# **Supplementary Information**

Synthesis of terminal ribose analogues of adenosine 5'-diphosphate ribose (ADPR) as probes for the Transient Receptor Potential (TRP) cation channel TRPM2

Ondřej Baszczyňski<sup>§2</sup>, Joanna M. Watt<sup>§1,2</sup>, Monika D. Rozewitz<sup>3</sup>, Andreas H. Guse<sup>3</sup>, Ralf Fliegert<sup>3</sup>¶ and Barry V. L. Potter<sup>1,2,¶</sup>\*

<sup>1</sup>Medicinal Chemistry & Drug Discovery, Department of Pharmacology, University of Oxford, Mansfield Road, Oxford, OX1 3QT, UK

<sup>2</sup>Wolfson Laboratory of Medicinal Chemistry, Department of Pharmacy and Pharmacology, University of Bath, Bath, BA2 7AY, UK

<sup>3</sup>The Calcium Signalling Group, Department of Biochemistry and Molecular Cell Biology, University Medical Center Hamburg-Eppendorf, Martinistrasse 52, 20246 Hamburg, Germany

 $\S =$  Equal contribution,  $\P =$  Equal contribution

#### **Contents:**

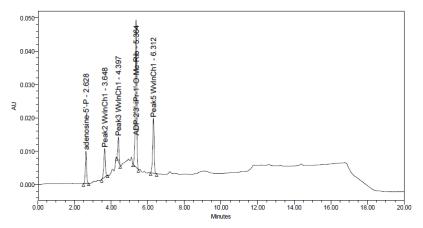
Supplementary Figure 1: Testing of suitable coupling procedure for pyrophosphate bond formation.	S2
Supplementary Figure 2: HPLC trace showing decomposition of 2"-deoxy-ADPR 3	S4
<sup>1</sup> H-NMR, <sup>13</sup> C-NMR and <sup>31</sup> P-NMR data for compounds <b>8-12</b> , <b>14-19</b> , <b>21-27</b> , and <b>30</b>	S5-S29
<sup>1</sup> H-NMR, <sup>13</sup> C-NMR and <sup>31</sup> P-NMR data for final compounds <b>1-6</b>	S30-S38
HPLC and MS data for final compounds 1-6	S39-S44

Supplementary Figure 1: Testing of suitable coupling procedure for pyrophosphate bond formation.<sup>1</sup>

a)

### Example of HPLC analysis of the coupling reaction (Peak 4 = product – compound 1)

	SAMPLE	INFORMAT	ION
Sample Name:	obb-57-own	Acquired By: Date Acquired: Acq. Method Set: Date Processed: Processing Method: Channel Name: Proc. Chnl. Descr.:	Joanna
Sample Type:	Unknown		09/02/2016 11:46:59 AM
Vial:	29		RP18 LC
Injection #:	1		25/02/2016 9:57:54 AM
Injection Volume:	10.00 ul		test protected ADPR
Run Time:	20.0 Minutes		WVInCh1
Sample Set Name:	2016_02_08_obb57own		PDA 259.1 nm

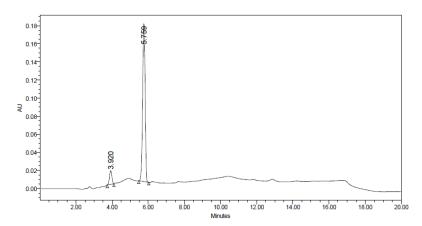


	Peak Name	RT	Area	% Area	Height
1	adenosine-5'-P	2.628	46143	10.11	9658
2	Peak2 WvlnCh1	3.648	50166	10.99	8606
3	Peak3 WvlnCh1	4.397	34789	7.62	7294
4	ADP-2'3'-iPr-1'-O-Me-Rib	5.364	235493	51.59	44239
5	Peak5 WvlnCh1	6.312	89866	19.69	16515

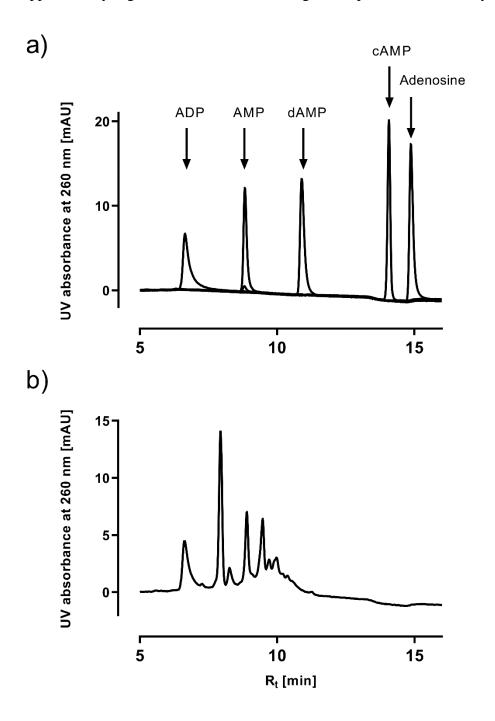
## b) Dabrowski-Tumanski procedure<sup>2</sup>

# Dabrowski-Tumanski procedure $^2$ : An example of HPLC analysis of the coupling reaction (Peak 4 = product - compound 1)

	SAMPLE	INFORMAT	ION
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time: Sample Set Name:	2016_02_24_obb59-own2 Unknown 63 1 10.00 ul 20.0 Minutes 2016_02_24_obb59_test	Acquired By: Date Acquired: Acq. Method Set: Date Processed: Processing Method: Channel Name: Proc. Chnl. Descr.:	WvlnCh1

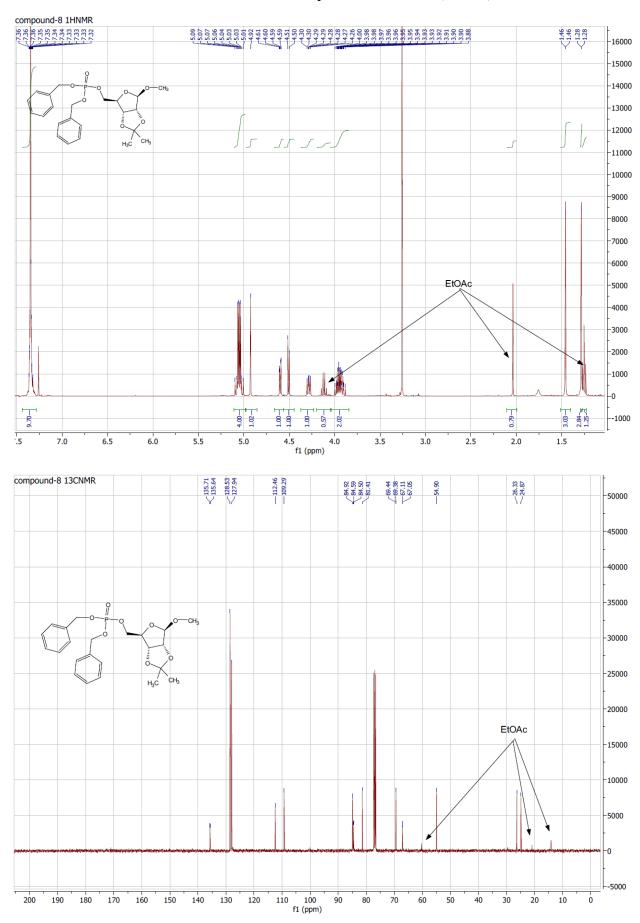


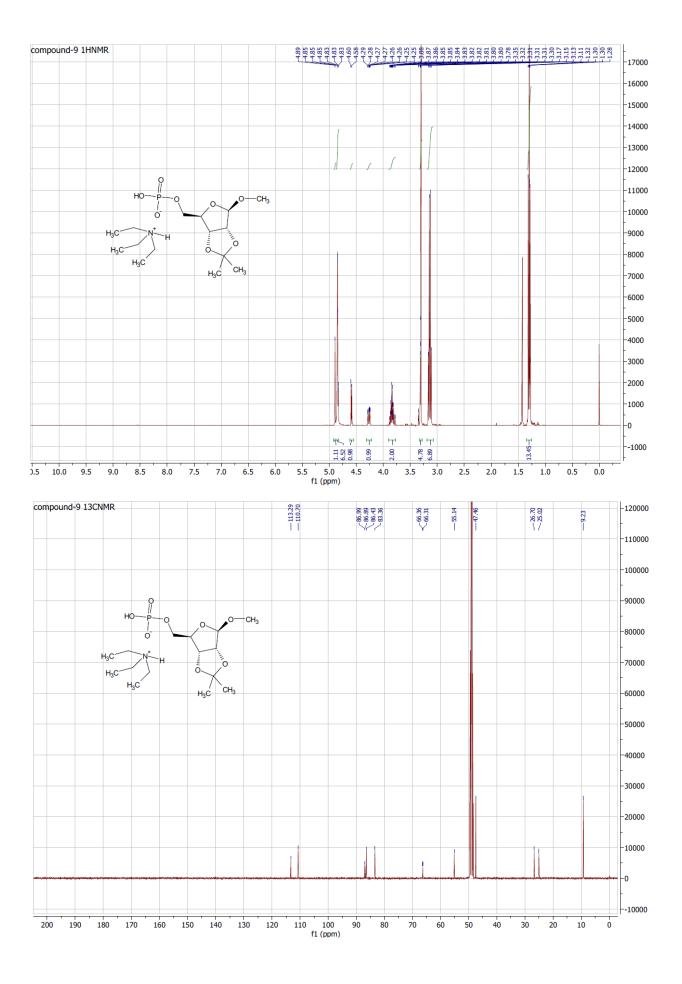
	Peak Name	RT	Area	% Area	Height
1		3.920	139613	7.63	15110
2	adenosine-5'-P-imidazolide	4.226			
3		5.759	1690715	92.37	174335
4	ADP-2'3'-iPr-1'O-Me-Rib	6.401			

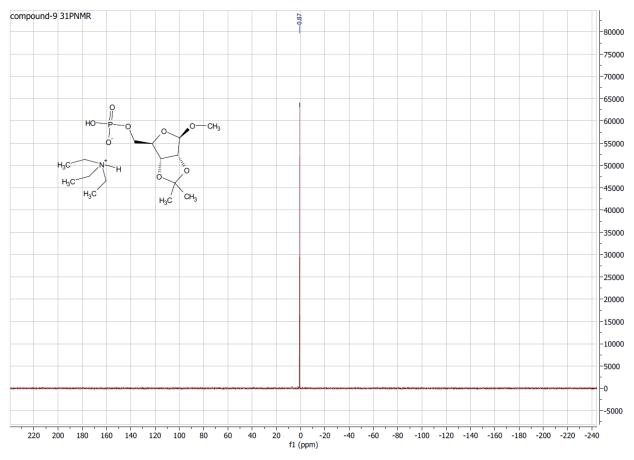


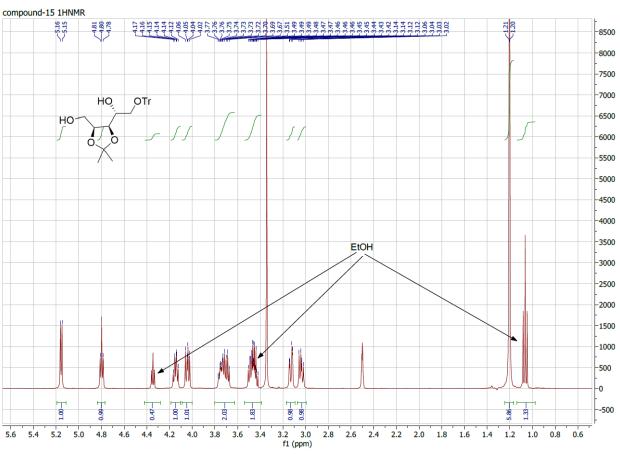
Integrity of 2"-deoxy-ADPR after transport and reconstitution was checked by RP-HPLC analysis on a 1260 Infinity system (Agilent Technologies). Samples and standards (ADP, AMP, dAMP, cAMP, Adenosine 250 pmol each) were run on a 250 mm × 4.6 mm Multohyp C18 5-µm column (Chromatographie Service) with a 4.0 mm × 3.0 mm guard cartridge containing a C18 ODS filter element (Phenomenex) at a flow rate of 0.8 mL/min with buffer (20 mmol/l KH<sub>2</sub>PO<sub>4</sub>, pH 6) with a linear gradient of methanol from 0 to 50% Methanol over 22.5 min. Adenine nucleotides were detected at 260 nm. Peaks were integrated using the ChemStation Software (Rev. C.01.05; Agilent Technologies). a) Chromatograms of standards (250 pmol each). b) Chromatogram of a preparation of 2"-deoxy-ADPR after freeze drying, transport and reconstitution in 10 mmol/l HEPES pH7.2. The sample should have contained 1 nmol of 2"-deoxy-ADPR, but eluted from the column as multiple peaks, one of the fragments co-elutes with ADP.

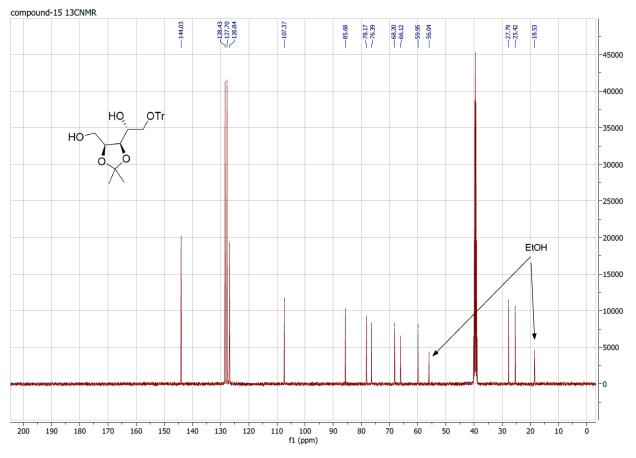
<sup>1</sup>H-NMR, <sup>13</sup>C-NMR and <sup>31</sup>P-NMR data for compounds **8-12**, **14-19**, **21-27**, and **30** 

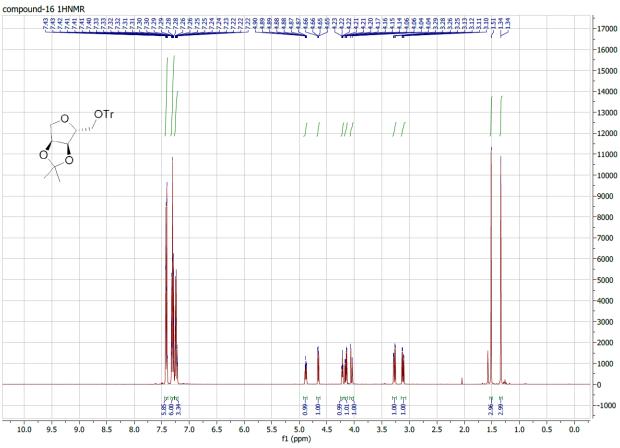


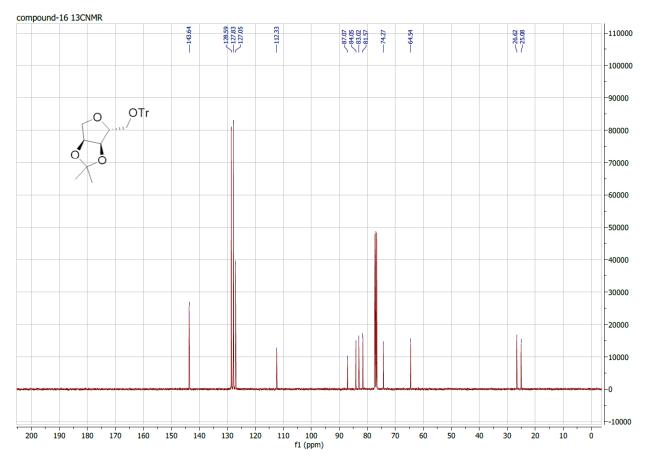


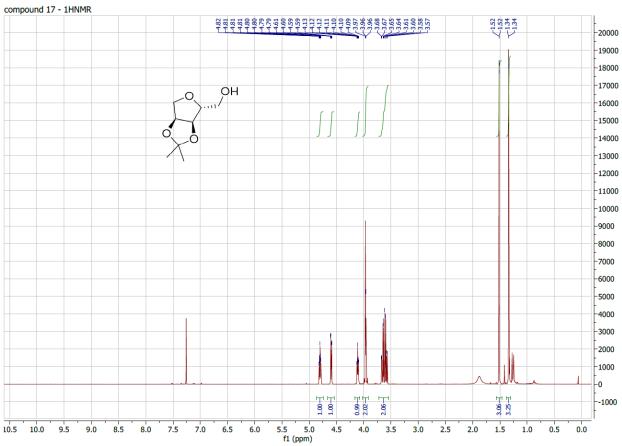


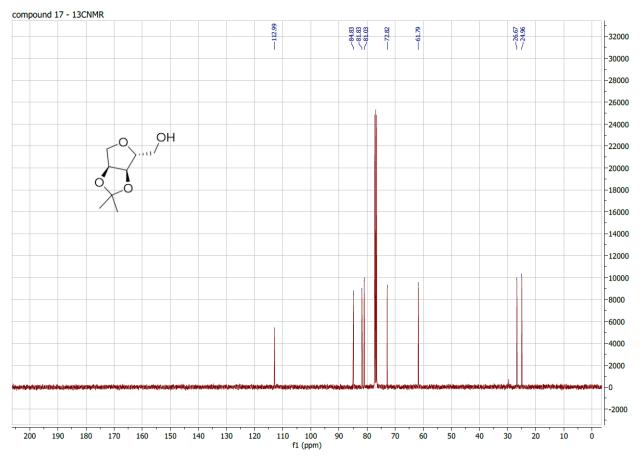


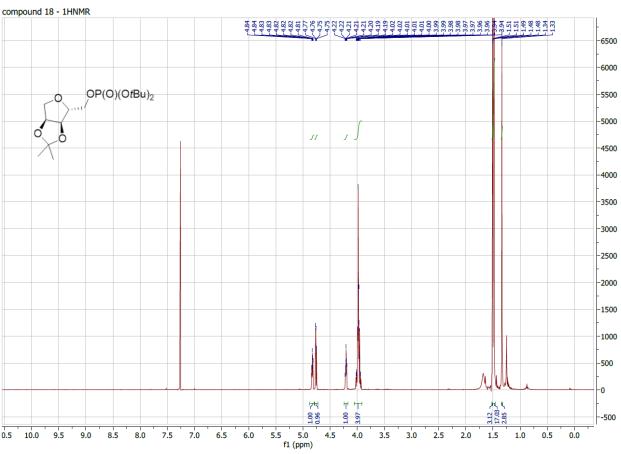


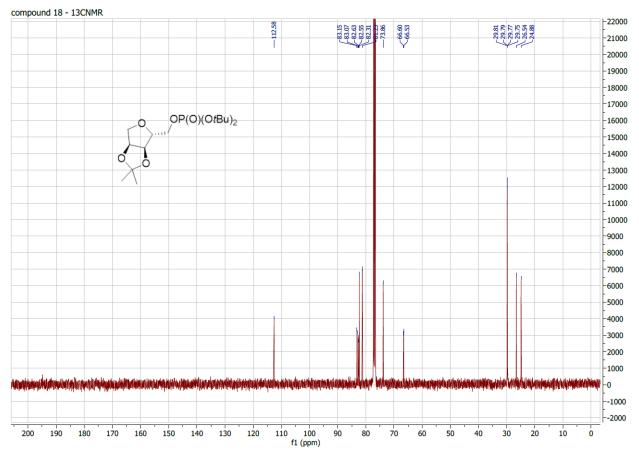


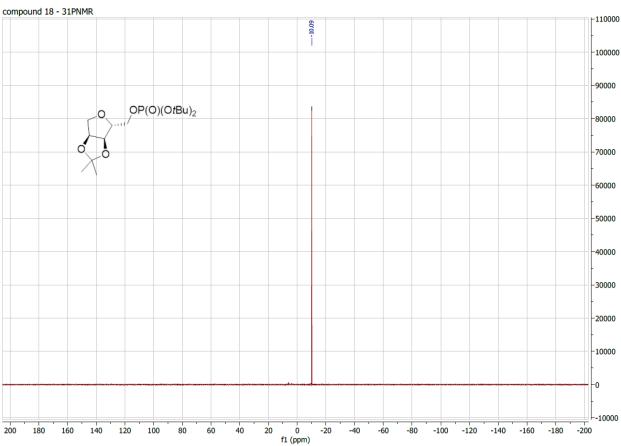


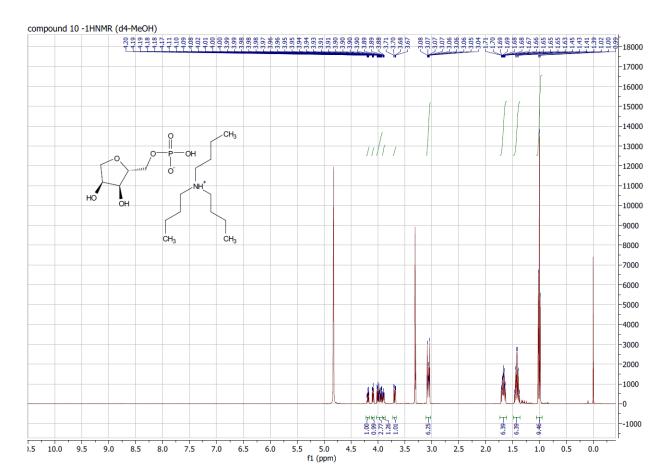


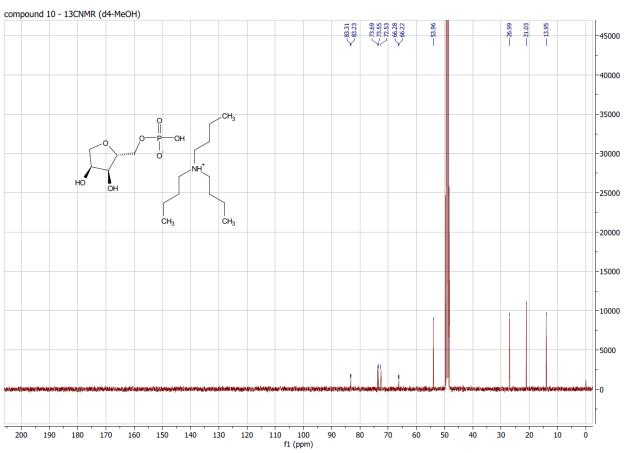


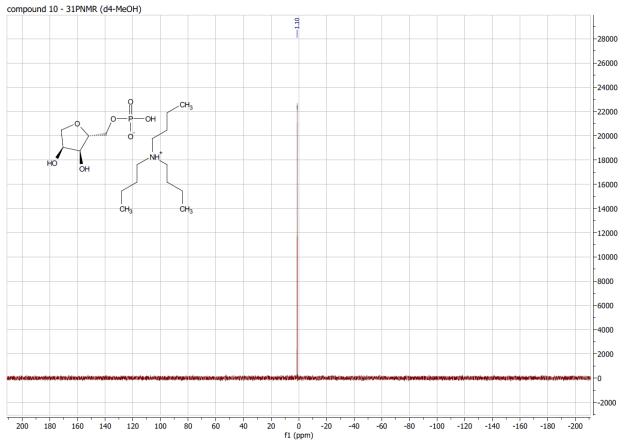


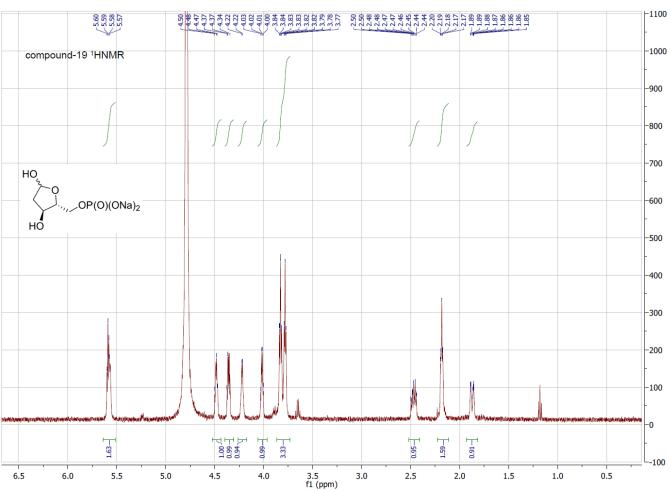


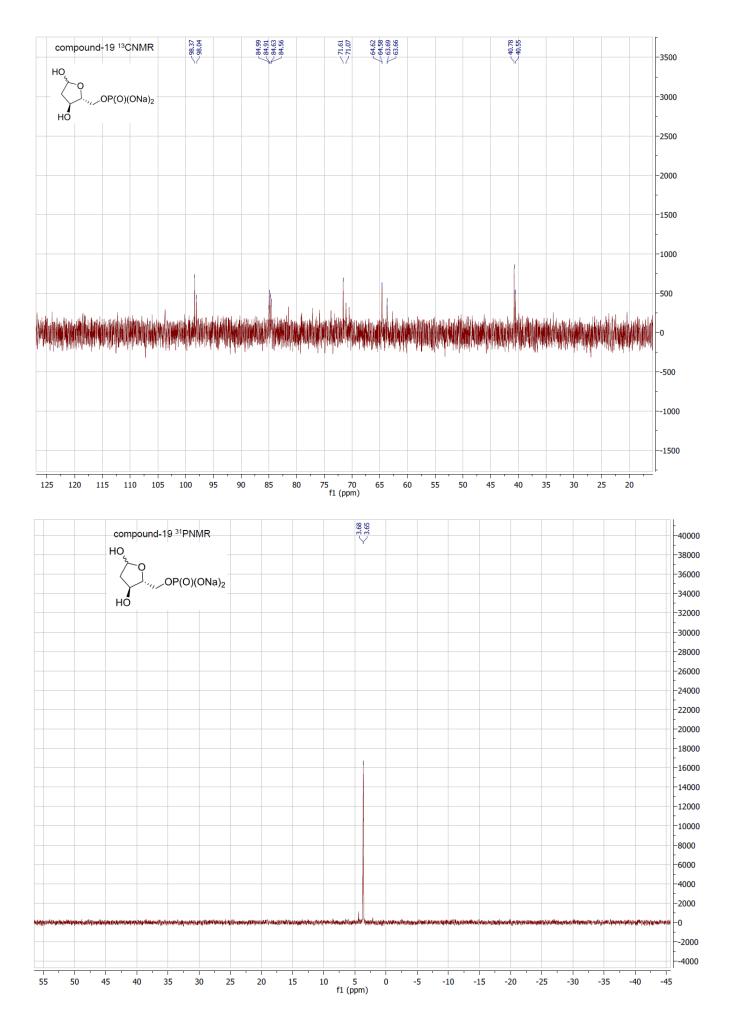


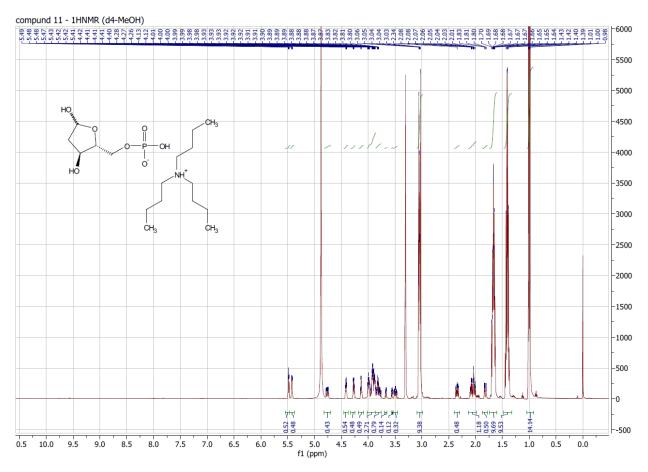


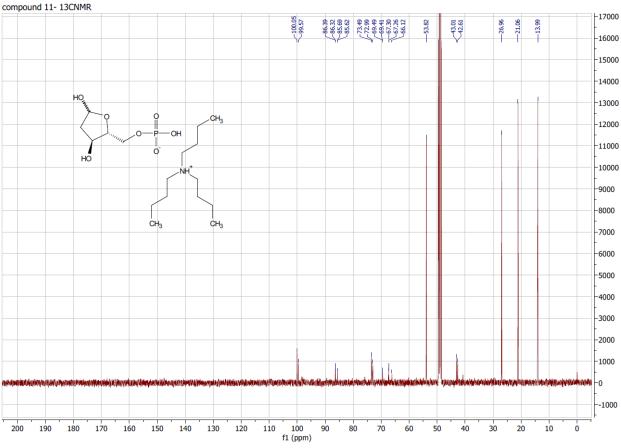


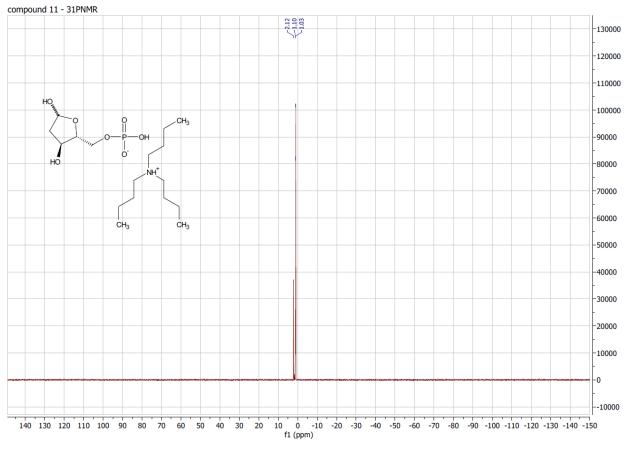


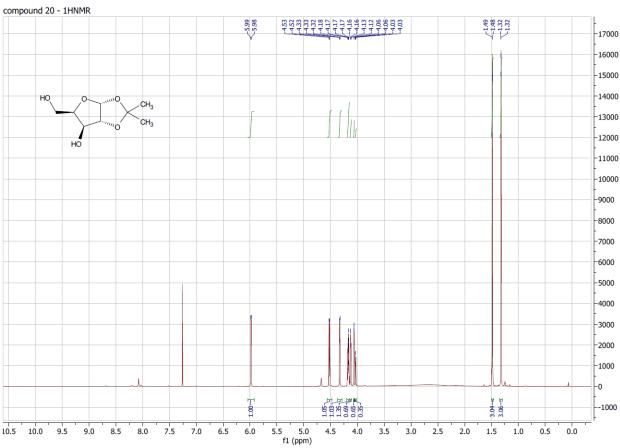


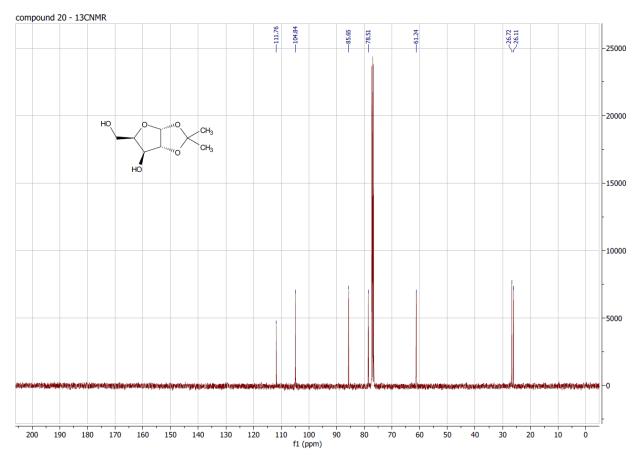


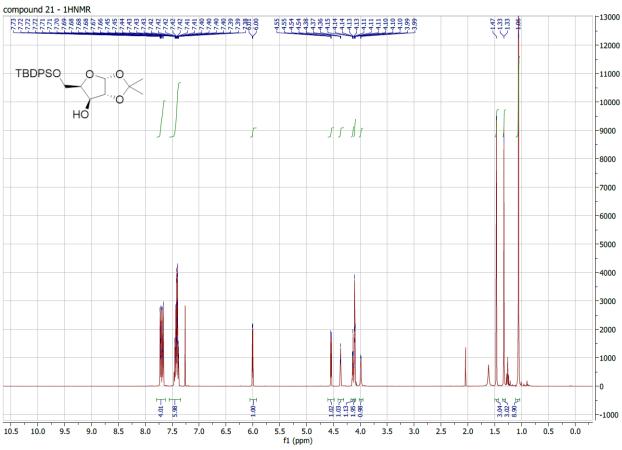


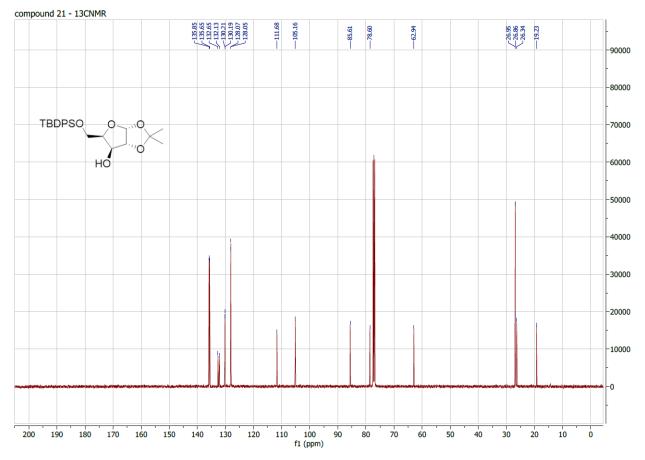


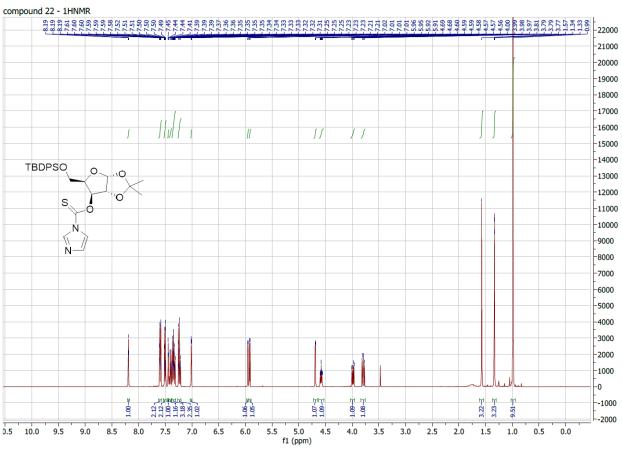


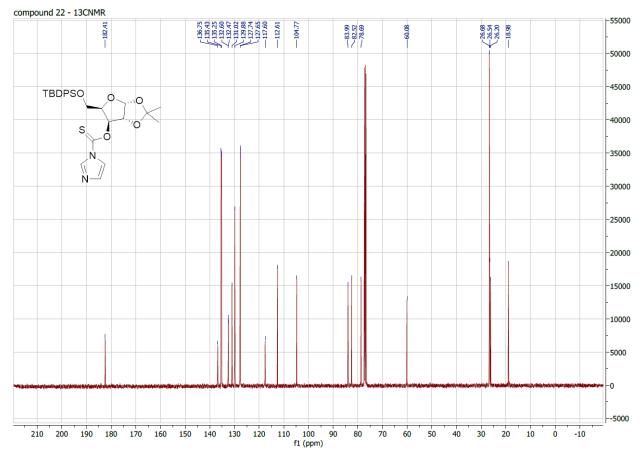


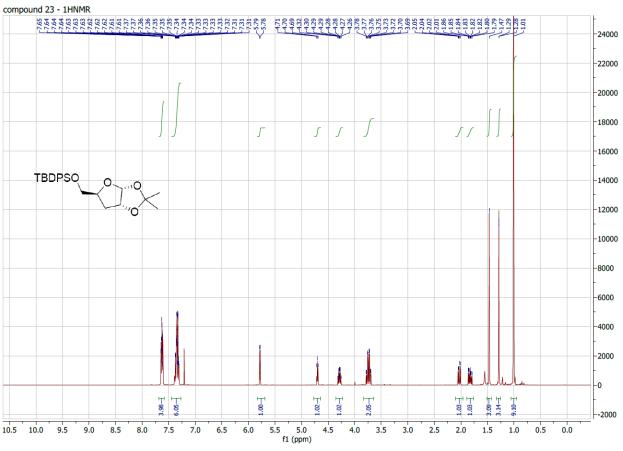


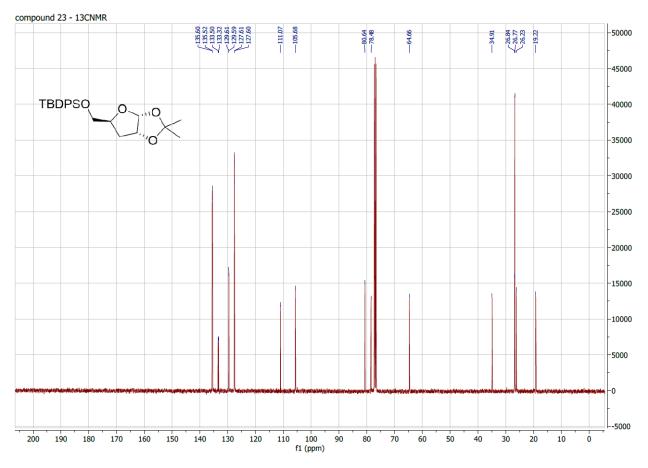


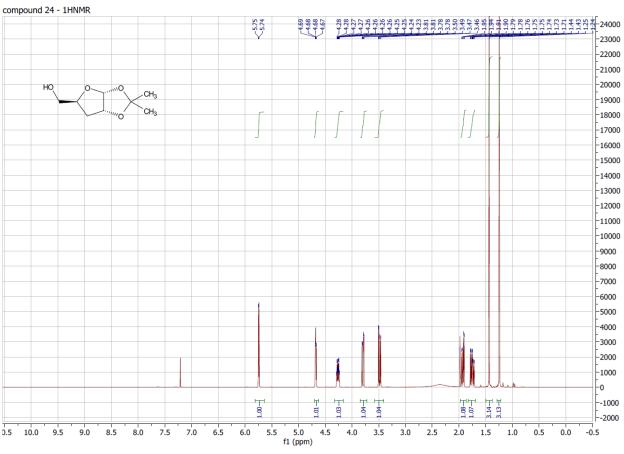


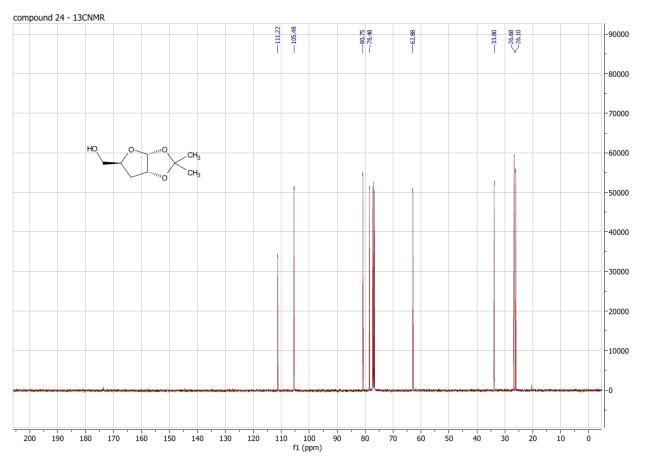


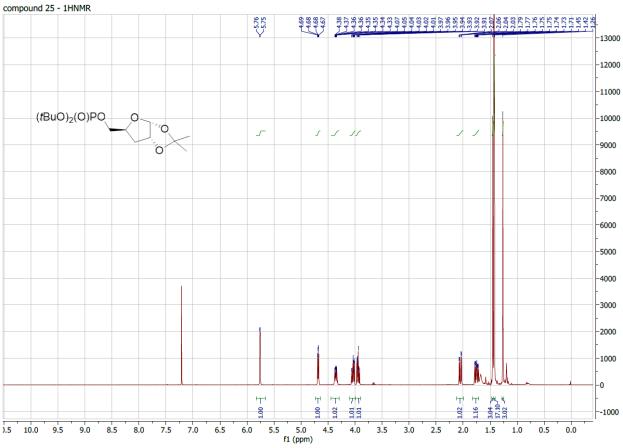


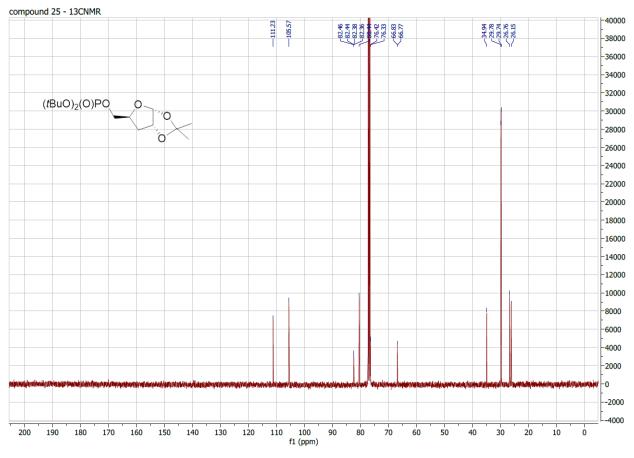


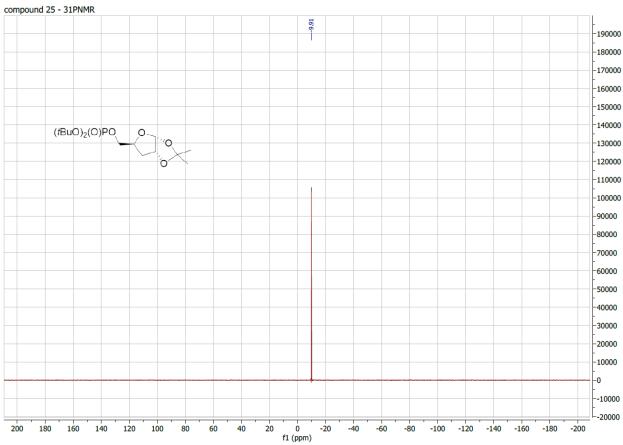


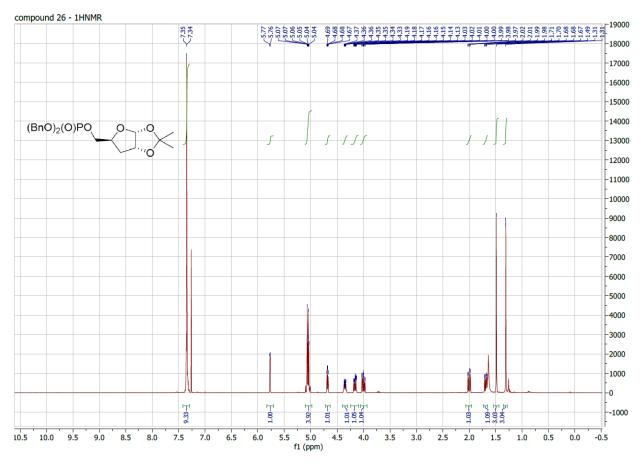


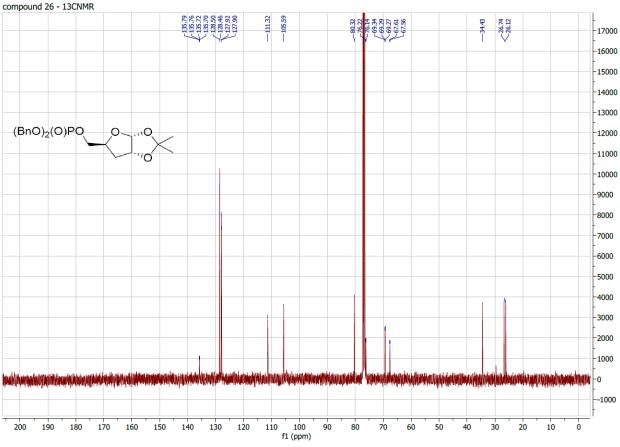


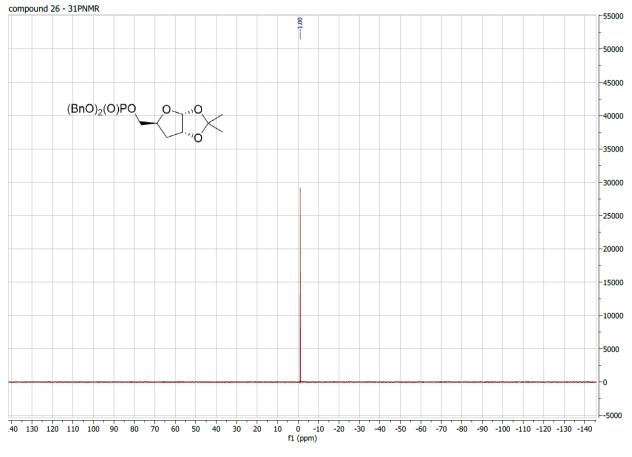


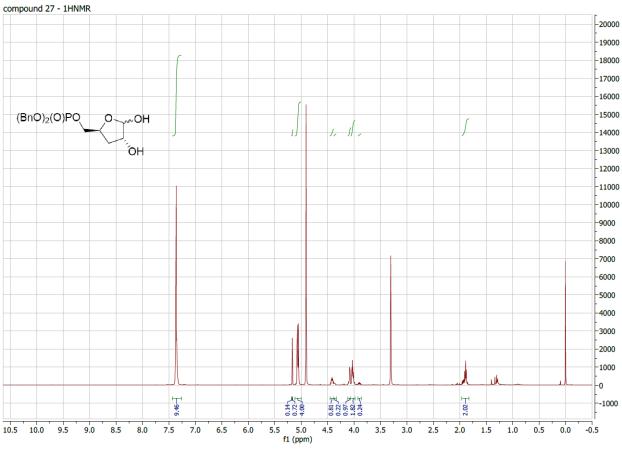


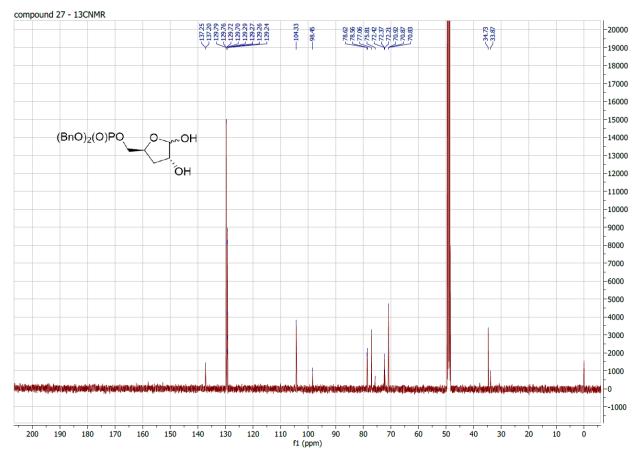


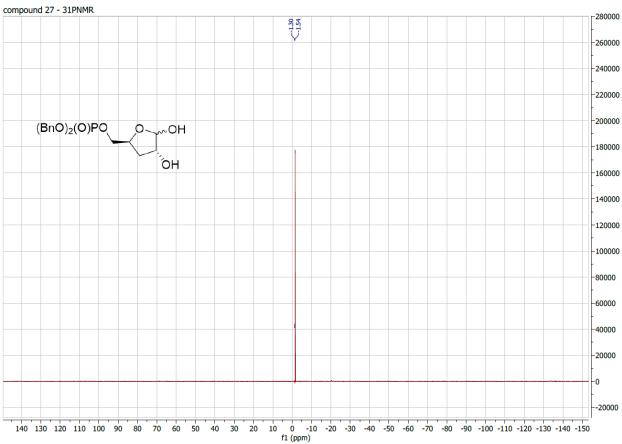


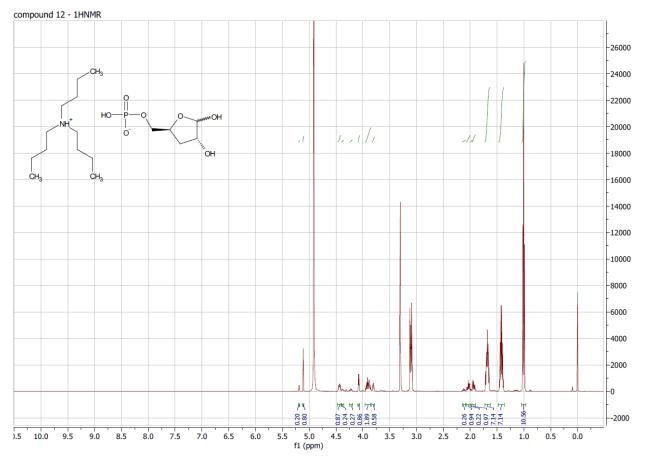


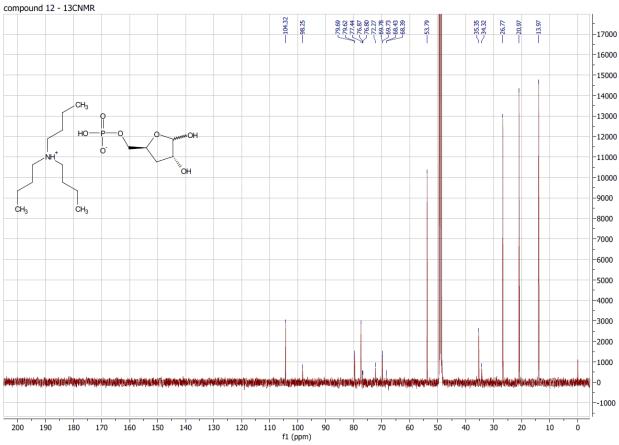


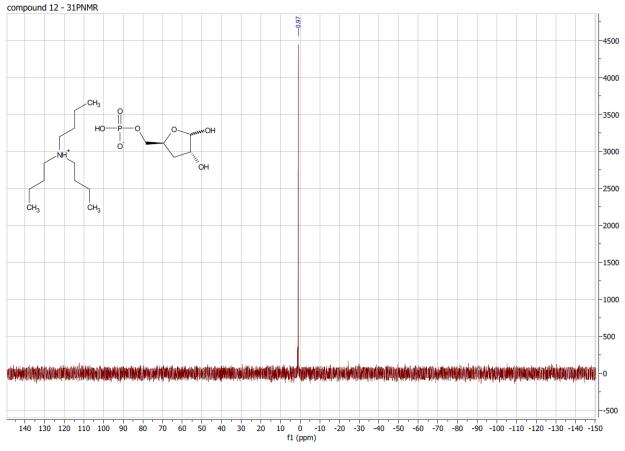


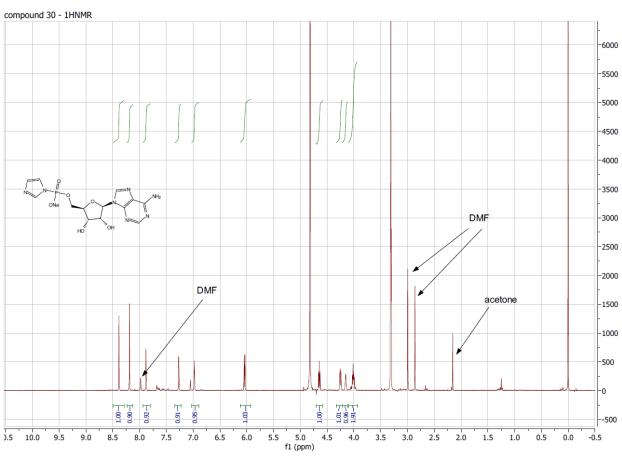


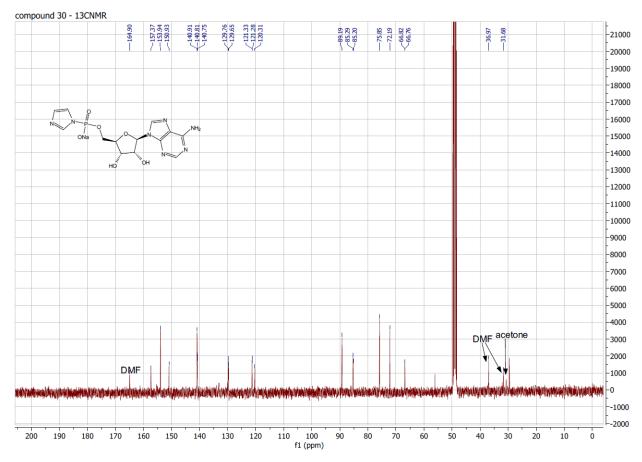


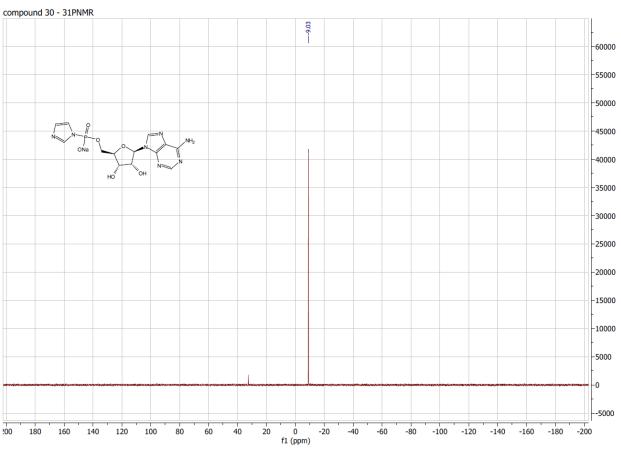


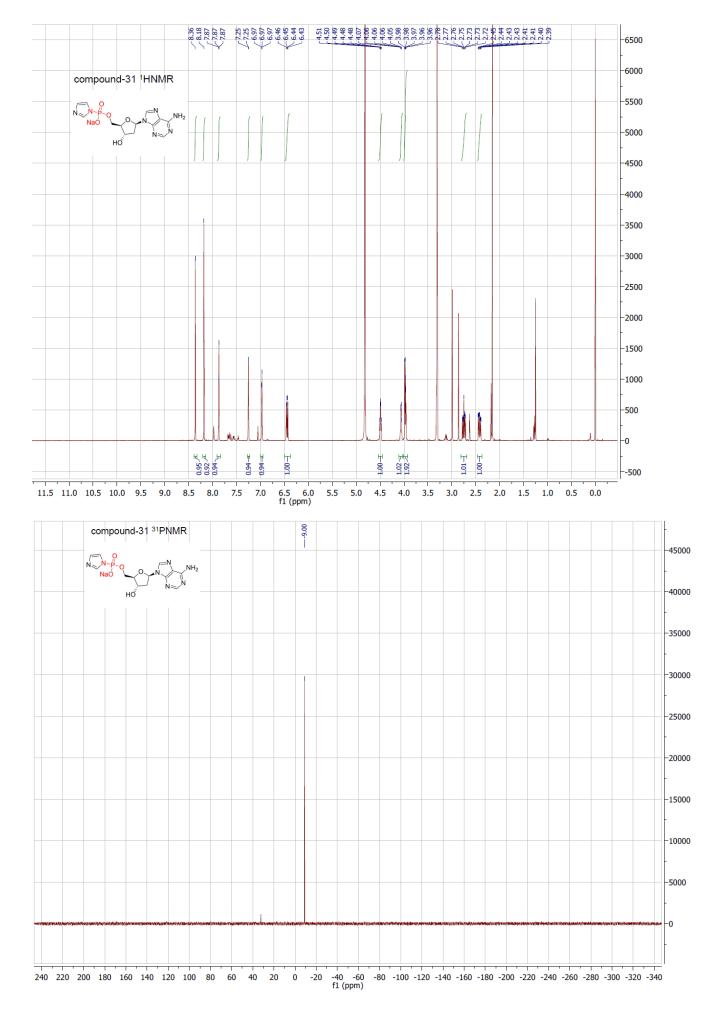


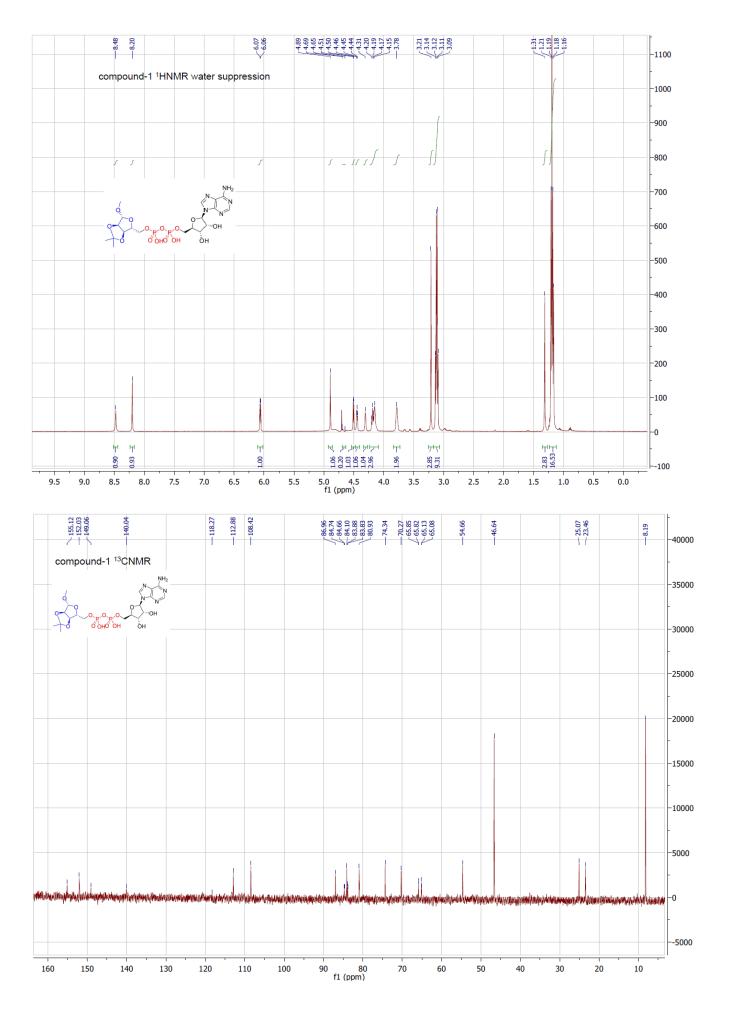


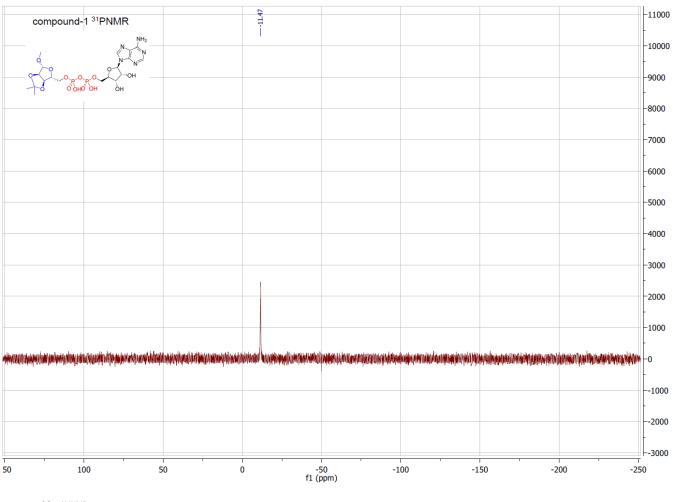


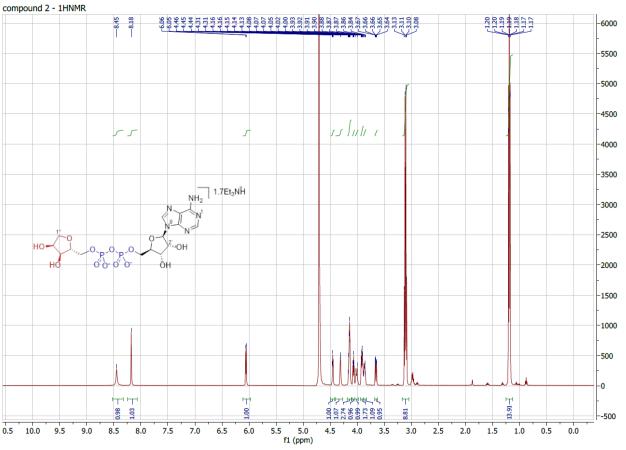


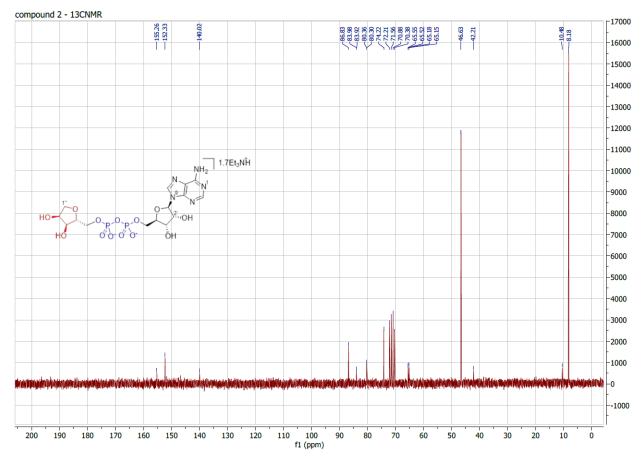


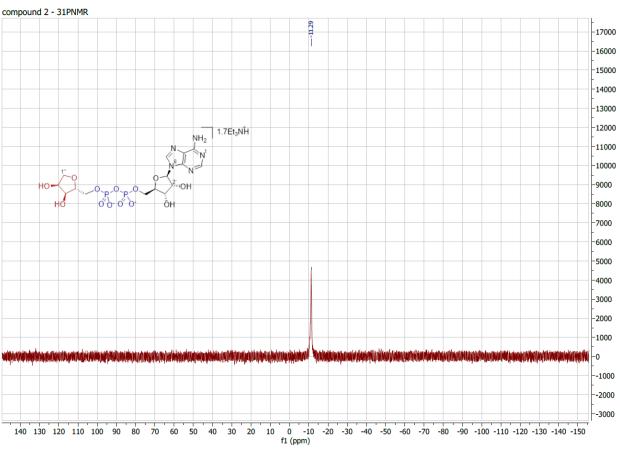


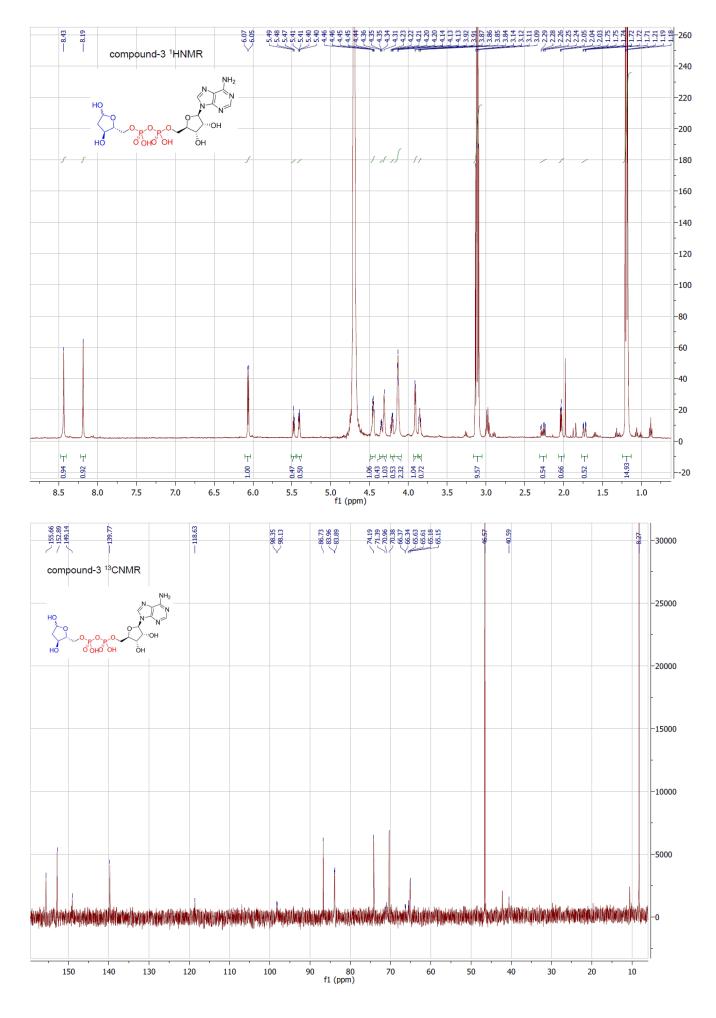


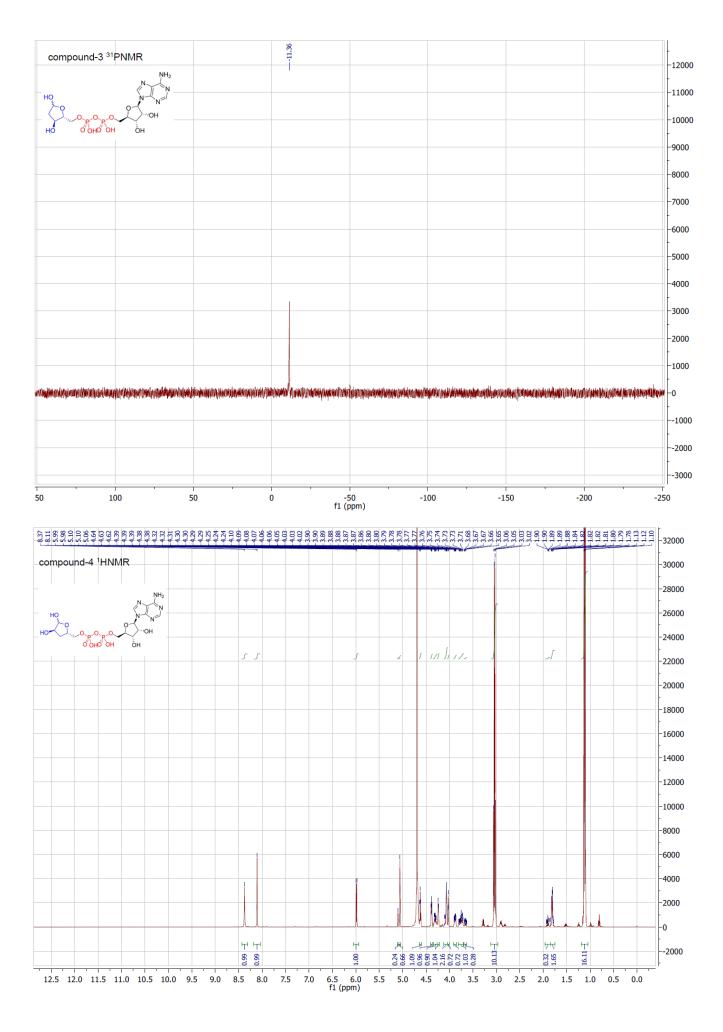


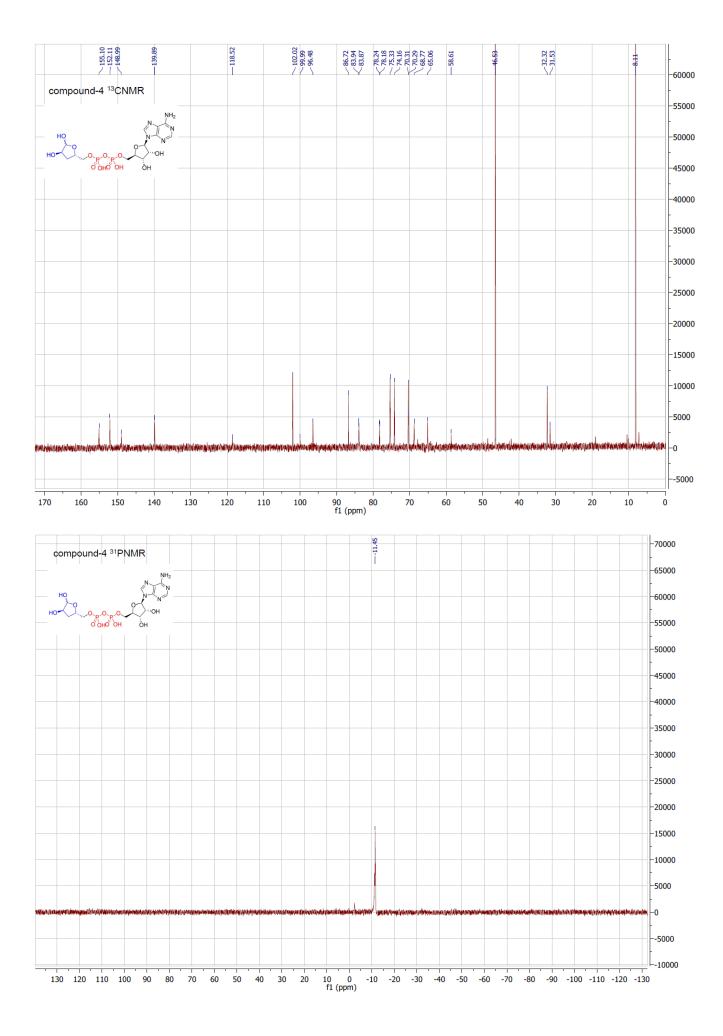


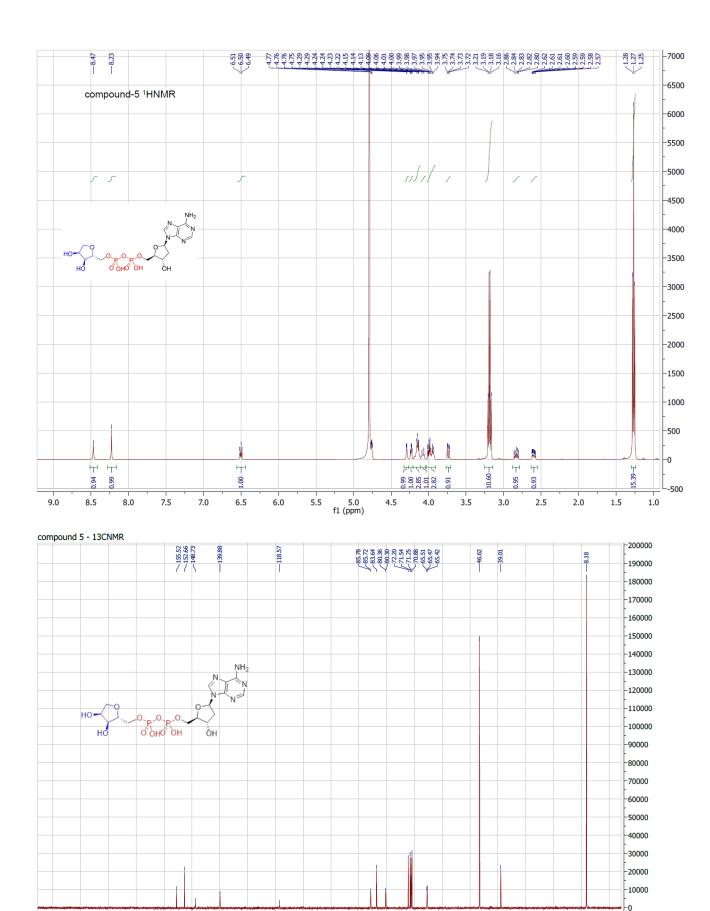










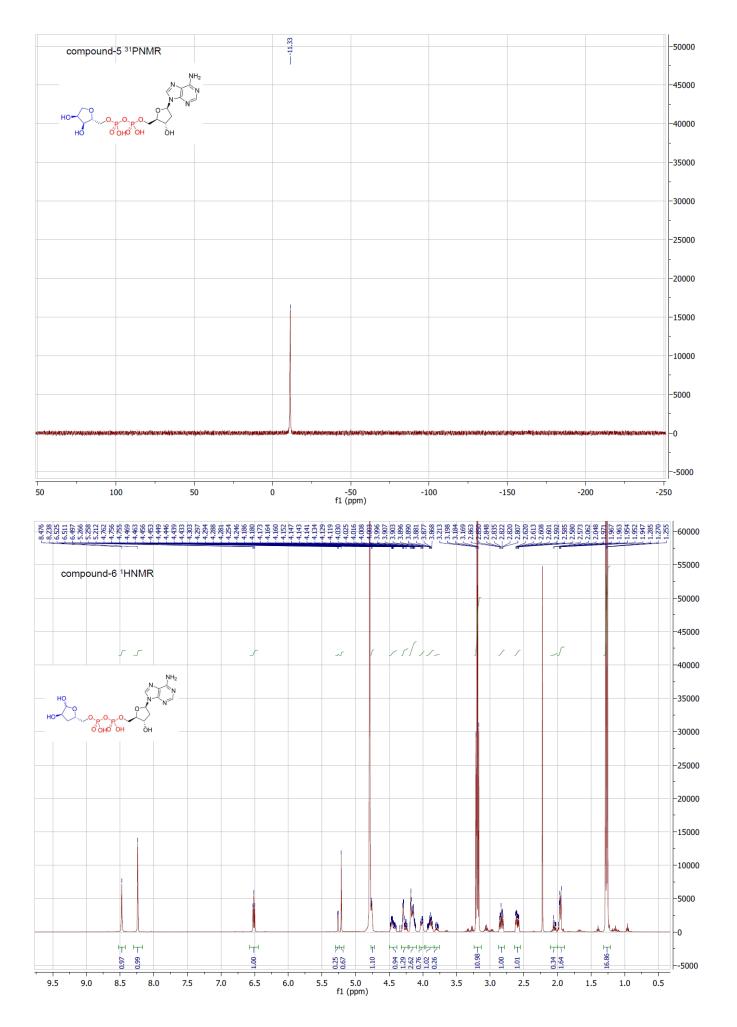


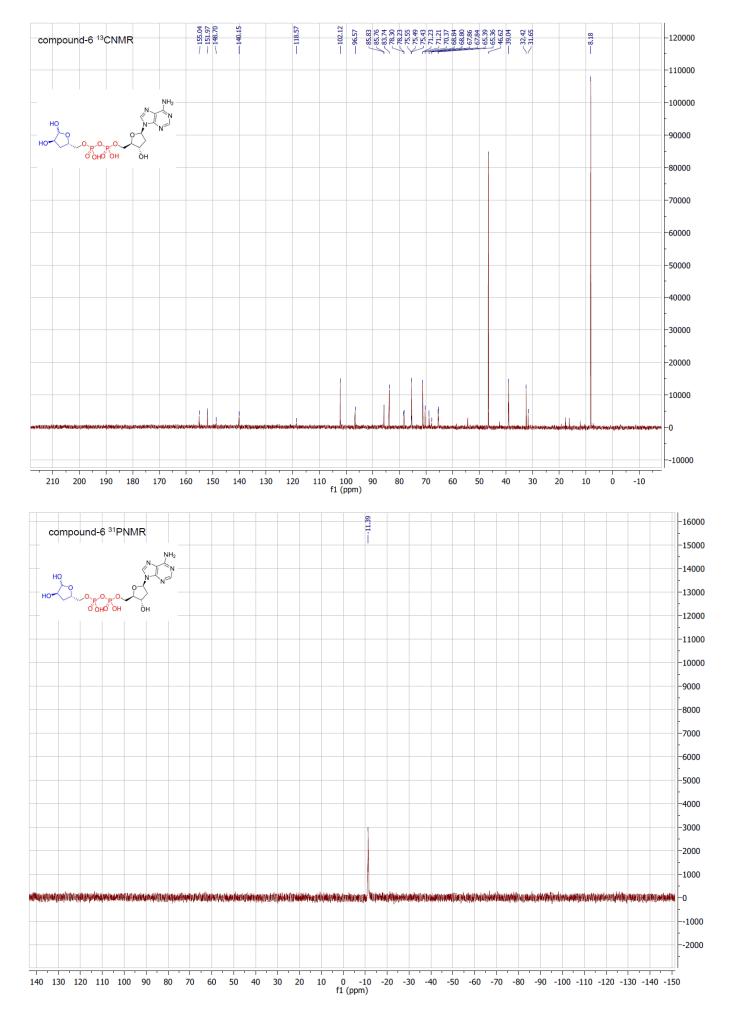
100 f1 (ppm)

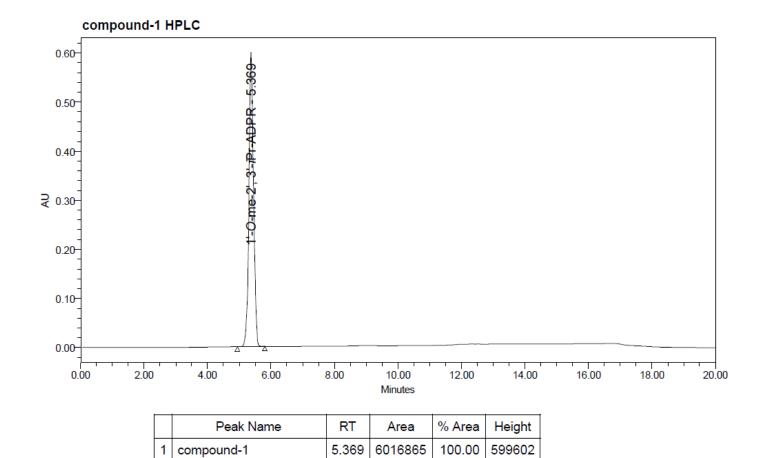
140

130

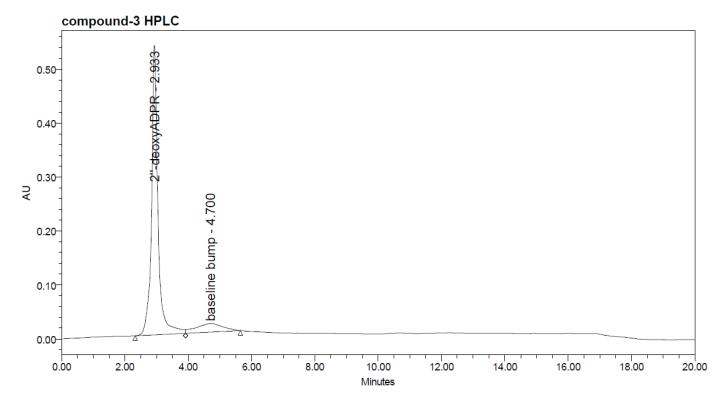
-10000



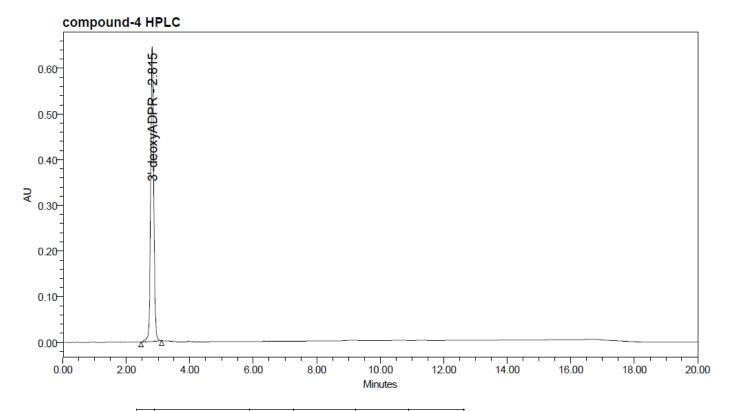




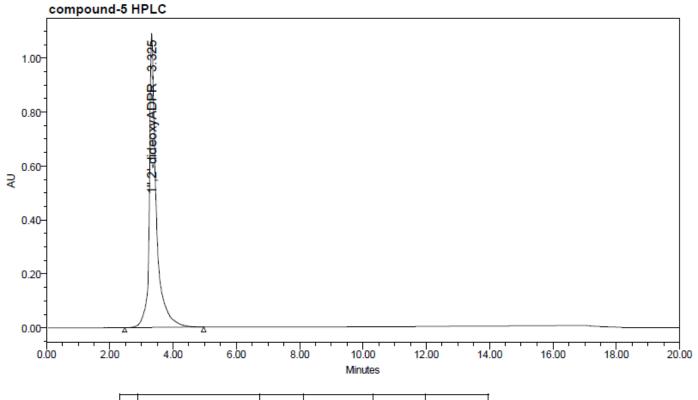
comp	pound-2 HPLC								
0.70-	<del>2.966</del> -								
0.60	DPR-								
0.50	"-deoxyADPR								
⊋ <sup>0.40</sup>	#	10							
0.30		baseline bump - 4.775							
0.20		dung e							
0.10		aseline							
0.00									
0.00	2.00 4.00	6.00	8.00	10.00 Minutes	12.00	14.00	16.00	18.00	20.00



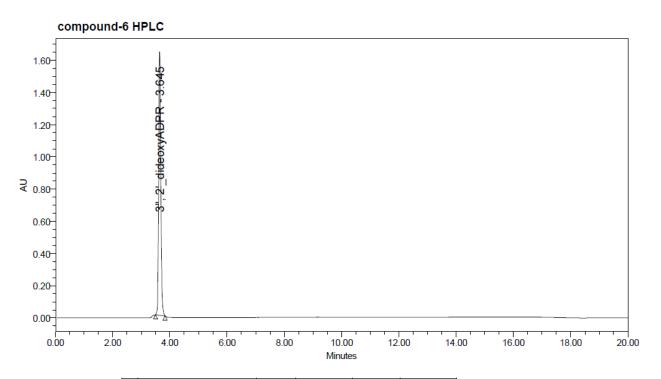
	Peak Name	RT	Area	% Area	Height
1	2"-deoxyADPR	2.933	7595191	88.97	536539
2	baseline bump	4.700	941594	11.03	15903



	Peak Name	RT	Area	% Area	Height	
1	compound-4	2.815	4332210	100.00	645567	

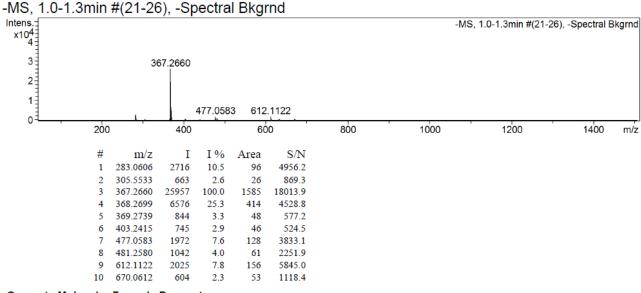


	Peak Name	RT	Area	% Area	Height
1	compound-5	3.325	16556203	100.00	1090063



	Peak Name	RT	Area	% Area	Height
1	lmp_1	2.996			
2	2d_AMP	3.282			
3	compound-6	3.645	8580452	100.00	1642392
4	Bis_2d_AMP	4.044			
5	2d_AMP_imidazolide	4.346			

#### compound-1 MS

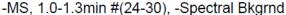


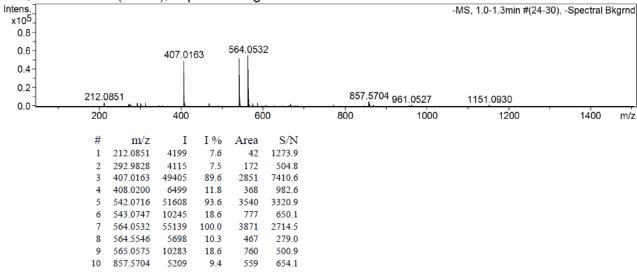
#### Generate Molecular Formula Parameters

Char	ge Toleran	ce SearchR	adius H/C	Ratio min.	H/C Ratio max.	Electron Conf.	Nitrogen Rule	sigma limit	
negat	tive 10 pp	om 0.0	)5 m/z	0	3	both	true	0.05	
Expected Formula C19 H29 N5 O14 P2 Adduct(s): H, Na									
#	meas. m/z	theo. m/z	Err[ppm]	Sigma	Form	nula			
1	612.1122	612.110799	-1.40	0.0278	C 19 H 28 N 5 O 14	P 2			

Note: Sigma fits < 0.05 indicates high probability of correct MF, and mass accuracy of 5ppm or better is generally acceptable for publication

#### compound-2 MS





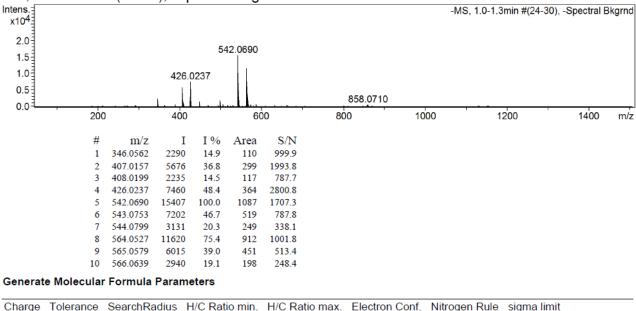
#### Generate Molecular Formula Parameters

Cha	irge Toleran	ce SearchR	adius H/C l	Ratio min.	H/C Ratio max.	Electron Conf.	Nitrogen Rule	sigma limit	
nega	ative 10 pp	om 0.0	05 m/z	0	3	both	true	0.05	
Exp	ected Formu	la C15 H	123 N5 O13 I	P2		Adduct(s):	: H, Na		
#	meas. m/z	theo. m/z	Err[ppm]	Sigma	Form	nula			
1	542.0716	542.068934	-4.00	0.0066	C 15 H 22 N 5 O 13	P2			

Note: Sigma fits < 0.05 indicates high probability of correct MF, and mass accuracy of 5ppm or better is generally acceptable for publication

#### compound-3 MS

#### -MS, 1.0-1.3min #(24-30), -Spectral Bkgrnd

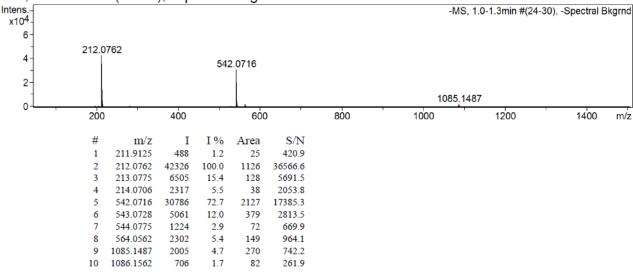


Cha	rge Toleran	ce SearchR	adius H/C l	Ratio min.	H/C Ratio max.	Electron Conf.	Nitrogen Rule	sigma limit	
nega	itive 10 pp	om 0.0	5 m/z	0	3	both	true	0.05	
Exp	ected Formu	la C15 H	23 N5 O13 I	P2		Adduct(s)	: H, Na		
#	meas. m/z	theo. m/z	Err[ppm]	Sigma		Formula			
-	566 0639	566 066529	5 60		C 15 H 23 N 5 Na 1				

Note: Sigma fits < 0.05 indicates high probability of correct MF, and mass accuracy of 5ppm or better is generally acceptable for publication

#### compound-4 MS

-MS, 1.0-1.3min #(24-30), -Spectral Bkgrnd



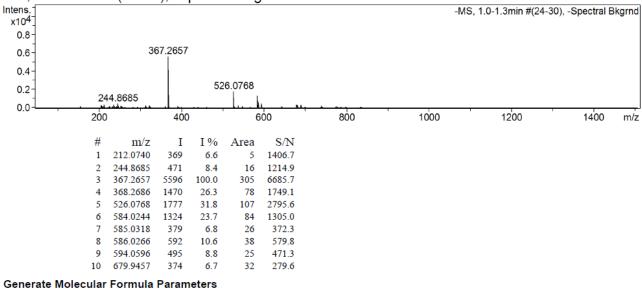
#### Generate Molecular Formula Parameters

Charge	Tolerance	e SearchRa	adius H/C	Ratio min.	H/C Ratio max.	Electron Conf.	Nitrogen Rule	sigma limit	
negative	10 ppr	n 0.0	5 m/z	0	3	both	true	0.05	
Expecte	ed Formula	C15 H	23 N5 O13	P2		Adduct(s)	: H, Na		
# me	eas. m/z	theo. m/z	Err[ppm]	Sigma	Forr	nula			
-	E 40 0740	542.068934	-3.90	0.0400	C 15 H 22 N 5 O 13	- D O			

Note: Sigma fits < 0.05 indicates high probability of correct MF, and mass accuracy of 5ppm or better is generally acceptable for publication

#### compound-5 MS

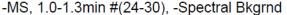
#### -MS, 1.0-1.3min #(24-30), -Spectral Bkgrnd

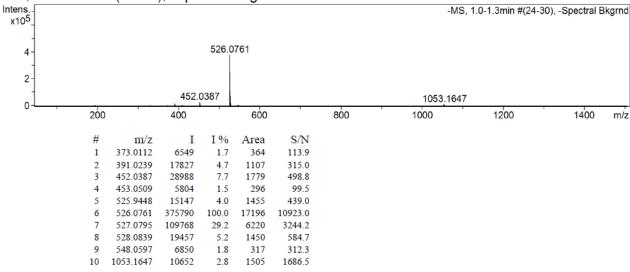


Charg	e Toleranc	e SearchRa	dius H/C l	Ratio min.	H/C Ratio max.	Electron Conf.	Nitrogen Rule	sigma limit	
negativ	е 10 ррг	m 0.05	5 m/z	0	3	both	true	0.05	
Expec	ted Formul	a C15 H2	23 N5 O12 F	2		Adduct(s)	: H, Na		
# n	neas. m/z	theo. m/z	Err[ppm]	Sigma	Forn	nula			
1	526.0768	526 074019	-4 20	0.0397	C 15 H 22 N 5 O 12	P2			

Note: Sigma fits < 0.05 indicates high probability of correct MF, and mass accuracy of 5ppm or better is generally acceptable for publication

#### compound-6 MS





#### Generate Molecular Formula Parameters

Char	ge Toleran	ce SearchF	Radius H	I/C Ratio min.	H/C Ratio max.	Electron Conf.	Nitrogen Rule	sigma limit
negat	ive 10 pp	om 0.	.05 m/z	0	3	both	true	0.05
Expe	cted Formu	la C15 l	H23 N5 O	12 P2		Adduct(s)	: H, Na	
	meac m/z	theo m/z	Errinna	n] Sigma	Formula			

Note: Sigma fits < 0.05 indicates high probability of correct MF, and mass accuracy of 5ppm or better is generally acceptable for publication

#### **References:**

- (1) Ko, H.; Das, A.; Carter, R. L.; Fricks, I. P.; Zhou, Y.; Ivanov, A. A.; Melman, A.; Joshi, B. V.; Kováč, P.; Hajduch, J.; Kirk, K. L.; Harden, T. K.; Jacobson, K. A. Molecular Recognition in the P2Y14receptor: Probing the Structurally Permissive Terminal Sugar Moiety of Uridine-5'-Diphosphoglucose. *Bioorganic Med. Chem.* **2009**, *17* (14), 5298–5311.
- (2) Dabrowski-Tumanski, P.; Kowalska, J.; Jemielity, J. Efficient and Rapid Synthesis of Nucleoside Diphosphate Sugars from Nucleoside Phosphorimidazolides. *European J. Org. Chem.* **2013**, 2013 (11), 2147–2154.