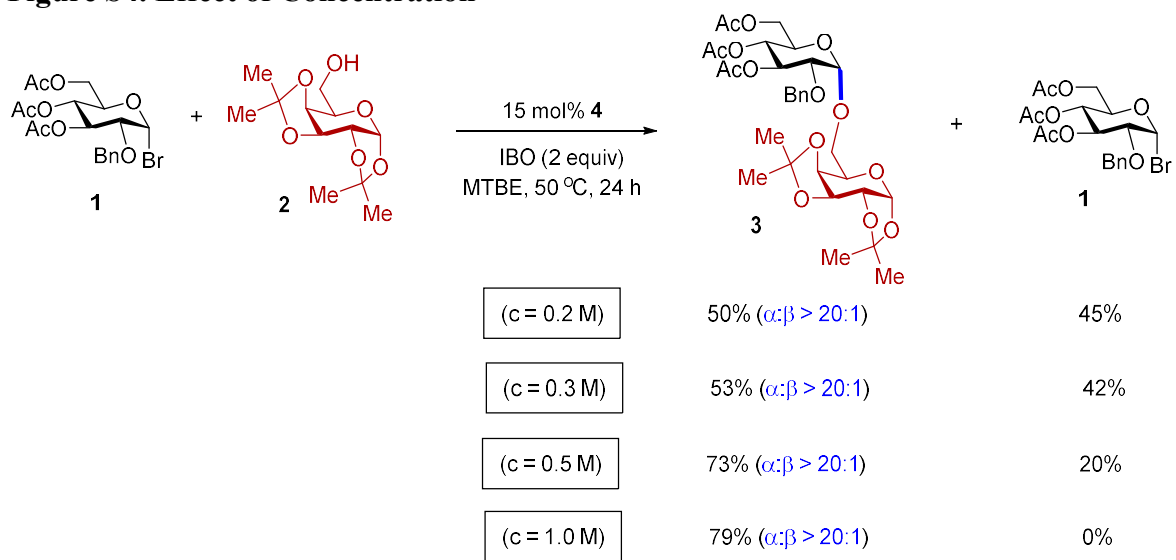
**Figure S2. Screening of Hydrogen Bromide (HBr) Scavengers**



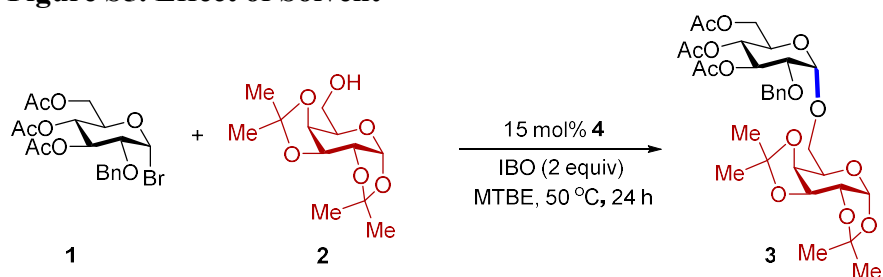
**Figure S3. Increasing Catalyst Loading**



**Figure S4. Effect of Concentration**

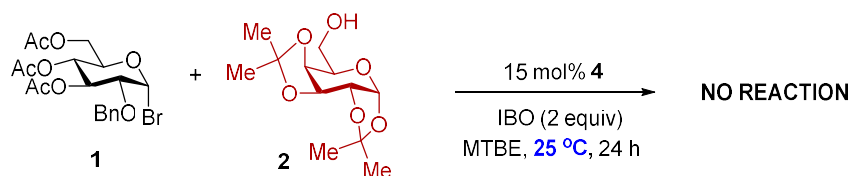


**Figure S5. Effect of Solvent**



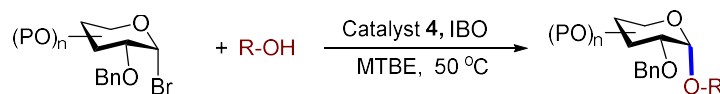
Entry	Solvent	Recovered Donor <b>1</b> (%)	<b>3</b> Yield (%)	$\alpha:\beta$
1	MTBE	20	73	>20:1
2	CPME	13	72	>20:1
3	Toluene	30	63	>20:1
4	DCE	-	71	>20:1

**Figure S6. Effect of Reaction Temperature**



### 3. Phenanthroline-Catalyzed Glycosylation Reactions

#### General Procedure



#### Standard Conditions A:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.2 mmol, 1.0 equiv), alcohol (0.4 mmol, 3.0 equiv), catalyst **4** (0.03 mmol, 15 mol%), IBO (0.4 mmol, 2.0 equiv.) and MTBE (0.4 mL). The resulting solution was stirred at 50 °C for 24 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 5/1→3/1) to give the desired product.

#### Standard Conditions B:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.4 mmol, 2.0 equiv), alcohol (0.2 mmol, 1.0 equiv), catalyst **4** (0.06 mmol, 30 mol%), IBO (0.4 mmol, 2.0 equiv.) and MTBE (0.2 mL). The resulting solution was stirred at 50 °C for 48 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 5/1→3/1) to give the desired product.

#### Standard Conditions B':

A 10 mL Schlenk flask was charged with glycosyl bromide (0.4 mmol, 2.0 equiv), alcohol (0.2 mmol, 1.0 equiv), catalyst **4** (0.06 mmol, 30 mol%), IBO (0.4 mmol, 2.0 equiv.) and MTBE (0.4 mL). The resulting solution was stirred at 50 °C for 24 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 9/1→4/1) to give the desired product.

#### Standard Conditions C:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.6 mmol, 3.0 equiv), alcohol (0.2 mmol, 1.0 equiv), catalyst **4** (0.1 mmol, 50 mol%), IBO (0.6 mmol, 3.0 equiv.) and MTBE (0.2 mL). The resulting solution was stirred at 50 °C for 48 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 5/1→3/1) to give the desired product.

#### Standard Conditions D:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.2 mmol, 2.0 equiv), alcohol (0.1 mmol, 1.0 equiv), catalyst **4** (0.02 mmol, 20 mol%), IBO (0.2 mmol, 2.0 equiv.) and MTBE (0.2 mL). The resulting solution was stirred at 25 °C for 24 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 9/1→4/1) to give the desired product.

#### Standard Conditions D':

A 10 mL Schlenk flask was charged with glycosyl bromide (0.2 mmol, 2.0 equiv), alcohol (0.1 mmol, 1.0 equiv), catalyst **4** (0.02 mmol, 20 mol%), IBO (0.2 mmol, 2.0 equiv.) and MTBE (0.2 mL). The resulting solution was stirred at 25 °C for 48 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 9/1→4/1) to give the desired product.

#### Standard Conditions E:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.2 mmol, 1.0 equiv), alcohol (0.6 mmol, 3.0 equiv), catalyst **4** (0.04 mmol, 20 mol%), IBO (0.4 mmol, 2.0 equiv.) and MTBE

(0.4 mL). The resulting solution was stirred at 25 °C for 24 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 9/1→4/1) to give the desired product.

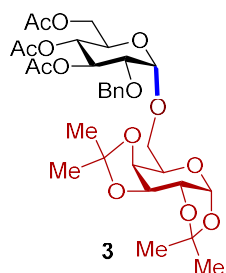
#### Standard Conditions F:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.2 mmol, 1.0 equiv), alcohol (0.6 mmol, 3.0 equiv), catalyst **4** (0.04 mmol, 20 mol%), IBO (0.4 mmol, 2.0 equiv.) and MTBE (0.4 mL). The resulting solution was stirred at 50 °C for 24 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 9/1→4/1) to give the desired product.

#### Standard Conditions G:

A 10 mL Schlenk flask was charged with glycosyl bromide (0.22 mmol, 1.1 equiv), alcohol (0.2 mmol, 1.0 equiv), catalyst **4** (0.03 mmol, 15 mol%), IBO (0.4 mmol, 2.0 equiv.) and MTBE (0.4 mL). The resulting solution was stirred at 50 °C for 24 h, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 33/1→9/1) to give the desired product.

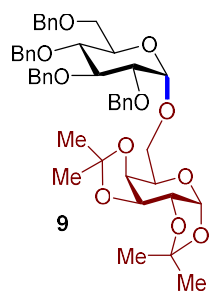
The  $\alpha/\beta$  ratio of the desired products were determined by  $^1\text{H}$  NMR analysis based on the ratio of the anomeric protons of both  $\alpha$ - and  $\beta$ -anomers. When the anomeric protons are overlapped, other protons of both anomers were analyzed. In some cases, we utilized prep-TLC to separate the  $\alpha$ -anomer from the  $\beta$ -anomer so that we can accurately determine the  $\alpha/\beta$  ratio of the mixture.



Conditions A: 73% (117 mg),  $\alpha:\beta >20:1$

$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **3** has been reported.<sup>1</sup>

**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.30-7.27 (m, 5 H), 5.49 (d,  $J$  = 5.2 Hz, 1 H), 5.43 (t,  $J$  = 10.0 Hz, 1 H), 5.00-4.90 (m, 2 H), 4.70-4.55 (m, 3 H), 4.34-4.28 (m, 3 H), 4.12-4.06 (m, 1 H), 4.04-4.00 (m, 2 H), 3.80-3.72 (m, 2 H), 3.55 (dd,  $J$  = 10.0, 3.6 Hz, 1 H), 2.07 (s, 3 H), 2.01 (s, 3 H), 2.00 (s, 3 H), 1.56 (s, 3 H), 1.43 (s, 3 H), 1.33 (s, 3 H), 1.28 (s, 3 H).  $^1\text{H}$  NMR matches with the literature report.<sup>1</sup>

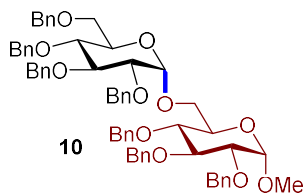


Conditions D: 95% (74.6 mg),  $\alpha:\beta = 14:1$



$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **9** has been reported.<sup>2</sup>

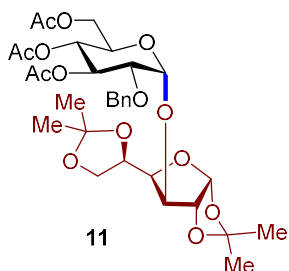
**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.40 – 7.09 (m, 20H), 5.53 (d,  $J$  = 5.0 Hz, 1H), 4.99 (m, 2H), 4.82 (m, 2H), 4.73 (m, 2H), 4.66 – 4.57 (m, 2H), 4.48 (m, 2H), 4.40 – 4.29 (m, 2H), 4.09 – 3.96 (m, 2H), 3.88 – 3.56 (m, 7H), 1.54 (s, 3H), 1.46 (s, 3H), 1.35 (s, 3H), 1.34 (s, 3H).  $^1\text{H}$  NMR matches with the literature report.<sup>2</sup>



Conditions E: 63% (124.2 mg),  $\alpha:\beta$  = 14:1

$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **10** has been reported.<sup>2</sup>

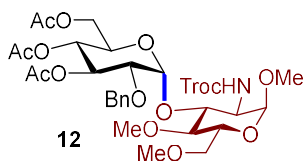
**$^1\text{H}$  NMR (400 MHz, CDCl<sub>3</sub>):**  $\delta$  7.44 – 7.16 (m, 35H), 5.08 – 4.97 (m, 4H), 4.93 – 4.83 (m, 3H), 4.81 – 4.70 (m, 4H), 4.67 – 4.61 (m, 3H), 4.56 – 4.46 (m, 2H), 4.10 – 4.01 (m, 2H), 3.93 – 3.83 (m, 3H), 3.82 – 3.66 (m, 4H), 3.65 – 3.58 (m, 2H), 3.52 (dd,  $J$  = 9.6, 3.6 Hz, 1H), 3.43 (s, 3H).  $^1\text{H}$  NMR matches with the literature report.<sup>2</sup>



Conditions B: 63% (100 mg),  $\alpha$  only

$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **11** has been reported.<sup>3</sup>

**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.38-7.22 (m, 5 H), 5.94 (d,  $J$  = 3.6 Hz, 1 H), 5.39 (t,  $J$  = 10.0 Hz, 1 H), 5.31 (d,  $J$  = 3.6 Hz, 1 H), 4.92 (t,  $J$  = 10.0 Hz, 1 H), 4.71 (d,  $J$  = 12.0 Hz, 1 H), 4.60-4.52 (m, 2 H), 4.47-4.41 (m, 1 H), 4.26-4.4.02 (m, 2 H), 4.13-3.97 (m, 5 H), 3.57 (dd,  $J$  = 10.0, 3.6 Hz, 1 H), 2.09 (s, 3 H), 2.02 (s, 3 H), 1.97 (s, 3 H), 1.49 (s, 3 H), 1.41 (s, 3 H), 1.32 (s, 3 H), 1.24 (s, 3 H).  $^1\text{H}$  NMR matches with the literature report.<sup>3</sup>



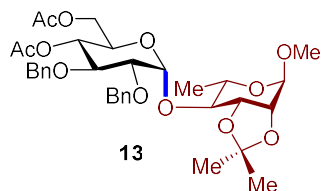
Conditions B: 50% (87 mg),  $\alpha$  only

**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.31-7.27 (m, 5 H), 5.47-5.35 (m, 3 H), 4.91 (t,  $J$  = 10.0 Hz, 1 H), 4.93 (d,  $J$  = 12.0 Hz, 1 H), 4.77-4.60 (m, 4 H), 4.33-4.19 (m, 2 H), 4.08 (dd,  $J$  = 12.0, 2.0 Hz,

1 H), 3.98-3.90 (m, 1 H), 3.85 (t,  $J = 10.0$  Hz, 1 H), 3.65 (dd,  $J = 10.0, 4.0$  Hz, 1 H), 3.62-3.59 (m, 3 H), 3.57-3.40 (m, 1 H), 3.39 (s, 3 H), 3.35 (s, 6 H), 2.10 (s, 3 H), 2.01 (s, 3 H), 1.91 (s, 3 H).

$^{13}\text{C}$  NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta = 170.3, 169.8, 154.0, 137.6, 128.4, 128.2, 127.8, 98.3, 98.2, 95.3, 90.9, 80.2, 74.6, 73.7, 71.9, 71.7, 70.7, 70.6, 68.9, 67.1, 62.9, 60.4, 59.1, 55.1, 54.4, 20.74, 20.71, 20.6$ .

HRMS (ESI): calc. for C<sub>31</sub>H<sub>43</sub>NO<sub>15</sub>Cl<sub>3</sub> (M+H): 774.1698; found: 774.1703.

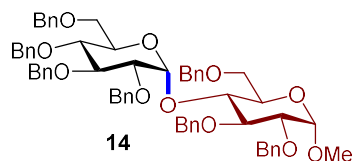


Conditions B: 73% (141 mg),  $\alpha$  only

$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.40-7.22$  (m, 10 H), 5.14 (t,  $J = 10.0$  Hz, 1 H), 4.96 (d,  $J = 4.0$  Hz, 1 H), 4.90-4.60 (m, 5 H), 4.36 (dd,  $J = 12.0, 2.8$  Hz, 1 H), 4.22-4.09 (m, 3 H), 4.00-3.88 (m, 2 H), 3.8-3.60 (m, 1 H), 3.63 (dd,  $J = 10.0, 3.2$  Hz, 1 H), 3.34 (s, 3 H), 3.38-3.30 (m, 1 H), 2.08 (s, 3 H), 1.91 (s, 3 H), 1.51 (s, 3 H), 1.33 (d,  $J = 4.4$  Hz, 3 H), 1.32 (s, 3 H).

$^{13}\text{C}$  NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta = 170.9, 169.5, 138.3, 137.5, 128.4, 128.3, 128.2, 128.0, 127.7, 127.6, 109.1, 98.3, 97.7, 81.2, 79.5, 79.4, 76.8, 75.9, 75.3, 74.3, 69.3, 67.6, 64.6, 61.6, 54.6, 28.1, 26.3, 20.74, 20.72, 17.3$ .

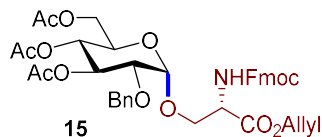
HRMS (ESI): calc. for C<sub>34</sub>H<sub>44</sub>O<sub>12</sub>Na (M+Na): 667.2730; found: 667.2735.



Conditions B': 55% (54.3mg)  $\alpha:\beta = 7:1$

$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **14** has been reported.<sup>2</sup>

$^1\text{H}$  NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.37 – 7.05 (m, 35H), 5.69 (d,  $J = 3.5$  Hz, 1H), 5.03 (d,  $J = 11.6$  Hz, 1H), 4.91 – 4.39 (m, 13H), 4.27 (d,  $J = 12.2$  Hz, 1H), 4.11 – 4.01 (m, 2H), 3.93 – 3.80 (m, 3H), 3.74 – 3.69 (m, 1H), 3.67 – 3.56 (m, 3H), 3.51 – 3.46 (m, 2H), 3.41 – 3.39 (m, 1H), 3.37 (s, 3H).  $^1\text{H}$  NMR matches with the literature report.<sup>2</sup>

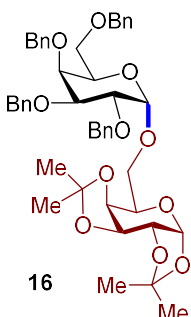


Conditions B: 79% (147 mg),  $\alpha:\beta = 20:1$

$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta = 7.76$  (d,  $J = 7.6$  Hz, 2 H), 7.63 (dd,  $J = 7.6, 3.2$  Hz, 2 H), 7.40-7.23 (m, 9 H), 6.04 (d,  $J = 8.8$  Hz, 1 H), 5.94-5.83 (m, 1 H), 5.40 (t,  $J = 9.6$  Hz, 1 H), 5.32 (d,  $J = 16.0$  Hz, 1 H), 5.24 (d,  $J = 9.6$  Hz, 1 H), 4.95 (t,  $J = 10.0$  Hz, 1 H), 4.77 (d,  $J = 3.2$  Hz, 1 H), 4.68-4.38 (m, 5 H), 4.45-4.40 (m, 2 H), 4.26-3.95 (m, 5 H), 3.90 (dd,  $J = 10.0, 3.2$  Hz, 1 H), 3.57 (dd,  $J = 10.0, 3.6$  Hz, 1 H), 2.05 (s, 3 H), 2.02 (s, 6 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 170.5, 170.0, 169.7, 169.4, 155.9, 143.70, 143.67, 141.2, 137.5, 131.4, 128.9, 129.5, 128.4, 128.1, 128.0, 127.7, 127.6, 127.0, 125.1, 119.9, 119.0, 98.1, 76.6, 72.8, 71.6, 70.2, 68.4, 67.8, 67.2, 66.4, 61.9, 54.5, 47.0, 20.7, 20.59, 20.57.

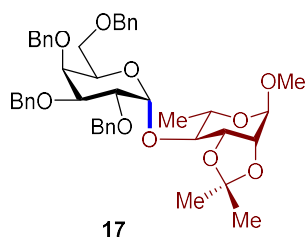
**HRMS (ESI):** calc. for C<sub>40</sub>H<sub>44</sub>NO<sub>13</sub> (M+H): 746.2813; found: 746.2810.



Conditions E: 77% (120.4mg)  $\alpha:\beta$  = 10:1

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **16** has been reported.<sup>4</sup>

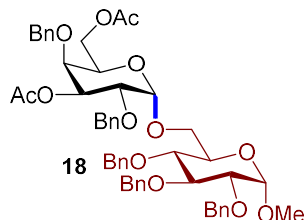
**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):**  $\delta$  7.43 – 7.10 (m, 20H), 5.53 (d,  $J$  = 5.0 Hz, 1H), 5.03 (d,  $J$  = 3.6 Hz, 1H), 4.95 (d,  $J$  = 11.4 Hz, 1H), 4.85 (d,  $J$  = 11.7 Hz, 1H), 4.78 – 4.72 (m, 3H), 4.62 – 4.56 (m, 2H), 4.52 – 4.40 (m, 2H), 4.35 – 4.29 (m, 2H), 4.10 – 3.95 (m, 5H), 3.84 – 3.73 (m, 2H), 3.62 – 3.51 (m, 2H), 1.54 (s, 3H), 1.45 (s, 3H), 1.35 – 1.29 (m, 6H). <sup>1</sup>H NMR matches with the literature report.<sup>4</sup>



Conditions F: 58% (86.4mg),  $\alpha$  only

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **17** has been reported.<sup>2</sup>

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.39 – 7.21 (m, 20H), 4.98 – 4.92 (m, 2H), 4.87 – 4.81 (m, 2H), 4.75 – 4.68 (m, 3H), 4.59 (d,  $J$  = 11.3 Hz, 1H), 4.48 (d,  $J$  = 11.9 Hz, 1H), 4.39 (d,  $J$  = 11.9 Hz, 1H), 4.24 (dd,  $J$  = 9.2, 4.5 Hz, 1H), 4.16 – 4.04 (m, 4H), 3.96 (dd,  $J$  = 10.2, 2.7 Hz, 1H), 3.77 – 3.60 (m, 2H), 3.50 (dd,  $J$  = 8.3, 4.6 Hz, 1H), 3.36 – 3.27 (m, 4H), 1.37 (s, 3H), 1.30 (d,  $J$  = 6.3 Hz, 3H), 1.25 (s, 3H). <sup>1</sup>H NMR matches with the literature report.<sup>2</sup>

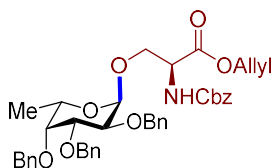


Conditions A: 85% (151 mg),  $\alpha$  only

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.41-7.21 (m, 25 H), 5.30 (dd,  $J$  = 10.8, 3.2 Hz, 1 H), 5.05 (d,  $J$  = 3.2 Hz, 1 H), 5.01 (d,  $J$  = 11.2 Hz, 1 H), 4.96 (d,  $J$  = 11.2 Hz, 1 H), 4.85 (d,  $J$  = 11.2 Hz, 1 H), 4.78-4.60 (m, 7 H), 4.53 (d,  $J$  = 11.6 Hz, 1 H), 4.17-4.00 (m, 6 H), 3.85-3.70 (m, 3 H), 3.61 (t,  $J$  = 9.6 Hz, 1 H), 3.45 (dd,  $J$  = 9.6, 3.6 Hz, 1 H), 3.41 (s, 3 H), 2.05 (s, 3 H), 1.96 (s, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 170.3, 170.2, 138.6, 138.3, 138.2, 138.0, 137.5, 128.3, 128.25, 128.21, 128.18, 128.0, 127.9, 127.82, 127.77, 127.7, 127.6, 127.5, 127.4, 97.7, 97.3, 81.9, 79.8, 77.8, 75.5, 74.98, 74.95, 74.9, 73.6, 73.1, 72.2, 70.1, 67.8, 66.1, 62.7, 55.0, 20.9, 20.6.

**HRMS (ESI):** calc. for C<sub>52</sub>H<sub>58</sub>O<sub>13</sub>Na (M+Na): 913.3775; found: 913.3787.



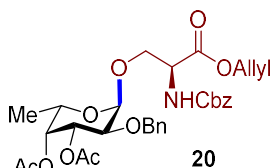
19

Conditions D: 80% (55.7 mg),  $\alpha$ : $\beta$  = 6:1

**<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):**  $\delta$  = 7.42 – 7.19 (m, 20H), 6.08 (d,  $J$  = 9.0 Hz, 1H), 5.90 – 5.80 (m, 1H), 5.29 (d,  $J$  = 17.2 Hz, 1H), 5.21 – 5.12 (m, 3H), 4.97 (d,  $J$  = 11.6 Hz, 1H), 4.85 – 4.77 (m, 2H), 4.73 – 4.53 (m, 7H), 4.20 (dd,  $J$  = 9.9, 2.2 Hz, 1H), 4.01 (dd,  $J$  = 10.1, 3.6 Hz, 1H), 3.80 (dd,  $J$  = 10.1, 2.7 Hz, 1H), 3.73 (q,  $J$  = 6.4 Hz, 1H), 3.60 – 3.52 (m, 2H), 1.07 (d,  $J$  = 6.4 Hz, 3H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 170.0, 156.2, 138.8, 138.5, 138.4, 136.3, 131.6, 128.5, 128.4, 128.3, 128.2, 128.1, 127.8, 127.6, 127.5, 118.6, 98.9, 79.0, 77.6, 76.4, 74.8, 73.3, 73.2, 69.0, 67.0, 66.8, 66.0, 54.4, 16.5.

**HRMS (ESI):** calc. for C<sub>41</sub>H<sub>45</sub>NO<sub>9</sub>Na (M+Na): 718.2987; found: 718.2967.



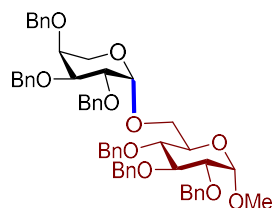
20

Conditions B: 88% (107 mg),  $\alpha$ : $\beta$  = 20:1

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.38-7.20 (m, 10 H), 6.00-5.93 (m, 2 H), 5.40-5.07 (m, 6 H), 4.74-4.52 (m, 6 H), 4.25 (dd,  $J$  = 10.0, 6.4 Hz, 1 H), 4.04-3.98 (m, 1 H), 3.81 (dd,  $J$  = 10.0, 3.6 Hz, 1 H), 3.57 (dd,  $J$  = 10.0, 3.2 Hz, 1 H), 2.13 (s, 3 H), 1.97 (s, 3 H), 1.08 (d,  $J$  = 6.4 Hz, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 170.3, 169.8, 169.5, 156.0, 137.9, 136.2, 131.4, 128.39, 128.36, 128.0, 127.8, 127.6, 119.1, 98.5, 73.4, 73.1, 71.2, 69.9, 69.1, 67.0, 66.2, 64.6, 54.3, 20.7, 20.6, 15.7.

**HRMS (ESI):** calc. for C<sub>31</sub>H<sub>37</sub>NO<sub>11</sub>Na (M+Na): 622.2264; found: 622.2265.



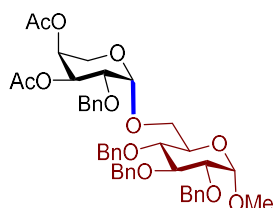
21

Conditions D': 47% (61mg),  $\alpha$  only

**$^1\text{H NMR}$  ( $\text{CDCl}_3$ , 400 MHz):**  $\delta$  = 7.42-7.22 (m, 30 H), 5.00-4.60 (m, 14 H), 4.03-3.96 (m, 2 H), 3.88-3.58 (m, 8 H), 3.46 (dd,  $J$  = 12.0, 4.0 Hz, 1 H), 3.32 (s, 3 H).

**$^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 100 MHz):**  $\delta$  = 138.82, 138.76, 138.6, 138.4, 138.3, 138.1, 128.32, 128.26, 128.24, 128.18, 127.91, 127.89, 127.8, 127.6, 127.4, 98.3, 97.8, 82.0, 80.0, 77.9, 76.3, 76.2, 75.6, 74.9, 73.9, 73.3, 72.8, 72.4, 71.6, 70.2, 66.4, 60.5, 54.9.

**HRMS (ESI):** calc. for  $\text{C}_{54}\text{H}_{58}\text{O}_{10}\text{Na}$  ( $\text{M}+\text{Na}$ ): 889.3922; found: 889.3943.



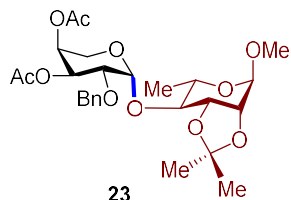
22

Conditions B: 84% (130 mg),  $\alpha$  only

**$^1\text{H NMR}$  ( $\text{CDCl}_3$ , 400 MHz):**  $\delta$  = 7.42-7.22 (m, 20 H), 5.39-5.32 (m, 2 H), 5.05 (d,  $J$  = 3.6 Hz, 1 H), 5.02 (d,  $J$  = 10.8 Hz, 1 H), 4.97 (d,  $J$  = 11.2 Hz, 1 H), 4.86 (d,  $J$  = 11.2 Hz, 1 H), 4.76 (d,  $J$  = 11.2 Hz, 1 H), 4.71-4.58 (m, 5 H), 4.01 (t,  $J$  = 10.0 Hz, 1 H), 3.96-3.53 (m, 7 H), 3.46 (dd,  $J$  = 9.6, 3.6 Hz, 1 H), 3.41 (s, 3 H), 2.13 (s, 3 H), 2.04 (s, 3 H).

**$^{13}\text{C NMR}$  ( $\text{CDCl}_3$ , 100 MHz):**  $\delta$  = 107.1, 169.9, 138.7, 138.3, 138.1, 138.0, 128.3, 128.21, 128.20, 127.9, 127.8, 127.7, 127.6, 127.5, 127.4, 127.2, 97.8, 97.7, 82.0, 79.9, 77.7, 75.5, 74.9, 73.7, 73.2, 72.2, 70.2, 69.4, 69.0, 66.2, 60.3, 55.0, 20.8, 20.7.

**HRMS (ESI):** calc. for  $\text{C}_{44}\text{H}_{50}\text{O}_{12}\text{Na}$  ( $\text{M}+\text{Na}$ ): 793.3200; found: 793.3211.



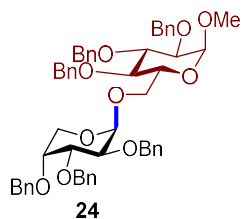
23

Conditions B: 73% (76 mg),  $\alpha$  only

**$^1\text{H NMR}$  ( $\text{CDCl}_3$ , 400 MHz):**  $\delta$  = 7.38-7.25 (m, 5 H), 5.36-5.30 (m, 2 H), 5.08 (d,  $J$  = 3.6 Hz, 1 H), 4.85 (s, 1 H), 4.72-4.63 (m, 2 H), 4.37 (d,  $J$  = 13.2 Hz, 1 H), 4.19-4.10 (m, 2 H), 3.73 (dd,  $J$  = 10.0, 6.4 Hz, 1 H), 3.77-3.55 (m, 2 H), 3.40 (dd,  $J$  = 10.0, 6.4 Hz, 1 H), 3.53 (s, 3 H), 2.11 (s, 3 H), 2.00 (s, 3 H), 1.50 (s, 3 H), 1.34 (s, 3 H), 1.31 (d,  $J$  = 6.4 Hz, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 170.2, 169.9, 137.8, 128.3, 127.83, 127.78, 109.0, 98.4, 97.7, 80.2, 76.8, 76.1, 74.0, 73.7, 69.8, 69.4, 64.8, 61.0, 54.6, 27.7, 26.3, 20.9, 20.8, 17.3.

**HRMS (ESI):** calc. for C<sub>26</sub>H<sub>36</sub>O<sub>11</sub>Na (M+Na): 547.2155; found: 547.2156.

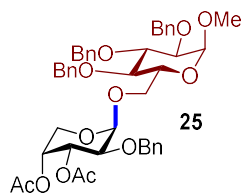


Conditions D': 48% (62mg),  $\alpha:\beta$  = 9:1

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.42-7.22 (m, 30 H), 5.00-4.60 (m, 14 H), 4.03-3.56 (m, 10 H), 3.50 (dd,  $J$  = 8.0, 4.0 Hz, 1 H), 3.32 (s, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 138.7, 138.62, 138.58, 138.4, 138.3, 138.2, 128.4, 128.31, 128.28, 128.2, 128.1, 128.794, 127.90, 127.83, 127.75, 127.7, 127.6, 127.5, 98.3, 97.9, 82.0, 80.0, 77.7, 76.2, 75.7, 74.9, 73.7, 73.4, 73.2, 72.3, 71.7, 70.0, 66.4, 60.4, 55.0.

**HRMS (ESI):** calc. for C<sub>54</sub>H<sub>58</sub>O<sub>10</sub>Na (M+Na): 889.3922; found: 889.3959.

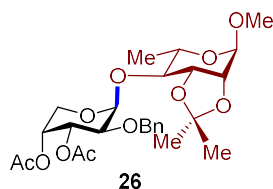


Conditions A: 83% (128 mg),  $\alpha$  only

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.43-7.20 (m, 20 H), 5.39-5.36 (m, 2 H), 5.02-4.94 (m, 2 H), 4.87-4.80 (m, 3 H), 4.75-4.60 (m, 5 H), 4.07-3.97 (m, 2 H), 3.95-3.88 (m, 2 H), 3.81 (dd,  $J$  = 10.0, 3.2 Hz, 1 H), 3.76-3.57 (m, 4 H), 3.37 (s, 3 H), 2.15 (s, 3 H), 2.03 (s, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta$  = 170.2, 170.0, 138.7, 138.5, 138.2, 137.9, 128.3, 128.22, 128.19, 128.0, 127.8, 127.7, 127.53, 127.46, 127.41, 127.38, 127.3, 98.0, 97.9, 81.8, 80.2, 77.4, 75.6, 74.8, 73.8, 73.4, 73.0, 69.6, 69.4, 66.6, 60.4, 55.0, 20.9, 20.8.

**HRMS (ESI):** calc. for C<sub>44</sub>H<sub>50</sub>O<sub>12</sub>Na (M+Na): 793.3200; found: 793.3204.



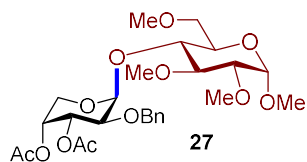
Conditions B: 71% (74 mg),  $\alpha$  only

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta$  = 7.39-7.28 (m, 5 H), 5.75 (d,  $J$  = 3.6 Hz, 1 H), 5.34-5.31 (m, 1 H), 5.25 (dd,  $J$  = 10.4, 3.6 Hz, 1 H), 4.86 (s, 1 H), 4.77 (d,  $J$  = 12.0 Hz, 1 H), 4.65 (d,  $J$  = 12.0 Hz, 1 H), 4.26 (dd,  $J$  = 6.8, 5.6 Hz, 1 H), 4.10 (d,  $J$  = 5.6 Hz, 1 H), 3.99 (dd,  $J$  = 12.8, 1.2 Hz, 1 H), 3.90 (dd,  $J$  = 10.4, 3.6 Hz, 1 H), 3.78-3.70 (m, 1 H), 3.67 (dd,  $J$  = 12.8, 2.0 Hz, 1 H), 3.55 (dd,  $J$  =

10.0, 6.4 Hz, 1 H), 3.36 (s, 3 H), 2.11 (s, 3 H), 2.03 (s, 3 H), 1.54 (s, 3 H), 1.36 (s, 3 H), 1.33 (d,  $J = 6.4$  Hz, 3 H).

**$^{13}\text{C}$  NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta = 170.2, 169.9, 138.0, 128.2, 127.7, 109.2, 97.9, 95.7, 78.4, 77.9, 76.0, 73.3, 72.6, 69.4, 68.8, 63.5, 60.7, 54.6, 27.9, 26.3, 20.9, 20.8, 18.2$ .

**HRMS (ESI):** calc. for C<sub>26</sub>H<sub>36</sub>O<sub>11</sub>Na (M+Na): 547.2155; found: 547.2150.

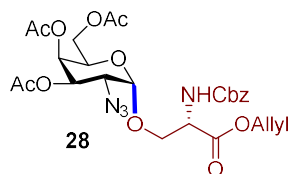


Conditions B: 82% (90 mg),  $\alpha$  only

**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta = 7.39\text{--}7.27$  (m, 5 H), 5.35–5.30 (m, 2 H), 5.08 (d,  $J = 4.0$  Hz, 1 H), 4.82 (d,  $J = 3.2$  Hz, 1 H), 4.75–4.63 (m, 2 H), 4.29 (d,  $J = 13.2$  Hz, 1 H), 3.95 (dd,  $J = 10.0, 4.0$ , Hz, 1 H), 3.76–3.40 (m, 6 H), 3.59 (s, 3 H), 3.49 (s, 3 H), 3.40 (s, 3 H), 3.27 (dd,  $J = 10.0, 4.0$  Hz, 1 H), 3.21 (s, 3 H), 2.12 (s, 3 H), 1.99 (s, 3 H).

**$^{13}\text{C}$  NMR (CDCl<sub>3</sub>, 100 MHz):**  $\delta = 170.3, 169.9, 137.8, 128.4, 128.0, 127.9, 98.5, 97.2, 82.5, 81.2, 75.2, 74.2, 74.1, 70.0, 69.8, 69.7, 60.9, 60.7, 58.8, 58.6, 55.1, 20.9, 20.8$ .

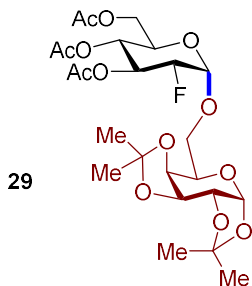
**HRMS (ESI):** calc. for C<sub>26</sub>H<sub>38</sub>O<sub>12</sub>Na (M+Na): 565.2261; found: 564.2260.



Conditions C: 50% (59 mg),  $\alpha$  only

$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **28** has been reported.<sup>5</sup>

**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta = 7.40\text{--}7.30$  (m, 5 H), 5.98–5.87 (m, 1 H), 5.82 (d,  $J = 8.0$  Hz, 1 H), 5.46–5.23 (m, 4 H), 5.20–5.10 (m, 2 H), 4.97 (d,  $J = 3.6$  Hz, 1 H), 4.70–4.56 (m, 3 H), 4.22–4.00 (m, 5 H), 3.62 (dd,  $J = 11.2, 3.6$  Hz, 1 H), 2.14 (s, 3 H), 2.05 (s, 3 H), 2.02 (s, 3 H).  $^1\text{H}$  NMR matches with the literature report.<sup>5</sup>



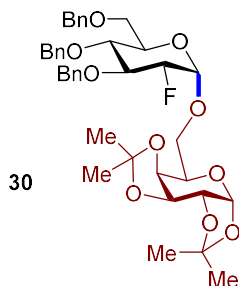
Conditions B: 61% (50 mg),  $\alpha:\beta > 20:1$

$^1\text{H}$  and  $^{13}\text{C}$  NMR of disaccharide **29** has been reported.<sup>6</sup>

**$^1\text{H}$  NMR (CDCl<sub>3</sub>, 400 MHz):**  $\delta = 5.58\text{--}5.45$  (m, 2 H), 5.11 (d,  $J = 4.0$  Hz, 1 H), 5.01 (t,  $J = 8.0$  Hz, 1 H), 4.60 (dd,  $J = 8.0, 4.0$  Hz, 1 H), 4.48 (ddd,  $J = 48.0, 8.0, 4.0$  Hz, 1 H), 4.34–4.25 (m, 3

H), 4.18-4.00 (m, 3 H), 3.90-3.73 (m, 2 H), 2.07(s, 3 H), 2.05 (s, 3 H), 2.02 (s, 3 H), 1.55 (s, 3 H), 1.41 (s, 3 H), 1.32 (s, 3 H), 1.31 (s, 3 H).  $^1\text{H}$  NMR matches with the literature report. <sup>6</sup>

$^{19}\text{F}$  NMR ( $\text{CDCl}_3$ , 376 MHz):  $\delta = -201.4$ .



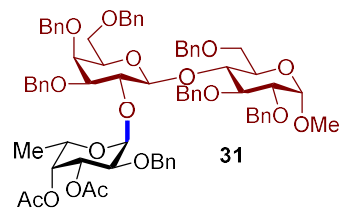
Conditions D': 83% (85 mg),  $\alpha:\beta = 16:1$

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta = 7.39$ -7.15 (m, 15H), 5.52 (d,  $J = 4.8$  Hz, 1H), 5.11 (d,  $J = 4.0$  Hz, 1H), 4.90 (d,  $J = 10.8$  Hz, 1H), 4.84 (d,  $J = 11.2$  Hz, 1H), 4.76 (d,  $J = 10.8$  Hz, 1H), 4.66-4.57 (m, 2.5H), 4.51-4.45 (m, 2.5H), 4.33-4.29 (m, 2H), 4.10 (dt,  $J = 12.4, 9.2$  Hz, 1H), 4.02 (t,  $J = 6.0$  Hz, 1H), 3.90 (dt,  $J = 10.0, 2.0$  Hz, 1H), 3.84 (dd,  $J = 10.4, 2.0$  Hz, 1H), 3.81-3.66 (m, 4H), 1.55 (s, 3H), 1.45 (s, 3H), 1.35 (s, 3H), 1.34 (s, 3H)

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz):  $\delta = 138.5, 138.2, 137.9, 128.4, 128.3, 127.9, 127.9, 127.8, 127.7, 127.7, 127.7, 109.3, 108.6, 96.8$  (d,  $J_{\text{C-F}} = 20.9$  Hz), 96.3, 91.1 (d,  $J_{\text{C-F}} = 191.0$  Hz), 80.6 (d,  $J_{\text{C-F}} = 16.1$  Hz), 76.8 (d,  $J_{\text{C-F}} = 8.3$  Hz), 75.1 (d,  $J_{\text{C-F}} = 2.7$  Hz), 75.0, 73.5, 70.74, 70.68, 70.6, 70.2, 68.1, 66.9, 66.2, 26.2, 26.0, 25.0, 24.5

$^{19}\text{F}$  NMR (376 MHz,  $\text{CDCl}_3$ ):  $\delta -199.09$  (dd,  $J = 49.5, 12.2$  Hz).

HRMS (ESI): calc. for  $\text{C}_{38}\text{H}_{47}\text{O}_{10}\text{FNa}$  ( $\text{M}+\text{Na}$ ): 717.3051; found: 713.3044.



Conditions B: 86% (213 mg),  $\alpha$  only

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz):  $\delta = 7.42$ -6.98 (m, 35 H), 5.72 (d,  $J = 2.4$  Hz, 1 H), 5.32 (dd,  $J = 10.8, 2.8$  Hz, 1 H), 5.18 (s, 1 H), 5.04 (d,  $J = 9.6$  Hz, 1 H), 4.90-4.58 (m, 8 H), 4.50-4.40 (m, 5 H), 4.36-4.25 (m, 3 H), 4.17-4.06 (m, 3 H), 3.95 (s, 1 H), 3.82-3.58 (m, 5 H), 3.55-3.45 (m, 2 H), 3.43 (s, 3 H), 3.39-3.30 (m, 2 H), 2.10 (s, 3 H), 1.99 (s, 3 H), 1.08 (d,  $J = 6.4$  Hz, 3 H).

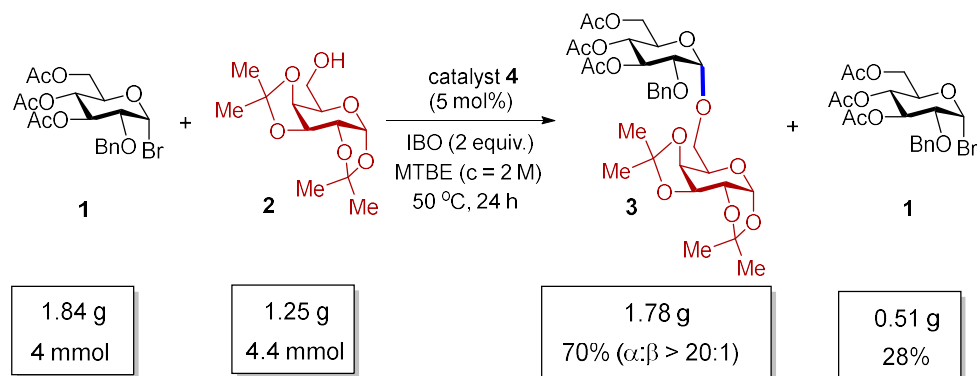
$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz):  $\delta = 170.5, 170.0, 140.0, 138.6, 138.3, 138.03, 138.01, 137.9, 128.5, 128.30, 128.27, 128.2, 128.1, 128.0, 127.8, 127.7, 127.64, 127.58, 127.56, 127.5, 127.31, 127.28, 127.2, 127.02, 126.97, 126.1, 100.2, 98.3, 96.8, 83.8, 80.3, 78.7, 75.6, 75.0, 74.6, 73.7, 73.3, 73.2, 73.03, 72.98, 72.8, 72.2, 71.7, 70.7, 69.7, 69.6, 67.9, 67.6, 64.2, 55.3, 20.7, 20.5, 15.5$ .

HRMS (ESI): calc. for  $\text{C}_{31}\text{H}_{37}\text{NO}_{11}\text{Na}$  ( $\text{M}+\text{Na}$ ): 622.2264; found: 622.2265.



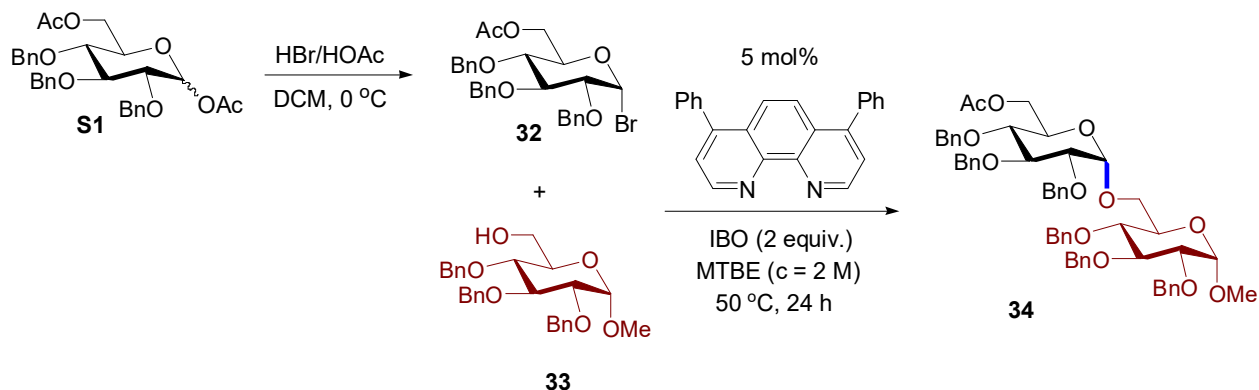
## Gram Scale synthesis

Figure S7. Gram Scale Synthesis of Disaccharide 3



A 50 mL round-bottom flask was charged with glycosyl bromide **1** (1.83 g, 4.0 mmol, 1.0 equiv), alcohol **2** (1.25 g, 4.8 mmol, 1.2 equiv), catalyst **4** (66 mg, 0.2 mmol, 15 mol%), IBO (0.7 mL, 8.0 mmol, 2.0 equiv.) and MTBE (2.0 mL). The resulting solution was stirred at 50 °C for 24 h under open-air atmosphere, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 5/1→3/1) to give the desired disaccharide **3** (1.784 g, 70%,  $\alpha:\beta > 20:1$ ) and recovered **1** (0.515 g, 28%).

## Synthesis of Octasaccharides 40

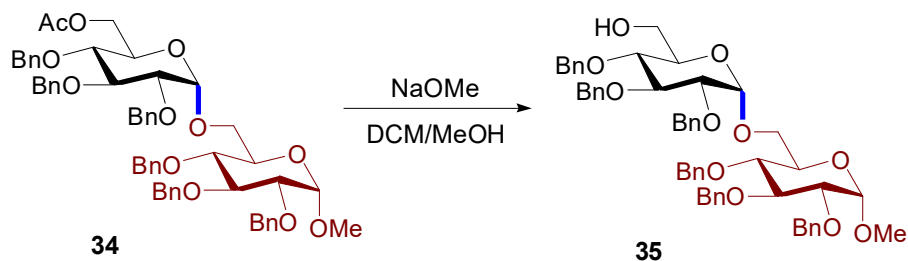


A 500 mL round-bottom flask was charged with **S1** (8.03 g, 15.0 mmol, 1.5 equiv.) and DCM (150 mL). The solution was cooled to 0 °C, then HBr/HOAc (33% wt, 15 mL) was added. The solution was stirred at 0 °C for 30 minutes till the reaction was complete as monitored by TLC. The solution was diluted with ethyl acetate, washed with saturated NaHCO<sub>3</sub> solution for two times, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated in vacuo, and the afforded glycosyl bromide **32** was used directly.

A 50 mL round-bottom flask was charged with glycosyl bromide **32** (15.0 mmol, 1.5 equiv), alcohol **33** (4.63 g, 10.0 mmol, 1.0 equiv), BPhen (166 mg, 0.5 mmol, 5 mol%), IBO (1.78 mL, 20.0 mmol, 2.0 equiv.) and MTBE (2.0 mL). The resulting solution was stirred at 50 °C for 24 h under open-air atmosphere, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 20/1→10/1) to give the desired disaccharide **34** (8.36 g, 89%, α:β > 20:1):

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **34** has been reported.<sup>7</sup>

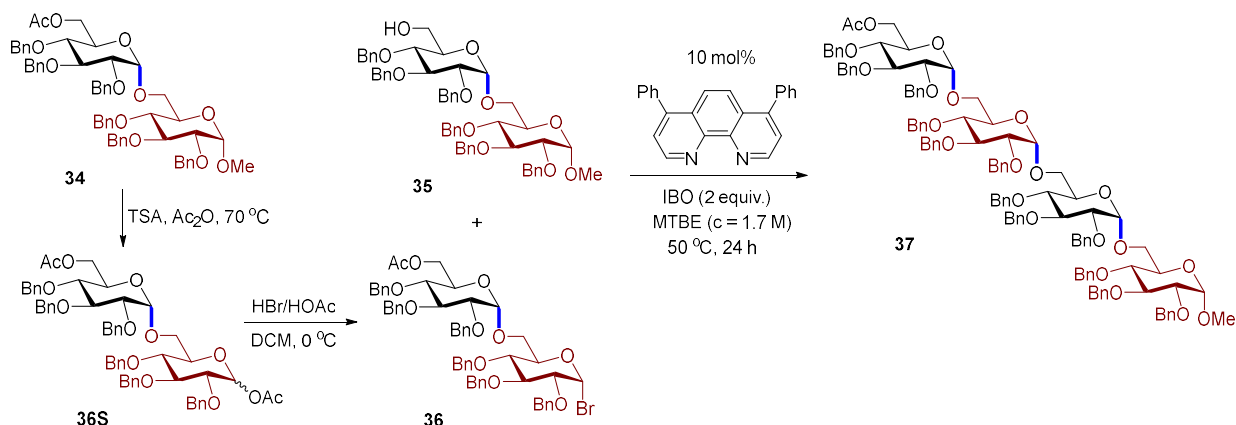
**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.40–7.28 (m, 30 H), 5.00–4.60 (m, 14 H), 4.28–4.22 (m, 2 H), 4.05–4.00 (m 2 H), 3.90–3.80 (m, 3 H), 3.78–3.66 (m, 2 H), 3.55–3.46 (m, 3 H), 3.40 (s, 3 H), 2.01 (s, 3 H).



A 50 mL oven-dried RBF was charged with **34** (350 mg, 0.37 mmol, 1.0 equiv.), MeONa (10 mg, 0.19 mmol, 0.5 equiv.), and CH<sub>2</sub>Cl<sub>2</sub>/MeOH (1 mL/1 mL). The solution was stirred at room temperature overnight. When the reaction was complete as monitored by TLC, the reaction mixture was evaporated, and purified by flash chromatography on silica gel (hexane/ethyl acetate: 2/1→1/1) to afford 341 mg (99%) of **35**:

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **35** has been reported.<sup>7</sup>

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.40–7.28 (m, 30 H), 5.00–4.52 (m, 14 H), 4.28–4.22 (m, 2 H), 4.05–3.46 (m, 10 H), 3.35 (s, 3 H).



A 50 mL round-bottom flask was charged with **34** (940 mg, 1.0 mmol, 1.0 equiv.), PTSA·H<sub>2</sub>O (248 mg, 1.3 mmol, 1.3 equiv.), and Ac<sub>2</sub>O (6 mL). The solution was stirred at 70 °C for 2 hours. The solution was diluted with ethyl acetate, washed with saturated NaHCO<sub>3</sub> (aq.) for three times, concentrated in vacuo, and the residue was purified by silica gel flash chromatography (hexane/ethyl acetate = 4/1 ~ 2/1) to afford 572 mg (61%) of **36S**:

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **36S** has been reported.<sup>7</sup>

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.40-7.28 (m, 30 H), 6.28 (d, *J* = 4.0 Hz, 1 H), 5.00-4.60 (m, 13 H), 4.28-4.22 (m, 2 H), 4.05-3.46 (m, 11 H), 2.13 (s, 3 H), 1.99 (s, 3 H).

A 50 mL round-bottom flask was charged with **36S** (500 mg, 0.51 mmol, 1.5 equiv.) and DCM (30 mL). The solution was cooled to 0 °C, then HBr/HOAc (33% wt, 0.5 mL) was added. The solution was stirred at 0 °C for 20 minutes till the reaction was complete as monitored by TLC. The solution was diluted with ethyl acetate, washed with saturated NaHCO<sub>3</sub> solution for two times, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated in vacuo, and the afforded glycosyl bromide **36** was used directly.

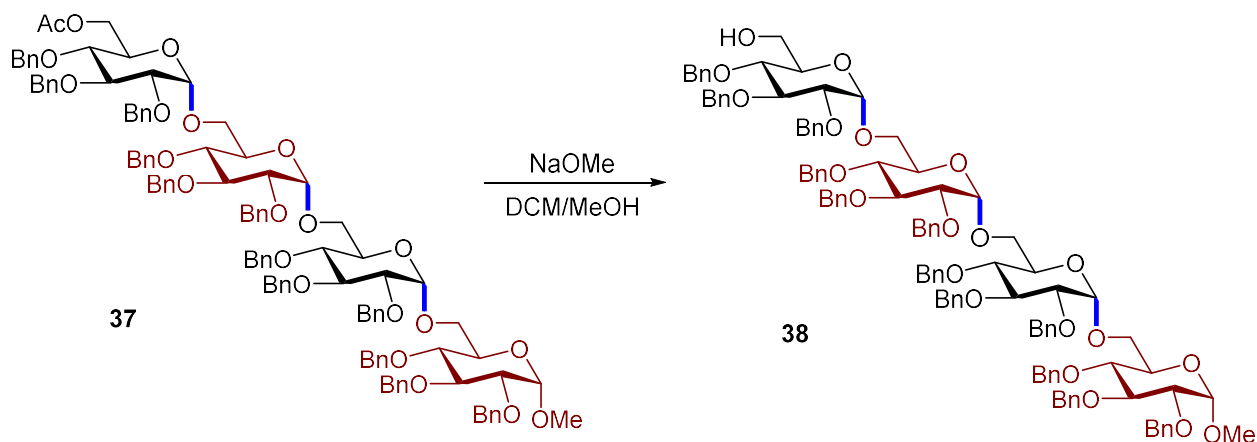
A 50 mL round-bottom flask was charged with glycosyl bromide **36** (0.51 mmol, 1.5 equiv), alcohol **33** (320 mg, 0.34 mmol, 1.0 equiv), BPhen (11 mg, 0.034 mmol, 10 mol%), IBO (0.06 mL, 0.68 mmol, 2.0 equiv.) and MTBE (0.2 mL). The resulting solution was stirred at 50 °C for 24 h under open-air atmosphere, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 20/1→10/1) to give the desired tetraccharide **37** (520 mg, 86%, α:β > 20:1):

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **37** has been reported.<sup>7</sup>

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.42-7.28 (m, 60 H), 5.11 (d, *J* = 4.0 Hz, 1 H), 5.05-4.60 (m, 27 H), 4.28-4.22 (m, 2 H), 4.08-4.00 (m, 4 H), 3.90-3.75 (m, 12 H), 3.60-3.42 (m, 6 H), 3.40 (s, 3 H), 2.03 (s, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):** δ = 170.6, 138.8, 138.6, 138.5, 138.4, 138.3, 138.1, 137.9, 128.30, 128.25, 128.2, 128.0, 127.94, 127.88, 127.73, 127.70, 127.65, 127.6, 127.5, 127.44, 127.41, 127.37, 127.32, 127.28, 97.9, 97.00, 96.95, 82.0, 81.5, 80.3, 80.2, 80.1, 80.0, 77.6, 77.4, 75.6, 75.5, 75.3, 74.9, 74.8, 73.3, 72.3, 72.2, 72.1, 70.71, 70.64, 70.5, 70.3, 68.6, 65.6, 65.5, 65.5, 62.9, 55.1, 20.8.

**HRMS (ESI):** calc. for C<sub>111</sub>H<sub>118</sub>O<sub>22</sub>Na (M+Na): 1825.8007; found: 1925.8009.

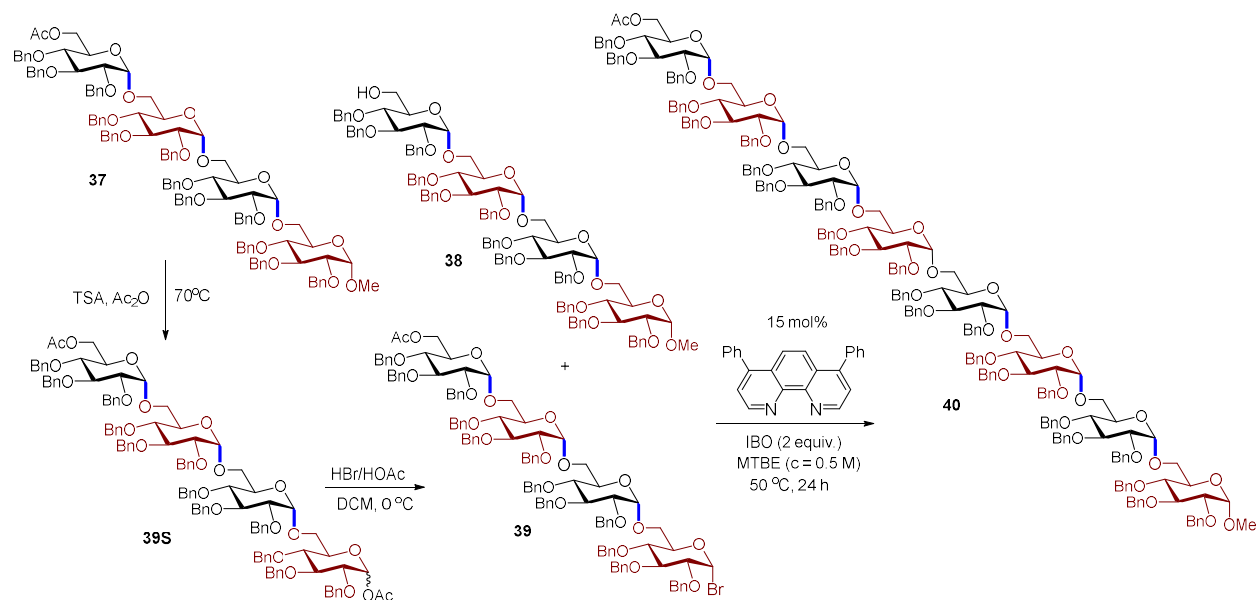


A 50 mL oven-dried RBF was charged with **37** (250 mg, 0.14 mmol, 1.0 equiv.), MeONa (4 mg, 0.07 mmol, 0.5 equiv.), and CH<sub>2</sub>Cl<sub>2</sub>/MeOH (1 mL/1 mL). The solution was stirred at room temperature. When the reaction was complete as monitored by TLC, the reaction mixture was evaporated, and purified by flash chromatography on silica gel (toluene/ethyl acetate: 5/1→3/1) to afford 170 mg (70%) of **38**:

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **38** has been reported.<sup>7</sup>

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.42-7.28 (m, 60 H), 5.05-4.60 (m, 28 H), 4.05-3.40 (m, 24 H), 3.37 (s, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):** δ = 138.8, 128.7, 138.6, 138.54, 138.45, 138.4, 138.3, 138.2, 138.1, 128.33, 128.30, 128.27, 128.2, 127.94, 127.91, 127.8, 127.64, 127.57, 127.5, 127.40, 127.35, 127.1, 98.0, 97.1, 97.0, 82.0, 81.5, 81.4, 77.7, 77.5, 75.6, 75.42, 75.37, 74.9, 73.3, 72.31, 72.25, 72.2, 70.82, 70.75, 70.7, 70.5, 65.8, 65.6, 65.4, 61.8, 55.1.



A 50 mL round-bottom flask was charged with **37** (500 mg, 0.27 mmol, 1.0 equiv.), PTSA·H<sub>2</sub>O (67 mg, 0.35 mmol, 1.3 equiv.), and Ac<sub>2</sub>O (3 mL). The solution was stirred at 70 °C for 2 hours. The solution was diluted with ethyl acetate, washed with saturated NaHCO<sub>3</sub> (aq.) for

three times, concentrated in vacuo, and the residue was purified by silica gel flash chromatography (toluene/ethyl acetate = 8/1 ~ 5/1) to afford 249 mg (51%) of **39S**:

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **S3** has been reported.<sup>7</sup>

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.40-7.28 (m, 60 H), 6.34 (d, *J* = 4.0 Hz, 1 H), 5.00-4.60 (m, 27 H), 4.28-4.22 (m, 2 H), 4.05-3.46 (m, 22 H), 2.08 (s, 3 H), 2.02 (s, 3 H).

A 25 mL round-bottom flask was charged with **39S** (110 mg, 0.06 mmol, 1.5 equiv.) and DCM (6 mL). The solution was cooled to 0 °C, then HBr/HOAc (33% wt, 0.06 mL) was added. The solution was stirred at 0 °C for 15 minutes till the reaction was complete as monitored by TLC. The solution was diluted with ethyl acetate, washed with saturated NaHCO<sub>3</sub> solution for two times, dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated in vacuo, and the afforded glycosyl bromide **39** was used directly.

A 50 mL round-bottom flask was charged with glycosyl bromide **39** (0.06 mmol, 1.5 equiv), alcohol **38** (70 mg, 0.04 mmol, 1.0 equiv), BPhen (2 mg, 0.006 mmol, 15 mol%), IBO (0.007 mL, 0.08 mmol, 2.0 equiv.) and MTBE (0.08 mL). The resulting solution was stirred at 50 °C for 24 h under open-air atmosphere, diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 20/1→10/1) to give the desired octasaccharide **40** (109 mg, 77%, α:β > 20:1):

<sup>1</sup>H and <sup>13</sup>C NMR of disaccharide **40** has been reported.<sup>7</sup>

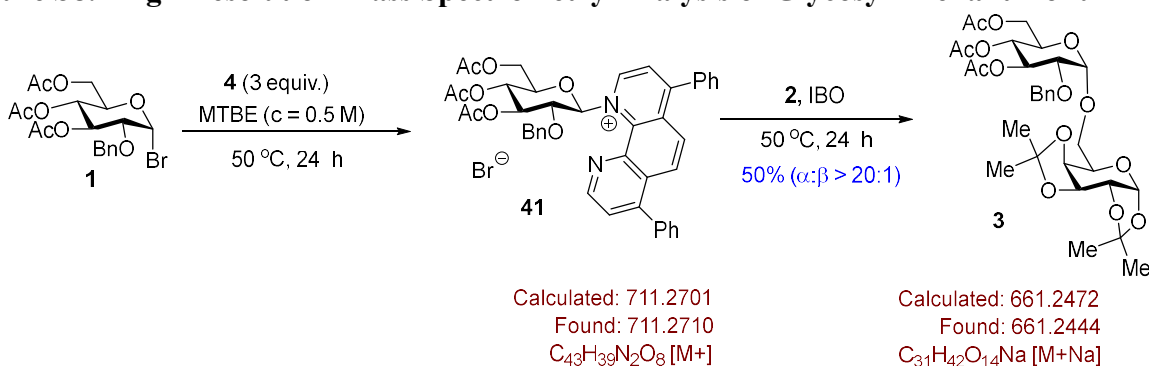
**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):** δ = 7.42-7.28 (m, 120 H), 5.05 (d, *J* = 4.0 Hz, 1 H), 5.05-4.40 (m, 54 H), 4.25-4.18 (m, 2 H), 4.08-4.00 (m, 8 H), 3.90-3.30 (m 39 H), 3.32 (s, 3 H), 1.98 (s, 3 H).

**<sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz):** δ = 170.7, 138.8, 138.6, 138.54, 138.46, 138.4, 138.2, 138.0, 128.38, 128.36, 128.3, 128.2, 128.04, 127.99, 127.9, 127.8, 127.6, 127.5, 127.4, 127.34, 127.27, 98.0, 97.30, 97.25, 97.18, 97.16, 97.1, 97.0, 82.1, 81.5, 80.4, 80.3, 80.2, 80.0, 75.7, 75.5, 75.4, 75.0, 74.9, 73.4, 72.2, 72.13, 72.07, 70.88, 70.87, 70.8, 70.7, 70.6, 68.7, 65.5, 63.0, 55.1, 20.8.

**HRMS (ESI):** calc. for C<sub>219</sub>H<sub>230</sub>O<sub>42</sub>Na (M+Na): 3554.5754; found: 3554.5867.

## 4. Mechanistic Studies

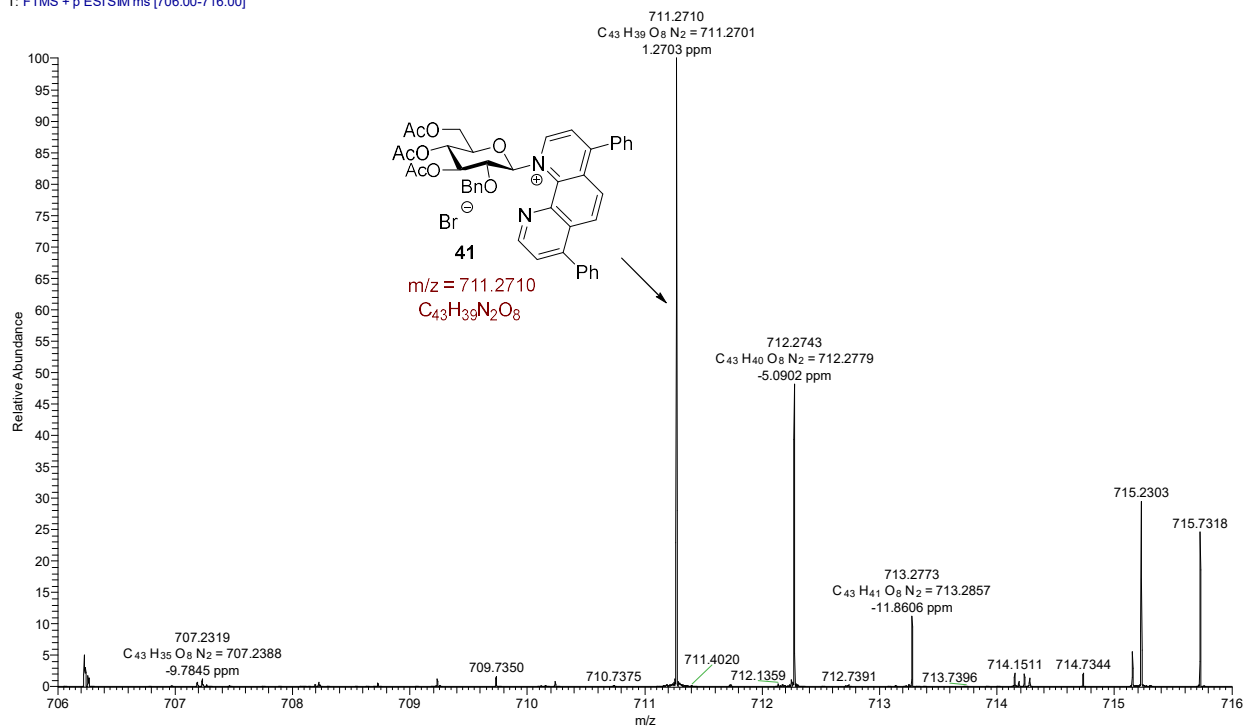
Figure S8. High Resolution Mass Spectrometry Analysis of Glycosyl Phenanthrolium **41**



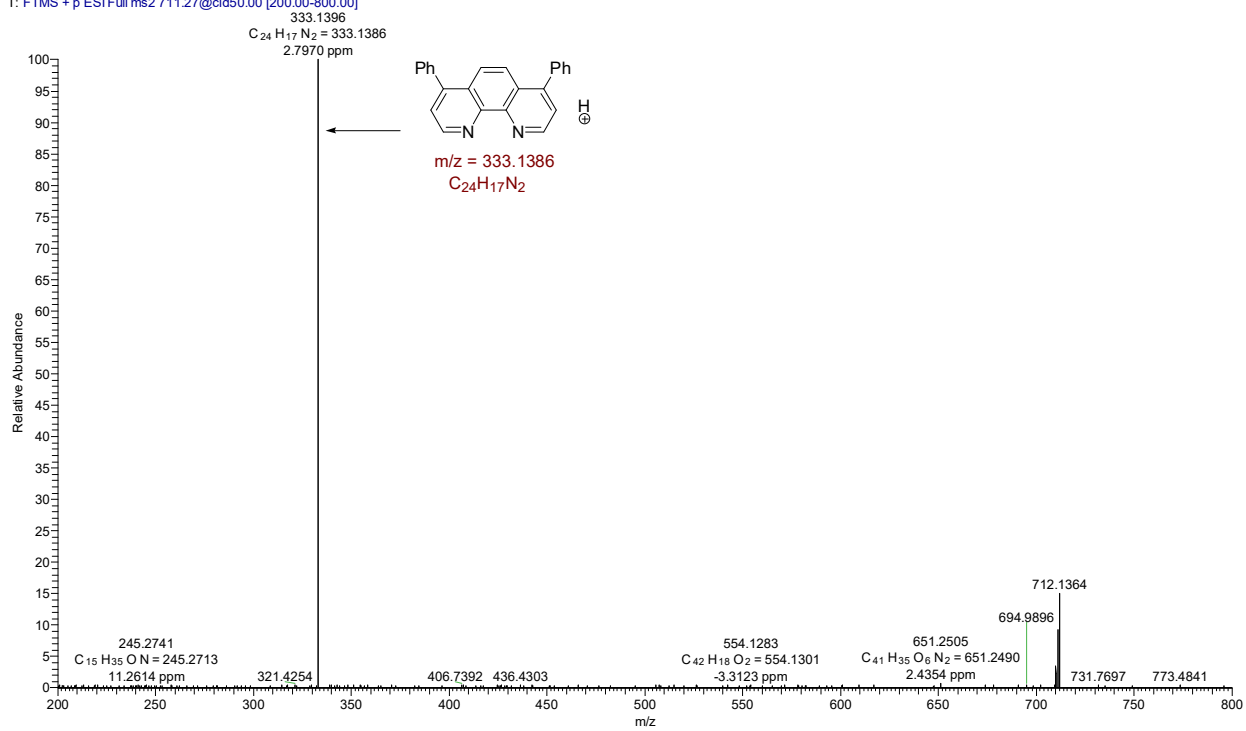
A 10 mL bottle was charged with glycosyl bromide **1** (46 mg, 0.1 mmol, 1.0 equiv), **4** (100 mg, 0.3 mmol, 3.0 equiv.), and MTBE (1.2 mL). The reaction mixture was stirred at 50 °C for 24 h. Formation of the glycosyl phenanthrolium ion **41** was confirmed using electrospray ionization (ESI) with an  $m/z$  ratio of 711.2710 (see below). Subsequent fragmentation of **41** using collision induced dissociation (CID) led to the formation of various fragment ions, most notably the phenanthroline species with an  $m/z$  ratio of 331.1396 (see below). The mixture was concentrated and dried *in vacuo*. The resulting residue was mixed with alcohol **2** (39 mg, 0.15 mmol, 1.5 equiv.), and MTBE (0.4 mL). The reaction mixture was stirred at 50 °C for 12 h, formation of the desired disaccharide **3** was confirmed by high resolution electrospray ionization (ESI), diluted with toluene, and purified by silica gel flash chromatography (toluene/ethyl acetate: 5/1→3/1) to give the desired disaccharide **3** (31 mg, 50%,  $\alpha:\beta > 20:1$ ).

Sample Preparation for ESI: 50  $\mu$ L of a solution of glycosyl bromide **1** (46 mg, 0.1 mmol, 1.0 equiv), catalyst **4** (100 mg, 0.3 mmol, 3.0 equiv.), and MTBE (1.2 mL) was diluted with 1 mL of dry acetonitrile (ACN). The resulting acetonitrile sample was infused at a rate of 5  $\mu$ L/min via syringe pump.

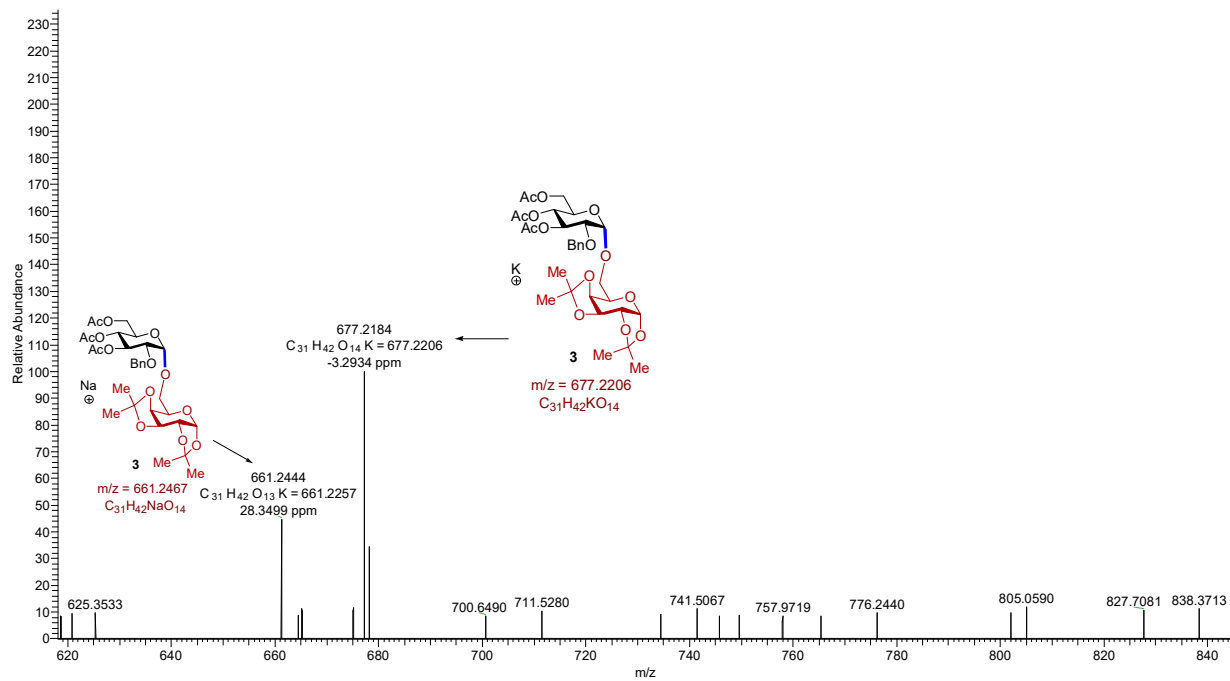
FY-87-26-1 Dry ACN#1 RT: 0.00 AV: 1 NL: 8.22E5  
T: FTMS + p ESI SIM ms [706.00-716.00]



FY-87-26-1\_711@60V #1 RT: 0.03 AV: 1 NL: 5.34E4  
T: FTMS + p ESI Full ms 2711.27@cid50.00 [200.00-800.00]

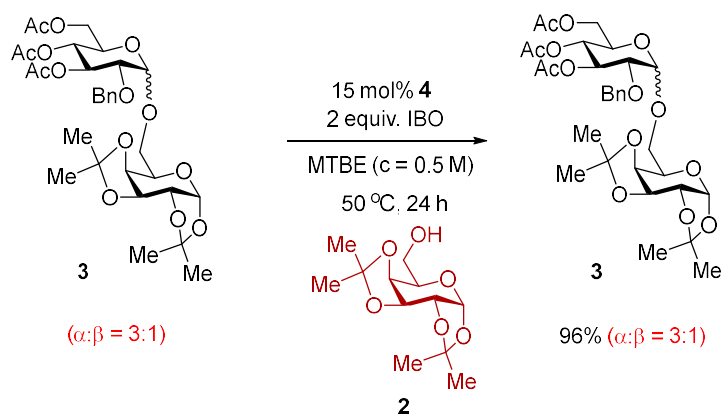


FY87-31 #1 RT: 0.01 AV: 1 NL: 4.79E3  
T: FTMS + p ESI Full ms [50.00-1500.00]

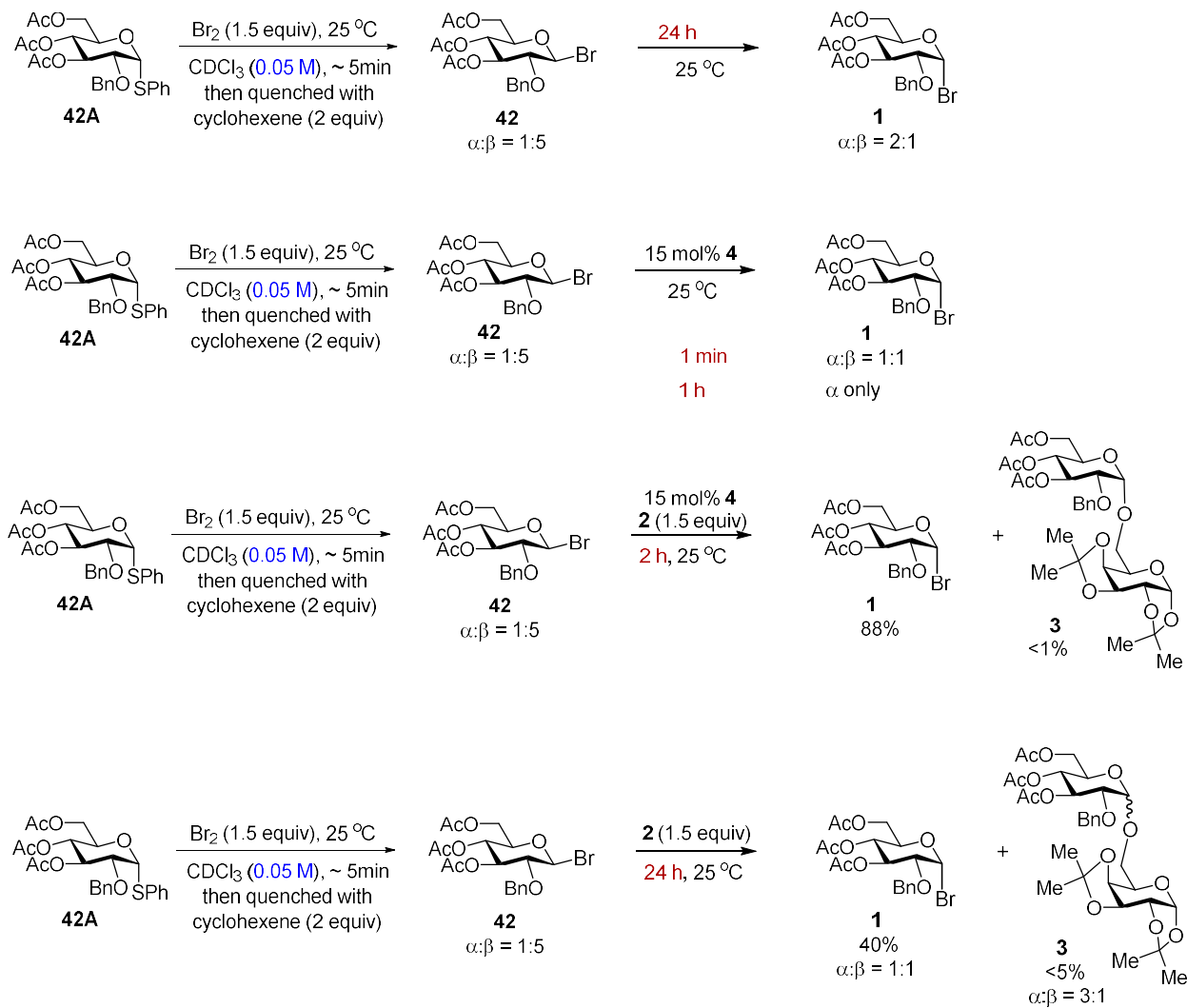




**Figure S9. Attempted Isomerization of Disaccharide 3.**

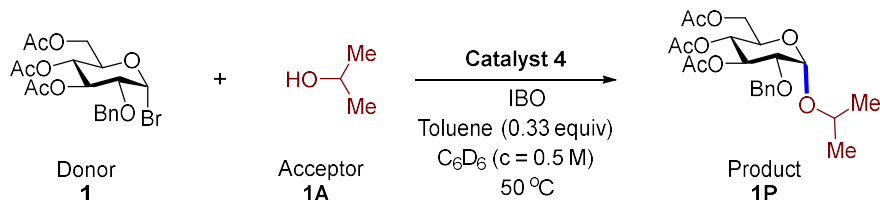


**Figure S10. Anomerization of  $\beta$ -Bromide to  $\alpha$ -Bromide**



## 5. Kinetic Study

### General Experimental Procedure for Kinetic Studies

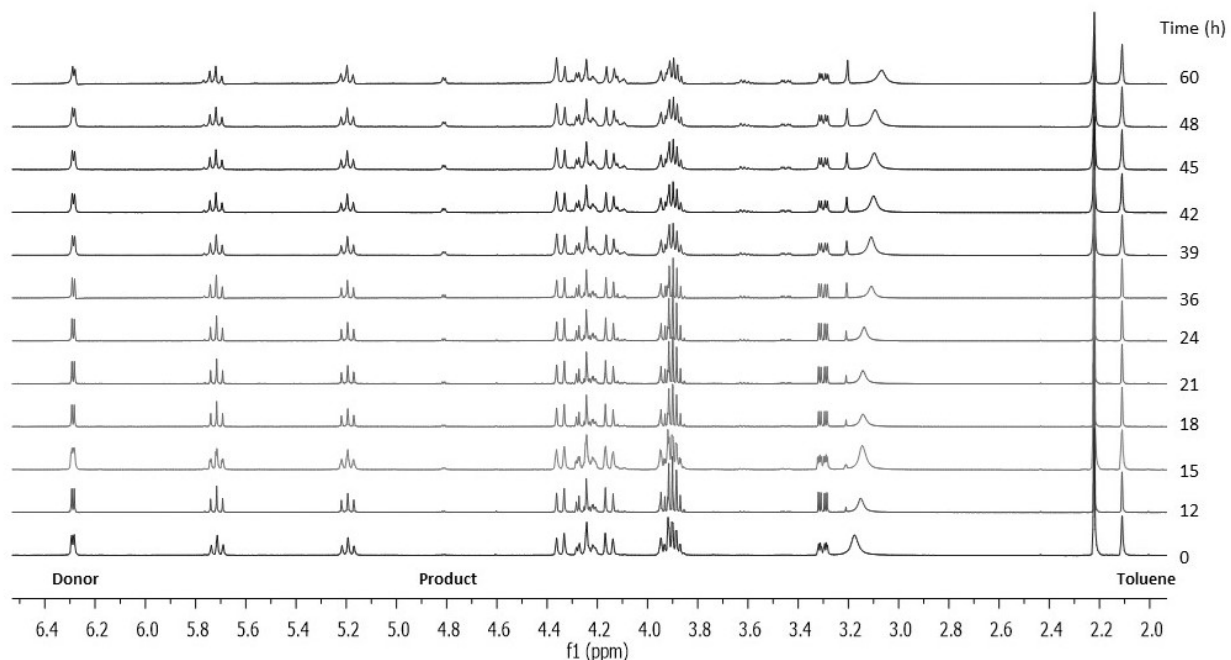


A 10 mL scintillation vial was charged with glycosyl bromide **1** (fixed amount, 0.25 mmol, 1.0 equiv), isopropanol acceptor **1A** (vary amount from 0.5 to 5 equiv), catalyst **4** (vary amount from 2 to 20 mol%), IBO (vary amount from 1.5 to 3 equiv), toluene (internal standard, 0.083mmol, 0.33 equiv), and  $\text{C}_6\text{D}_6$  (0.5 mL). The resulting solution was then transferred to a 5 mm NMR tube.  $^1\text{H}$  NMR spectrum was acquired on a 400 MHz instrument before heating. Then the mixture in NMR tube was then consistently shaken and heated in a 50 °C water bath. Between 3 and 60 h, spectra were obtained depending on the experiment. Example spectra and example rate plot were based on standard condition: 0.25 mmol glycosyl bromide **1** (1.0 equiv), 0.75 mmol acceptor (3.0 equiv), 15 mol% catalyst **4**, 0.5 mmol IBO (2 equiv), 0.083 mmol toluene (0.33 equiv) as an internal standard, and 0.5 mL  $\text{C}_6\text{D}_6$  (0.5 M). The kinetic runs were repeated twice.

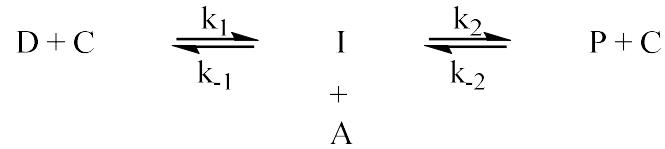
### Spectra Processing

The spectra for each kinetic experiment were processed using MestReNova (v. 6.0.2, Mestrelab Research S.L.). The concentrations of product were measured by integration of its H-1 proton against the toluene internal standard,  $\delta = 2.1$  ppm. Peak fitting or deconvolution algorithms were not used for integration.

Figure S11: Example Spectra Array for a Kinetic Experiment



## Rate Equation Derivation



D = Donor C = Catalyst I = Intermediate A = Acceptor P = Product

$$\frac{d[P]}{dt} = k_2[I][A] - k_{-2}[P][C]$$

$$\frac{d[I]}{dt} = k_1[D][C] - k_{-1}[I] - k_2[I][A] + k_{-2}[P][C]$$

Assume steady state,

$$\frac{d[I]}{dt} = 0$$

Therefore,

$$[I] = \frac{k_1[D][C] + k_{-2}[P][C]}{k_{-1} + k_2[A]}$$

Since  $k_{-2} \approx 0$ ,

$$\frac{d[P]}{dt} = k_2[I][A]$$

$$[I] = \frac{k_1[D][C]}{k_{-1} + k_2[A]}$$

Substitute [I] into rate equation,

$$\frac{d[P]}{dt} = \frac{k_1 k_2 [D][C][A]}{k_{-1} + k_2[A]}$$

For fixed donor and acceptor concentration,

$$\frac{d[P]}{dt} = k'[C]$$

$$\text{where } k' = \frac{k_1 k_2 [D][A]}{k_{-1} + k_2[A]}$$

For fixed donor and catalyst concentration,

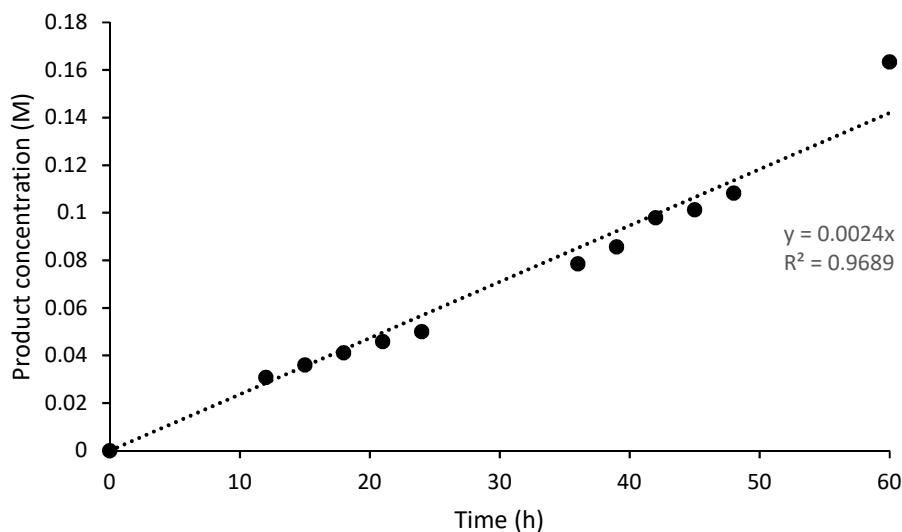
$$\frac{d[P]}{dt} = \frac{k''[A]}{k_{-1} + k_2[A]}$$

$$\text{where } k'' = k_1 k_2 [D][C]$$

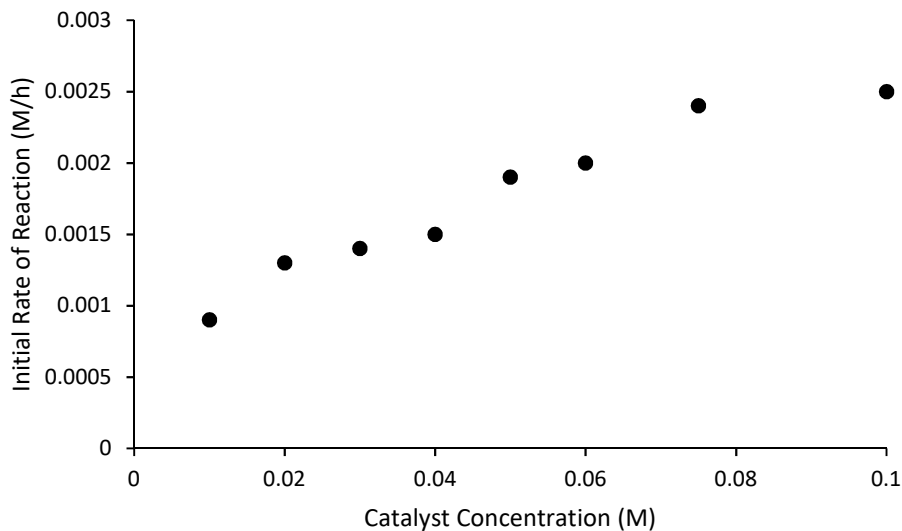
## Graphing

For each kinetic experiment, the concentration of product versus time were plotted on Excel 2016. Linear regression was obtained by best fitting with all points (Figure S12). Slope of the best-fit line represents the initial rate of reaction for each kinetic experiment. The initial rate was then graphed against catalyst concentration for fixed acceptor concentration (Figure S13), and against acceptor concentration for fixed catalyst concentration (Figure S14). The product formation versus time was also compared at different equivalent of IBO (Figure S15).

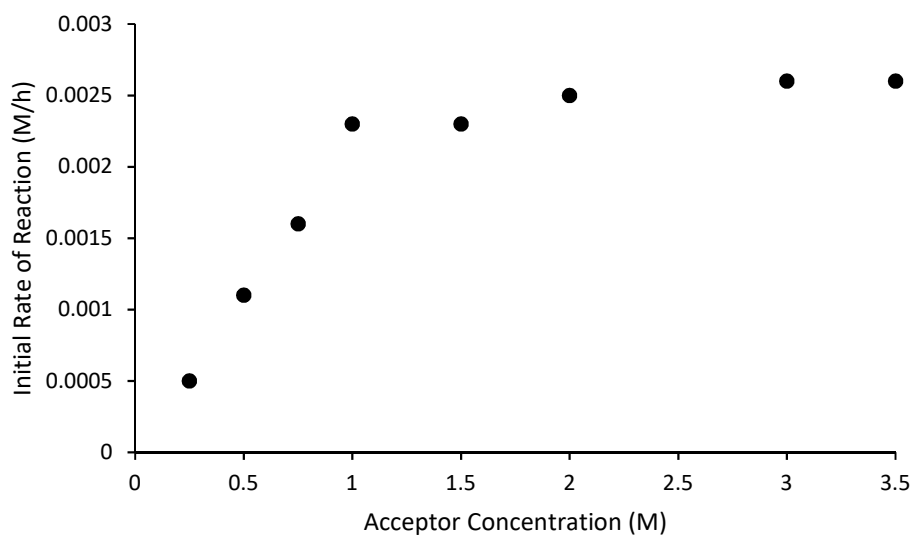
**Figure S12: Example Rate Plot: Product concentration versus time for a kinetic experiment**



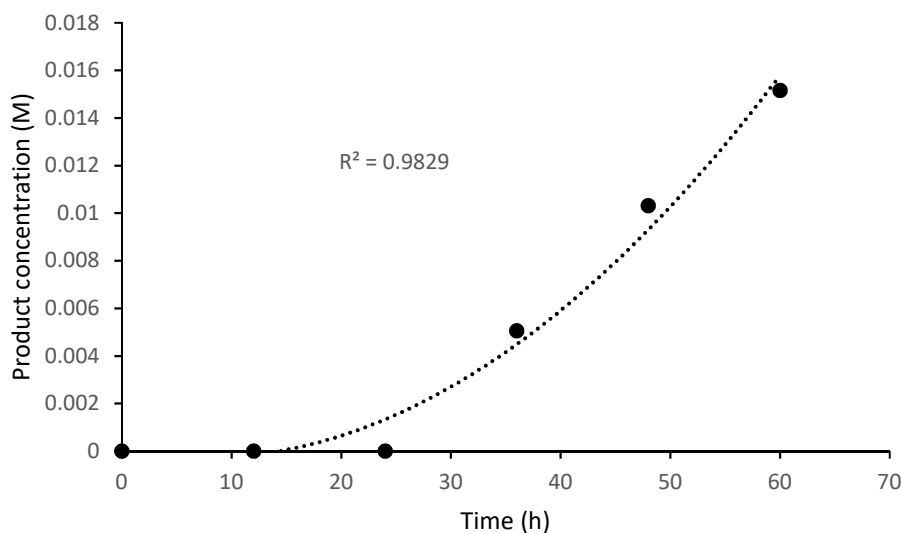
**Figure S13: Rate of Reaction versus Catalyst concentration**



**Figure S14: Rate of Reaction versus Acceptor concentration**

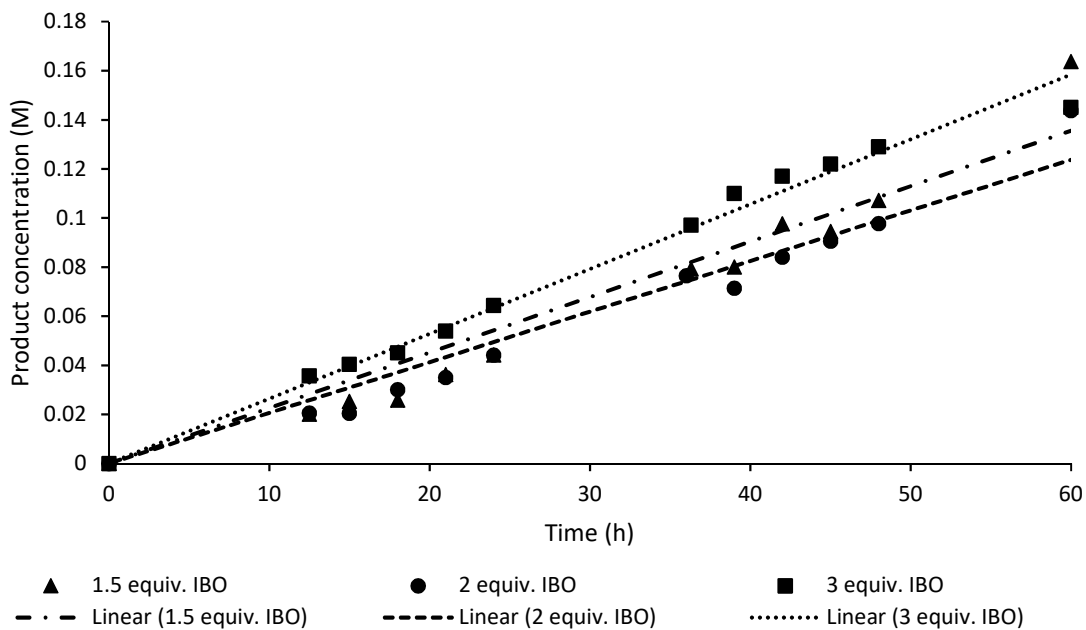


**Figure S15: Product formation versus time at 0 mol% catalyst**



As shown in Figure S15, the desired product was not observed even after 24 h. This result is consistent with our control experiment illustrated in Table 1 in the main text. After 24 h, the desired product was slowly formed in the reaction. Even after 60 h, only 3% conversion was observed. Collectively, these results suggest that the background reaction only takes place after a long period.

**Figure S16: Product formation versus time at different equivalent of IBO**



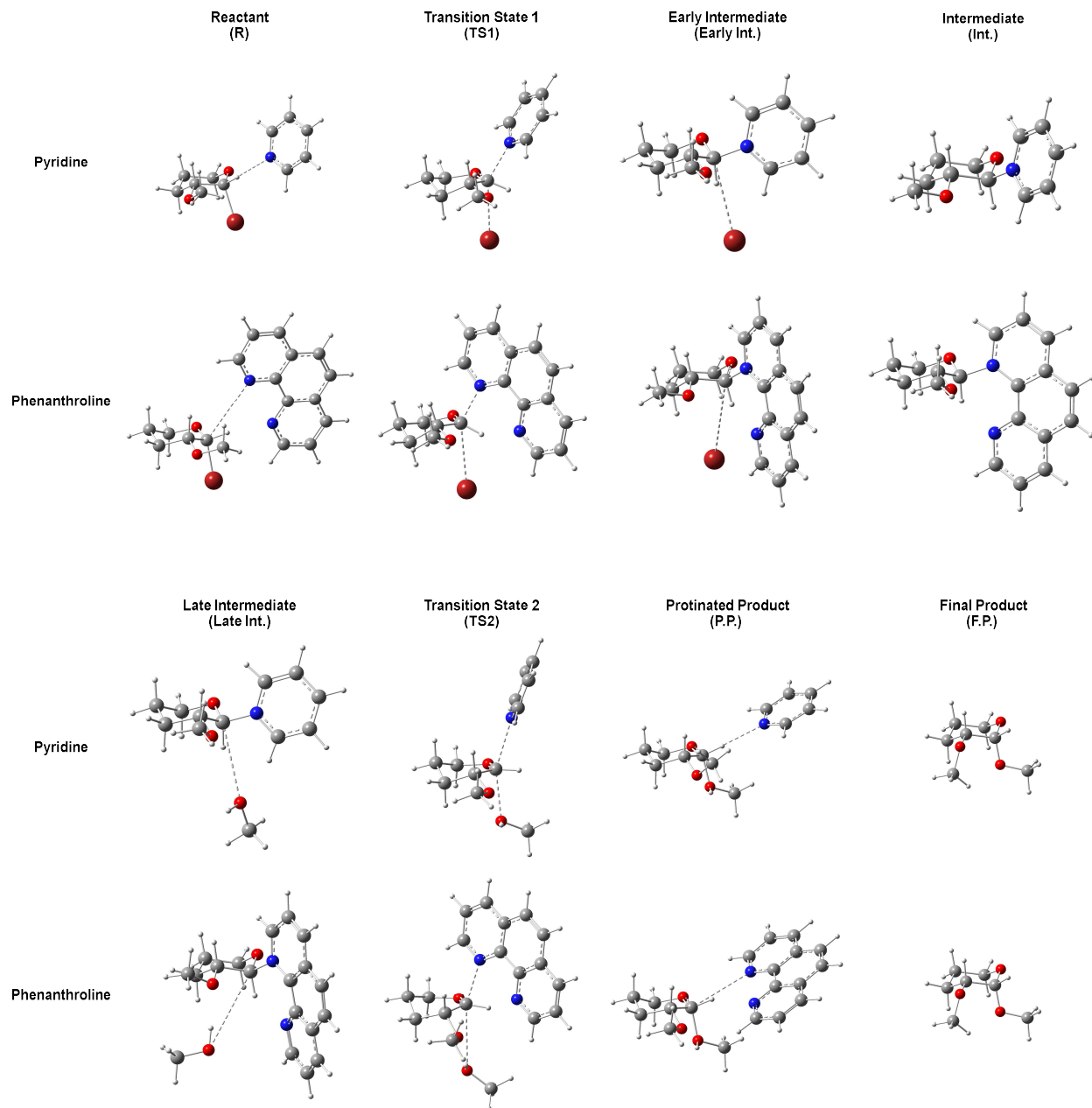
As shown in Figure S16, the rate of reactions does not change significantly with varying amounts of IBO.



## 6. DFT Calculations

All calculations were carried out with Gaussian 09.<sup>8</sup> Geometry optimization for reactant, intermediates, transition states, and products were computed at the B3LYP/6-31+G(d,p) level of theory<sup>9-19</sup> with the SMD implicit solvation model<sup>20</sup> in diethyl ether. There is only one imaginary frequency for transition state structures and no imaginary frequency for reactant, intermediates, and products. Non-covalent interactions (NCI) were calculated with the NCIPLOT program<sup>21</sup>.

Figure S17. Optimized Structures



### Cartesian coordinate

#### R\_PY

Charge = 0 Multiplicity = 1

E(RB3LYP) = -3205.781567 A.U.

C	3.165678	-0.862673	0.914227
C	1.702113	-0.529111	1.187095
C	0.918667	-0.302628	-0.120761
C	2.458617	-1.656448	-1.359637
C	3.264411	-1.998425	-0.113949
H	-0.154033	-0.165863	0.013853
H	1.206831	-1.407443	1.641965
H	3.665818	0.037347	0.538657
H	3.649743	-1.141440	1.856002
H	2.374567	-2.505276	-2.041685
H	2.905739	-0.813732	-1.901420
H	2.889110	-2.934871	0.318076
H	4.308677	-2.168287	-0.401121
O	1.090023	-1.309635	-1.038264
O	1.650421	0.552641	2.095004
C	0.352833	0.864031	2.594682
H	0.496524	1.572110	3.414703
H	-0.280182	1.333903	1.831964
H	-0.152285	-0.033922	2.979810
Br	1.469670	1.518986	-0.913602
C	-3.171264	-1.357526	-0.003801
C	-4.561412	-1.491805	0.013727
C	-5.343616	-0.339545	-0.084219
C	-4.702561	0.895954	-0.193114
C	-3.306148	0.925624	-0.199057
N	-2.543906	-0.175163	-0.103079
H	-6.428108	-0.402898	-0.075930
H	-2.532836	-2.235889	0.066227
H	-5.012868	-2.475585	0.098166
H	-5.267353	1.819697	-0.273660
H	-2.775699	1.871569	-0.283247

#### TS1\_Pyridine

Charge = 0 Multiplicity = 1

E(RB3LYP) = -3205.742135 A.U.

C	-5.165043	-1.057924	4.224031
C	-3.880020	-1.893345	4.131315
C	-3.157567	-1.632197	2.822409
C	-4.145048	0.569000	2.602085
C	-4.774478	0.404940	3.972188
H	-2.892909	-2.452994	2.172671

H	-3.212357	-1.536053	4.932933
H	-5.878555	-1.411047	3.474269
H	-5.604370	-1.165295	5.220105
H	-3.633380	1.524705	2.479966
H	-4.879408	0.431134	1.803687
H	-4.087127	0.754456	4.752987
H	-5.657945	1.052425	4.007178
O	-3.114332	-0.439298	2.296118
O	-4.024068	-3.295490	4.244385
C	-4.393226	-3.739654	5.545971
H	-4.330271	-4.830192	5.530396
H	-3.707183	-3.349535	6.313441
H	-5.419995	-3.451648	5.803900
Br	-5.236851	-2.441940	0.860721
C	-0.297338	-0.789112	3.133226
C	-0.651211	-2.860559	4.112630
C	1.065146	-0.831422	3.427556
H	-0.745591	0.052266	2.611427
C	0.698961	-2.987454	4.434549
H	-1.373753	-3.634712	4.355379
C	1.570872	-1.952079	4.088848
H	1.708896	-0.009216	3.133177
H	1.052359	-3.879117	4.941661
H	2.628145	-2.020504	4.326862
N	-1.128338	-1.774789	3.486910

### Early-Int Pyridine

Charge = 0 Multiplicity = 1

E(RB3LYP) = -3205.780059 A.U.

C	-0.977437	-0.014877	-0.234492
C	0.507013	-0.414415	-0.264780
C	1.157591	0.307525	-1.456687
C	-0.320577	2.144483	-1.351567
C	-1.118441	1.516693	-0.213707
H	0.708897	-0.038540	-2.399584
H	1.001389	-0.077003	0.663104
H	-1.461162	-0.430267	-1.127199
H	-1.465638	-0.453476	0.641522
H	-0.274356	3.232799	-1.271206
H	-0.746465	1.879807	-2.328632
H	-0.760918	1.916962	0.743750
H	-2.172137	1.802310	-0.312824
O	1.058957	1.705878	-1.311847
O	0.728302	-1.803501	-0.443279
C	0.474076	-2.595682	0.712863
H	0.783671	-3.614213	0.467393

H	1.057879	-2.240204	1.575075
H	-0.589903	-2.606685	0.980826
Br	0.142230	-1.083344	-4.614798
C	3.017332	-0.883664	-2.497979
C	4.354840	-1.246640	-2.572974
C	5.268691	-0.694632	-1.676783
C	4.821866	0.222686	-0.719592
C	3.479705	0.553053	-0.676767
N	2.610260	-0.006352	-1.551971
H	6.317988	-0.968214	-1.723704
H	2.248107	-1.239918	-3.185574
H	4.663093	-1.948971	-3.338823
H	5.502336	0.681639	-0.011709
H	3.063093	1.258113	0.030411

### Late-Int\_Pyridine

Charge = 1 Multiplicity = 1

E(RB3LYP) = -749.617112 A.U.

C	-0.730893	-0.389269	4.670408
C	-0.333098	-0.690589	3.216943
C	-0.094231	0.647053	2.492105
C	0.531658	1.775469	4.480249
C	0.303691	0.534413	5.332661
H	0.605764	-1.268423	3.200938
H	-1.718309	0.090559	4.665501
H	-0.823993	-1.324027	5.231621
H	1.364813	2.380695	4.842692
H	-0.369349	2.404585	4.443091
H	1.255809	0.005776	5.464295
H	-0.039576	0.838365	6.327928
H	-1.040014	1.196161	2.409346
O	0.898466	1.410611	3.126324
O	-1.335530	-1.372948	2.478171
C	-1.452830	-2.763861	2.782135
H	-2.185689	-3.174753	2.084416
H	-0.492994	-3.280352	2.641497
H	-1.805560	-2.931050	3.806916
C	1.666038	0.067618	0.873161
C	2.113625	-0.173183	-0.413657
C	1.222770	-0.045093	-1.482694
C	-0.097738	0.328148	-1.229975
C	-0.502734	0.563726	0.074513
N	0.378672	0.427699	1.093359
H	1.556730	-0.229354	-2.498762
H	2.302223	-0.002066	1.745215

H	3.148400	-0.456313	-0.567634
H	-0.817715	0.445097	-2.031551
H	-1.510158	0.868789	0.342437
O	-3.234962	1.622885	1.420355
H	-3.821605	0.921727	1.738478
C	-4.027480	2.773498	1.087036
H	-3.334761	3.544764	0.742493
H	-4.740076	2.550720	0.283639
H	-4.571612	3.151124	1.961002

### TS2\_Pyridine

Charge = 1 Multiplicity = 1

E(RB3LYP) = -749.576283 A.U.

C	-2.855387	0.118082	4.147442
C	-1.554162	-0.226211	3.414612
C	-1.131059	0.884767	2.481254
C	-2.524633	2.537315	3.580265
C	-2.721342	1.542013	4.702108
H	-0.741961	-0.296768	4.154747
H	-3.697911	0.049101	3.451550
H	-3.021321	-0.594358	4.958946
H	-2.159265	3.508415	3.912918
H	-3.419443	2.670430	2.968309
H	-1.888109	1.601405	5.411953
H	-3.627893	1.833705	5.242449
H	-0.467473	0.675293	1.650944
O	-1.493953	2.097752	2.592761
O	-1.580067	-1.411349	2.632948
C	-1.530213	-2.623084	3.399128
H	-1.458846	-3.438124	2.676809
H	-0.647442	-2.636196	4.050844
H	-2.435196	-2.754604	4.002588
C	2.109344	0.254125	3.708084
C	3.426568	0.499526	4.097540
C	3.872134	1.821972	4.146036
C	2.985934	2.844177	3.801244
C	1.687177	2.501101	3.422824
N	1.251901	1.233013	3.379777
H	1.725830	-0.762742	3.655572
H	4.080993	-0.327368	4.353543
H	3.289193	3.885915	3.822274
H	0.969768	3.269275	3.142570
O	-2.850613	0.222568	0.870337
H	-2.707990	-0.728504	1.005998
C	-2.607727	0.559128	-0.504016

H	-1.607524	0.246037	-0.827881
H	-3.358523	0.096501	-1.154871
H	-2.688353	1.645070	-0.587301
H	4.890686	2.051081	4.444885

### **P.P. Pyridine**

Charge = 1 Multiplicity = 1

E(RB3LYP) = -749.581689 A.U.

C	-3.870530	0.739542	4.458652
C	-2.660633	0.040344	3.845857
C	-2.189057	0.710469	2.541629
C	-3.365879	2.751078	3.057606
C	-3.640692	2.257309	4.465847
H	-1.807475	0.078033	4.534291
H	-4.765690	0.498941	3.872454
H	-4.028989	0.354954	5.470139
H	-3.064117	3.798323	3.029713
H	-4.232135	2.611828	2.402189
H	-2.800661	2.518573	5.120919
H	-4.525641	2.775177	4.850162
H	-1.230917	0.342825	2.159945
O	-2.229592	2.043806	2.450052
O	-2.986489	-1.322379	3.528731
C	-1.990378	-2.301681	3.885473
H	-2.372762	-3.265374	3.545084
H	-1.030785	-2.092770	3.400001
H	-1.860258	-2.323500	4.972479
C	1.572925	-1.253546	0.976549
C	2.828676	-1.277656	0.367203
C	3.405759	-0.066887	-0.019891
C	2.703447	1.115253	0.220561
C	1.453109	1.035796	0.836746
N	0.885017	-0.123190	1.212223
H	1.099192	-2.181772	1.289003
H	3.336675	-2.222856	0.203253
H	3.110720	2.081270	-0.060965
H	0.883305	1.940267	1.038472
O	-3.250482	0.081728	1.487456
H	-3.378996	-0.828706	1.876640
C	-2.852456	0.057015	0.071358
H	-2.697791	1.092985	-0.223977
H	-1.941053	-0.530584	-0.045306
H	-3.689622	-0.378341	-0.474093
H	4.381066	-0.044923	-0.497414

**R\_Phenanthroline**

Charge = 0 Multiplicity = 1

E(RB3LYP) = -3529.131672 A.U.

C	-3.993558	1.500050	0.470229
C	-2.569535	1.137649	0.879646
C	-1.958005	0.072863	-0.050844
C	-3.436644	0.676151	-1.834739
C	-4.043595	1.817929	-1.030854
H	-0.902486	-0.125992	0.130853
H	-1.913404	2.015235	0.733408
H	-4.651604	0.654809	0.703013
H	-4.330500	2.355153	1.065414
H	-3.317745	0.933174	-2.889613
H	-4.051955	-0.229490	-1.760809
H	-3.493841	2.744299	-1.240905
H	-5.078536	1.971915	-1.358092
O	-2.100055	0.350633	-1.385021
O	-2.572927	0.781270	2.247415
C	-1.283959	0.585559	2.825891
H	-1.444944	0.438945	3.896946
H	-0.778460	-0.298565	2.419823
H	-0.643028	1.465185	2.672583
Br	-2.805047	-1.765816	0.398445
C	1.714344	2.940687	0.211854
C	2.970910	3.567448	0.082207
C	4.080702	2.772231	-0.120422
C	3.926853	1.368636	-0.194964
N	1.530963	1.628708	0.149740
H	5.071582	3.206541	-0.225356
H	0.823998	3.545925	0.373971
H	3.050089	4.648305	0.142821
C	2.611964	0.835820	-0.054473
C	3.566797	-1.438132	-0.352504
C	5.045565	0.493954	-0.409042
C	4.872493	-0.855385	-0.483967
H	6.034435	0.933174	-0.511232
H	5.720335	-1.515626	-0.647084
C	2.427172	-0.609197	-0.137590
C	3.359376	-2.834547	-0.434978
C	1.022254	-2.437342	-0.087463
C	2.082447	-3.341283	-0.303166
H	4.209556	-3.491266	-0.601170
H	0.003282	-2.802317	0.022860
H	1.884973	-4.406934	-0.362760
N	1.180580	-1.122825	-0.003998

**TS1\_Phenanthroline**

Charge = 0 Multiplicity = 1

E(RB3LYP) = -3529.084393 A.U.

C	-4.832837	-1.136532	-0.140903
C	-4.303745	-0.769552	-1.514306
C	-2.122037	-0.282629	-0.532137
C	-2.599154	-0.723233	0.843558
C	-3.723997	-1.763530	0.711842
H	-4.108011	-1.663546	-2.112895
H	-4.962598	-0.090317	-2.058418
H	-5.247817	-0.254187	0.363888
H	-5.658730	-1.842705	-0.284324
H	-1.108411	-0.506712	-0.837674
H	-3.036928	0.157551	1.342279
H	-3.326255	-2.665176	0.236734
H	-4.104714	-2.017156	1.705850
O	-3.002586	-0.081185	-1.490781
O	-1.460566	-1.145939	1.567035
C	-1.664216	-1.224333	2.972681
H	-2.045852	-0.273438	3.375875
H	-2.354892	-2.031828	3.248503
H	-0.688564	-1.433368	3.417941
C	-0.263071	3.520834	0.601485
C	-1.431935	4.313269	0.617952
C	-2.633262	3.773092	0.208256
C	-2.655467	2.422931	-0.167697
N	-1.579505	1.647940	-0.171724
H	-3.547830	4.355343	0.186540
H	-1.366314	5.346314	0.947825
C	-0.358111	2.167247	0.156693
C	2.092672	1.960155	0.479007
C	1.000322	4.059662	1.019385
C	2.131222	3.302978	0.979323
H	1.028773	5.087483	1.369350
H	3.084715	3.710742	1.303531
H	-3.585596	1.949036	-0.467952
C	0.864090	1.385193	0.043225
C	1.915529	-0.567230	-0.594471
C	3.259025	1.165428	0.375887
C	3.174125	-0.105738	-0.153823
H	1.810356	-1.553311	-1.040913
H	4.210041	1.572092	0.709375
H	4.049257	-0.740782	-0.246443
N	0.807330	0.150323	-0.501538
Br	-1.509336	-3.098709	-1.937457



**Early-Int\_Phenanthroline**

Charge = 0 Multiplicity = 1

E(RB3LYP) = -3529.117038 A.U.

C	3.413047	-1.615412	0.157217
C	2.730066	-1.289559	-1.165587
C	0.621486	-1.036414	-0.116946
C	1.163775	-1.413635	1.273045
C	2.637886	-0.984231	1.326668
H	2.778037	-0.211542	-1.364888
H	3.163791	-1.839863	-2.004129
H	3.480130	-2.704501	0.284276
H	4.436301	-1.223479	0.133495
H	0.632982	0.042238	-0.253042
H	1.092668	-2.507866	1.404565
H	2.690083	0.108903	1.252686
H	3.079820	-1.286894	2.281772
O	1.336241	-1.705391	-1.134053
O	0.326505	-0.773834	2.227565
C	0.566309	-1.155354	3.578381
H	0.537703	-2.249383	3.694402
H	1.528206	-0.780345	3.948803
H	-0.235231	-0.713082	4.175176
C	-3.213101	-1.298790	-0.237435
C	-3.310406	-2.702530	-0.317783
C	-2.171879	-3.475019	-0.407028
C	-0.941174	-2.834135	-0.364426
N	-0.816108	-1.496803	-0.267639
H	-2.206728	-4.554814	-0.488265
H	-4.295584	-3.159589	-0.316734
C	-1.930940	-0.666277	-0.244183
C	-3.118578	1.514077	-0.170416
C	-4.417531	-0.523149	-0.160447
C	-4.374520	0.833649	-0.113672
H	-5.362310	-1.056986	-0.143424
H	-5.285944	1.421191	-0.053090
H	-0.012150	-3.384695	-0.409170
C	-1.888026	0.793759	-0.248401
C	-0.670384	2.763919	-0.365159
C	-3.053118	2.928412	-0.177673
C	-1.832211	3.560297	-0.273587
H	0.328258	3.192543	-0.449560
H	-3.977120	3.496562	-0.115464
H	-1.753229	4.642633	-0.283105
N	-0.709938	1.442268	-0.353247
Br	2.966011	2.469945	-0.561160

### Late-Int\_Phenanthroline

Charge = 1 Multiplicity = 1

E(RB3LYP) = -1072.953844 A.U.

C	3.948746	0.006459	-0.600737
C	2.738404	-0.753398	-0.036707
C	1.666931	-0.844725	-1.139819
C	3.268131	-0.775464	-2.888302
C	4.427835	-0.640844	-1.910023
H	1.284929	0.143133	-1.385831
H	3.047814	-1.765906	0.260604
H	3.652559	1.047386	-0.783395
H	4.754107	0.018587	0.140380
H	3.537440	-1.359901	-3.770562
H	2.907806	0.209865	-3.216492
H	4.851043	-1.633361	-1.710807
H	5.218884	-0.034976	-2.365866
O	2.169425	-1.498264	-2.283796
O	2.137065	-0.101851	1.083636
C	2.788461	-0.357602	2.335317
H	2.174019	0.106721	3.109131
H	2.848500	-1.436870	2.525286
H	3.795812	0.073725	2.364849
C	-1.674590	-2.016109	0.295172
C	-1.330875	-3.364414	0.524382
C	-0.117094	-3.858388	0.092077
C	0.780475	-2.971038	-0.484598
N	0.489658	-1.670838	-0.687550
H	-2.043836	-4.007321	1.031896
H	0.166742	-4.895859	0.221216
H	1.771238	-3.280822	-0.787921
C	-0.767191	-1.154374	-0.394254
C	-2.946512	-1.531671	0.745482
C	-2.507212	0.606390	-0.292660
C	-3.337579	-0.256391	0.489375
H	-3.585785	-2.216981	1.292473
H	-4.296324	0.115571	0.837841
C	-1.233404	0.175534	-0.772599
C	-0.952656	2.149067	-1.948778
C	-2.950053	1.899417	-0.660739
C	-2.174147	2.680677	-1.487706
H	-0.323896	2.725273	-2.623544
H	-3.910336	2.250639	-0.293896
H	-2.487028	3.673773	-1.791963
N	-0.501063	0.954106	-1.604767
C	2.460689	3.392179	1.902387
H	2.263773	4.467325	1.861927

H	3.485275	3.217565	1.544107
H	2.394301	3.071400	2.951962
O	1.487303	2.751086	1.081629
H	1.685955	1.798711	1.045207

### TS2\_Phenanthroline

Charge = 1 Multiplicity = 1

E(RB3LYP) = -1072.925835 A.U.

C	3.041997	-1.145325	1.339295
C	1.879416	-0.202137	1.004107
C	1.661776	-0.064409	-0.495591
C	3.128566	-1.944227	-1.018879
C	2.982154	-2.371789	0.423766
H	1.199705	0.848806	-0.859277
H	0.944991	-0.593364	1.423993
H	3.985838	-0.609668	1.190767
H	2.984313	-1.437963	2.390568
H	2.853009	-2.710168	-1.743885
H	4.129879	-1.571742	-1.247143
H	2.046096	-2.923772	0.569718
H	3.801693	-3.062246	0.648375
O	2.246623	-0.789953	-1.368239
O	2.097863	1.126162	1.468289
C	1.581499	1.393847	2.779342
H	1.816992	2.437283	2.996186
H	0.494537	1.252751	2.804039
H	2.057113	0.754587	3.532703
C	-2.980646	-0.856853	-0.780681
C	-2.946317	-2.239871	-1.067468
C	-1.731675	-2.878409	-1.206344
C	-0.560020	-2.121933	-1.031315
N	-0.555133	-0.826461	-0.745513
H	-3.880864	-2.781780	-1.182656
H	-1.663734	-3.935692	-1.438388
H	0.408258	-2.604693	-1.128247
C	-1.741801	-0.164190	-0.637365
C	-4.224652	-0.153918	-0.643256
C	-3.026414	1.924645	-0.260732
C	-4.247870	1.182742	-0.384936
H	-5.147608	-0.715865	-0.752666
H	-5.190724	1.711767	-0.279239
C	-1.767906	1.269674	-0.390458
C	-0.625666	3.250610	-0.085594
C	-3.019499	3.318584	-0.021693
C	-1.816516	3.988663	0.069210

H	0.338047	3.752282	-0.027570
H	-3.963945	3.845245	0.083682
H	-1.774974	5.057931	0.249485
N	-0.595692	1.943300	-0.307167
C	4.369886	3.221619	-0.792391
H	4.791029	3.908122	-0.045279
H	3.421306	3.636085	-1.161327
H	5.067002	3.153985	-1.631673
O	4.207772	1.902889	-0.269087
H	3.573140	1.924271	0.467067

### **P.P.\_Phenanthroline**

Charge = 1 Multiplicity = 1

E(RB3LYP) = -1072.934271 A.U.

C	-4.461771	0.118736	-0.668452
C	-2.978264	0.497048	-0.652404
C	-2.114274	-0.468339	0.171724
C	-3.833932	-2.158233	0.149110
C	-4.608942	-1.390854	-0.904615
H	-1.035916	-0.335866	0.048101
H	-2.564408	0.457458	-1.670845
H	-4.915734	0.392003	0.292270
H	-4.978064	0.683617	-1.449854
H	-3.807940	-3.230831	-0.045232
H	-4.235250	-1.988095	1.153859
H	-4.245808	-1.664524	-1.902618
H	-5.662653	-1.684133	-0.850087
O	-2.419447	-1.769544	0.158155
O	-2.722091	1.783634	-0.079002
C	-3.134058	2.910277	-0.866027
H	-2.813483	3.800659	-0.322860
H	-2.647884	2.885207	-1.848412
H	-4.221739	2.935473	-0.988627
C	1.175683	-2.790508	-0.223724
C	2.385232	-3.484857	-0.429725
C	3.555516	-2.756553	-0.501630
C	3.509066	-1.349521	-0.368291
N	1.093102	-1.472249	-0.093095
H	4.512323	-3.246483	-0.661196
H	0.238555	-3.340168	-0.164982
H	2.382239	-4.565578	-0.529185
C	2.235641	-0.744250	-0.162245
C	3.358826	1.468823	-0.109203
C	4.692921	-0.539909	-0.438446
C	4.620504	0.815012	-0.314947
H	5.648936	-1.032003	-0.594729

H	5.517349	1.425970	-0.370482
C	2.158604	0.704883	-0.024875
C	3.256544	2.873420	0.016207
C	0.891444	2.605158	0.285692
C	2.017536	3.449753	0.212342
H	4.155635	3.481058	-0.044065
H	-0.097703	3.032939	0.438384
H	1.900196	4.524253	0.310169
N	0.947608	1.283463	0.176168
O	-2.356588	0.093293	1.686307
H	-2.431456	1.070481	1.532070
C	-1.313559	-0.240655	2.671156
H	-1.636175	0.194337	3.617258
H	-1.288839	-1.326541	2.736050
H	-0.358028	0.163756	2.337060

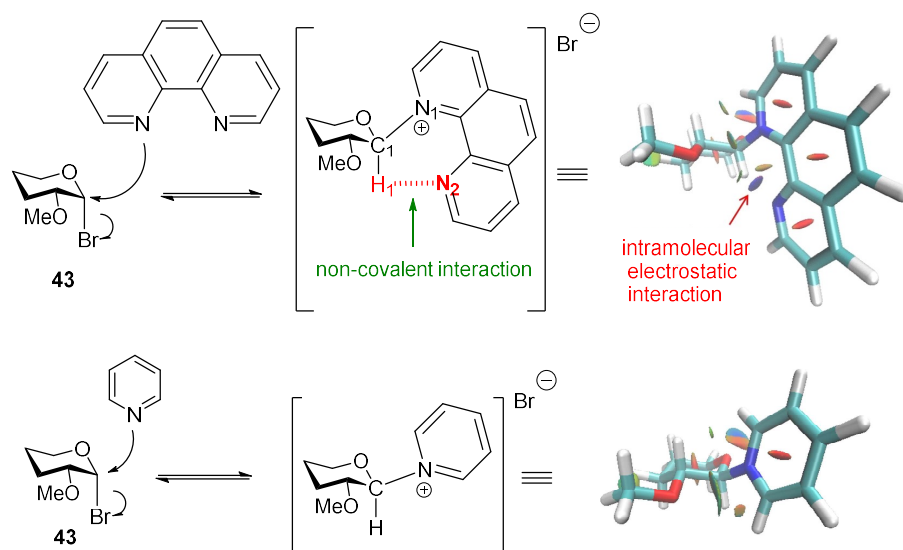
### F.P.

Charge = 0 Multiplicity = 1

E(RB3LYP) = -500.857189 A.U.

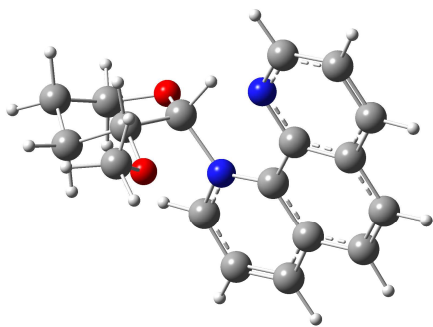
C	-0.971857	2.777809	0.206469
C	0.143537	1.891855	-0.340900
C	-0.402564	2.420275	-2.617915
C	-1.564834	3.322628	-2.158680
C	-1.289012	3.920576	-0.775019
H	1.093404	2.442269	-0.374092
H	0.286058	0.999275	0.274321
H	-1.869481	2.167448	0.370149
H	-0.669375	3.180538	1.180988
H	-0.679505	1.884649	-3.536240
H	-2.423899	2.646692	-2.069705
H	-0.440830	4.612784	-0.816608
H	-2.164644	4.487563	-0.438355
O	-0.171255	1.394694	-1.657549
O	-1.971671	4.247518	-3.169780
C	-1.274600	5.489851	-3.273648
H	-1.444181	6.128026	-2.396592
H	-0.199287	5.349351	-3.420718
H	-1.695065	5.991785	-4.149668
C	1.806428	2.500481	-3.498558
H	2.602142	3.227343	-3.678294
H	1.470061	2.088219	-4.460408
H	2.194024	1.685755	-2.876070
O	0.746794	3.202677	-2.847322

**Figure S18. Non-Covalent Interaction (NCI) Plot**



Non-covalent interactions plot (reduced density gradient isosurface = 0.3) for the optimized structure at B3LYP/6-31+G(d,p). Blue surfaces represent attractive interactions, and red surfaces represent repulsive interactions.

Figure S19.  $\alpha$ -Phenanthroline Ion Complex



**Cartesian coordinate**

Charge = 1 Multiplicity = 1

E(RB3LYP) = -957.199378997 A.U.

C	4.086881	-0.473795	1.063480
C	3.948378	-0.865978	-0.395503
C	1.507094	-0.314190	-0.297910
C	1.738606	0.372746	1.081503
C	3.200836	0.743180	1.338816
H	4.416635	-0.108746	-1.038505
H	4.424230	-1.824800	-0.613053
H	3.796730	-1.311996	1.709529
H	5.139878	-0.252583	1.269229
H	1.434340	-0.381216	1.822433
H	3.491811	1.579591	0.690013
H	3.304992	1.084245	2.373517
O	2.566786	-1.066868	-0.802839
O	0.873471	1.496010	1.162405
C	0.396212	1.774747	2.477277
H	-0.183311	0.930033	2.873686
H	1.214702	2.008021	3.170599
H	-0.253776	2.648778	2.394501
C	-2.744152	0.654767	-1.468700
C	-3.848522	0.038739	-0.833144
C	-3.642805	-0.826952	0.217777
C	-2.315528	-1.061228	0.627755
N	-1.262677	-0.498310	0.056632
H	-4.467824	-1.314480	0.726314
H	-4.850053	0.264515	-1.187833
C	-1.432556	0.344288	-0.991055
C	-0.549776	1.895644	-2.710343
C	-2.950416	1.566943	-2.549713
C	-1.889094	2.172815	-3.139651
H	-3.965843	1.768384	-2.877275
H	-2.023079	2.877420	-3.953941
H	-2.109883	-1.731477	1.459715
C	-0.287031	0.968310	-1.654181

C	2.038401	1.368866	-1.983280
C	0.513801	2.557314	-3.357233
C	1.815517	2.302823	-2.987059
H	3.042471	1.118257	-1.682457
H	0.285304	3.264720	-4.148495
H	2.660303	2.792718	-3.456058
N	1.047694	0.724056	-1.345547
H	0.661735	-0.983060	-0.219510

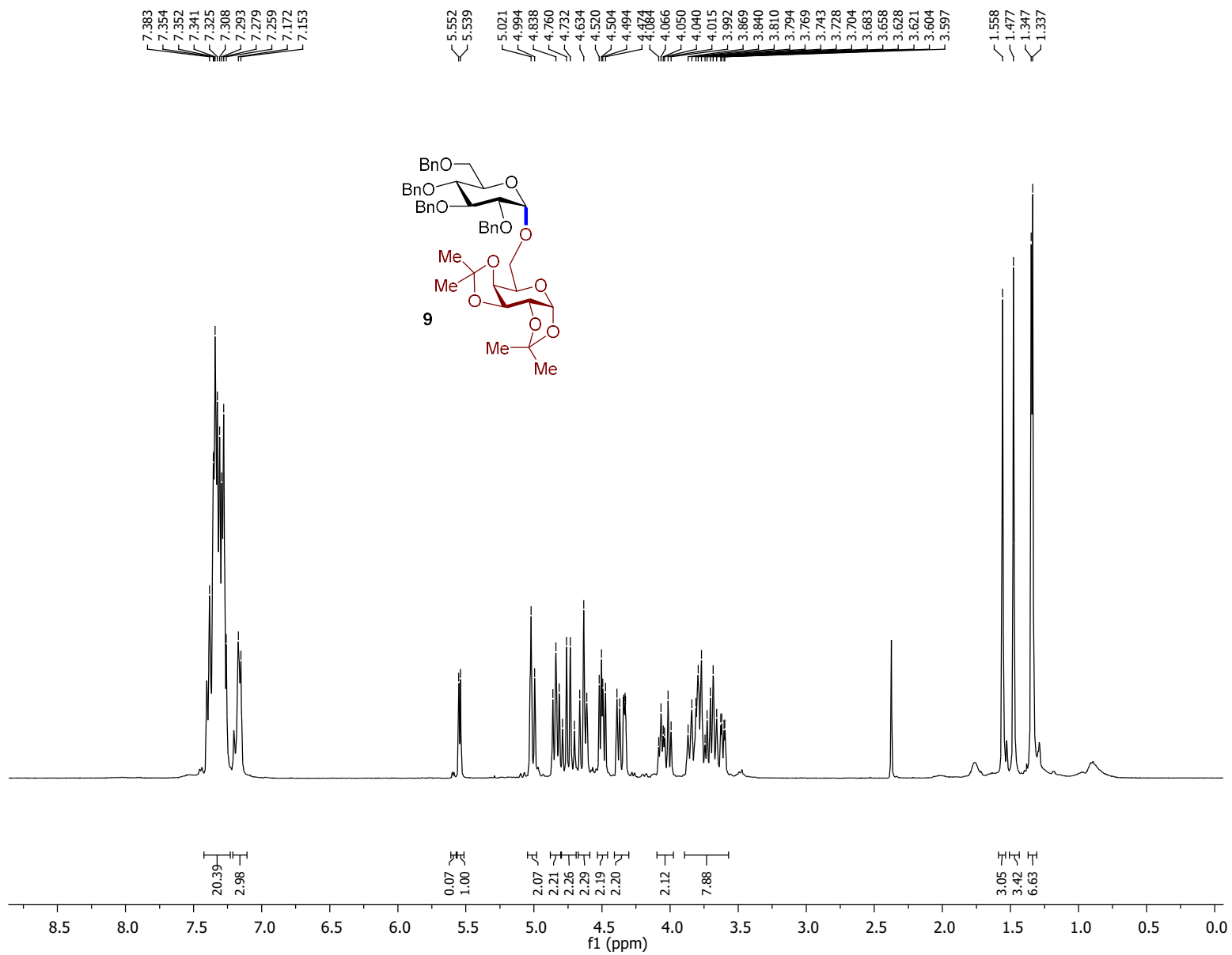


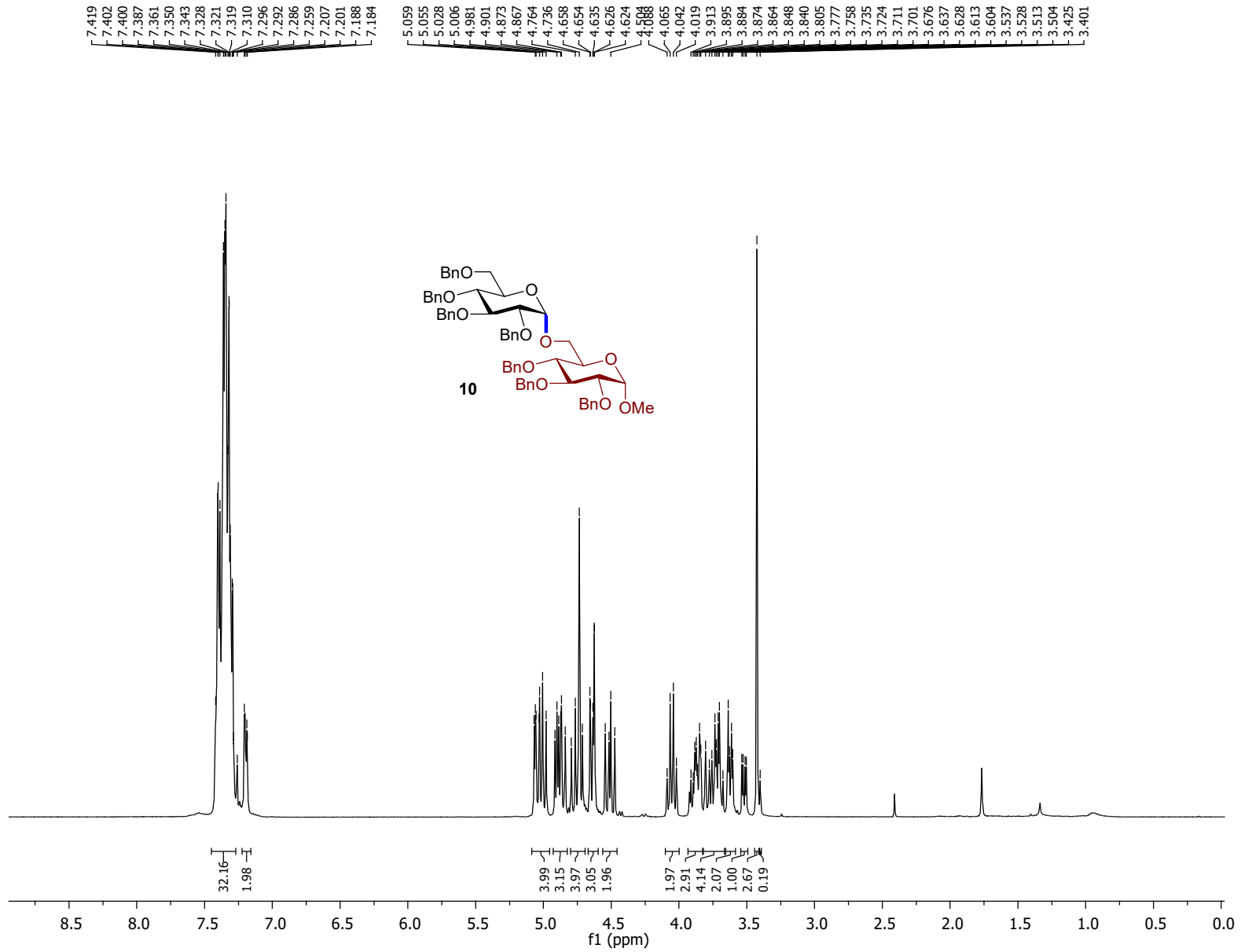
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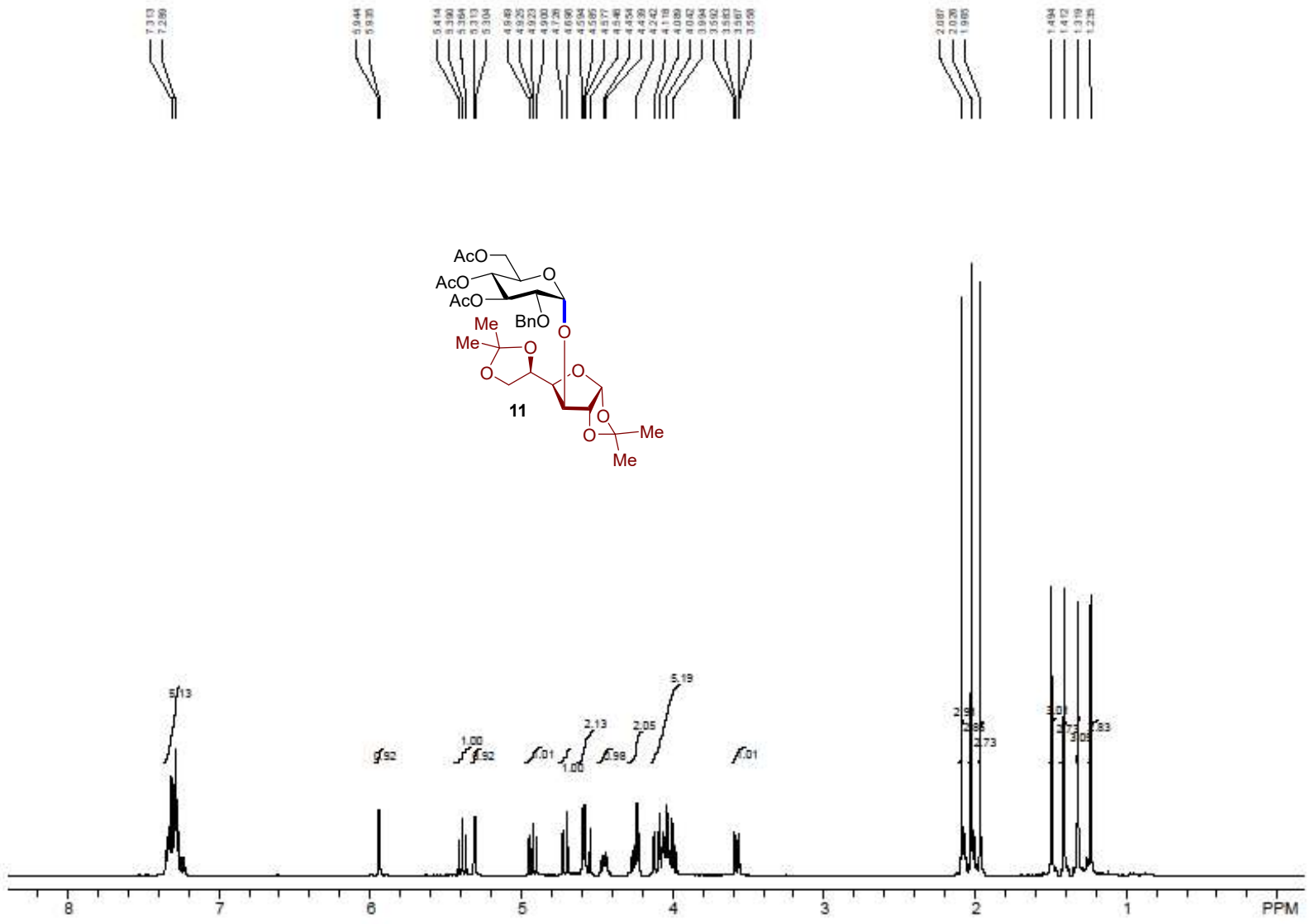
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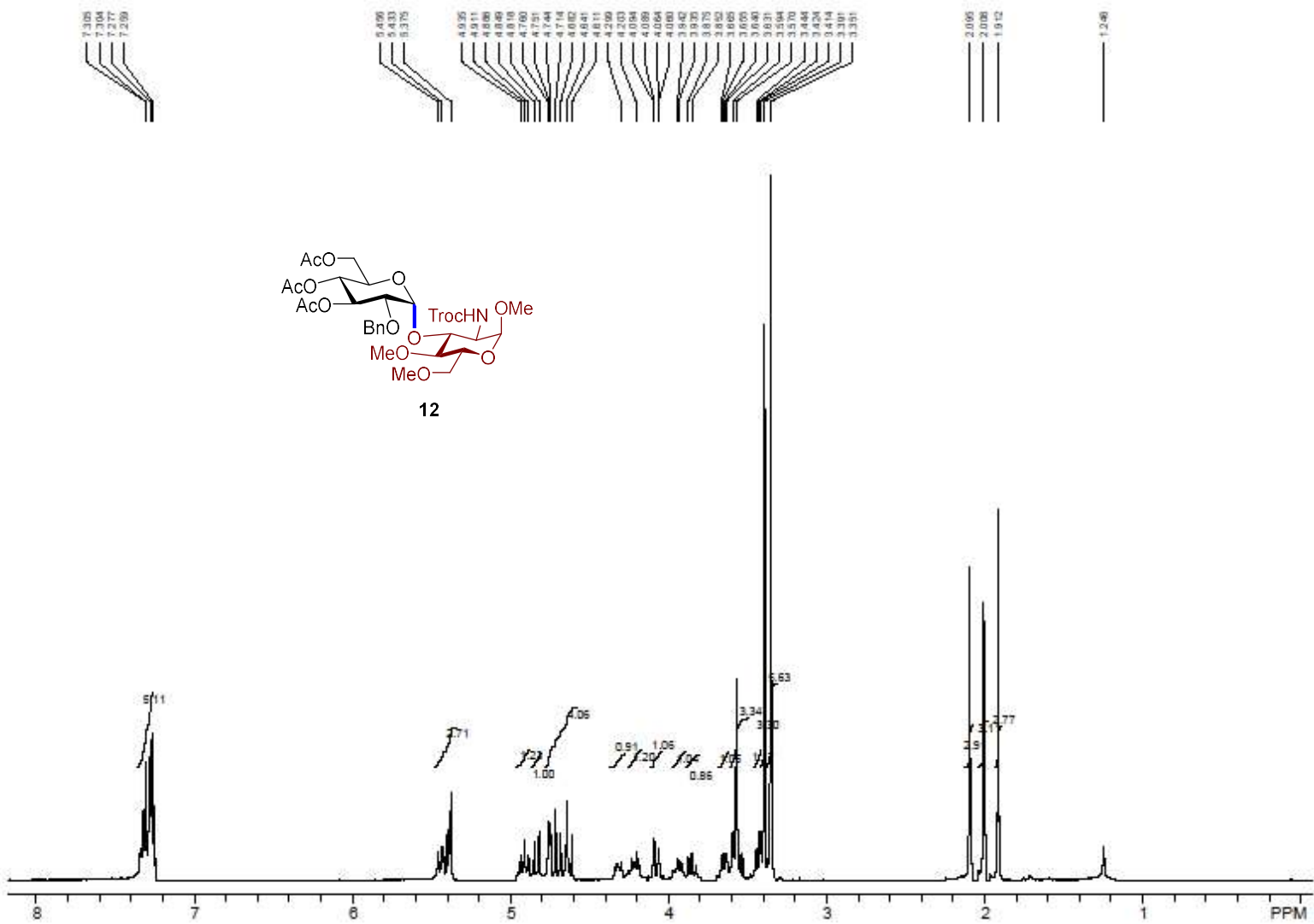
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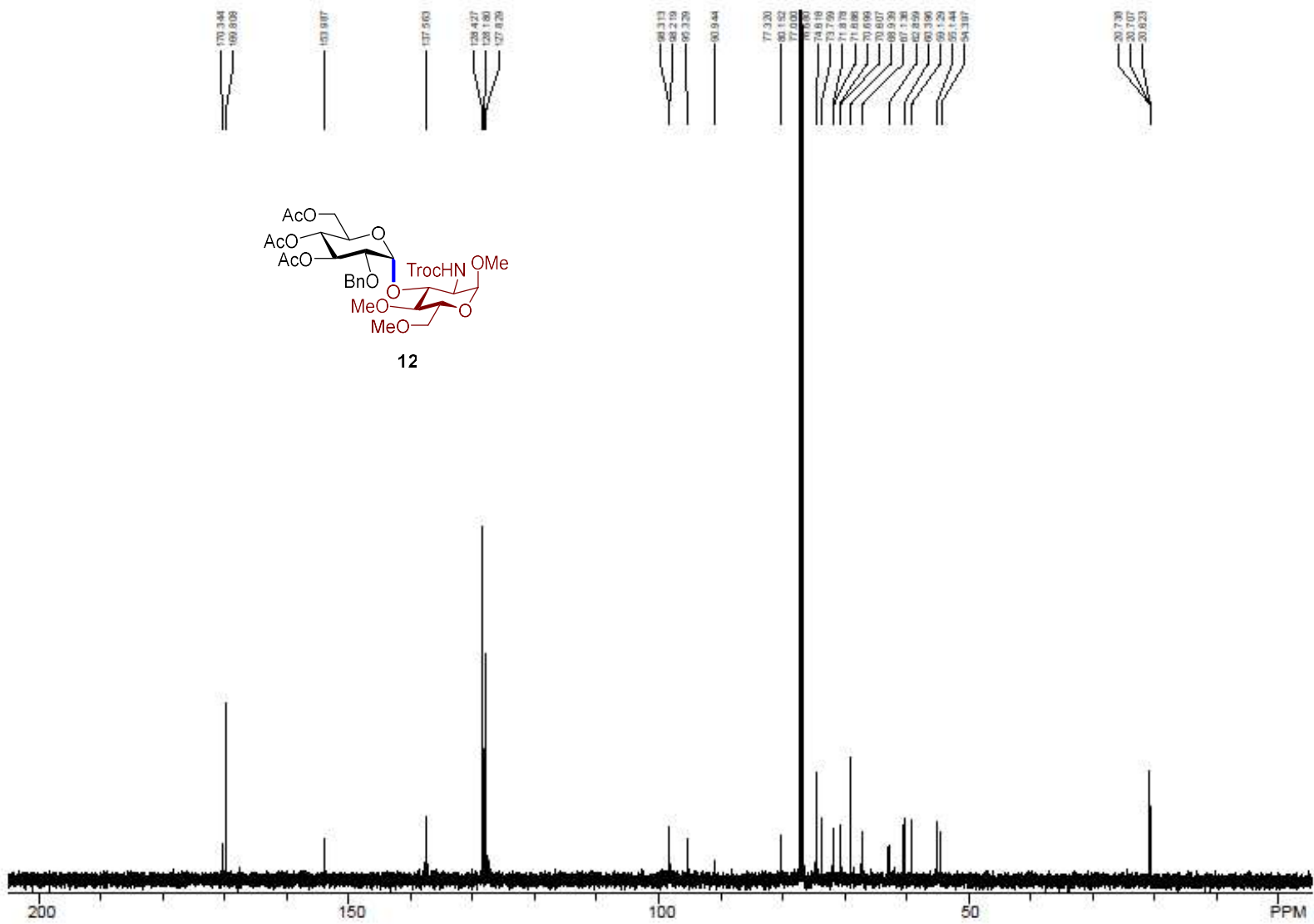




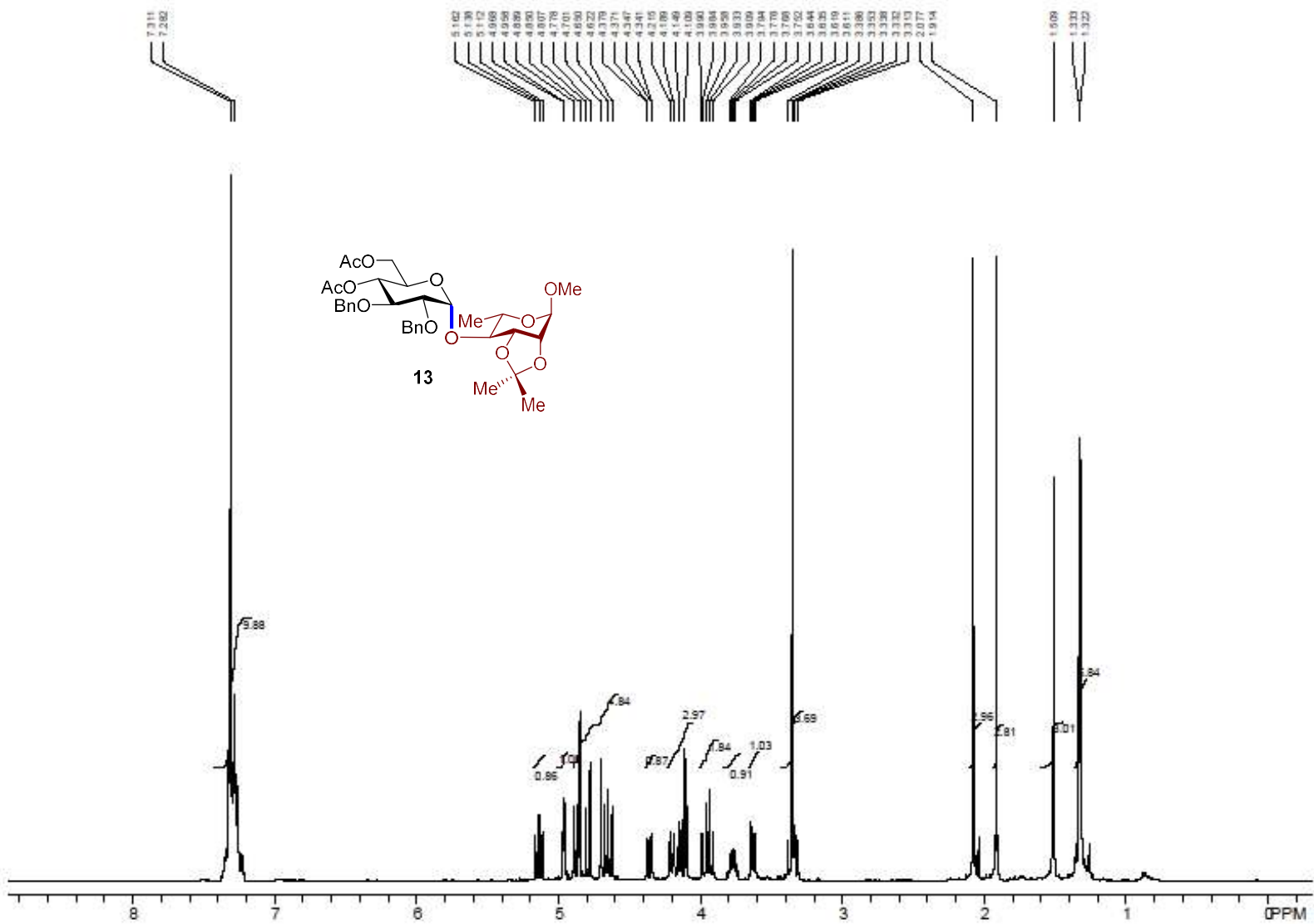


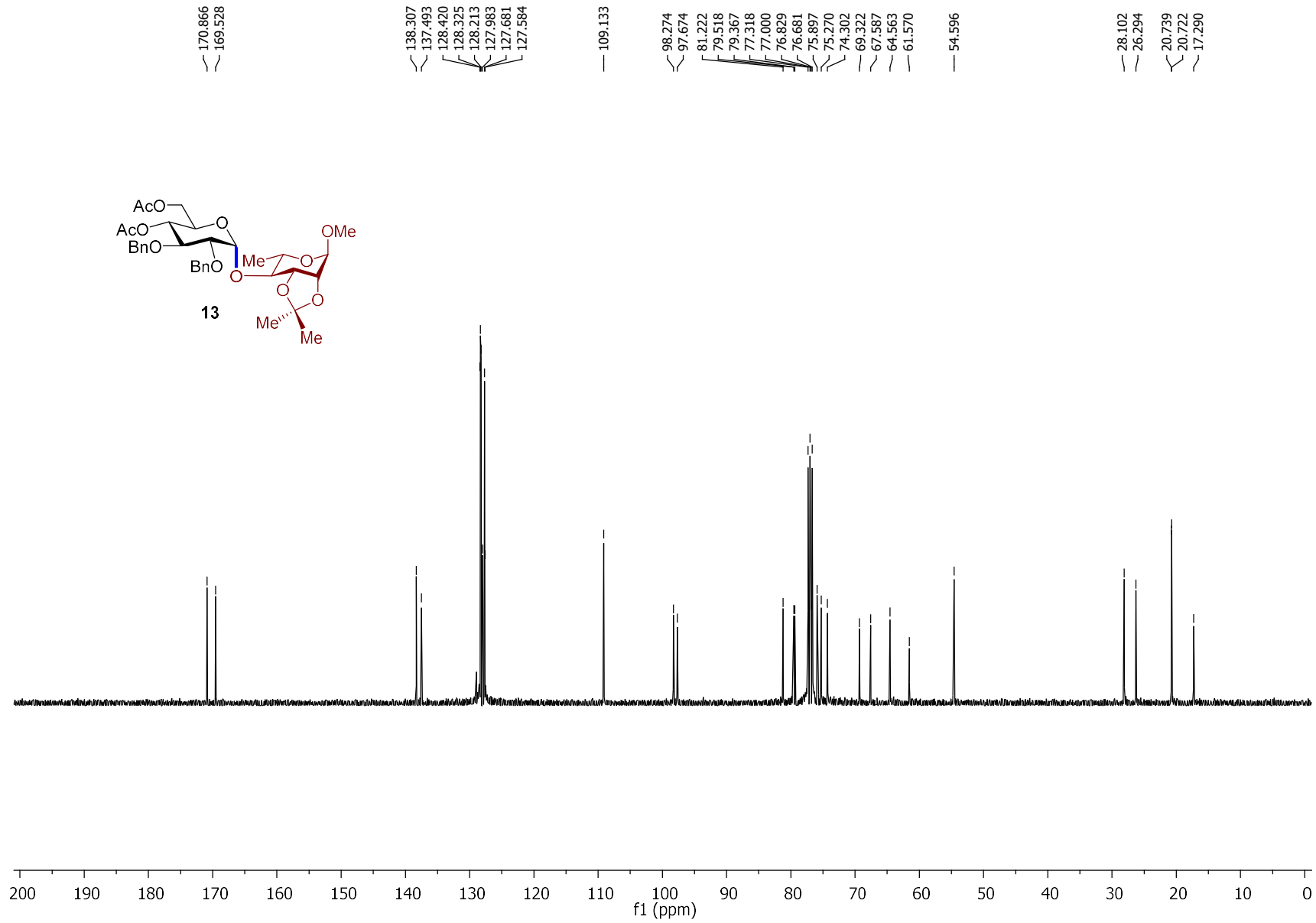


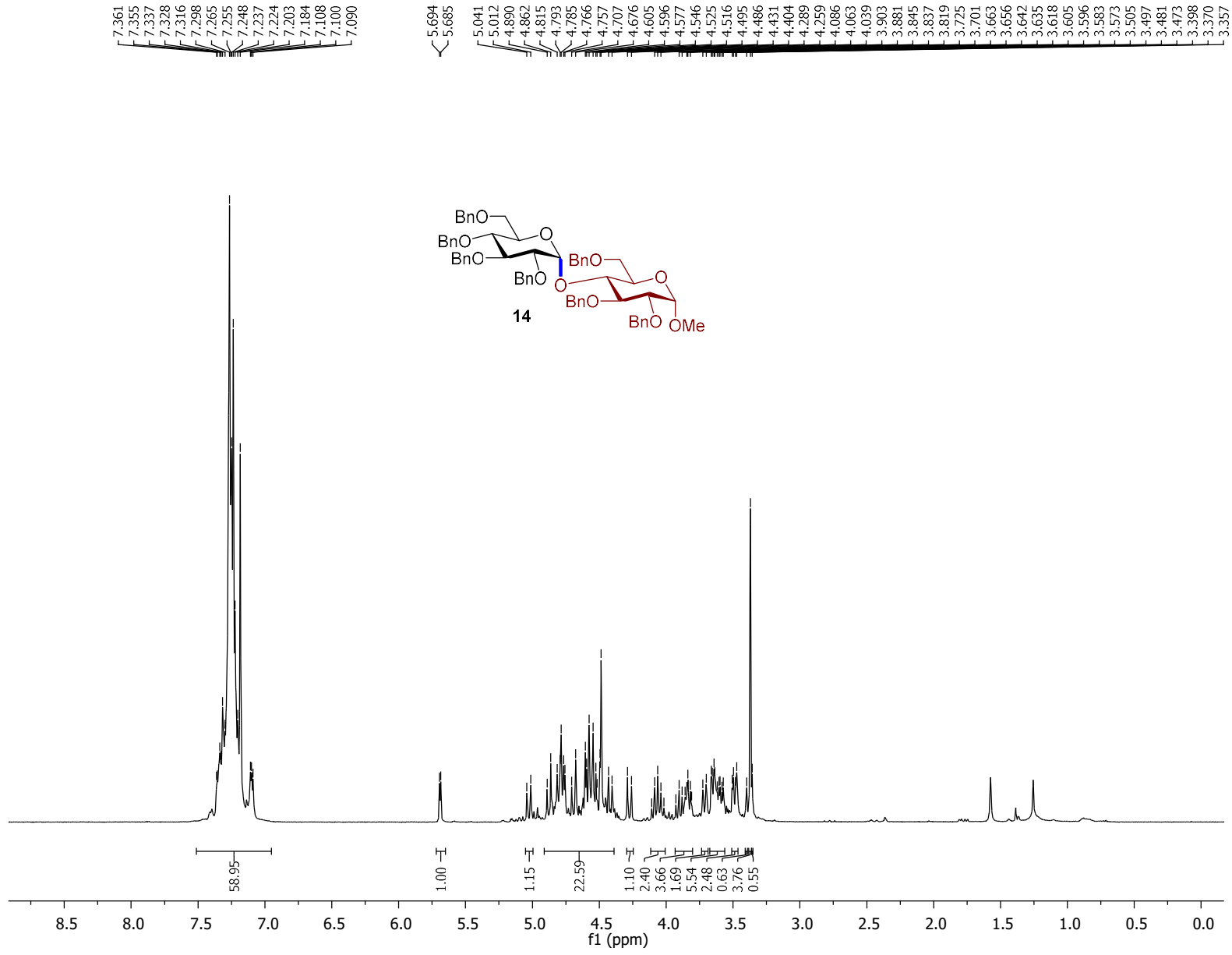


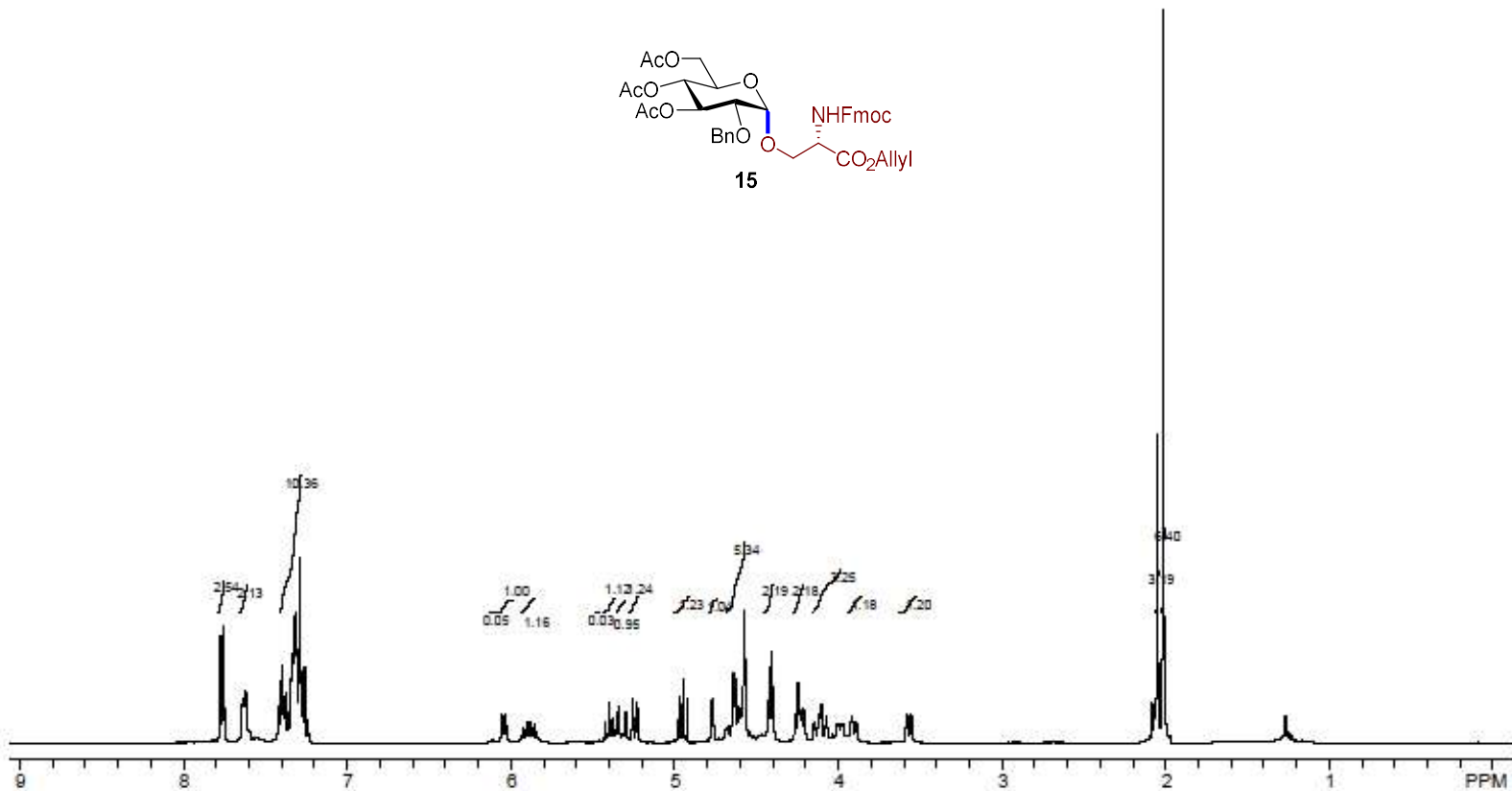
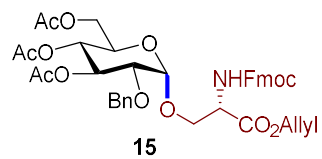
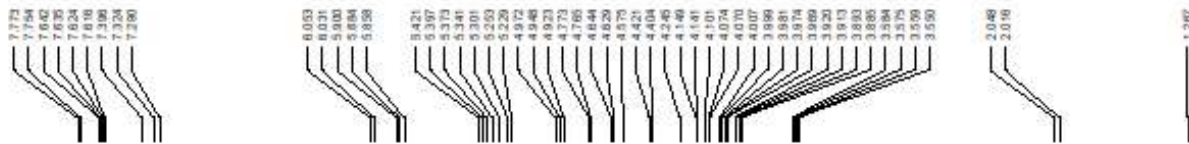


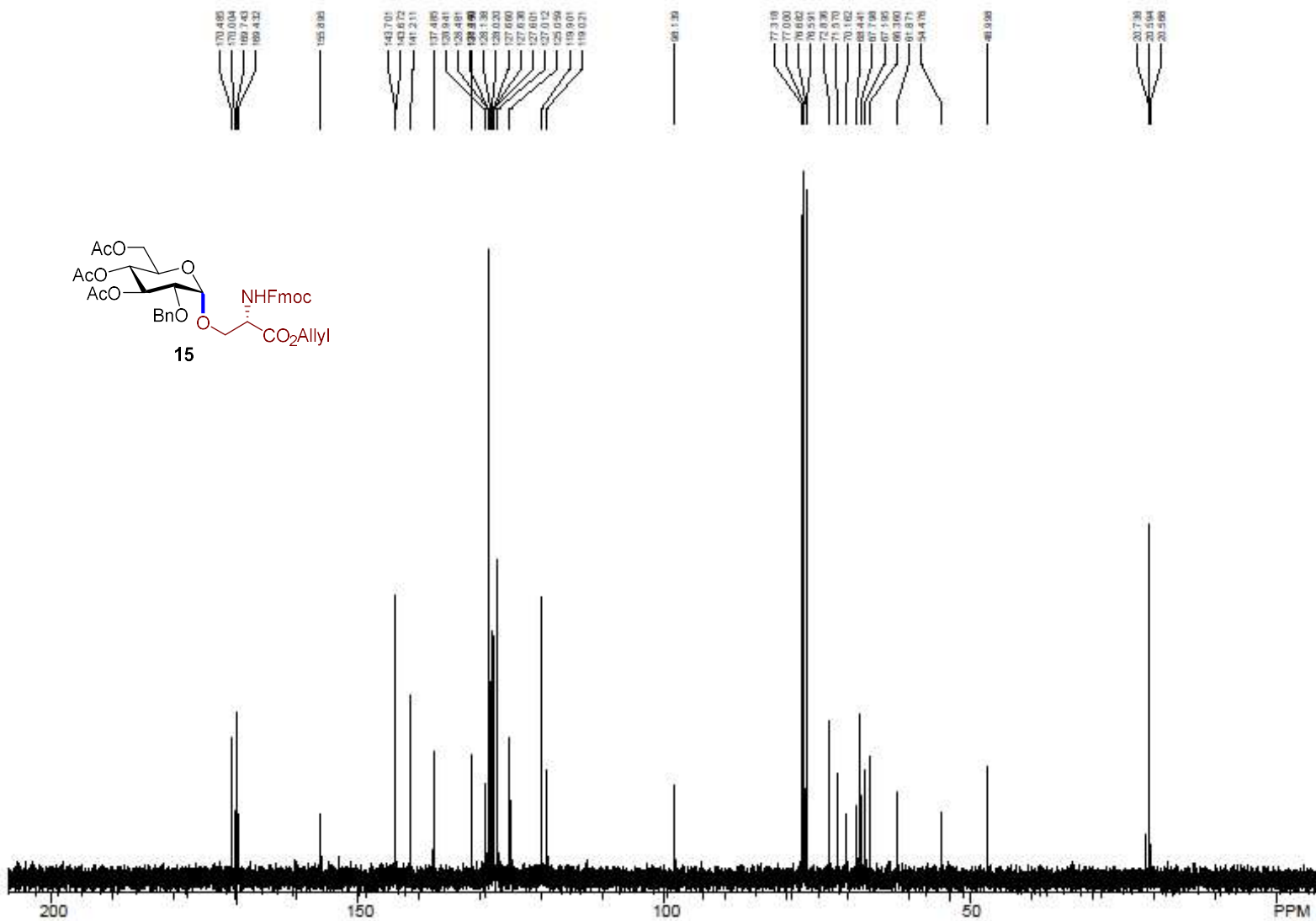


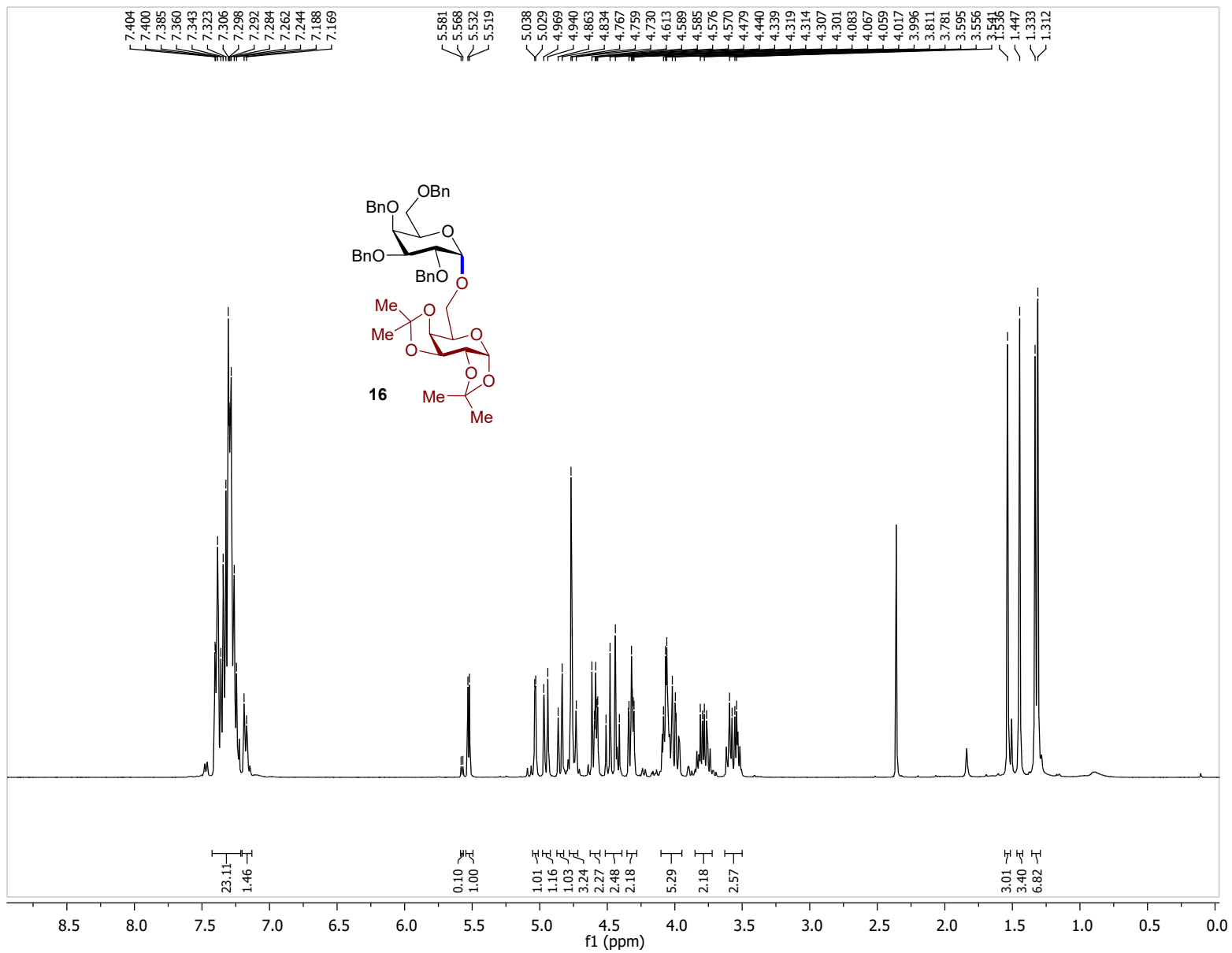


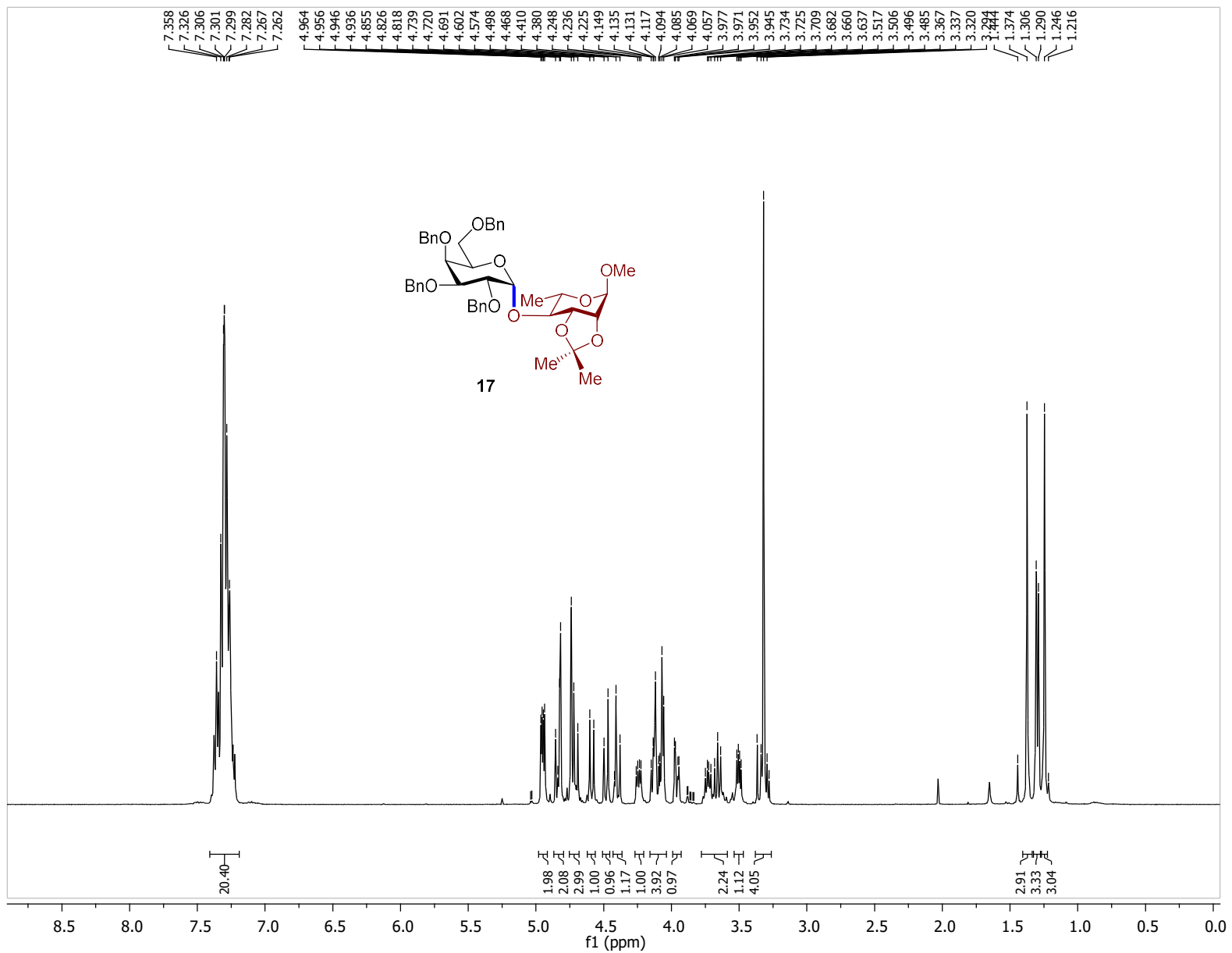


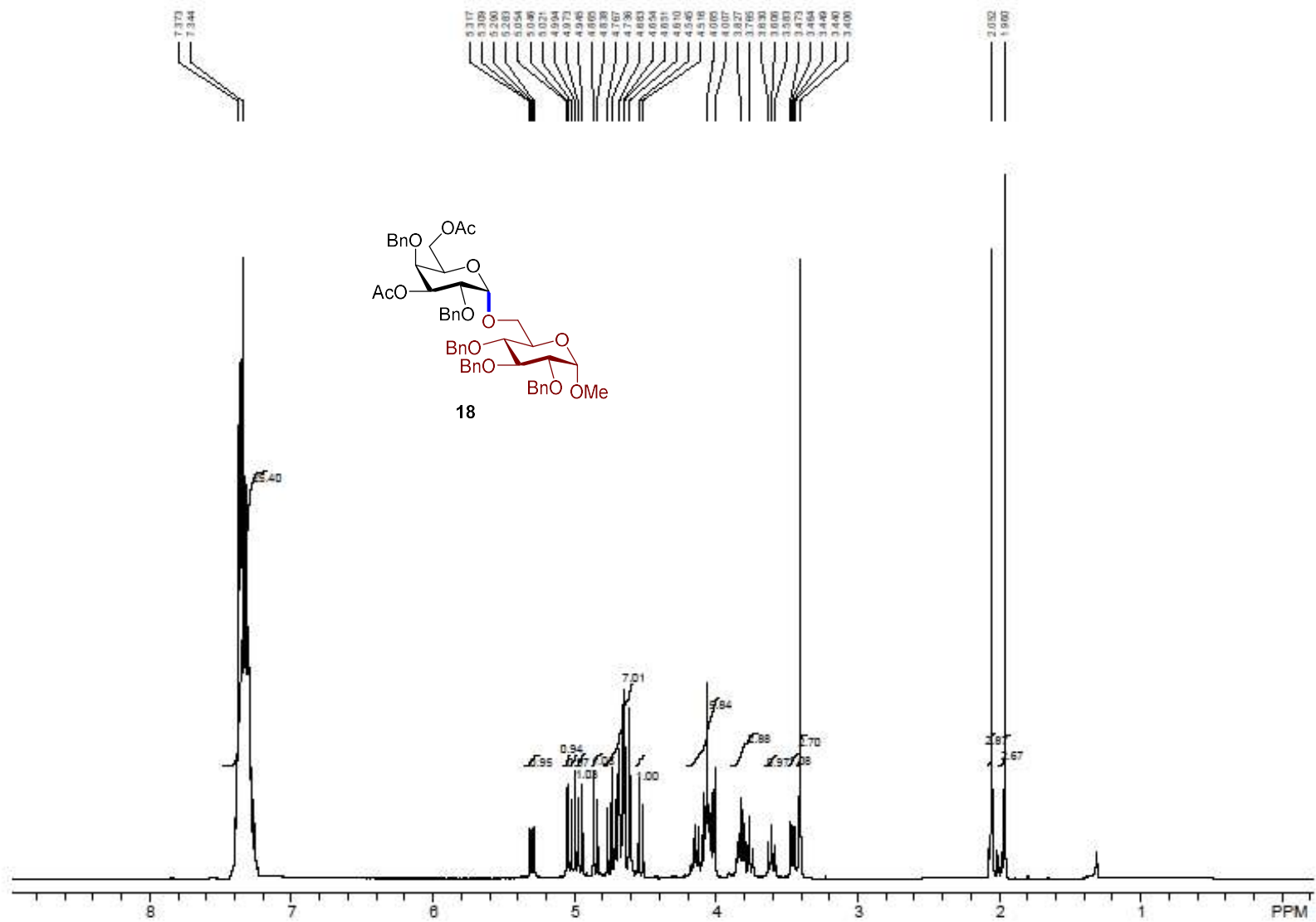




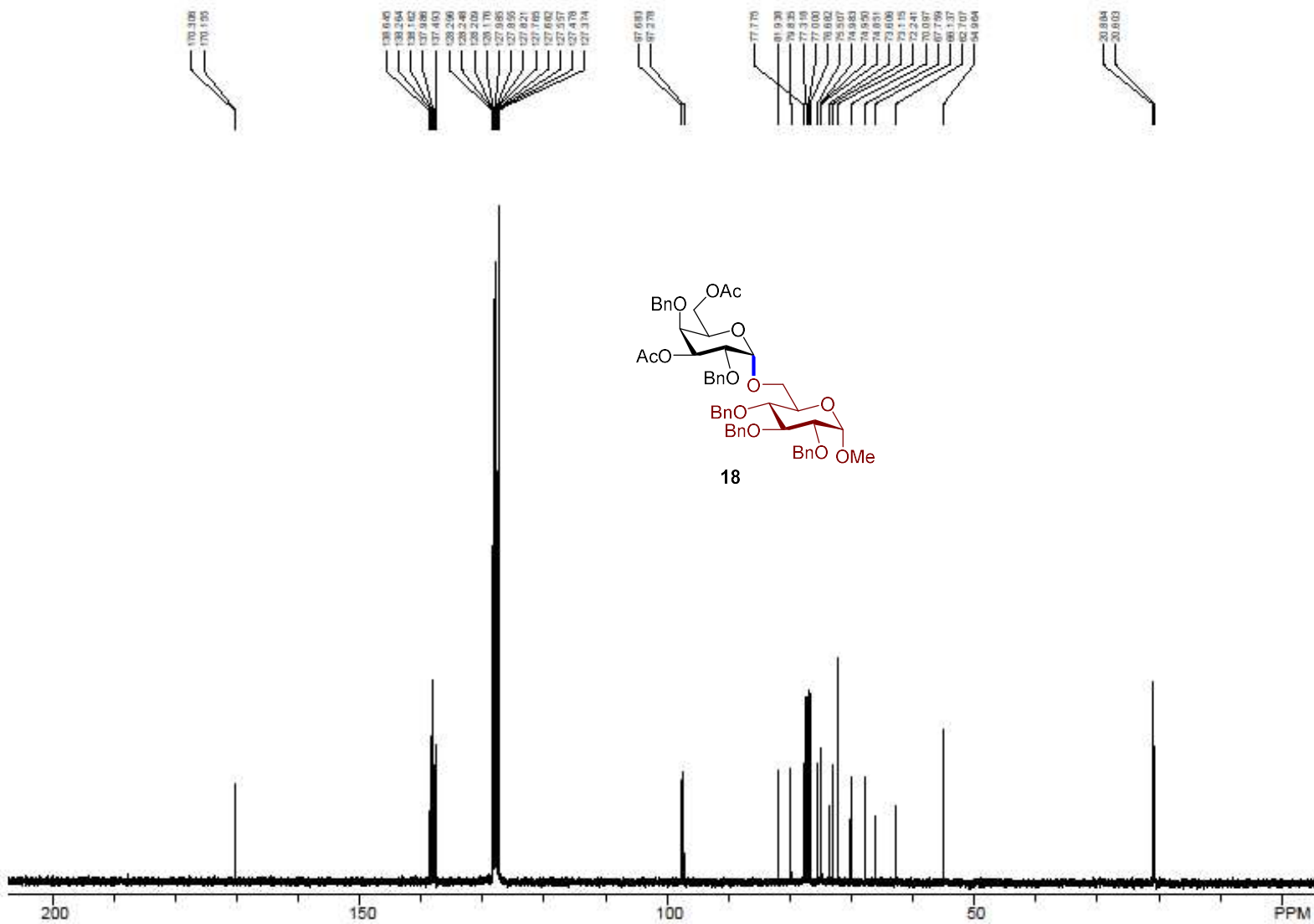


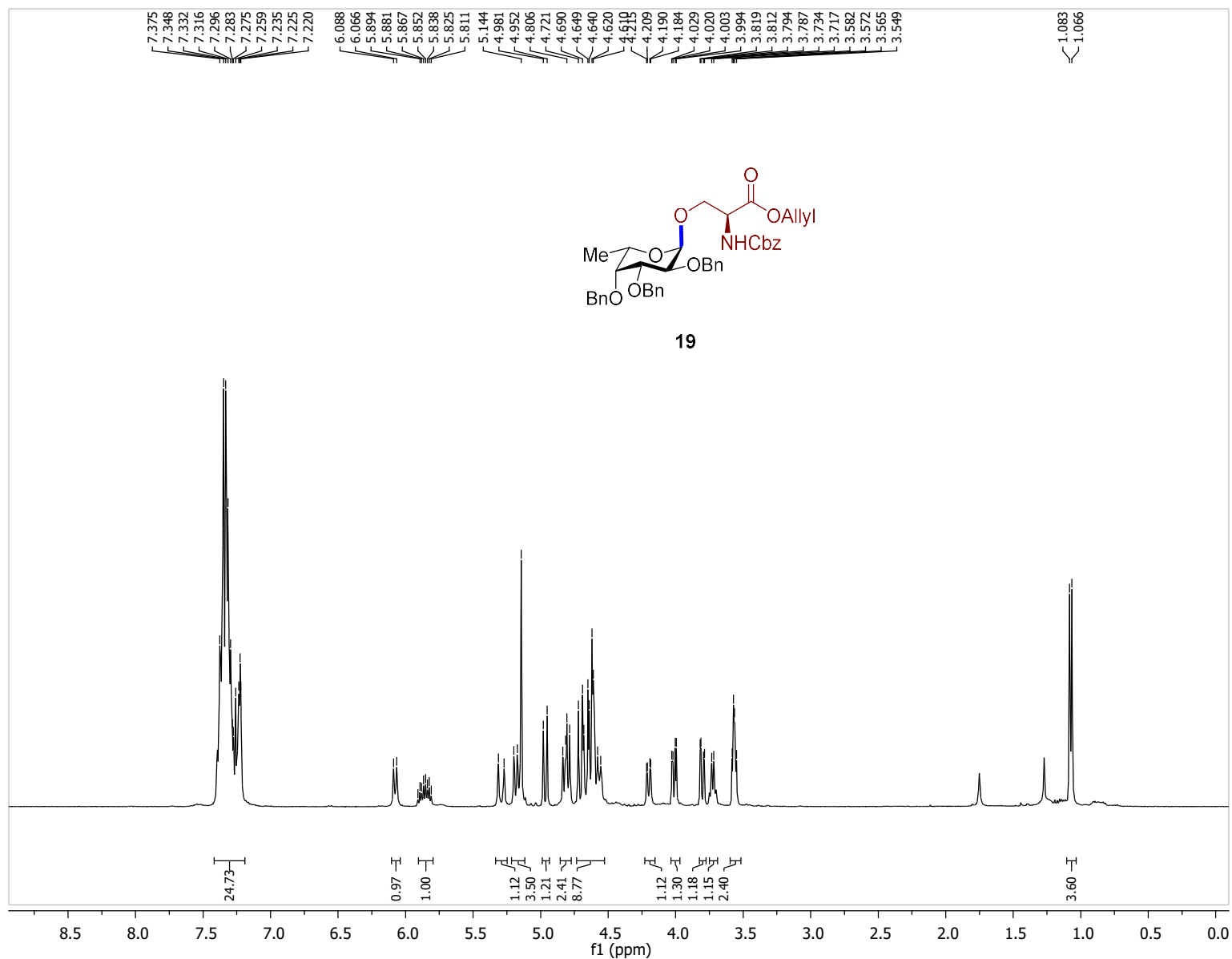


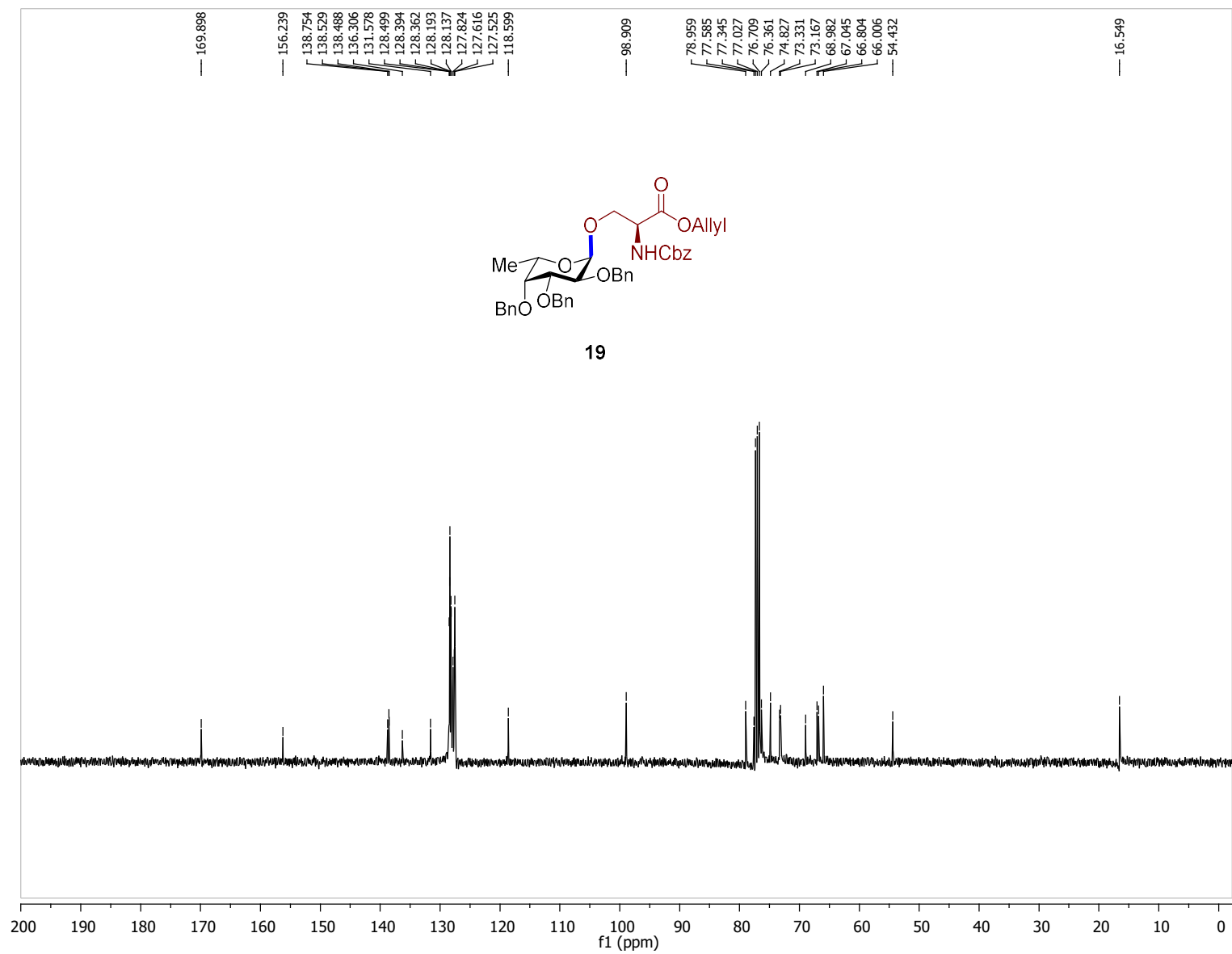


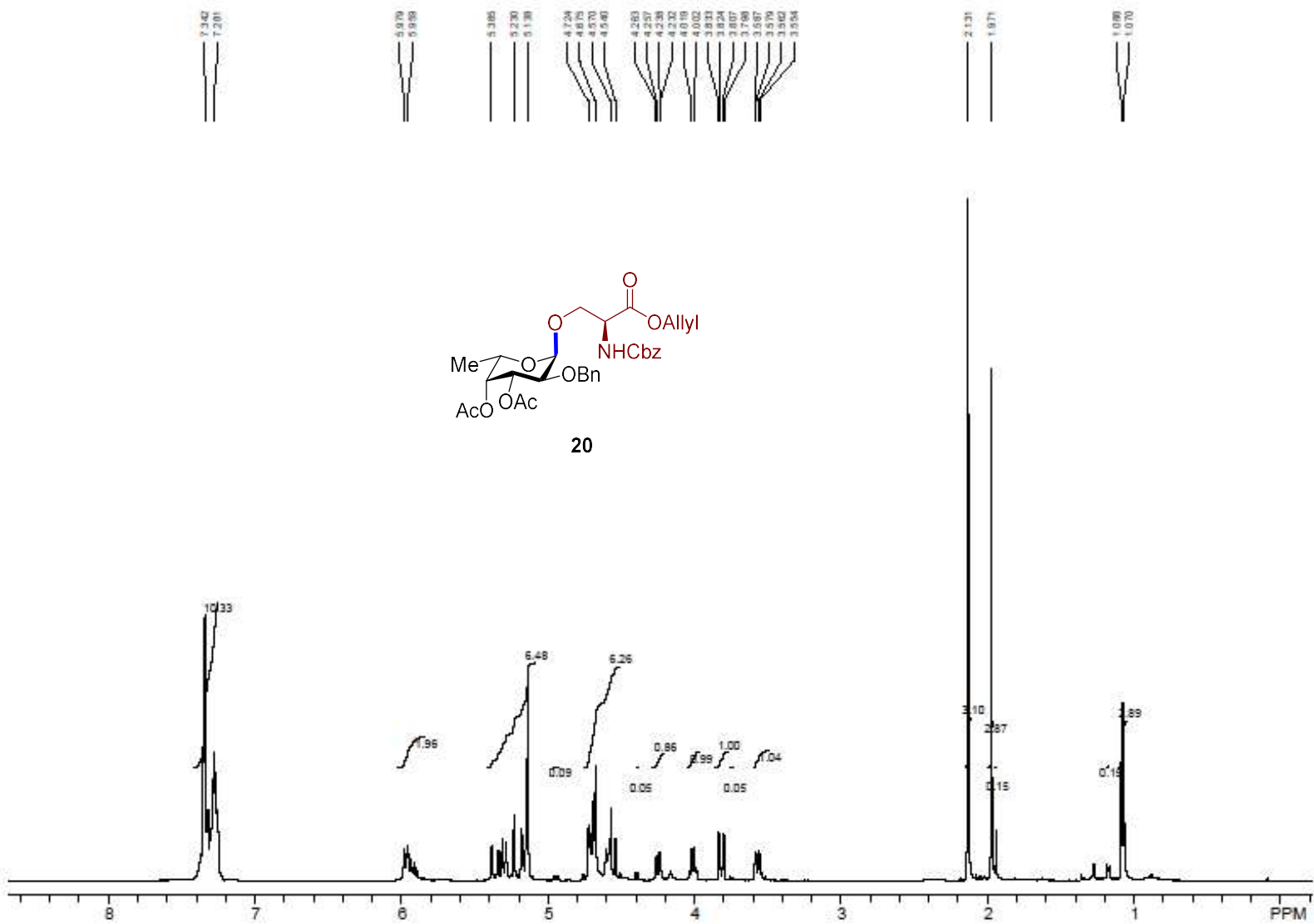


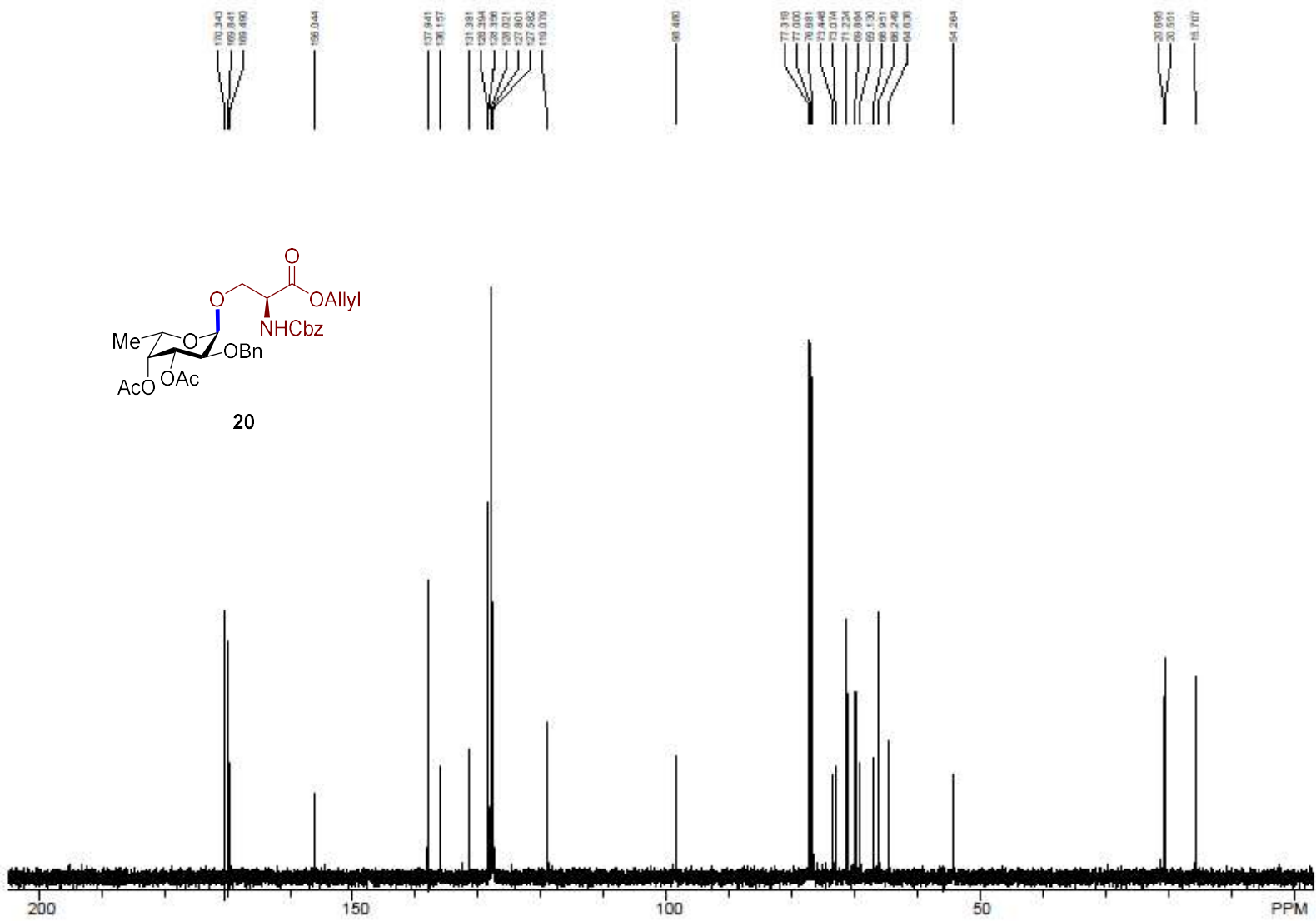




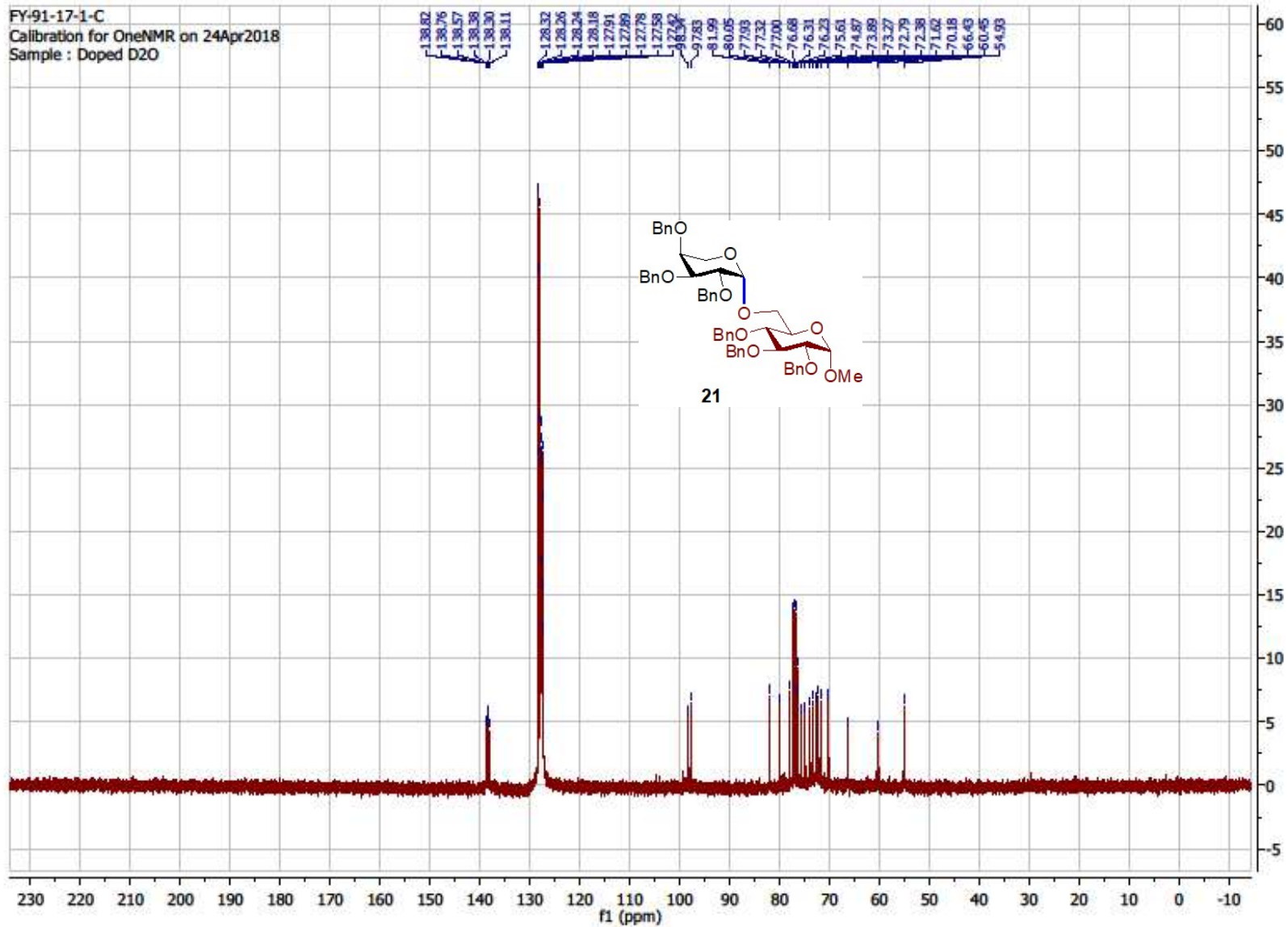


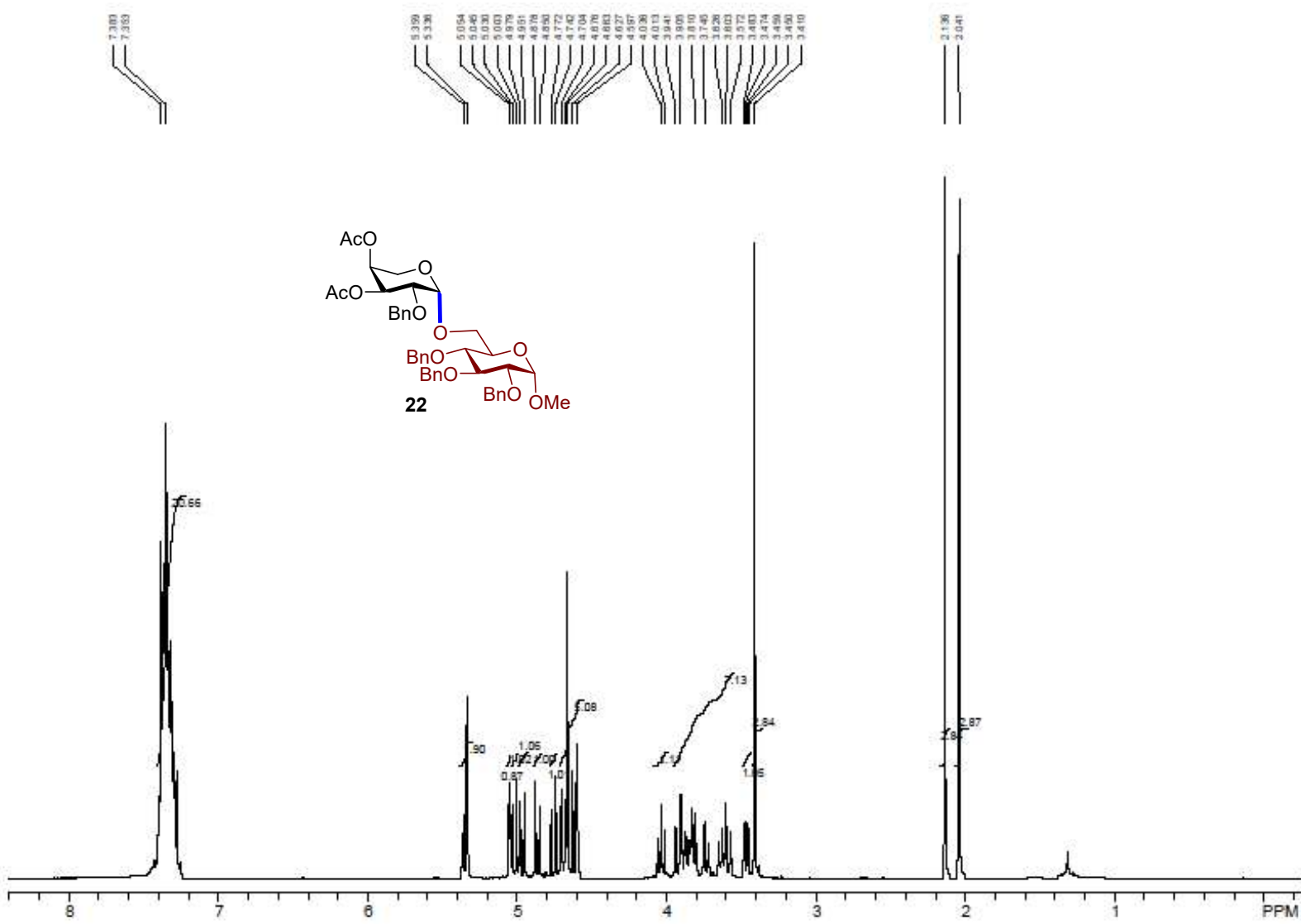






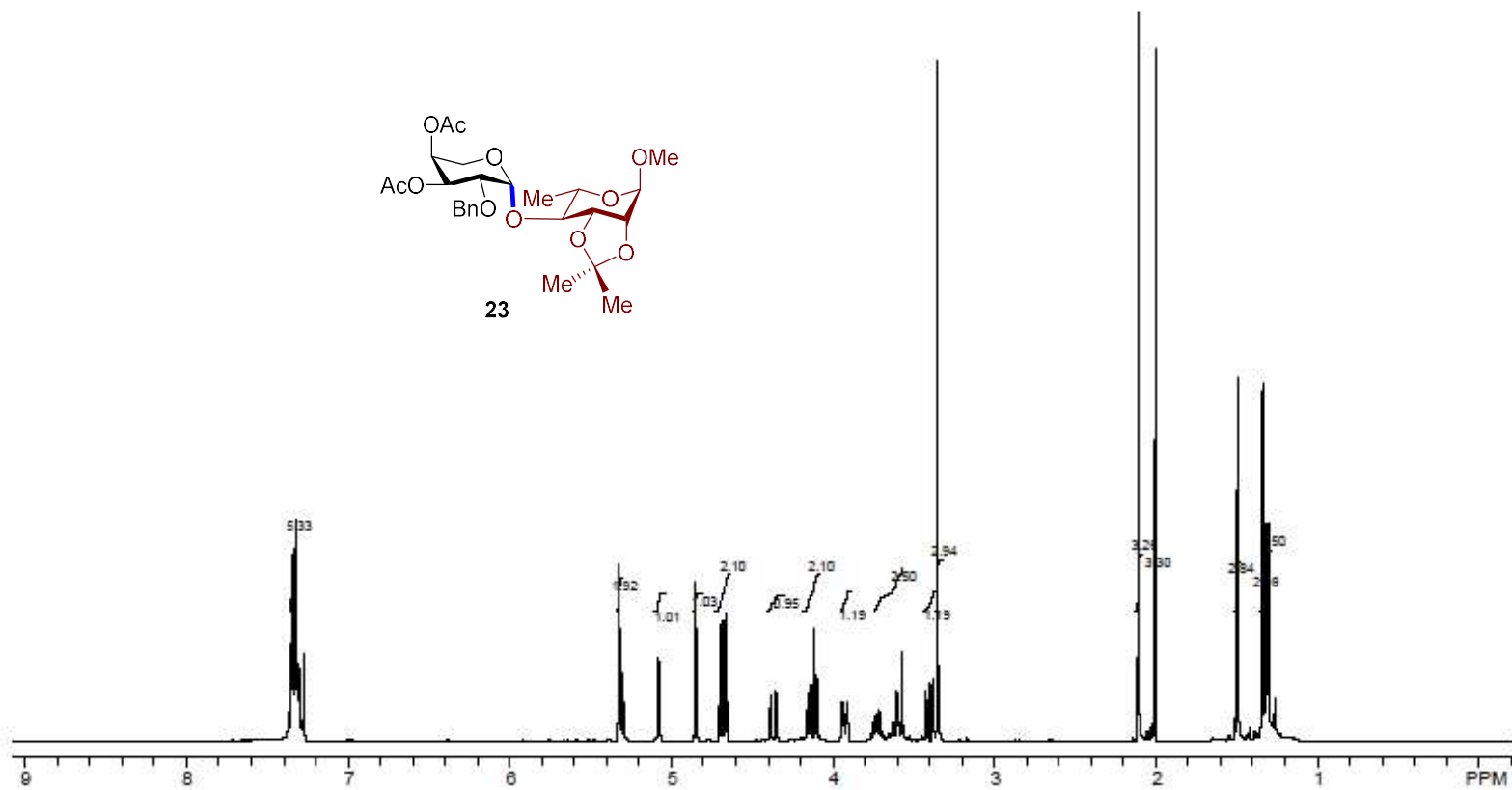
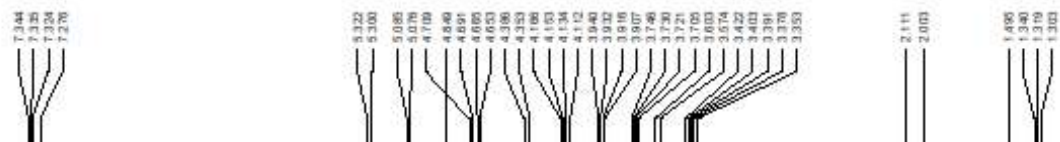


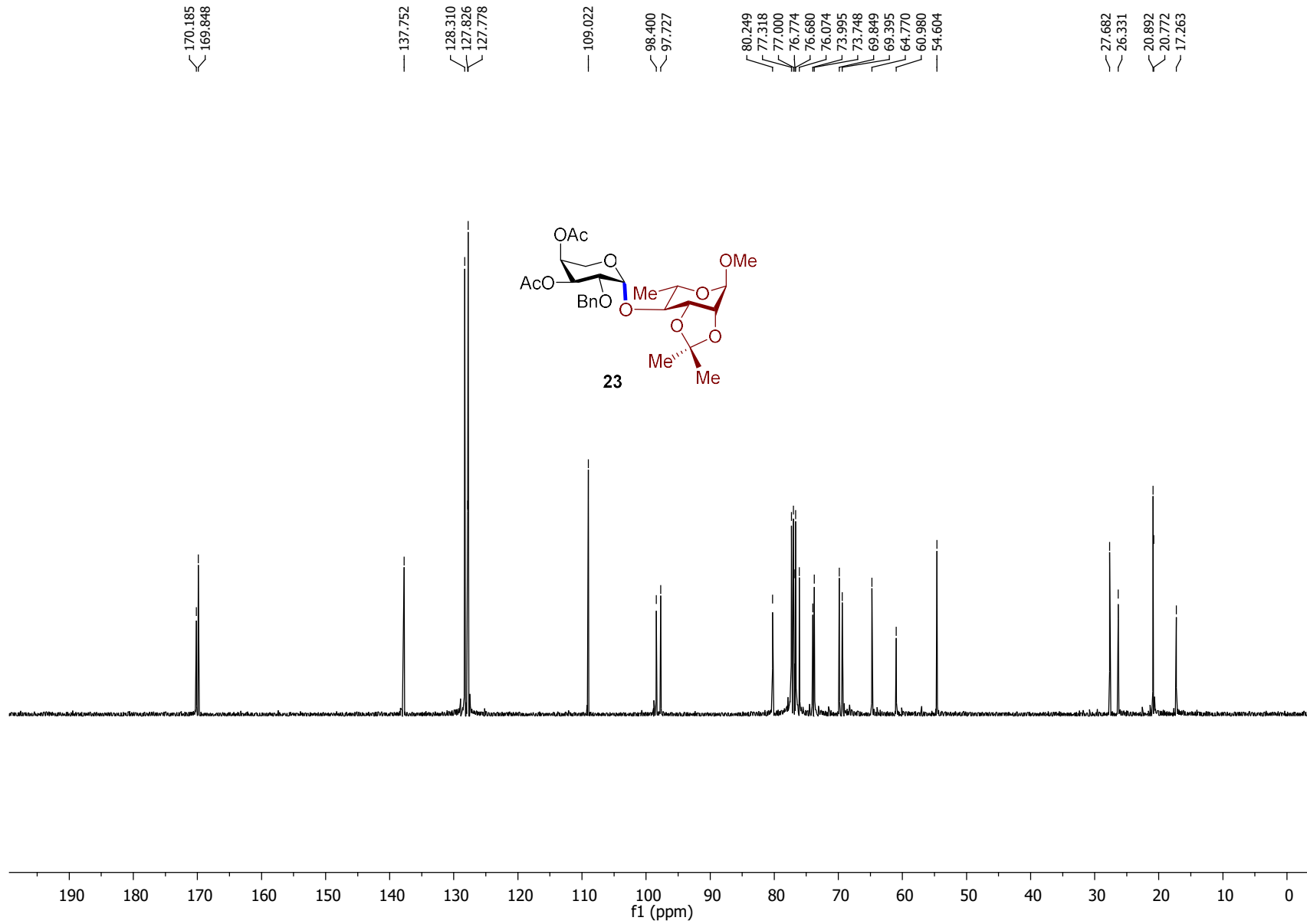














FY-91-16-1-C  
Calibration for OneNMR on 24Apr2018  
Sample : Doped D2O

