## Table of Contents

Appendix Figure S1. Generation of OTUB1 <sup>fl/fl</sup> mice and phenotypes	p.2
of OTUB1 <sup>wt/<math>\Delta</math></sup> , and Nestin-Cre OTUB1 <sup>wt/fl</sup> mice	
Appendix Figure S2. OTUB1 directly interacts with and stabilizes SOCS1	p.4



B Breeding: OTUB1<sup>wt/Δ</sup> X OTUB1<sup>wt/Δ</sup>

mice	OTUB1 <sup>wt/wt</sup>	OTUB1 <sup>wt/∆</sup>	OTUB1
number	11	18	0
percentage	38%	60%	0%

C Breeding: Nestin-Cre OTUB1<sup>wt/fl</sup> X OTUB1<sup>wt/fl</sup>

Cre OTUB1	Nestin-Cre <sup>-</sup> (number/percentage)	Nestin-Cre <sup>*</sup> (number/percentage)
OTUB1 <sup>wt/wt</sup>	19 (11.9%)	24 (15%)
OTUB1 <sup>wt/ll</sup>	42 (26.3%)	50 (31.3%)
OTUB1 <sup>n/n</sup>	25 (15.6%)	0 (0%)

Appendix Figure S1. Generation of  $OTUB1^{fl/fl}$  mice and phenotypes of  $OTUB1^{wt/\Delta}$ , and Nestin-Cre  $OTUB1^{wt/fl}$  mice

(A) Schematic representation of the gene targeting strategy for the generation of OTUB1<sup>fl/fl</sup> mice. The neomycin and puromycin resistance markers were removed from the conditional

allele by FLP-mediated gene recombination. The resulting OTUB1<sup>fl/fl</sup> mice were further crossed with mice containing different Cre transgenes to generate global/conditional OTUB1 knockout mice.

(B) OTUB1<sup>wt/fl</sup> mice were crossed with Rosa 26-Cre mice to delete exons 2 and 3 of OTUB1, resulting in out-of-frame translation and germline inactivation of OTUB1. Rosa 26-Cre OTUB1<sup>wt/fl</sup> mice were crossed with C57BL/6 mice to remove the Rosa 26-Cre transgene, resulting in the heterogeneous OTUB1<sup>wt/ $\Delta$ </sup> mice, in which one OTUB1 allel was inactivated. For the generation of OTUB1<sup> $\Delta/\Delta$ </sup> mice, OTUB1<sup>wt/ $\Delta$ </sup> mice were crossed with OTUB1<sup>wt/ $\Delta$ </sup> mice. (C) Nestin-Cre OTUB1<sup>wt/fl</sup> mice were generated by crossing OTUB1<sup>fl/fl</sup> mice with Nestin-Cre mice.





Appendix Figure S2. OTUB1 directly interacts with and stabilizes SOCS1

(A) Primary astrocytes from GFAP-Cre OTUB1<sup>fl/fl</sup> mice were cotransfected with SOCS1-MYC and K48 ubiquitin-HA plasmids or a combination of SOCS1-MYC, OTUB1-GFP, and K48 ubiquitin-HA plasmids. Twenty-four hours after transfection, cells were left untreated or stimulated with IFN- $\gamma$  (10 ng/ml) for 30 min before lysation. Proteins from whole cell lysates were immunoprecipitated with anti-MYC antibody and analyzed by WB for MYC, GFP, and HA.

(B) Primary astrocytes from GFAP-Cre OTUB1<sup>fl/fl</sup> mice were cotransfected with SOCS1-MYC and GFP plasmids or SOCS1-MYC and OTUB1-GFP plasmids. Twenty-four hours after transfection, cells were left untreated or stimulated with IFN- $\gamma$  (10 ng/ml) for 30 min

before lysation. Proteins from whole cell lysates were immunoprecipitated with anti-MYC antibody and analyzed by WB for MYC and GFP.

(C) Primary astrocytes from GFAP-Cre OTUB1<sup>fl/fl</sup> mice were cotransfected with SOCS1-MYC and GFP plasmids or SOCS1-MYC and OTUB1-GFP plasmids. Twenty-four hours after transfection, cells were treated with CHX (100  $\mu$ g/ml) for indicated times. SOCS1 protein levels were analyzed by WB.

(D) Primary astrocytes from GFAP-Cre OTUB1<sup>fl/fl</sup> mice were transfected with GFP, OTUB1-GFP, OTUB1- $\Delta$ N-GFP, and OTUB1-C91S-GFP plasmids, respectively. Twenty-four hours after transfection, cells were stimulated with IFN- $\gamma$  (10 ng/ml) for 4 h followed by treatment with CHX (100 µg/ml) for indicated times. SOCS1 protein abundance in the whole cell lysates was analyzed by WB.