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Supplementary Materials for

ALBA protein complex reads genic R-loops to maintain genome stability in *Arabidopsis*

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Table S1 (Microsoft Excel format). Primers and substrates used in this study. Table S2 (Microsoft Excel format). List of AtALBA1-bound loci.



Fig. S1. Domain structure of ALBA proteins in *Arabidopsis.* (**A**) Domain structure of AtALBA proteins from the Pfam database. AtALBA1, AtALBA2 and AtALBA3 belong to the Rpp20-like subfamily and contain only an Alba-like DNA/RNA-binding domain. AtALBA4, AtALBA5 and AtALBA6 belong to the Mdb2-like subfamily. In addition to an Alba-like domain at the N-terminal, Mdb2-like subfamily members contain DUF390 and RGG motifs at the C-terminal. The numbering refers to the amino acid position. (**B**) Amino acid sequence alignment of the Alba-like DNA/RNA-binding domains of AtALBA5.



Fig. S2. Purification of AtALBA1 and AtALBA2 wild-type and mutant proteins and diagram of probes used in EMSAs. (A) Coomassie blue-stained gel showing purified AtALBA1 and AtALBA2 wild-type and mutant proteins. Recombinant AtALBA1-His and AtALBA2-His were purified by Ni-NTA affinity chromatography. (B) Amino acid sequence alignment of the Alba-like DNA/RNA-binding domains from Alba proteins in different species. The conserved lysine (K), which is critical for DNA binding, is indicated in a black box. (C) Diagram of different substrates used in EMSA.





Fig. S3. Characterization of the nucleic acid binding properties of AtALBA1 and AtALBA2. (A) EMSA gel showing AtNDX (16 nM) binding to single-stranded DNA probes (5 nM) and artificial R-loops with 5'-biotin-labeled DNA (1) (5 nM). 80 nM of GST proteins were used for negative control. (B) EMSA gel showing AtALBA1 (25, 50, 75 nM) binding to single-stranded RNA probes (5 nM) and competition by unlabeled RNA probes (2 μ M). Two randomly selected sequences were used in this assay. (C) EMSA gel showing AtALBA1 (75 nM) binding to the DNA-RNA hybrid probe (5 nM). The DNA-RNA hybrid probes were incubated with RNase H1 for 0 min or 10 min. S9.6 antibody was used as a positive control. (D) EMSA gel showing AtALBA2 (25, 50, 75 nM) binding to dsDNA probes (5 nM) and competition by unlabeled DNA probes (2

 μ M). Four randomly selected sequences were used in this assay. (**E-G**) Quantification of the affinities of proteins (25, 50, 75 nM) towards different types of nucleic acids (2 μ M) by Agilent 2100 BioAnalyzer. Sharp peaks (blue arrow) indicate free probe and broad peaks (red arrow) indicate formation of nucleic acids/protein complexes. 75 nM of K30E mutant proteins were used as negative controls. For the right lane in S3G, AtALBA1 and AtALBA2 were mixed in the mole ratio 1:1.



Fig. S4. Subcellular localization and interaction of AtALBA1 and AtALBA2. (A) Localization of AtALBA1-GFP and AtALBA2-GFP in Arabidopsis protoplasts. NLS-mCherry (red) was used to label the nuclei. Bars = $10 \mu m.$ (B) Western blot analysis of AtALBA1-Flag and AtALBA2-Flag in total protein extract, nuclear and cytoplasmic fractions. Transgenic AtALBA1-Flag/alba1-1 AtALBA2-Flag/alba2-1 AtALBA1-Flag and plants were used. and AtALBA2-Flag expression were under the control of their respective native promoters. Histone H3 and Tubulin were used as nuclear and cytoplasmic markers, respectively. T: total protein extract, N: nuclear fraction, C: cytoplasmic fraction. (C) Split luciferase complementation results showing the interaction between AtALBA1 and AtALBA2 in Nicotiana benthamiana leaves. Standard errors were calculated from three biological replicates, *p<0.05, **p<0.01, two-tailed Student's *t*-test. (**D**) Co-immunoprecipitation (co-IP) results showing the interaction between AtALBA1 and AtALBA2. Transgenic plants expressing AtALBA1-Flag, AtALBA2-Flag and AtALBA1-Myc under their native promoters and their F1 offspring were used for co-IP. The asterisk indicates a non-specific band. (E) Co-localization of AtALBA1 and AtALBA2 in the nucleus. AtALBA1 and AtALBA2 were stained using anti-Myc (red) and anti-Flag (green), respectively, in Col-0 plants (upper panel) or F1 heterozygous plants expressing AtALBA1-Myc and AtALBA2-Flag under their native promoters (lower panel). DNA was stained with DAPI (blue). The frequency of nuclei displaying each interphase pattern is shown on the right. Bars = $2.5 \,\mu m$.



Fig. S5. Characterization of AtALBA1-bound loci. (A) Venn diagram showing the overlap of AtALBA1 ChIP-seq peaks in two biological replicates. (B) Box plots showing expression levels of non-AtALBA1-bound genes and AtALBA1-bound genes. RPGC: reads per genomic content (1 x normalization). (C) Association of AtALBA1 and AtALBA2 with specific genic regions. Transgenic *AtALBA1-Flag/alba1-1, AtALBA1-Flag/alba1-1alba2-1, AtALBA2-Flag/alba2-1* and *AtALBA2-Myc/alba1-1alba2-1* plants were used for ChIP assays. Expression of *AtALBA1-Flag* and *AtALBA2-Flag/Myc* were under the control of their respective native promoters. ChIP experiments were performed with anti-Flag or anti-Myc. Two biological replicates were performed,

and very similar results were obtained. Standard errors were calculated from three technical replicates, *p<0.05, **p<0.01, ***p<0.001 (two tailed Student *t*-Test). (**D**) Association of AtALBA1 and AtALBA2 with specific genic regions. Transgenic *AtALBA1-Flag/alba1-1* and *AtALBA2-Flag/alba2-1* plants were used. Expression of *AtALBA1-Flag* and *AtALBA2-Flag* was under the control of their respective native promoters. ChIP experiments were performed with anti-Flag antibody. The RNase III treatment was performed before cross-linking. Two biological replicates yielded very similar results. Standard errors were calculated from three technical replicates, *p<0.05, **p<0.01, ***p<0.001, two-tailed Student's *t*-test.



Fig. S6. Characterization of the T-DNA insertion mutants for *AtALBA1* **and** *AtALBA2.* (**A**) Schematic diagram showing the positions of T-DNA insertion mutations at the *AtALBA1* and *AtALBA2* loci. Black rectangles represent exons. Gray rectangles represent UTRs. (**B**) RT-PCR analysis of *AtALBA1* and

AtALBA2 mRNA levels in the alba1 and alba2 mutants, respectively. The yellow arrow indicates a mutant transcript of AtALBA2 in alba2-1. (**C**) Relative expression levels of AtALBA2 in alba2-1. Standard divisions were calculated from three technical replicates, ***p<0.001 (two tailed Student *t*-Test). (**D**) Sequence alignment of the mutant transcript of AtALBA2 in alba2-1 with wild-type AtALBA2 CDS. The nucleotide insertion carried by the mutant transcript is from the T-DNA vector backbone. (**E**) Amino acid sequence alignment showing a 9-amino acid insertion in the Alba-like DNA/RNA-binding domain of AtALBA2 in alba2-1. (F) Developmental phenotype ofCol-0 and the indicated mutants. Bars = 0.5 cm.





Fig. S7. Detection of R-loop levels in Col-0 and alba1-1alba2-1 by immunostaining and ssDRIP-seq. (A) Immunostaining detection of R-loops in Col-0 and alba1-1alba2-1. R-loops in individual cells were stained with S9.6 antibodies (green). DNA was stained with DAPI (blue). At least 30 cells were stained for each sample. Bars = $2.5 \mu m$. (B) Metaplot of R-loop levels in Col-0 and alba1-1alba2-1 as determined by ssDRIP-seq. 0 on the X-axis indicates the R-loop peak center. The Y-axis indicates ssDRIP-seq read density. (C) R-loop levels on five chromosomes in Col-0 and alba1-1alba2-1. Chr. 1, Chr. 2, Chr. 3, Chr. 4, and Chr. 5 represent chromosomes 1, 2, 3, 4 and 5, respectively. (D) Metaplot of R-loop levels on AtALBA1-bound genes in Col-0 and

alba1-1alba2-1. The gray bar on the X-axis indicates the genes. -500 bp and +500 bp represent 500 bp upstream of the TSS and 500 bp downstream of the TTS, respectively. (**E**) Snapshots of genomic regions with AtALBA1 enrichment and R-loops.







Fig. S8. Molecular phenotypes of Col-0 and alba1-1alba2-1 without and with MMS treatment. (A) Representative microscopic images showing γ H2AX foci formation (green) in Col-0, *alba1-1*, *alba1-2*, *alba2-1*, *alba1-1alba2-1*, and *alba1-2alba2-1* plants under normal growth conditions. γ H2AX foci were detected by immunostaining using anti- γ H2AX antibody. Nuclei were stained with DAPI (blue). Bars = 5 µm. (B) Immunofluorescence showing elevated γ H2AX signal in Col-0 after 5 min of γ -irradiation (100 Gy). Bars = 2 µm. (C and D) Relative expression levels of *RAD51* and *BRCA1* in Col-0 and the indicated mutants. After growth under normal conditions for 5 d, plants were treated with 50 ppm MMS for 48 h. Two biological replicates were performed, and very similar results were obtained. The multiple comparison in S8C was calculated with Tukey. The level by alpha default is 0.05. Standard errors in S8D were calculated from three technical replicates, *p<0.05, **p<0.01, ***p<0.001,

two-tailed Student's *t*-test. **(E)** Relative expression levels of *AtALBA1* and *AtALBA2* in Col-0 plants grown on 1/2 MS medium supplemented with 0 or 50 ppm MMS. Three biological replicates were performed. Standard errors were calculated from three technical replicates, **p<0.01, ***p<0.001, two-tailed Student's *t*-test. **(F)** Localization pattern of AtALBA1 and AtALBA2 in the nucleus without and with MMS treatment. AtALBA1 and AtALBA2 were stained using anti-Flag (green), under normal conditions (upper panel) or with 50 ppm MMS treatment (lower panel). DNA was stained with DAPI (blue). Bars = 2.5 μ m. **(G)** Slot blot showing total R-loop levels in Col-0 and the indicated mutants without and with MMS treatment. 100 ng of genomic DNA were used for blotting. Results from two replicates were presented. **(H)** Information for γ H2AX ChIP-seq data, including number of reads after adapter cut, unique mapped reads and rate, average length and quality of reads. **(I)** Scatterplot of replicates using bigwig files and deepTools. Spearman's correlation coefficients are presented.