

Supplementary Figure Legends

Figure S1: Differentiation ability of young and aged MSC.

(a-c) Gene expression levels of *PPAR γ* , *FABP4* and *LPL* adipogenic genes at steady state in young and aged MSC (young, n=4; aged, n=8). *p*-value was determined by Mann-Whitney test; **p* < 0.05. (d-f) Quantitative Real time PCR for *PPAR γ* , *FABP4* and *LPL* adipogenic genes in young and aged MSC at basal level and post-differentiation with adipogenic medium for 21 days. Gene expression data are represented as $2^{-\Delta\text{CT}}$ relative to *ACTNB* housekeeping. Histograms indicate median $2^{-\Delta\text{CT}}$ values (young, n=3; aged, n=4). *p*-value was determined by Mann-Whitney test; **p* < 0.05. (g) Representative pictures of differentiated young and aged MSC stained with Oil Red O; Red signal indicates lipid droplets. (h-j) Gene expression levels of *RUNX2*, *SPARC* and *COL1A2* osteogenic genes at steady state in young and aged MSC (young, n=4; aged, n=8). (k-m) Quantitative Real time PCR for early *RUNX2*, *SPARC* and *COL1A2* osteogenic genes in young and aged MSC at basal level and post-differentiation with osteogenic medium for 21 days. Gene expression data are represented as $2^{-\Delta\text{CT}}$ relative to *ACTNB*. Bars indicate median $2^{-\Delta\text{CT}}$ values (young, n=3; aged, n=3). *p*-value was determined by Mann-Whitney test; **p* < 0.05. (n) Representative pictures of differentiated young and aged MSC stained with Alizarin Red. In all panels, each squared dot represents MSC from an individual donor (red=young; blue=aged).

Figure S2: Analysis of lipofuscin, ROS levels, oxidized DNA and DDR markers in young and aged MSC.

(a) Representative picture for SenTraGor staining in young (n=4; Y1-4) and aged (n=6; A1-6) MSC. White arrows indicate positive SenTraGor Staining. Nuclei were counterstained with DAPI. Senescent BJ fibroblasts were used as a positive control while young MSC (Y3) stained in the absence of anti-biotin antibody or SenTraGor were used as negative controls. Scale bar= 20 μ m. (b) ROS levels as measured by CellROX staining in young and aged MSC. Lines indicate mean values \pm SEM. ROS levels were calculated as DMFI relative to unstained controls (young, n=4; aged, n=4); *p*-value was determined by Mann-Whitney test; **p* < 0.05. (c) Relative mRNA expression of *FOXO4* measured by quantitative Real-Time PCR. Gene expression data are represented as $2^{-\Delta\text{CT}}$ relative to *GUSB* as scatter dot plots, bars indicate median $2^{-\Delta\text{CT}}$ values (young, n=6; aged, n=8); *p*-value was determined by Mann-Whitney test; **p* < 0.05. (d) FACS analysis for 8-oxo-dG levels in young and aged MSC expressed as percentage of positive cells and represented as scatter dot plots (young, n=4; aged, n=7). Each squared dot represents MSC from an individual donor (red=young; blue=aged); (e-f) Representative confocal images for pATM (pATMS1981) and γ H2A.X in young and aged MSC at early passages. Human BJ cells induced into senescence by irradiation (20Gy)

were used as positive control for DDR foci staining. For senescent cells numbers indicate the percentage of DDR-positive cells. For young and aged MSC numbers indicate the median of the percentage of foci positive cells for γ H2A.X (3 young and 4 aged MSC) and for pATM staining (3 young and 3 aged). Nuclei were counterstained with DAPI. Scale bar=20 μ m.

Figure S3: Aged MSC display reduced immunomodulatory properties.

(a) Schematic representation of the *in vitro* model to assess MSC immune-modulatory properties in suppressing PBMC proliferation. **(b)** *In vitro* immunomodulatory effect of young and aged MSC on human healthy donor peripheral blood mononuclear cells (PBMC). The graph shows the percentage of residual proliferation of PHA-stimulated PBMC in presence of young and aged MSC at different ratios (1:2, 1:20 and 1:200), calculated by measuring 3 H-thymidine incorporation after 72 hours of co-culture. Dash line represents PBMC proliferation in the absence of MSC considered as 100% and used to normalize PBMC proliferation in presence of young or aged MSC (young, n=3; aged, n=4). Each squared dot represents MSC from an individual donor (red=young; blue=aged). *p*-value was determined by Mann-Whitney test; **p* < 0.05. **(c-f)** Gene expression analysis for *CXCL10*, *PTGS2*, *TGF- β 1* and *IDO1* by quantitative Real Time PCR in young and aged MSC at early passages in culture. Gene expression data are represented as $2^{-\Delta CT}$ relative to housekeeping genes in scatter dot plots, bars indicate median $2^{-\Delta CT}$ values (young, n=4; aged, n=8); *p*-values was determined by Mann-Whitney test; **p* < 0.05.

Figure S4: SASP activation in human bone marrow samples and MSC aging.

(a-f) Plasma levels of IL6, IL8, MCP1, IL1 α , TNF α , and IL1 β in human bone marrow samples from young and aged donors measured by Luminex technology (young, n=6; aged, n=8); Values of individual donors are represented as scatter dot plot, lines indicate median values. *p*-value was determined by Mann-Whitney test; ***p* < 0.01; **p* < 0.05. **(g-i)** Relative mRNA expression of *IL6*, *MCP1* and *IL1 β* late passages young (n=3) and aged MSC (n=3). Gene expression data are represented as $2^{-\Delta CT}$ relative to *GUSB* as scatter dot plots, histograms indicate median $2^{-\Delta CT}$ values. *p*-value was determined by Mann-Whitney test; **p* < 0.05. **(j-m)** Relative mRNA expression of HSPC supportive genes *CXCL12*, *VCAM*, *VEGF* and *ANGPT1* measured by quantitative Real-Time PCR in young (n=6) and aged (n=8) MSC. Gene expression data are represented as $2^{-\Delta CT}$ relative to *GUSB* as scatter dot plots, bars indicate median $2^{-\Delta CT}$ values.

Figure S5: Paracrine effects of CM from aged MSC of subjects under chronic steroids treatment on HSPC functionality.

(a) Percentage of CD34⁺ HSPC measured by FACS after 4 days in culture with 1:1 StemSpan/CM derived from young or aged MSC. CD34⁺ cells grown without CM in StemSpan were used as control. **(b)** Histograms represent the mean percentage \pm SD of primitive (CD90⁺/CD133⁺), early (CD90⁺/CD133⁻) and committed (CD90⁻/CD133⁻) progenitors within total CD34⁺ cells for each donor analyzed at 96 hours; (CM from young, n=4; aged, n=4). HSPC from two independent CB-derived donors were employed. **(c)** Number of HSPC colonies in methylcellulose analyzed at 96 hours post exposure to CM derived from healthy young, aged MSC or MSC derived from aged donors under steroids treatment. Red, white and light grey bars represent erythroid, myeloid and mixed colonies, respectively. CD34⁺ cells grown without CM (CTRL) were used as control. **(d-e)** Gene expression analysis of *IL1 α* and *IL6* by quantitative Real Time PCR in CB-derived CD34⁺ cells cultured for 96 hours either in absence (CTRL) or in presence of CM derived from young MSC, aged MSC and aged MSC under steroids treatment. Gene expression data are represented as $2^{-\Delta CT}$ relative to *GUSB* housekeeping, histograms indicate median $2^{-\Delta CT}$ values (CD34⁺ CTRL n=3; young, n=5; aged, n=5; aged under steroids treatment, n=3). *p*-value was determined by Mann-Whitney test; **p* < 0.05. **(f-i)** Relative mRNA expression of *IL8*, *IL6*, *MCP1* and *IL1 α* in healthy aged or in MSC derived from aged donors under steroids treatment (aged, n=8, aged under steroids treatment, n=3). Gene expression data are represented as $2^{-\Delta CT}$ relative to *GUSB* as scatter dot plots, histograms indicate median $2^{-\Delta CT}$ values. *p*-values was determined by Mann-Whitney test; **p* < 0.05. **(j-l)** Levels of IL8, IL6 and MCP1 measured by Luminex assay in CM collected from early passages aged MSC and aged MSC of subjects under steroids treatment. Values are represented as scatter dot plot, lines indicate median values; (aged, n=6; aged under steroids treatment, n=3). *p*-value was determined by Mann-Whitney test; **p* < 0.05. **(m)** Relative mRNA expression of *CDKN1A* in healthy aged or in MSC derived from aged donors under steroids treatment (aged, n=8, aged under steroids treatment, n=3). Gene expression data are represented as $2^{-\Delta CT}$ relative to *GUSB* as scatter dot plots, bars indicate median $2^{-\Delta CT}$ values.

Supplementary Table 1.

List of pediatric, young and aged MSC samples employed in each figure panel.

Figures	Number of donors		
	Pediatric MSC	Young Adults MSC	Aged MSC
1b (day7, day14)	4	6	12; 7
1d-e	2	4	8
1g-h	2	4	8
2c	2	2	4
2d-f	1	-	2
2g	3	3	8
2h	3	3	8
2j	3	1	8
3b	3	2	5
3d	3	2	5
3f	3	2	5
3g	3	4	7
3i	2	1	3
3j	2	1	3
4a	2	4	8
4b-d	3	3	8
4e	2	4	8
4f-k	2	3	6
5b-e	2	3	5
6a-c	-	-	3
6d-f	-	-	3
S1a-c	-	4	8
S1d-f	-	3	4
S1h-j	-	4	8
S1k-m	-	3	3
S2a	2	2	6
S2b	3	1	4
S2c	3	3	8
S2d	2	2	7
S2e	2	2	4
S2f	2	1	3
S3b	1	2	4
S3c	2	2	8
S3d-f	-	4	8
S4a-f	-	6	8
S4g-i	2	1	3
S4j-m	3	3	8
S5a-b	2	2	5
S5c-e	2	3	5
S5f-i	-	-	8
S5j-l	-	-	6
S5m	-	-	8

Supplementary Table 2

Sequences of RT-qPCR primers employed.

	<i>Forward</i>	<i>Reverse</i>
<i>ACTNB</i>	ACAGAGCCTCGCCTTTGC	ACTCCATGCCAGGAAGGAA
<i>PPARG</i>	TCAGAAATGCCTTGCAGTGG	TATCACTGGAGATCTCCGCCAA
<i>LPL</i>	CCGCCGACCAAAGAAGAGAT	TAGCCACGGACTCTGCTACT
<i>FABP4</i>	AAACTGGTGGTGGAAATGCGT	GCGAACTTCAGTCCAGGTCA
<i>RUNX2</i>	CCGGAATGCCTCTGCTGTTA	TGTCTGTGCCTTCTGGGTTT
<i>SPARC</i>	ATTGACGGGTACCTCTCCCA	GAAAAGCGGGTGGTGAAT
<i>COL1A2</i>	ACAAGGCATTCGTGGCGATA	ACCATGGTGACCAGCGATAC
<i>TGF-β1</i>	CGATGAGCAGCTTTTCCAGA	AGTGAACCCGTTGATGTCCA
<i>IDO1</i>	TGGCCAGCTTCGAGAAAGAG	TGGCAAGACCTTACGGACATC
<i>PTGS2</i>	CAAATTGCTGGCAGGGTTGC	AGGGCTTCAGCATAAAGCGT
<i>IL8</i>	CATCTCACTGTGTGTAAACATGAC	CCTTGGCAAACTGCACCTTAC
<i>IL1β</i>	GCTCAAGTGTCTGAAGCAGCC	CAGCTTCAAAGAACAAGTCATCCT
<i>IL6</i>	GATTCAATGAGGAGACTTGCCTGG	CTCACTACTCTCAAATCTGTTCTGG
<i>IL1α</i>	GGTTGAGTTTAAGCCAATCCA	TGCTGACCTAGGCTTGATGA
<i>MCPI</i>	CTGTGATCTTCAAGACCATTGTG	AGTTTGGGTTTGCTTGTCAG
<i>CDKN2A</i>	CCAACGCACCGAATAGTTACG	GCGCTGCCATCATCATG
<i>CDKN1A</i>	CAGCATGACAGATTTCTACCACTC	CTCGCGCTTCCAGGACTG
<i>GUSB</i>	CTGACACCTCCAAGTATCCCAAG	GTCGTGTACAGAAGTACAGACCGC
<i>FOXO4</i>	GAGCCAGATCCCTGAGTCATG	GGTAGCTCTAAAGCAGGGTCC
<i>VEGFA</i>	CATCTTCAAGCCATCCTGTG	GGAAGCTCATCTCTCCTATG
<i>CXCL12</i>	TGCCCTTCAGATTGTAGCCC	CGAGTGGGTCTAGCGGAAAG
<i>VCAM1</i>	GACCTTCATCCCTACCATTG	TTCACAGAACTGCCTTCCCTCCA
<i>ANGPT1</i>	ACATGGGCAATGTGCCTACA	TCTCAAGTTTTTGCAGCCACTG
<i>CXCL10</i>	GCAGAGGAACCTCCAGTCTCAGC	GAGAGAGGTACTIONCTTGAATGC