

## ***Supporting Information***

ELF4 facilitates innate host defenses against *Plasmodium* by activating transcription of *Pf4* and *Ppbp*

**Dandan Wang<sup>1†</sup>, Zeming Zhang<sup>1†</sup>, Shuang Cui<sup>2</sup>, Yingchi Zhao<sup>1</sup>, Samuel Craft<sup>3#</sup>, Erol Fikrig<sup>4\*</sup>, Fuping You<sup>1\*</sup>**

This article contains Figs. S1–S4 and Tables S1–S3:

Fig. S1. Elf4 deficiency results in compensatory hemolytic anemia.

Fig. S2. Heat maps of functional groups for up- and down-regulated transcripts in bone marrow and spleens.

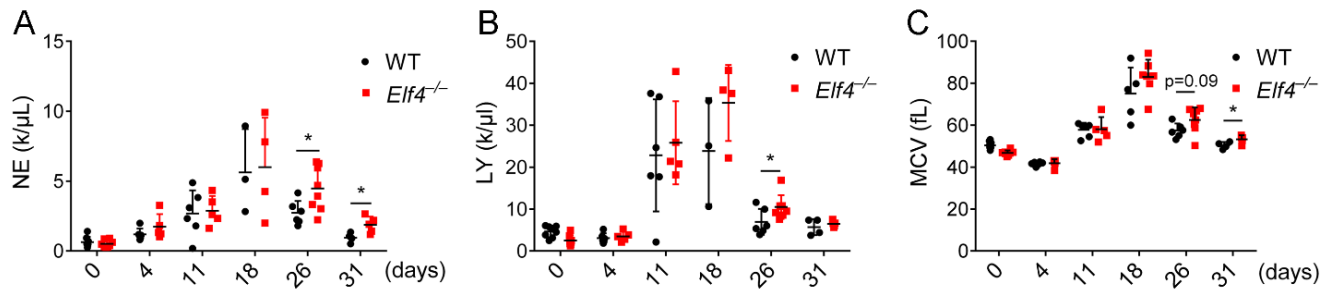
Fig. S3. ELF4 targets the promoter of *Pf4* and *Ppbp*

Fig. S4. ELF4 activates transcription of *PF4* and *PPBP*

Table S1. List of oligonucleotide primers used in this study.

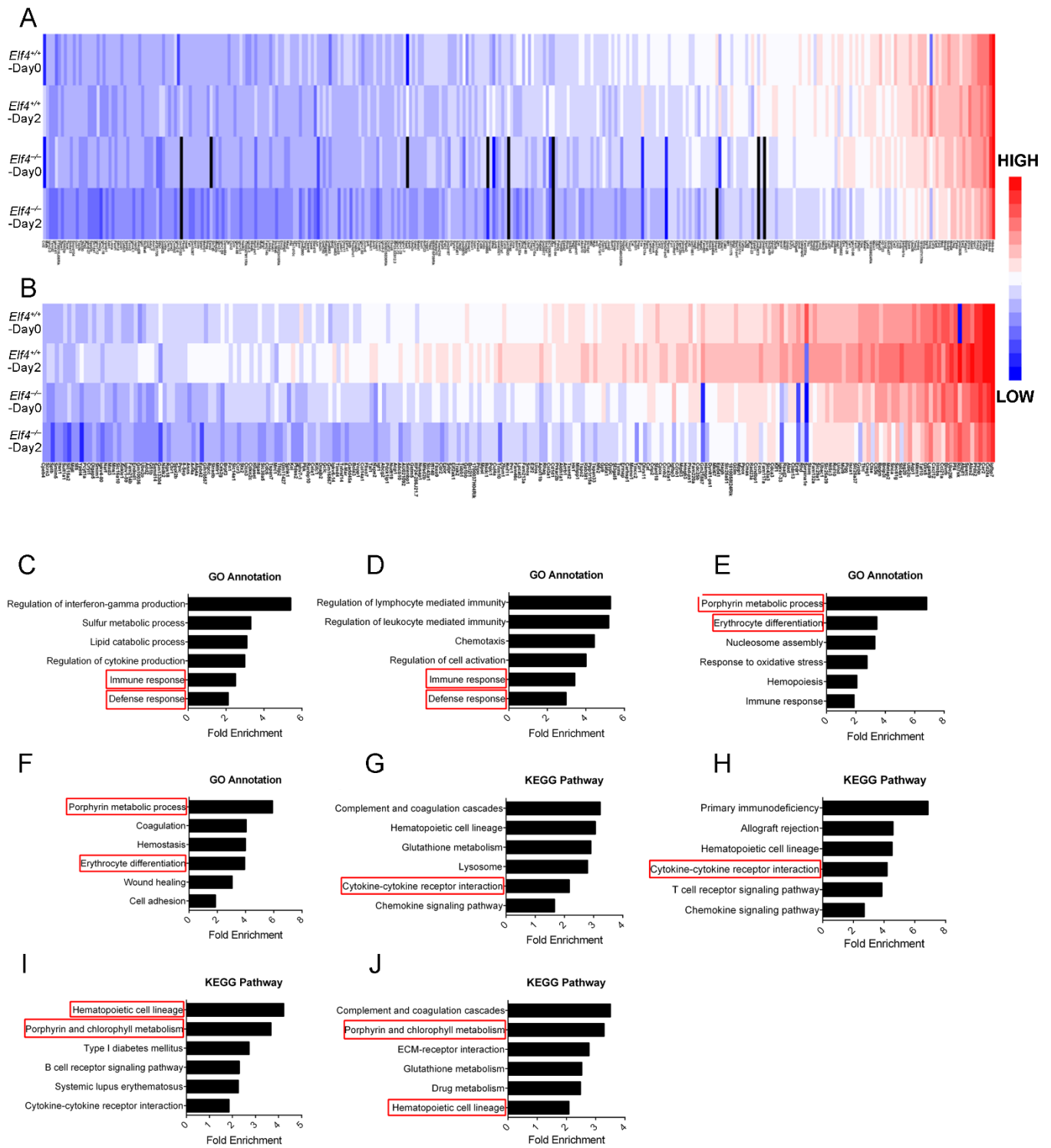
Table S2. Gene list and analysis results of bone marrow RNA-seq data.

Table S3. Gene list and analysis results of spleen RNA-seq data.



**Fig. S1. *Eif4* deficiency results in compensatory hemolytic anemia.**

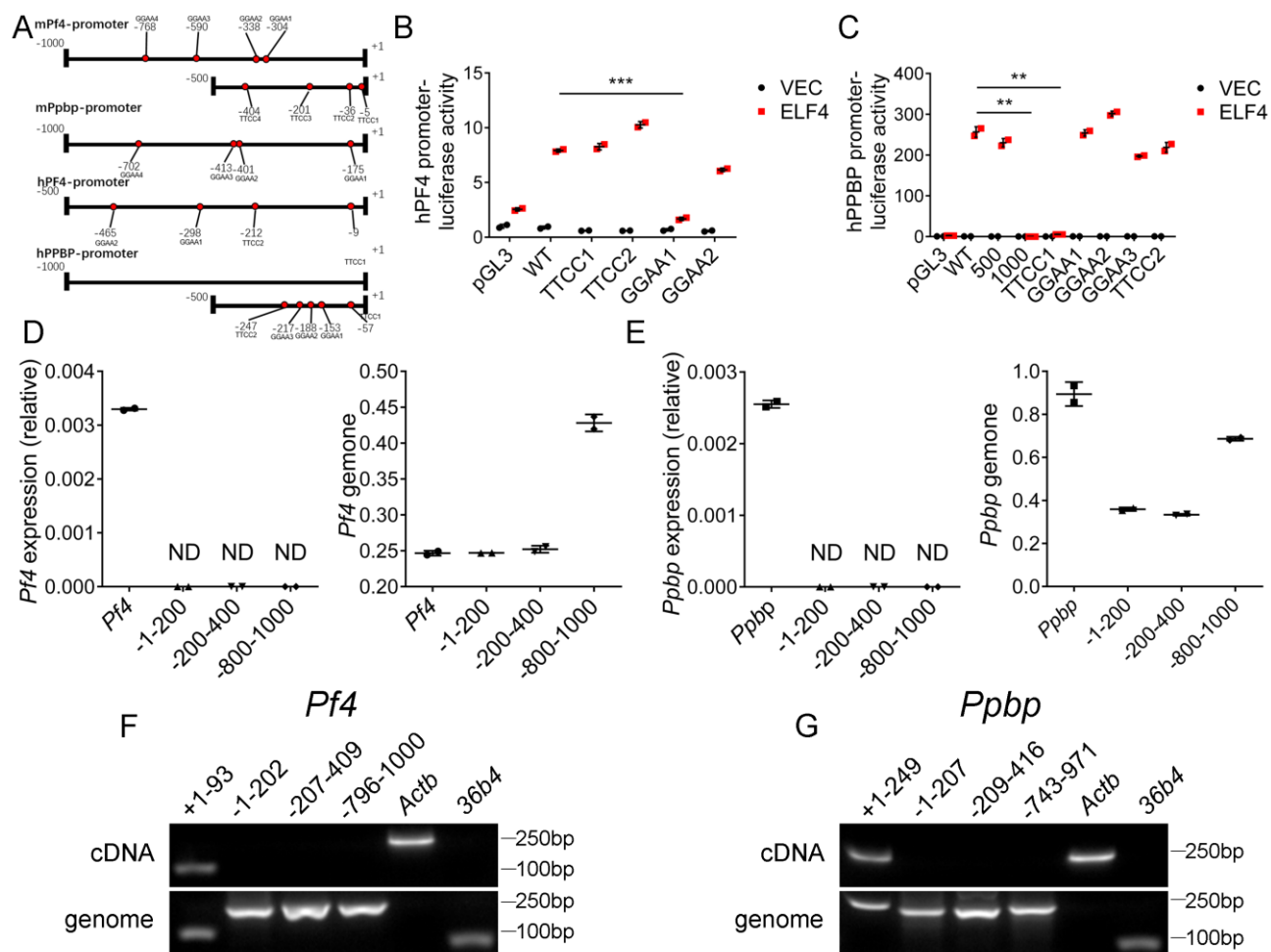
(A-C) Neutrophilicgranulocyte (NE) (A), lymphocyte (LY) (B), and mean corpuscular volume (MCV) (C) were measured and compared between *Eif4*<sup>+/+</sup> and *Eif4*<sup>-/-</sup> mice at indicated days post *P. yoelii* 17XNL infection. All data are means  $\pm$  s.d. from three experiments; *t*-test, \**P* < 0.05.



**Fig. S2. Heat maps of functional groups for up- and down-regulated transcripts in bone marrow and spleens.**

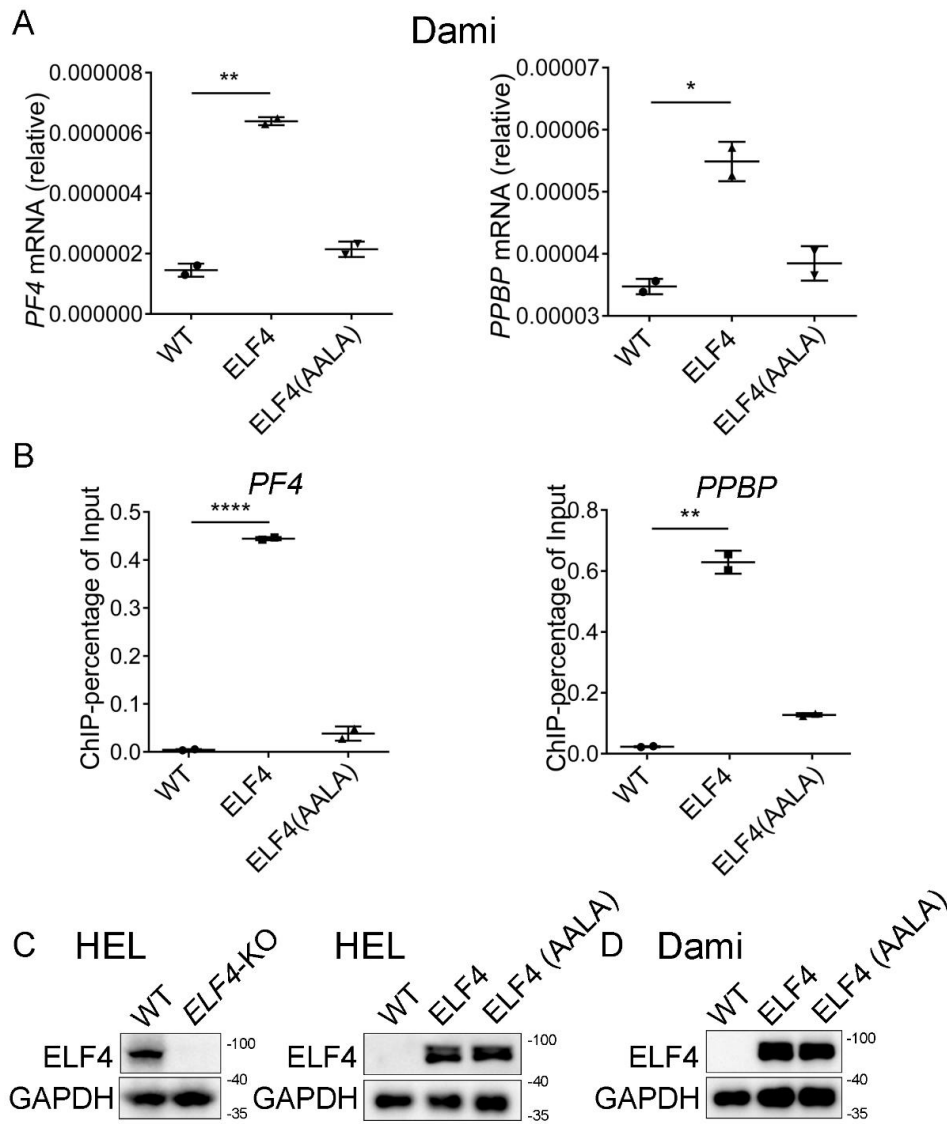
(A-B) Heat map showing differentially expressed transcripts in bone marrow (A) and spleens (B) of *Eif4*<sup>+/+</sup> and *Eif4*<sup>-/-</sup> mice 2 days after infection with *P. yoelii* 17XNL ( $1 \times 10^5$  infected red blood cells). Uninfected control mice (Day0) were injected with an equivalent amount of uninfected RBC. Columns represent

individual mice, and rows represent differentially expressed genes in *P. yoelii* 17XNL-infected *Elf4*<sup>-/-</sup> mice compared with *Elf4*<sup>+/+</sup> mice. The color bar indicates relative expression of genes, with red indicating higher expression and blue indicating lower expression. **(C-J)** Functional enrichment analysis showing overrepresentation of genes upregulated in mouse bone marrow **(C, D)** and spleens **(G, H)** after infection with *P. yoelii* 17XNL or downregulated in mouse bone marrow **(E, F)** and spleens **(I, J)** after infection with *P. yoelii* 17XNL. The notable genes and related biological pathways are highlighted. Technical replication was performed twice independently and data are representative of a single experiment.



**Fig. S3. ELF4 targets the promoter of *Pf4* and *Ppbp*.**

(A) Illustration of human *PF4* and *PPBP*, mouse *Pf4* and *Ppbp* promoter and their mutants-driven luciferase plasmids constructs. (B-C) Luciferase activity in HEK293T cells transfected with human *PF4* (B) or *PPBP* (C) promoter or the different GGAA (or TTCC) -mutated promoters-driven luciferase reporters, together with *Elf4* expression plasmid or empty vectors. (D) Relative quantification of +1-93 region and several upstream regions of *Pf4* was assessed by quantitative PCR (qPCR) in mouse megakaryocytes. (E) Relative quantification of +1-249 region and several upstream regions of *Ppbp* was assessed by quantitative PCR (qPCR) in mouse megakaryocytes. (F) RT-PCR analysis assessing the quantity of +1-93 region and several upstream regions of *Pf4* in total RNA reverse-transcribed cDNA or genome of megakaryocytes. (G) RT-PCR analysis assessing the quantity of +1-249 region and several upstream regions of *Ppbp* in total RNA reverse-transcribed cDNA or genome of megakaryocytes. For (D) and (E), quantity was relative to *Actb* in cDNA and *36b4* in genome. For (F) and (G), *Actb* primer overlaps the second exon and the first and second intron of *Actb* gene. *36b4* primer was designed within the second intron of mouse *36b4* gene. Thus, *Actb* was a positive control in cDNA but a negative control in genome and *36b4* was a positive control in genome but a negative control in cDNA. All data are means  $\pm$  s.d. from three independent experiments; *t*-test, \*\**P* < 0.01; \*\*\**P* < 0.001; ND, no detected.



**Fig. S4. ELF4 activates transcription of *PF4* and *PPBP*.**

(A) Expression of *PF4* and *PPBP* in wild-type (WT) Dami cells or Dami cells stably transfected with ELF4 or ELF4 (AALA) was assessed by quantitative PCR (qPCR) analysis. (B) Binding ability of ELF4 on *PF4* and *PPBP* promoters was assessed by chromatin immunoprecipitation followed by quantitative PCR (ChIP-qPCR) analysis in WT Dami cells or Dami cells stably transfected with ELF4 or ELF4 (AALA). (C-D) ELF4 expression in WT and modified HEL cells (C) or Dami cells (D) was assessed by immunoblot. GAPDH was used as control. Data in (A-B) are means  $\pm$  s.d. from three independent experiments; *t*-test, \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.0001$ .

**Table S1. List of oligonucleotide primers used in this study.**

Gene	Forward (5' to 3')	Reverse (5' to 3')
<i>36b4</i>	TTGTGCAAAGGGCTGAGACT	GTCACTCAACAGCAGCCTGA
<i>Actb</i>	CACTGTTCGAGTCGCGTCCA	TGACCCATTCCCACCATCAC
<i>GAPDH</i>	GAAGGTGAAGGTCGGAGTC	GAAGATGGTGATGGGATTTC
<i>Hprt</i>	CATTATGCCGAGGATTTGGA	AATCCAGCAGGTCAGCAAAG
<i>Ifnb1</i>	TCCGAGCAGAGATCTTCAGGAA	TGCAACCACCACTCATTCTGAG
<i>Isg15</i>	GAGCTAGAGCCTGCAGCAAT	AGACCCAGACTGGAAAGGGT
<i>Ifng</i>	AGACAATCAGGCCATCAGCA	TGGACCTGTGGGTTGTTGAC
<i>mPf4</i>	CGCTGCGGTGTTTCGAGG	TCACCTCCAGGCTGGTGA
<i>Pf4(+1-93)</i>	CCTGTATCCTGGGTTTCCGGAC	CTAGGGCTCTTAAGTGCAGGC
<i>hPF4</i>	CGAGTTTCCCATCGCACTGA	AGCGCTGGCGAAGGC
<i>mPpbp</i>	TGCCCACTTCATAACCTC	GGGTCCAGGCACGTTTT
<i>Ppbp(+1-249)</i>	AACATAGCAGCAGTGTTCTGG	GGGAAGCAGCCTTCATACTCA
<i>hPPBP</i>	TGCAGACTTGTAGGCAGCAA	TGCAAGGCATGAAGTGGTCT
<i>ChIP-PF4</i>	CATGCTTCACACACAAAGCCA	TCTCTGCTGTCATCATGCAACT
<i>ChIP-PPBP</i>	TTGGGCACAGCTTCAAATGC	TGGCTTCTCATGCCCTGAAG

**Table S2. Gene list and analysis results of bone marrow RNA-seq data.****Table S3. Gene list and analysis results of spleen RNA-seq data.**