

Figure S1. Related to Figure 1.

(A) List of tumors, xenograft models, and cell lines used in this study and the datasets generated.

(B) Expression (RPKM) of *LIN28B* across a panel of high-grade gliomas (H3.3K27M, H3.3WT and H3.3G34R/V) and normal brain tissues (fetal and adult brain).

(C) Western blot of LIN28B, MYC and MYCN proteins across a panel of H3K27WT and H3.3K27M pHGG cell lines.

(**D**) Single cell profiling of *IRX2* and *PAX3* across GBM and HGG-H3K27M samples derived from Filbin et al. (Filbin et al., 2018).



Figure S2. Related to Figure 3.

(A) The sequence of the gRNA used to target the K27M mutant allele of H3F3A is shown (top). The tracks of MiSeq data at the target locus demonstrate the insertions and deletions present on the mutant allele, while WT reads are not affected (bottom).

(B-F) Western blot (B) and mass spectrometry (C-F) of H3WT (C), H3K27M (D), H3K27ac (E), and H3K27me3 (F) in BT245 and KO.

(G-H) Pie graphs and heatmaps illustrating number and proportion of gained, lost, and retained H3K27ac sites within DIPG-XIII (G) and BT245 (H) H3K27M driven models as compared to isogenic KO lines.

(I-J) Dot plot illustrating ChIP-Rx signal (Log2) at H3K27M models BT245 (I) and DIPG-XIII (J) and the minimal to no observable shift in H3K27ac density upon KO.

(K) ATAC-seq signal demonstrating loss of coherence with H3K27ac peaks between the K27M and WT groups (left) and between the K27M and K27M-KO groups (right). p<0.001=***.















Figure S3. Related to Figure 3.

(A) Cell viability of BT245 and isogenic KO lines to bromodomain inhibition by JQ-1.

(B-C) Cell viability of pHGG lines in a 7-day growth assay under treatment with bromodomain inhibitors JQ-1 and I-BET762.

(D-E) Cell viability of BT245 **(D)** and DIPG-XIII **(E)** and corresponding KO control lines to CBP/EP300 inhibitor CBP-30.

(F) Viability of pHGG lines in a 7-day growth assay under treatment with the CBP/EP300 inhibitor CBP-30.

(G) Immunoblot of an immortalized human astrocyte cell line that over-express H3K27M or empty vector control, along with quantification of H3K27me3 and H3K27ac levels.

(H) Quantification of H3K27ac in NSC models that over-express H3K27M as compared to empty vector controls.

(I) Cell viability of astrocytes that over-express H3K27M following treatment with C646 as compared to empty vector controls.



Figure S4. Related to Figure 4.

(A) Quantitative ChIP-Rx scaled reads between BT245 K27M (red) and KO (blue) cell cultures across chromosome 8.

(B) Enrichment of H3K27ac across the human genome in one megabase windows comparing BT245 versus KO clones using a Wilcoxon rank sum test. p<0.001=***. The horizontal line inside the box corresponds to the median, the lines above and below the box delimit the interquartile range (IQR), and the end of the whiskers delimit values up to 1.5 times the IQR.

(C) Enrichment of H3K27ac across repetitive regions (LINE, DNA, LTR, SINE) of the human genome annotated by the RepeatMasker database comparing BT245 vs KO using a Wilcoxon rank sum test. p<0.001=***.

(D-F) Changes in deposition of H3K27ac over repetitive elements in BT245 (n=2) relative to KO (n=1) **(D)** and in expression of repetitive elements in in BT245 (n=5) relative to early-passage KO lines that represent the effect of K27M-KO (n=3) **(E)** and late-passage KO that may have lost the effect of H3K27me3 restoration through passaging (n=2) **(F)**. Histograms showing the distributions of the log2 fold-change (LFC) of genomic features in K27M relative to KO. Orange: H3K27ac deposition over repetitive elements (Repbase). Green: transcription of repetitive elements (Repbase). Grey: H3K27ac (left) and transcription (middle and right) of Ensembl genes (GRCh37). Elements with sufficiently high expression (baseMean > 100) and LFCs in [-3,3] range shown here.

(G) Measurement of expression of a panel of ERV families by droplet digital PCR (ddPCR) in H3K27M lines compared to respective KO lines. Plots show mean +SEM of 3 experimental replicates, BT245 cell line. Y-axis is in log2 scale.

 Treatment
 - 0.rug

 Cell line
 - 0.rug

 HXC27 status
 - 0.rug

 - 0.rug
 - 0.rug

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Figure S5. Related to Figure 6.

(A) Hierarchical clustering of samples based on global transcriptomic profiles. Color key: low (blue) to high (red) similarity between pairs of samples. Similarity between samples is calculated using the Euclidean distance over their variant-stabilizing-transformed transcriptomic profiles.

(B-C) Changes in expression of repetitive elements in K27M relative to KO after combination treatment (panobinostat and 5-azacytidine) for both DIPG-XIII (n=3) relative to KO (n=2) (B) and BT245 (n=2) relative to KO (n=2) (C). Histograms showing the distributions of the log2 fold-change (LFC) of transcription of repetitive elements (green; Repbase) and Ensembl genes (grey; GRCh37) in K27M relative to KO. Elements with sufficiently high expression (baseMean > 100) and LFCs in [-3,3] range shown here. (D-E) Measurement of ERV family expression by ddPCR upon 48 hour treatment with panobinostat (50 nM) or 5-azacytdine 5 μ M), normalized to vehicle control in BT245 (D) and corresponding KO lines (E). Plots show mean +SEM of 3 experimental replicates of the BT245 line. Two-way analysis of variance compares vehicle to each drug, with Bonferroni posttest significance shown by p<0.001=***, p<0.01=**, p<0.05=*. Y-axis is in log2 scale.











Figure S6. Related to Figure 6.

(A) Dose response curves of H3K27M and KO lines of BT245 to 5-azacytidine in isogenic context. Plots show mean +/-SEM of 3 experimental replicates.

(B-C) Cell viability of H3.3K27M and WT cells in a 7-day growth assay across a panel of pHGG lines **(B)** and corresponding IC50 values **(C)**.

(**D**) Dose response curves of H3K27M and KO lines of DIPG-XIII to 5-azacytidine in isogenic context. Plots show mean +/-SEM of 3 experimental replicates.

(E-F) Cell viability in 7-day growth assay under treatment of lines with sub-IC50 doses of panobinostat (15 nM), 5-azacytidine (1.5 μ M) and their combination in BT245 versus KO (E) and to H3.3K27M compared to WT cell lines (F). Plots show mean +SEM of 3 experimental replicates. A two-tailed t-test compares combination treated lines, with p<0.001=***.













DIPG XIII				
	DMSO	Panobinostat	5-aza	combo
Median survival	86	95	107	125
BT245				
	DMSO	Panobinostat	5-aza	combo
Median survival	110	131	143	155
pcGBM2				
	DMSO	Panobinostat	5-aza	combo
Median survival	85	101	90	96

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Figure S7. Related to Figure 7.

(A-B) Waterfall plot illustrating changes in the expression levels of the interferon response genes in H3.3K27M HGG tumors (n=17) compared to H3K27WT (n=15) (A) and in BT245 cell line (n=5) relative to KO (n=5) at baseline (B). Y-axis: log2 fold-change of expression in K27M relative to K27WT or KO. Red: upregulated genes in K27M. Blue: downregulated genes. Significantly deregulated genes highlighted (p \leq 0.05; baseMean \geq 100).

(C-D) Waterfall plots illustrating the induction of interferon response genes in DIPG-XIII relative to KO upon treatment with panobinostat and 5-azacytidine (C) and panobinostat alone (D). Y-axis: log2 fold-change of expression in K27M relative to KO (combination DIPG-XIII n=3, DIPG-XIII-KO n=2; panobinostat DIPG-XIII n=3, DIPG-XIII n=2).

(E) Western blot demonstrates decreased expression of MAVS protein (57 kD band) by shRNA targeting. (F-I) Dose response curves to 5-azacytidine in a 7-day growth assay of H3.3K27M and KO lines of DIPG-XIII (F-G) and BT245 (H-I) transduced with lentiviral vectors expressing short hairpin RNA (shRNA) controls (non-targeting, NT) or a pool of 4 shRNAs targeting MAVS. Plots show mean +/-SEM of 3 experimental replicates.

(J) Western blot of phosphorylated eIF2-alpha upon 48 hours treatment with panobinostat (50 nM) or 5-azacytidine (5 μ M) in BT245 and KO lines.

(K) Summary of median survivals of mice bearing PDX models of DIPGXIII, BT245, and pcGBM2 following treatment with DMSO, panobinostat, 5-azacytidine, and combination as related to Figures 7E-F.