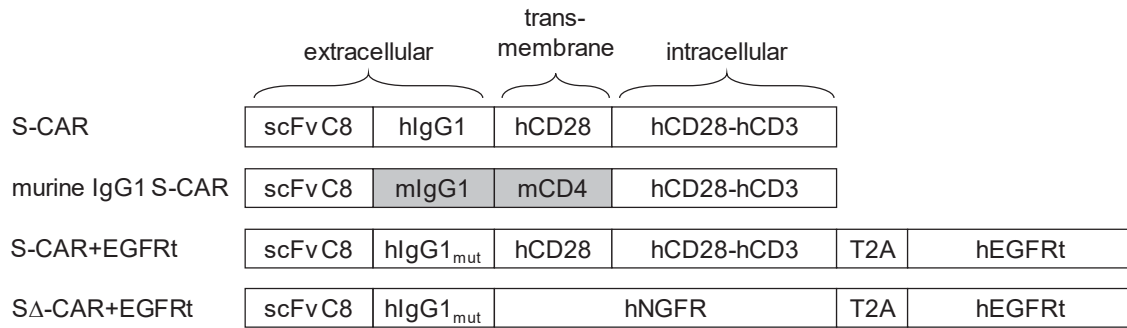


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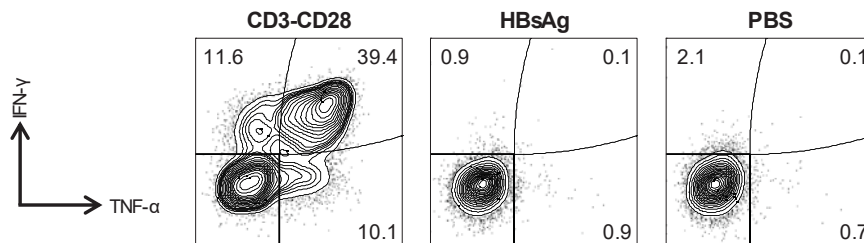
## **Supplemental Information**

### **Evaluation of a Fully Human, Hepatitis B Virus-Specific Chimeric Antigen Receptor in an Immunocompetent Mouse Model**

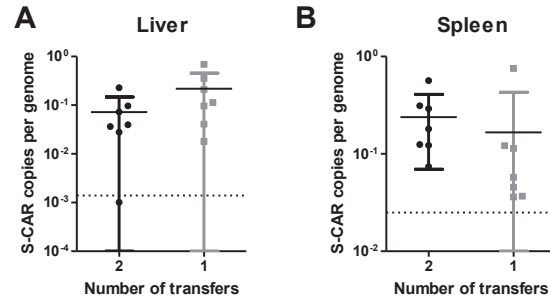
**Marvin M. Festag, Julia Festag, Simon P. Fräßle, Theresa Asen, Julia Sacherl, Sophia Schreiber, Martin A. Mück-Häusl, Dirk H. Busch, Karin Wisskirchen, and Ulrike Protzer**



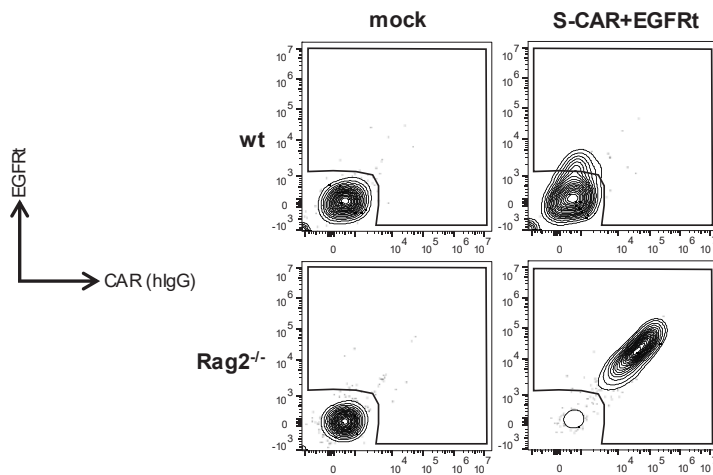
**Figure S1: Scheme of CAR constructs.** S-CAR and murine IgG1 S-CAR harbor the identical scFv C8 and intracellular signaling domains of human CD28 and CD3 $\zeta$ . The S-CAR has a human IgG1 spacer and a human CD28 transmembrane domain, the murine construct harbors a murine IgG1 spacer and the murine CD4 transmembrane domain (grey boxes). The human IgG1 S $\Delta$ -CAR harbors the domain of human nerve growth factor receptor (NGFR) instead of CD3 $\zeta$  and CD28 and serves as negative control. If indicated, human EGFRt is co-expressed utilizing a T2A element. If EGFRt is co-expressed, the S-CAR and S $\Delta$ -CAR contain a human IgG1 spacer with decreased Fc-receptor binding capacity (hIgG1<sub>mut</sub>).<sup>19</sup>



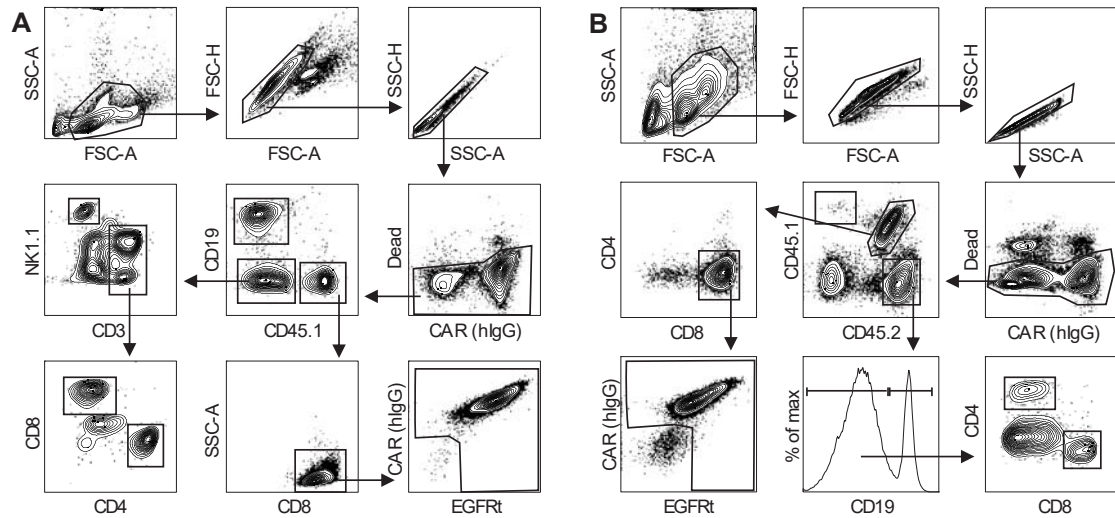
**Figure S2: Intracellular cytokine staining of stimulated T cells.** Exemplary flow cytometry plots of spleen-derived T cells cultured on HBsAg-, anti-CD3/anti-CD28- or PBS-coated control plates overnight (see also Figure 1E). Expression of IFN- $\gamma$  and TNF- $\alpha$  was analyzed on transferred CD45.1<sup>+</sup> CD8<sup>+</sup> T cells.



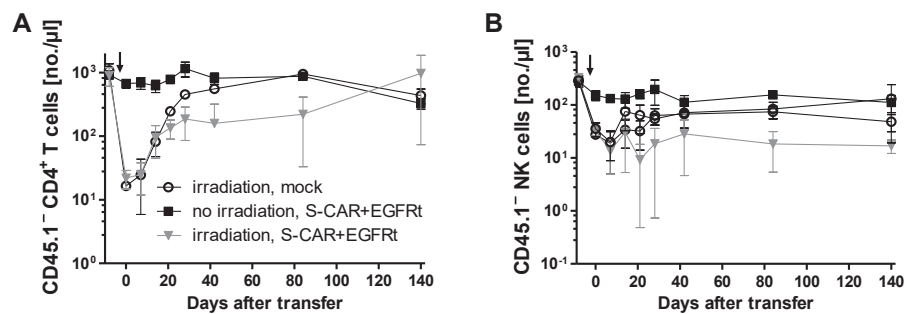
**Figure S3: Quantitative PCR for S-CAR integrates.** HBV-transgenic mice were injected once or twice with S-CAR T cells (n=7 per group, see also Figure 1). The number of S-CAR integrates normalized to the single copy gene *PRNP* in (A) liver and (B) spleen tissue was analyzed by quantitative PCR. A plasmid containing both the S-CAR and *PRNP* coding sequenced served as standard. Dotted line represents the background determined in untreated mice. Data points represent individual animals, mean values  $\pm$  SD are indicated.



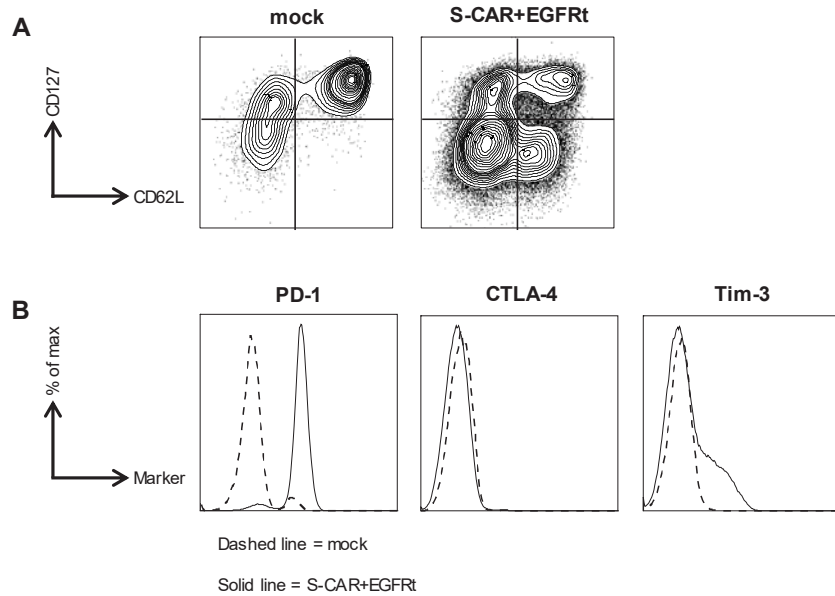
**Figure S4: S-CAR and EGFRt-expression on transferred cells.** Exemplary flow cytometry plots of S-CAR and EGFRt expression on CD45.1<sup>+</sup> T cells isolated from blood of HBV-naïve wildtype or Rag2<sup>-/-</sup> mice on day 10 after transfer (see also Figure 3A).



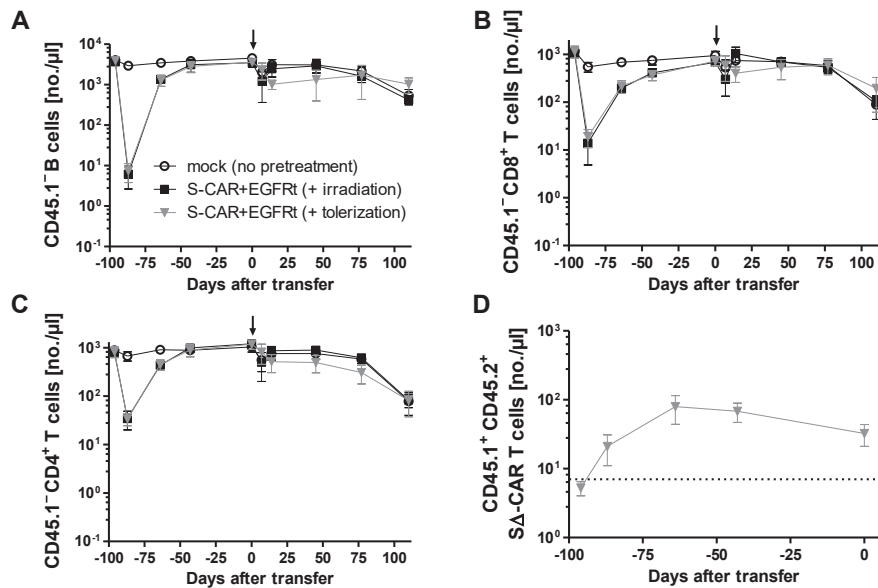
**Figure S5: Gating strategy to differentiate between endogenous and transferred immune cells. A)** Gating strategy applied to calculate results of figures 4C-E and S6. Lymphocytes were gated according to their size (FSC) and granularity (SSC). Subsequently duplets were excluded (FSC-A vs. FSC-H and SSC-A vs. SSC-H). After removal of dead cells, B cells (CD19<sup>+</sup>) were identified. Moreover CD45.1<sup>+</sup> cells (transferred cells) were distinguished from CD45.1<sup>-</sup> cells (endogenous cells). All transferred cells were identified to be CD8<sup>+</sup> as well as CAR<sup>+</sup> (hIgG<sup>+</sup>) and EGFRt<sup>+</sup>. Within the population of CD19<sup>-</sup> CD45.1<sup>-</sup> cells, NK cells (CD3<sup>+</sup> NK1.1<sup>+</sup>) as well as CD3<sup>+</sup> T cells were detected, whereby CD3<sup>+</sup> T cells were further analyzed according to the expression of CD4 and CD8. **B)** Gating strategy applied to calculate results of Figures 6D and S8. Lymphocytes were gated according to their size (FSC) and granularity (SSC). Subsequently duplets were excluded (FSC-A vs. FSC-H and SSC-A vs. SSC-H). After removal of dead cells, CD45.1<sup>+</sup> cells (cells of first transfer), CD45.1<sup>+</sup>/CD45.2<sup>+</sup> (cells of second transfer) and CD45.2<sup>+</sup> cells (endogenous cells) were identified. All CD45.1<sup>+</sup>/CD45.2<sup>+</sup> cells were identified to be CD8<sup>+</sup> as well as CAR<sup>+</sup> (hIgG<sup>+</sup>) and EGFRt<sup>+</sup>. Within the endogenous cells, CD19<sup>+</sup> B cells were distinguished from CD19<sup>-</sup> cells, which were used to further gate on CD4<sup>+</sup> and CD8<sup>+</sup> T cells.



**Figure S6: Concentration of CD4<sup>+</sup> T cells and NK cells in peripheral blood after irradiation.** AAV-HBV infected CD45.2<sup>+</sup> wildtype mice were irradiated one day before transfer of S-CAR<sup>+</sup>/EGFRt<sup>+</sup> T cells (see also Figure 4). **A)** Amount of (endogenous) CD45.1<sup>-</sup> CD4<sup>+</sup> T cells and **B)** NK1.1<sup>+</sup> NK cells per  $\mu$ l peripheral blood at indicated time points. Arrows mark time point of irradiation. All data are presented as mean values  $\pm$  SD. (n=4 per group)



**Figure S7: Memory and exhaustion marker expression of transferred T cells.** Exemplary flow cytometry plots pregated on transferred CD45.1<sup>+</sup> T cells in splenocyte population. **A)** Memory marker expression (see also Figure 4F). **B)** Exhaustion marker expression (see also Figure 4G).



**Figure S8: Concentration of endogenous immune cells and SΔ-CAR T cells after tolerization.** AAV-HBV infected CD45.2<sup>+</sup> wildtype mice were irradiated one day before SΔ-CAR<sup>+</sup>/EGFRt<sup>+</sup> T-cell transfer (see also Figure 6). Cell concentrations in peripheral blood monitored over time. Arrows mark time of S-CAR<sup>+</sup>/EGFRt<sup>+</sup> T-cell transfer on day 0. **A)** Concentration of CD45.1<sup>-</sup> CD19<sup>+</sup> B cells. **B)** Concentration of CD45.1<sup>-</sup> CD8<sup>+</sup> T cells. **C)** Concentration of CD45.1<sup>-</sup> CD4<sup>+</sup> T cells. **D)** Concentration of CD45.1<sup>+</sup> CD45.2<sup>+</sup> SΔ-CAR<sup>+</sup>/EGFRt<sup>+</sup> T cells. The dotted line represents the background determined in untreated mice. All data are presented as mean values ± SD. (n=5 per group)