# Red Blood Cell Membrane Bioengineered Zr-89 Labelled Hollow Mesoporous Silica Nanosphere for Overcoming Phagocytosis

Jun Young Lee, Chirag K. Vyas, Gun Gyun Kim, Pyeong Seok Choi, Min Goo Hur, Seung Dae Yang, Young Bae Kong, Eun Je Lee, Jeong Hoon Park\*

#### Jun Young Lee et al., Supplementary Method

#### Production of <sup>89</sup>Zr (Ref. 1)

<sup>89</sup>Zr was produced by <sup>89</sup>Y(p, n)<sup>89</sup>Zr reaction at RFT-30 an indigenous 30 MeV prototype cyclotron (Advanced Radiation Technology Institute, Korea Atomic Energy Research Institute) using the optimized irradiation conditions mounting the <sup>89</sup>Y metallic target on solid target irradiation station. In a nutshell, <sup>89</sup>Y (99.95%) was mounted at the irradiation station cooled with water jet. The initial 30 MeV of proton energy was degraded using 3.1 mm of aluminum degrader and the calculated incident proton energy on target was ~12.8 MeV. The target was irradiated using 12.8 MeV protons at 30 μA current for 2 hours producing 1.6 mCi/μA.h of <sup>89</sup>Zr with ≥ 99.99 % of radionuclidic purity. Irradiated target was allowed to cool for one hour and manually demounted and shifted to the hot-cell. The bulk target was dissolved in 6 M HCl (trace metal basis) after which hydroxamate based extraction chromatographic resin was employed for separation and purification of <sup>89</sup>Zr in oxalate and chloride form with ≥99.9% of radiochemical purity. The purified <sup>89</sup>Zr was employed for further labelling and biodistribution studies.

#### Preparation of <sup>18</sup>F-HMSN and <sup>68</sup>Ga-HMSN

<sup>18</sup>F was produced by <sup>18</sup>O(p, n)<sup>18</sup>F reaction at RFT-30 an indigenous 30 MeV prototype cyclotron (Advanced Radiation Technology Institute, Korea Atomic Energy Research Institute). The produced <sup>18</sup>F was separated and solvent exchanged through a QMA cartridge with K2.2.2/K<sub>2</sub>CO<sub>3</sub>. 370 MBq of <sup>18</sup>F was stirred in acetonitrile with HMSN of 1 mg at 80 °C for 3 hours. After the addition of trifluoroacetic acid (20  $\mu$ L), the reaction was allowed to proceed at 80 °C for 1 hour. To remove "free <sup>18</sup>F", the supernatant was removed by centrifugation at 10,000 rpm for 5 min and washed twice with DW.

<sup>68</sup>Ga was eluted from a <sup>68</sup>Ge/<sup>68</sup>Ga generator (Eckert&Ziegler, Obninsk, Germany) with 407 - 481 MBq activity per elution. After elution in 1 mL of 0.1M HCl, the <sup>68</sup>GaCl<sub>3</sub><sup>+</sup> was

neutralized with 1 M NaOH solution, immediately added to HMSN suspension (500  $\mu$ L of 0.1 M HEPES buffer) and incubated at 70 °C for 1 hour.

# Reference

 Holland, J. P., Sheh, Y. & Lewis, J. S. Standardized methods for the production of high specific-activity zirconium-89. *Nuclear medicine and biology* 36, 729-739 (2009).



Supplementary Figure S1. FT-IR spectrum of prepared SNP and HMSN.



**Supplementary Figure S2.** Powder X-ray diffraction pattern of HMSN. A disordered mesostructure at 2-theta angle of 21°.



**Supplementary Figure S3.** TGA curve of HMSN in nitrogen atmosphere, heating rate: 10 °C/min.



**Supplementary Figure S4.** Representative TEM images of (A, B) HMSNs and (C, D) Rm-HMSNs with well-dispersed biomimetic nanoparticles



**Supplementary Figure S5.** Western blot results demonstrating that CD47 protein in intact with Rm-HMSNs



**Supplementary Figure S6.** Cytotoxic effect of Rm-HMSN and HMSN on the CT-26 cancer cell line measured by the MTT assay. Data are shown as mean  $\pm$  SEM of three separate experiments.

SEM; standard error of mean



**Supplementary Figure S7.** Optimized labelling efficiency of chelator free <sup>89</sup>Zr-SNP and <sup>89</sup>Zr-HMSN using neutralized (A) <sup>89</sup>Zr-oxalate and (B) <sup>89</sup>Zr-chloride. Data are shown as mean ± SEM of three separate experiments.





**Supplementary Figure S8.** Cellular uptake and Internalization of Rm-<sup>89</sup>Zr-HMSNs into various cancer cell lines (CT-26, KB, and A549 cells) at a series of time points (0.5, 1, 4, 24, and 48 hour). Data are shown as mean  $\pm$  SEM of three separate experiments.

	<sup>18</sup> F-HMSN	<sup>68</sup> Ga-HMSN
Labelling efficiency	2.93 ± 0.12	99.8 ± 1.64
Stability in human serum (Incubation time: 30 min)	28.7 ± 1.40	85.0 ± 1.22

**Supplementary Table S1.** Comparison of the labeling efficiency and stability of <sup>18</sup>F-HMSN and <sup>68</sup>Ga-HMSN as a positron emitter for PET tracer (n=3).

	<sup>89</sup> Zr-HMSNs			Rm- <sup>89</sup> Zr-HMSNs		
Incubation Time (hour)	Human serum	Cell culture media	0.9% NaCl	Human serum	Cell culture media	0.9% NaCl
0.25	91.57 ±	89.69 ±	91.61 ±	98.62 ±	98.64 ±	98.88 ±
	1.28	1.75	2.47	0.67	0.24	1.08
0.5	86.41 ±	69.87 ±	92.31 ±	95.44 ±	92.44 ±	98.90 ±
	2.14	1.21	2.14	0.34	1.22	0.94
2	89.84 ±	60.82 ±	92.00 ±	93.15 ±	89.65 ±	98.89 ±
	2.45	1.33	2.03	0.47	1.44	0.12
24	57.22 ±	44.23 ±	89.28 ±	91.25 ±	85.39 ±	98.87 ±
	1.45	2.04	1.45	1.54	1.34	0.34
48	43.62 ±	41.25 ±	88.12 ±	88.45 ±	81.24 ±	98.62 ±
	0.98	1.48	0.54	0.54	1.46	1.34
72	42.31 ±	38.09 ±	86.37 ±	86.15 ±	76.98 ±	98.76 ±
	1.26	0.87	1.58	1.66	0.87	0.87
144	26.62 ±	29.33 ±	80.97 ±	82.22 ±	75.55 ±	98.69 ±
	2.00	1.62	1.23	1.24	0.95	0.94
168	18.33 ±	21.24 ±	79.22 ±	80.09 ±	67.03 ±	98.77 ±
	1.45	1.34	0.36	1.63	1.45	0.62

**Supplementary Table S2.** Data were calculated from iTLC measurement and are presented as average  $\pm$  maximum error (n=3).



**Supplementary Figure S9.** *In vivo;* Small-animal PET images of <sup>18</sup>F-HMSN (top) and <sup>68</sup>Ga-HMSN (bottom).

	1 hr	24 hr	48 hr	72 hr	96 hr
blood	3.87 ± 0.26	7.11 ± 0.22	3.21 ± 0.15	3.85 ± 1.04	3.49 ± 0.34
heart	3.54 ± 0.53	4.18 ± 0.43	4.55 ± 0.11	4.34 ± 0.27	4.88 ± 0.69
lung	17.1 ± 1.61	5.93 ± 0.80	7.28 ± 1.42	5.31 ± 0.67	6.40 ± 0.50
liver	11.3 ± 0.93	14.2 ± 3.91	14.3 ± 1.24	16.3 ± 1.53	17.0 ± 1.58
spleen	13.8 ± 1.00	26.7 ± 4.93	29.8 ± 4.15	27.9 ± 7.01	41.5 ± 0.75
stomach	0.21 ± 0.04	0.23 ± 0.06	0.16 ± 0.01	0.14 ± 0.01	0.15 ± 0.01
intestine	0.09 ± 0.01	0.09 ± 0.01	0.08 ± 0.01	$0.07 \pm 0.03$	0.05 ± 0.01
pancreas	0.40 ± 0.12	0.34 ± 0.16	0.31 ± 0.07	0.34 ± 0.13	0.56 ± 0.05
kidney	0.22 ± 0.02	0.19 ± 0.04	$0.25 \pm 0.03$	0.18 ± 0.03	0.20 ± 0.01
muscle	0.12 ± 0.01	0.14 ± 0.01	$0.25 \pm 0.02$	0.17 ± 0.04	0.22 ± 0.06
fat	1.27 ± 0.10	0.19 ± 0.03	0.52 ± 0.39	0.28 ± 0.13	$0.20 \pm 0.04$
bone	1.21 ± 0.17	1.60 ± 0.40	3.14 ± 0.32	2.89 ± 0.61	3.41 ± 0.43
skin	0.28 ± 0.04	0.12 ± 0.01	0.11 ± 0.01	0.10 ± 0.04	0.06 ± 0.01
tail	2.37 ± 0.31	$0.45 \pm 0.02$	$0.55 \pm 0.06$	0.51 ± 0.22	$0.50 \pm 0.04$
tumor	1.90 ± 0.07	3.47 ± 0.09	2.66 ± 0.15	2.13 ± 0.13	1.47 ± 0.17

Jun Young Lee et al., Supplementary Table. S3

**Supplementary Table S3.** Uptake value (% ID/g) (n = 3 per group) from a biodistribution study in which mice received  $\text{Rm}^{-89}$ Zr-HMSN 1 hr to 96 hr prior to tracer administration.

	1 hr	24 hr	48 hr	72 hr	96 hr
blood	$0.49 \pm 0.02$	0.48 ± 0.01	0.83 ± 0.05	0.57 ± 0.08	0.42 ± 0.01
heart	0.54 ± 0.05	0.83 ± 0.04	0.58 ± 0.02	0.49 ± 0.03	0.30 ± 0.03
lung	0.11 ± 0.01	0.59 ± 0.03	0.37 ± 0.04	$0.40 \pm 0.04$	0.23 ± 0.02
liver	0.16 ± 0.01	0.25 ± 0.04	0.18 ± 0.01	0.13 ± 0.01	0.08 ± 0.01
spleen	0.13 ± 0.01	0.13 ± 0.01	0.09 ± 0.01	0.07 ± 0.01	0.03 ± 0.01
stomach	9.22 ± 0.76	15.1 ± 1.94	15.9 ± 0.30	14.9 ± 0.97	9.56 ± 0.79
intestine	20.3 ± 1.20	36.0 ± 2.16	30.3 ± 1.30	35.0 ± 9.21	25.0 ± 0.85
pancreas	4.98 ± 0.81	11.8 ± 2.99	8.82 ± 1.36	7.25 ± 2.12	2.60 ± 0.21
kidney	8.44 ± 0.78	18.2 ± 2.03	10.5 ± 0.69	11.8 ± 1.77	7.22 ± 0.79
muscle	14.70.70	24.2 ± 1.79	10.7 ± 0.51	12.3 ± 1.47	6.88 ± 1.32
fat	1.50 ± 0.08	18.2 ± 1.56	7.10 ± 2.56	8.69 ± 2.19	7.53 ± 1.20
bone	1.58 ± 0.09	2.26 ± 0.34	$0.85 \pm 0.06$	0.76 ± 0.10	0.43 ± 0.04
skin	6.84 ± 0.71	29.0 ± 1.23	22.5 ± 0.76	24.3 ± 6.38	21.7 ± 2.69
tail	0.81 ± 0.06	7.69 ± 0.11	4.79 ± 0.28	4.80 ± 1.21	2.95 ± 0.24
tumor	-	-	-	-	-

Jun Young Lee et al., Supplementary Table. S4

**Supplementary Table S4.** Tumor to blood and tissue ratios of mice (n=3 per group) from biodistribution results