

SUPPLEMENTAL MATERIAL

Methods

Data sources and study population

The individual level data will not be made available to other researchers without IRB approval from the VA.

Participants of multiple ethnicities were recruited from approximately 50 VA healthcare facilities across the United States.¹ Individuals consented to a blood draw and to have their DNA extracted for genomic profiling and linked to their full electronic health record within the VA. Both MVP biobank and this analysis were approved by the VA institutional review boards.

Phenotypic measures

We obtained demographic variables such as age and sex from the MVP enrollment questionnaire and participants' electronic health record (EHR) data. We secured all measures of LDL-C performed in VA laboratories using standardized assays for up to 15 years. The MVP participants had a median of 11 measurements (1st quartile of 6 and 3rd quartile of 19) of LDL-C, with 4.4% having only one measurement of LDL-C. When more than one measure was available, we extracted the maximal level of LDL-C to approximate the most likely level of untreated LDL-C.² We also analyzed the single outpatient LDL-C value obtained closest to the time of enrollment into MVP during 2011-2016 and statin prescriptions going back to 2002. Evidence of statin use was obtained from prescribing and pharmacy records in the EHR.

Genotypic measures and identification of FH variants

Blood samples drawn from consenting MVP participants were shipped to a central biorepository in Boston, Massachusetts, where DNA was extracted and shipped to two external genotyping centers for genotyping on an Affymetrix biobank array designed specifically for the

MVP. Genotyping was performed at two sites using the same Affymetrix best practices pipeline including several quality control (QC) procedures targeting genotyping sites and batches. Genotype calling was performed on all samples together in batches grouped by site and sample processing date. Standard Axiom genotyping quality matrix (dish quality control (DQC) >0.95, QC call rate >0.97) were comparable across the batches and sites. Furthermore, an advanced marker and sample QC procedure was used to clean and harmonize genotype calls. Probe sets were analyzed for any inconsistencies across all batches and removed. QC metrics like the final sample call rates, non-missingness, minor allele frequencies, Ratio A Allele frequencies were all determined to be consistent across sites and batches. The MVP genomics working group applied genotype calling algorithms to the data in batches using the Affymetrix Power Tools Suite (v1.18). Standard quality control pipelines were used to exclude duplicate samples, samples with more heterozygosity than expected, or discordance between sex inferred by genotyping versus self-report. We also excluded related individuals (halfway between 2nd and 3rd degree relatives or closer) as measured by the KING³ software.

We queried the ClinVar database⁴ (archive of June 2017) for “pathogenic” or “likely pathogenic” variants linked to familial hypercholesterolemia. We considered variants with conflicting interpretations of pathogenicity but at least one “pathogenic” or “likely pathogenic” assertion. These ClinVar variants were matched by unique identifiers (*i.e.*, rs ID) and chromosomal position to variants genotyped on the MVP biobank array. Additional annotation was obtained from the Human Gene Mutation Database⁵ to further understand the molecular and functional effects of selected FH variants.

Statistical analysis

We used ADMIXTURE⁶ analysis with the 1000 Genome phase 3 samples and known

continental ancestries to assign genotyped individuals to European (>80% European Ancestry) and admixed African Americans (>50% African Ancestry). Global and ethnicity-specific principal component analyses were performed using flashPCA software.⁷ We then calculated carrier frequencies within each of two main ancestry groups.

We used linear regression models to estimate the effects of FH variants on untransformed LDL-C, adjusted for age, age² and sex for FH variants with a carrier count of 30 or greater (a carrier frequency of ~0.009%). For these variants, we considered associations to be significant if the p was < 0.05/the number of tested variants. FH variants with fewer than 30 carriers were combined into total burden and gene-specific scores (i.e., *LDLR*, *APOB* and *PCSK9*) to assess associations with maxLDL but they were not considered individually. All association analyses were performed in the R statistical environment version 3.2.5 (<http://www.r-project.org/>).

Phenome-wide association analysis of FH variants with clinical outcomes

Genotyped MVP participants were included in the phenome-wide association study (PheWAS) if the electronic health record (EHR) reflected two or more separate encounters in the VA Healthcare System in each of the two years prior to enrollment in the MVP. We identified 277,531 veterans and 21,209,658 prevalent ICD-9 diagnosis codes were available for analysis. ICD-9 diagnosis codes were collapsed to clinical disease groups and corresponding controls.⁸ Diseases were required to have a prevalence of more than 400 cases to be included in the PheWAS analysis. Among participants with European ancestry, carriers of FH variants were compared to the other participants using logistic regression adjusting for age, sex, and ten principal components using the PheWAS R package.⁹ A total of 1,171 disease phenotypes were available for analysis. Additionally, we identified individuals with clinical CHD or peripheral artery disease (PAD) using inpatient and outpatient ICD-9 and Current Procedural Terminology

codes available in EHR data from up to 15 years prior to enrollment in the MVP.

References

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Supplemental Table 1: Distribution of P/LP FH Variants (obtained from ClinVar database) among multi-ethnic MVP participants.

