

**Table S1: Sec22 SNARE homologues used during this study**

Organism	E-Value	NCBI reference number	Longin domain (aa)	Coil coiled region (aa)	Low complexity region (aa)	TMD length (aa)
<i>Saccharomyces cerevisiae</i>	8e-161	NP_013370.1	33-116	127-150	159-172	189-211
<i>Sordaria macrospora</i>	9e-163	XP_003346158.1	34-119			197-216
<i>Colletotrichum orbiculare</i>	4e-149	BAO27797.1	34-118	129-152		191-213
<i>Fusarium graminearum</i>	7e-163	XP_011323737.1	34-118	129-151		191-213
<i>Magnaporthe oryzae</i>	2e-161	XP_003719791.1	34-119	130-153		192-214
<i>Arabidopsis thaliana</i>	1e-165	NP_172653.1	35-119			193-212
<i>Nicotiana tabacum</i>	7e-165	XP_016484743.1	35-119			193-215
<i>Drosophila melanogaster</i>	5e-159	NP_569863.1	36-118			189-210
<i>Homo sapiens</i>	3e-162	NP_004883.3	36-118		162-179	191-213
<i>Rattus norvegicus</i>	1e-162	NP_001020857.1	36-118		162-179	191-213
<i>Mus musculus</i>	2e-162	NP_035472.1	36-118		162-179	191-213
<i>Caenorhabditis elegans</i>	7e-161	NP_508198.1	37-118		160-186	191-312
<i>Plasmodium falciparum</i>	7e-162	XP_001351271.1	31-123			194-216

\*Blue color represents absence of coil coiled region or low complexity region

Organism	Functions of Sec22	Sec22 interacting proteins	Function of Sec22 interacting proteins	References
<i>Saccharomyces cerevisiae</i>	Anterograde and retrograde trafficking, uptake of caesium ions, cellulase secretion, maintenance of ER morphology, autophagy	Bet1, Bos1, Sed5, Sec20, Ufe1, Slt1, Snc1/2, Sey1, Rtn1, Tip20, atg9,	Bet1, Bos1, Sed5 collaborate with sec22 during anterograde transport. Bos1 or Bet1 overproduction suppresses the secretory and growth defects of Sec22 mutant. <u>Sec20</u> , <u>Ufe1</u> , <u>Slt1</u> , <u>Snc1/2</u> the ER proteins cooperate with Sec22 in the retrograde Golgi to ER trafficking. Sey1, and Sec22 are involved in the homotypic ER membrane fusion. Sey1, Rtn1 and Sec22 are involved in the normal ER morphology. Tip20 could be responsible for binding or uncoating of COPI coated retrograde transport vesicles. A subset of <i>tip20</i> mutants was found to be lethal in combination with Sec22. Sec22 enables atg9 recruitment to the phagophore assembly site and helps in autophagosome	(Newman et al., 1990; Lewis and Pelham, 1996, Lewis et al., 1997, Cosson et al., 1997; Frigerio, G., 1998; Tsui et al., 2001; Liu and Barlowe, 2002; Liu et al., 2004; Hu et al., 2009; Nair et al., 2011; Anwar et al., 2012; Dr äxl et al., 2013; Lee, et al., 2015; Van Zyl et al., 2016)

			formation during autophagy.	
<i>Sordaria macrospora</i>	Effects vegetative growth, ascospore production, maturation, pigmentation, and germination. melanin biosynthesis, ER associated proteins and development related genes regulation	Melanin biosynthesis genes, development related genes		(Traeger and Nowrousian, 2015).
<i>Colletotrichum orbiculare</i>	Transport of virulence related effectors.			(Irieda et al., 2014).
<i>Fusarium graminearum</i>	cell wall integrity, mycelial growth, conidial production, infection of the host plant, endocytosis (unpublished data)			(Boenisch et al., 2017)
<i>Magnaporthe oryzae</i>	cell wall integrity, mycelial growth, conidial production, infection of the host plant, normal chitin deposition, regulation of ROS level, endocytosis, efficient expression of the extracellular enzymes peroxidases and laccases	Need to study song et al paper		(Song et al., 2010).
<i>Arabidopsis thaliana</i>	Gametophyte development. Regulation of uptake of non-			(El- Kasmi et al., 2011; Dr äxl et al., 2013).

	essential cation caesium (Cs b)			
<i>Nicotiana tabacum</i>	Overexpression causes collapse of Golgi membrane proteins into ER	Memb11, Phytolongins (Phyl)	Phyl1.1 export from the ER involves Sec22 along with other factors.	(Chatre et al., 2005; de Marcos Lousa et al., 2016)
<i>Drosophila melanogaster</i>	Its mutation causes ER proliferation, expanded ER lumens, abnormal Golgi morphology, enlargement of late endosomes. Essential for eye morphogenesis and Wingless (Wg)/Wnt signaling pathway	Wingless/Wnt secretion	Wingless (Wg)/Wnt signaling plays significant role in development, tissue homeostasis, and disease.	(Zhao et al., 2015; Li et al., 2015).
Mammals ( <i>Homo sapiens</i> , <i>Rattus norvegicus</i> , <i>Mus musculus</i> )	Membrane fusion, autophagy, regulate cell motion. Protein trafficking, translocation, and downregulation in the hippocampus of aging and Alzheimer's disease brains.	Sl1 Use1, Sec20, Syntaxin-18	Sl1 plays role in efficient protein secretion ER. Use1, Sec20, Syntaxin-18 play role in retrograde transport	(Burri et al., 2003; Dilcher et al., 2003; Chagoyen et al., 2006; Malsam and Söllner, 2011; Berchtold et al., 2013; Zhao et al. 2016).
<i>Caenorhabditis elegans</i>	Plays crucial role during import of RNAi silencing signals or cell autonomous RNAi	Sid5	Sec22 interacts with RNA transport protein Sid5 associated with late endosome	(Zhao et al., 2016).
<i>Plasmodium falciparum</i>	Encompasses signals for ER/Golgi recycling and fractional export beyond the ER/Golgi interface			(Ayong et al., 2009).

<b>Organism</b>	<b>Functions of Sec22</b>	<b>Reference</b>
<i>Saccharomyces cerevisiae</i>	Anterograde and retrograde trafficking, uptake of caesium ions, cellulase secretion, maintenance of ER morphology, autophagy	
<i>Sordaria macrospora</i>	Effects vegetative growth, ascospore production, maturation, pigmentation, and germination. melanin biosynthesis, ER associated proteins and development related genes regulation	
<i>Colletotrichum orbiculare</i>	Transport of virulence related effectors.	
<i>Fusarium graminearum</i>	cell wall integrity, mycelial growth, conidial production, infection of the host plant, endocytosis (unpublished data)	
<i>Magnaporthe oryzae</i>	cell wall integrity, mycelial growth, conidial production, infection of the host plant, normal chitin deposition, regulation of ROS level, endocytosis, efficient expression of the extracellular enzymes peroxidases and laccases	
<i>Arabidopsis thaliana</i>	Gametophyte development. Regulation of uptake of non-essential cation caesium (Cs <sup>þ</sup> )	
<i>Nicotiana tabacum</i>	Overexpression causes collapse of Golgi membrane proteins into ER	
<i>Drosophila melanogaster</i>	Its mutation causes ER proliferation, expanded ER lumens, abnormal Golgi morphology, enlargement of late endosomes. Essential for eye morphogenesis and Wingless (Wg)/Wnt signaling pathway	
<b>Mammals</b> ( <i>Homo sapiens</i> , <i>Rattus norvegicus</i> , <i>Mus musculus</i> )	Membrane fusion, autophagy, regulate cell motion. Protein trafficking, translocation, and downregulation in the hippocampus of aging and Alzheimer's disease brains.	
<i>Caenorhabditis elegans</i>	Plays crucial role during import of RNAi silencing signals or cell autonomous RNAi	

<i>Pasmodium falciparum</i>	Encompasses signals for ER/Golgi recycling and fractional export beyond the ER/Golgi interface	
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Sec22 interacting proteins	Function	References
Bet1	Bet1, Bos1, Sed5 collaborate with sec22 during anterograde transport. Bos1 or Bet1 overproduction suppresses the secretory and growth defects of Sec22 mutant.	
Bos1	Bet1, Bos1, Sed5 collaborate with sec22 during anterograde transport. Bos1 or Bet1 overproduction suppresses the secretory and growth defects of Sec22 mutant.	
Sed5		
Sec20	<u>Sec20, Ufe1, Slt1, Snc1/2</u> the ER proteins cooperate with Sec22 in the retrograde Golgi to ER trafficking	
Ufe1		
Slt1		
Snc1/2		
Sey1	Sey1, and Sec22 are involved in the homotypic ER membrane fusion. Sey1, Rtn1 and Sec22 are involved in the normal ER morphology.	
Rtn1		
Tip20	Tip20 could be responsible for binding or uncoating of COPI coated retrograde transport vesicles. A subset of <i>tip20</i> mutants was found to be lethal in combination with Sec22. Sec22 enables atg9 recruitment to the phagophore assembly site and helps in autophagosome formation during autophagy.	
atg9		
Melanin biosynthesis genes, development related genes		

Memb11,		
Wingless/Wnt secretion		
Syntaxin-17		
Syntaxin-18		
Sid5		
Phytolongins (Phyl)		

Table 1. The complete set of yeast SNAREs. Each of the yeast SNAREs is listed according to its structural classification as Q- or RSNARE.

The Q-SNAREs have been further divided as Qa-, Qb- and Qc-SNAREs (7) with one of each type of SNARE being required for a functional SNAREpin. Intracellular location (PM-plasma membrane, ER-endoplasmic reticulum) is shown: Golgi-vacuole denotes those

SNAREs localized to the prevacuolar compartment but also to Golgi or vacuole. The four SNAREs not anchored via a transmembrane segment are designated with an asterisks. The number of SNARE motifs is shown; note Sec9 and Spo20 have two motifs and fulfill the

requirement of both Qb- and Qc-SNARE within the plasma membrane SNAREpin. The approximate size of the N-terminal domain proximal

to the SNARE motif is indicated with references for the functional and/or structural analyses of the N-terminal domains given

Name	Location	SNARE motifs	N-term domain	Function of N-terminal domain	References
<b>Qa</b>					
Ufe1	ER	1	253	SM (Sly1) recruitment	(54)
Sed5	Golgi	1	248	SM (Sly1) recruitment	(54,59)
Tlg2	Golgi	1	242	SM (Vps45) recruitment	(55,56)
Pep12	Golgi-vacuole	1	194	SM (Vps45) recruitment	(53)
Vam3	vacuole	1	185	SM (Vps33) recruitment	(67)
Sso1	PM	1	188	SM (Sec1) recruitment	(68,69)
Sso2	PM	1	193	SM (Sec1) recruitment	(68,69)
<b>Qb</b>					
Sec20	ER	1	201	Tip20 recruitment.	(64)
Bos1	ER-Golgi	1	150	?	
Gos1	ER-Golgi	1	133	?	
Vti1	Golgi-vacuole	1	124	?	(70)
Sec9*	PM	2	432	?	
Spo20*	PM	2	178	?	(62)
<b>Qc</b>					
Slt1	ER	1	148	Frq1 recruitment?	
Sft1	Golgi	1	4	None	
Bet1	Golgi	1	50	?	
Tlg1	Golgi	1	130	?	
Syn8	Golgi-vacuole	1	163	Glc7 recruitment?	
Vam7*	vacuole	1	247	PHOX domain, PI-binding	(50)
Sec9*	PM	2	432	?	
Spo20*	PM	2	178	?	(62)
<b>R</b>					
Sec22	ER-Golgi	1	128	?	(58)
Ykt6*	Golgi-vacuole	1	135	Regulation/sorting of Ykt6?	(49,60)
Nyv1	Golgi-vacuole	1	160	?	
Snc1	PM-vesicles	1	23	None?	
Snc2	PM-vesicles	1	22	None?	

Table 1 Comparative analysis of Sec22 orthologs of different species	Query coverage	E value	Max Identity (%)	NCBI Reference number
<i>Fusarium graminearum</i>	100	6e-159	100	XP_011323737.1
<i>Alternaria alternata</i>	100	7e-154	97	KIL91621.1
<i>Saccharomyces cerevisiae</i>	99	3e-67	46	NP_013370.1
<i>Schizosaccharomyces pombe</i>	100	1e-82	55	NP_596218.2

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<i>Trichoderma harzianum</i>	100	1e-124	79	KKP06674.1
<i>Sclerotinia sclerotiorum</i>	100	2e-122	75	XP_001590003.1
<i>Neurospora crassa</i>	100	4e-116	75	XP_960888.1
<i>F. verticillioides</i>	100	3e-152	96	XP_018749557.1
<i>Arabidopsis thaliana</i>	98	5e-48	40	NP_172653.1
<i>Drosophila melanogaster</i>	96	3e-37	37	NP_001259103.1
<i>Plasmodium falciparum</i>	86	6e-28	31	XP_001351271.1
<i>Neonectria ditissima</i>	100	4e-136	84	KPM41255.1
<i>Cordyceps confragosa</i>	100	1e-129	80	OAA75842.1
<i>Botrytis cinerea</i>	100	3e-122	75	CCD48716.1
<i>Colletotrichum gloeosporioides</i>	100	2e-128	80	EQB50290.1
<i>Homo sapiens</i>	75	9e-39	43	XP_016858345.1
<i>Caenorhabditis elegans</i>	97	1e-34	34	NP_508198.1
<i>Glycine max</i>	98	8e-44	37	XP_003547819.1
<i>Mus musculus</i>	100	5e-45	39	NP_035472.1

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