

Supplemental Information

Circadian Clock Regulation of Hepatic Lipid

Metabolism by Modulation of m⁶A mRNA Methylation

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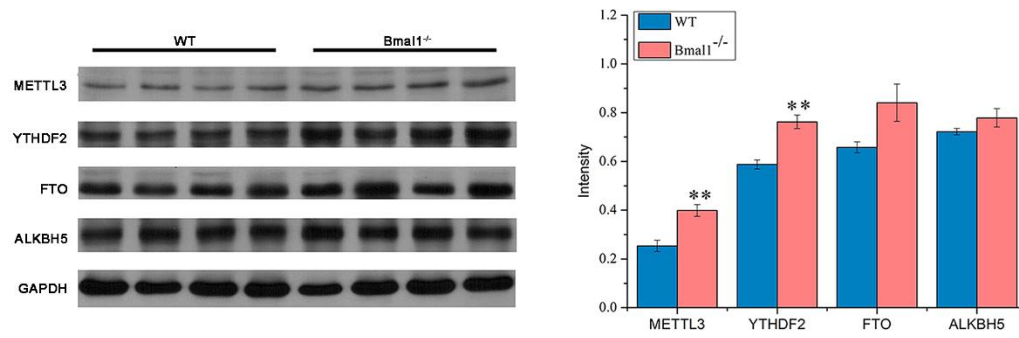


Figure S1. Liver-specific *Bmal1*^{f/f}-AlbCre-knockout affects m⁶A mRNA methylation in mice.

Related to Figure 1. Western blotting was used to measure METTL3, YTHDF2, ALKBH5, and FTO protein levels in the liver of WT and *Bmal1*^{-/-} mice at ZT0 ($n = 4$ per group). WT: *Bmal1*^{f/f}; *Bmal1*^{-/-}: *Bmal1*^{f/f}-AlbCre. All data are representative of three independent experiments. Data are represented as mean \pm SEM. * $P < 0.05$, ** $P < 0.01$ by two-tailed t -test.

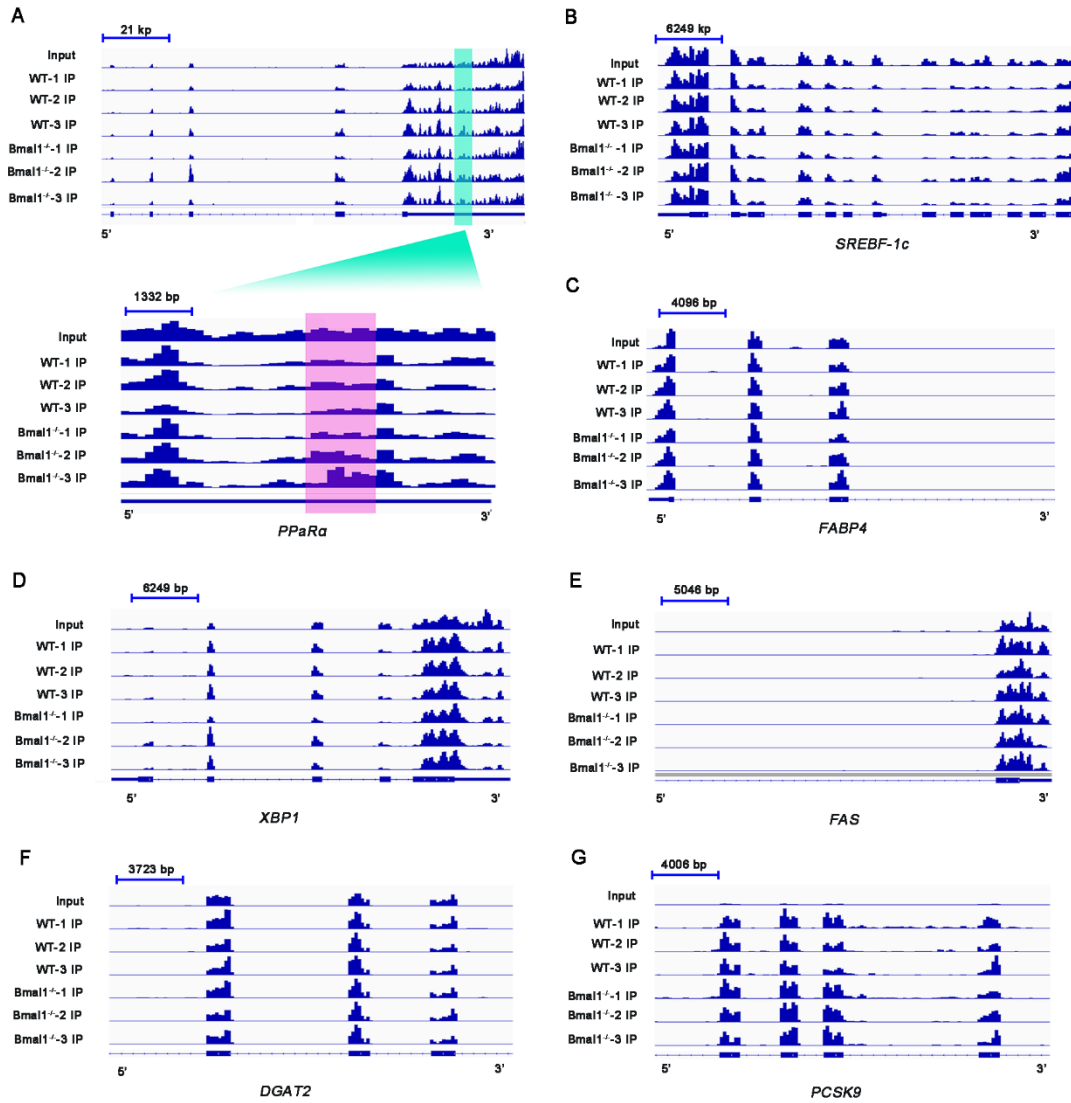


Figure S2. Liver-specific Bmal1^{f/f}-AlbCre-knockout changes *PPaRa* mRNA methylation in mice.

Related to Figure 2. (A-G) Integrative genomics viewer (IGV) plots showing a increase in m⁶A methylated peaks for *PPaRa* mRNA, but no change for other lipid metabolism genes among the input samples, WT IP, and *Bmal1*^{-/-} IP. Blue boxes represent exons ($n = 3$ per group). WT: Bmal^{f/f}, Bmal^{-/-}: Bmal^{f/f}-AlbCre.

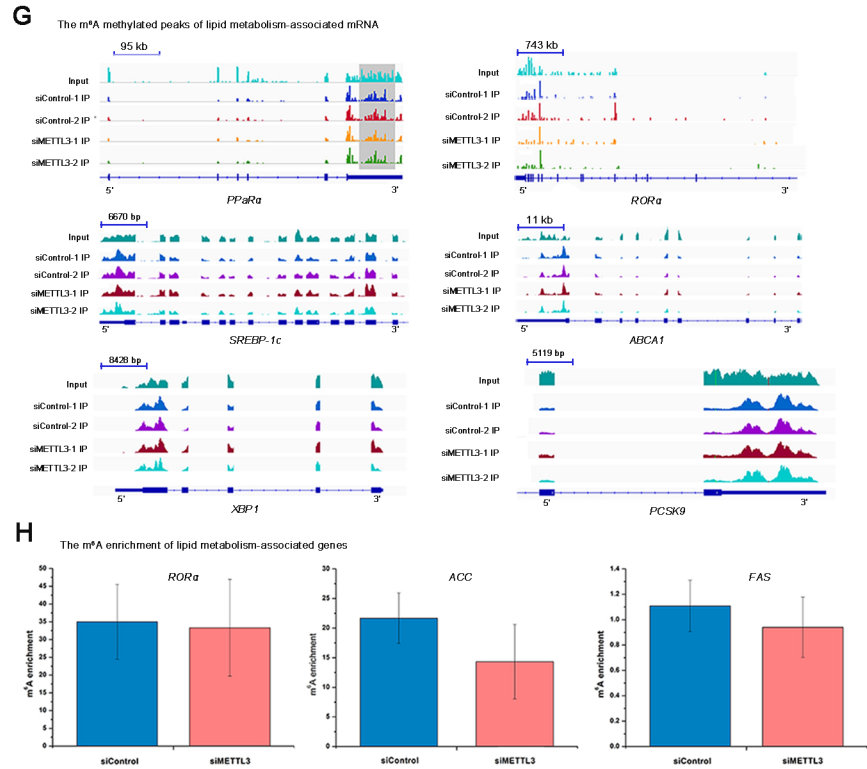
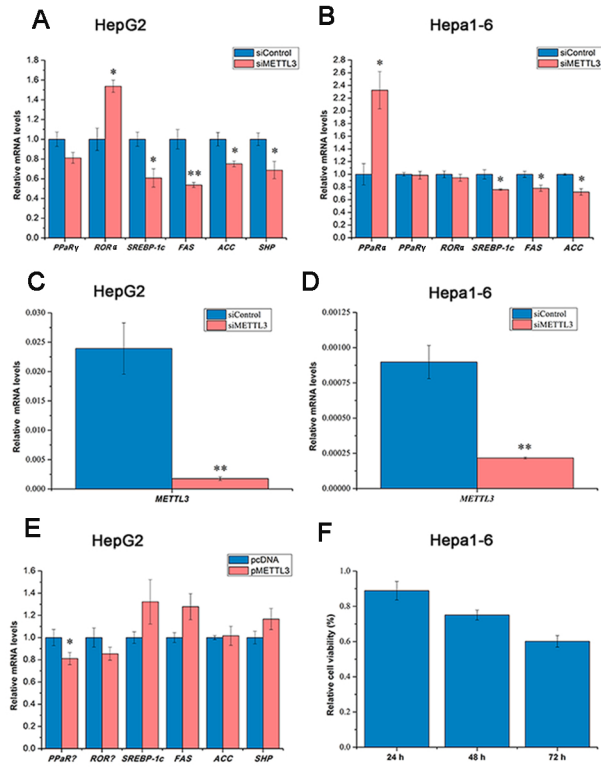


Figure S3. Lipid metabolism gene expression, m⁶A methylated peaks and enrichment following knockdown or overexpression of *METTL3*. Related to Figure 3. **(A)** Knockdown of *METTL3* changed mRNA expression of genes associated with lipid metabolism. Expression of *RORα* mRNA was significantly increased, however, expression of *SREBP-1c*, *Fas*, *ACC*, *SHP* mRNA were significantly decreased following si*METTL3* versus siControl in HepG2 cells ($n = 3$). **(B)** *METTL3* knockdown significantly up-regulated expression of *PPaRα* mRNA and decreased expression of *SREBP-1c*, *Fas*, *ACC* mRNA in Hepa1-6 cells ($n = 3$). **(C-D)** Expression of *METTL3* mRNA in HepG2 (C) and Hepa1-6 (D) cells after transfection of si*METTL3* versus siControl, respectively. The mRNA expression of *METTL3* in HepG2 cells is significantly higher than in Hepa1-6 cells ($n = 3$). **(E)** Lipid metabolism genes expression following overexpression of *METTL3* in HepG2 cells. Difference of *RORα*, *SREBP-1c*, *Fas*, *ACC*, *SHP* mRNA were not observed by overexpression of *METTL3*, with the exception of decreased *PPaRγ* mRNA ($n = 3$). **(F)** Knockdown of *METTL3* in Hepa1-6 cells reduced the relative cell viability determined by MTT at 24, 48, and 72 h after transfection ($n = 6$). **(G)** Integrative genomics viewer (IGV) plots showing increase of *PPaRα* (A) in m⁶A methylated peaks, but no difference for, *RORα* (B), *SREBP-1c* (C), *ABCA1* (D), *XBPI* (E), and *PCSK9* (F) mRNA among the siControl IP and si*METTL3* IP. Blue boxes represent exons and blue lines represent introns. $n = 2$. **(H)** The m⁶A enrichment of *RORα*, *ACC*, and *FAS* mRNA in HepG2 cells at 48 h post-transfection ($n = 3$). Data are representative of three independent experiments **(A-F, H)**. Data are represented as mean \pm SEM. * $P < 0.05$, ** $P < 0.01$ by two-tailed t -test.

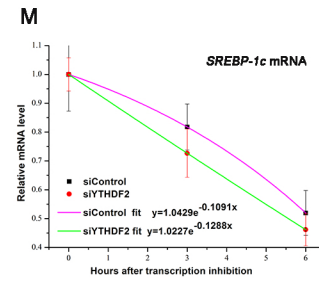
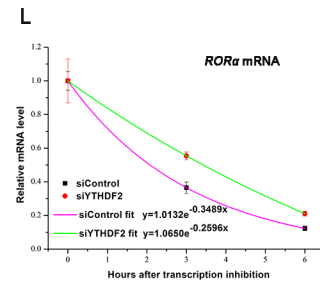
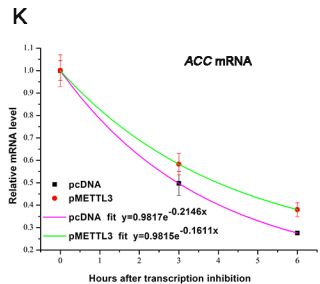
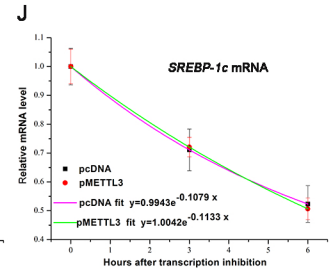
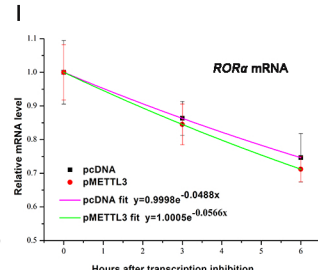
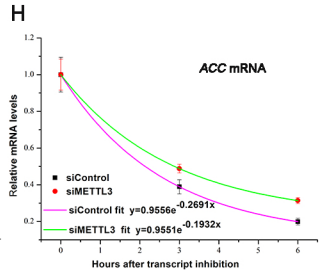
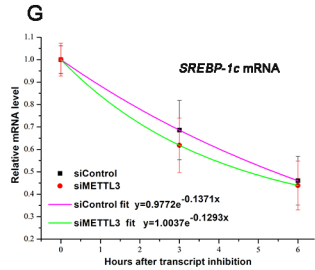
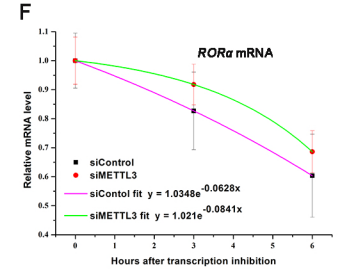
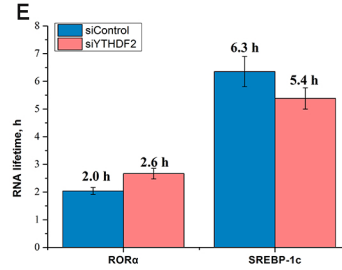
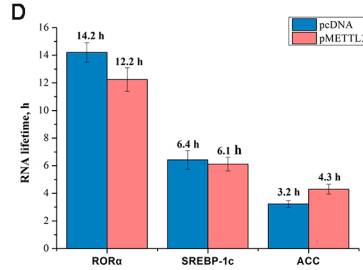
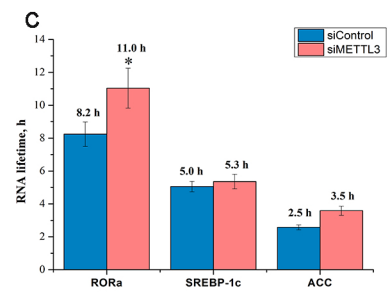
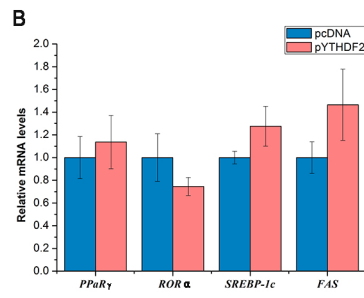
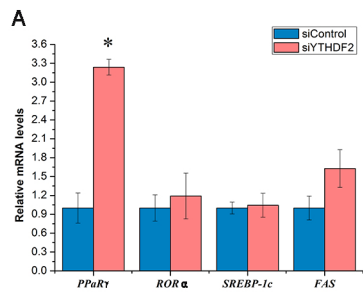


Figure S4 Lifetimes of nascent mRNAs in HepG2 cells. Related to Figure 4. **(A)** Effect of YTHDF2 knockdown on mRNA expression of genes associated with lipid metabolism in HepG2 cells. YTHDF2 knockdown did not affect *RORα*, *SREBP-1c*, and *Fas* mRNA expression, but increased expression of *PPaRγ* mRNA in HepG2 cells ($n = 3$). **(B)** Expression of lipid metabolism genes in the sample of pYTHDF2 versus pcDNA in HepG2 cells. We did not observe any differences in *PPaRγ*, *RORα*, *SREBP-1c*, and *Fas* mRNA expression between pYTHDF2 and pcDNA ($n = 3$). **(C-E)** Lifetime of *RORα*, *SREBP-1c*, and *ACC* mRNA in samples following knockdown of *METTL3* **(C)**, overexpression of *METTL3* **(D)**, and knockdown of *YTHDF2* **(E)** in HepG2 cells, respectively. HepG2 cells were treated with actinomycin for 0 h, 3 h, and 6 h before collection. Knockdown of *METTL3* significantly increased the lifetime of *PPaRα* and *RORα* mRNA, but had no impact on the lifetime of *SREBP-1c* and *ACC* mRNA in HepG2 cells. No differences were observed for the lifetime of *RORα*, *SREBP-1c*, and *ACC* mRNA following knockdown of YTHDF2, or overexpression of *METTL3* and YTHDF2 ($n = 3$). **(F-M)** Decay of *RORα*, *SREBP-1c*, and *ACC* mRNA in samples following knockdown of *METTL3*, overexpression of *METTL3*, or knockdown of *YTHDF2* in HepG2 cells, respectively ($n = 3$). All data are representative of three independent experiments. Data are represented as mean \pm SEM. * $P < 0.05$, ** $P < 0.01$ by two-tailed t -test.

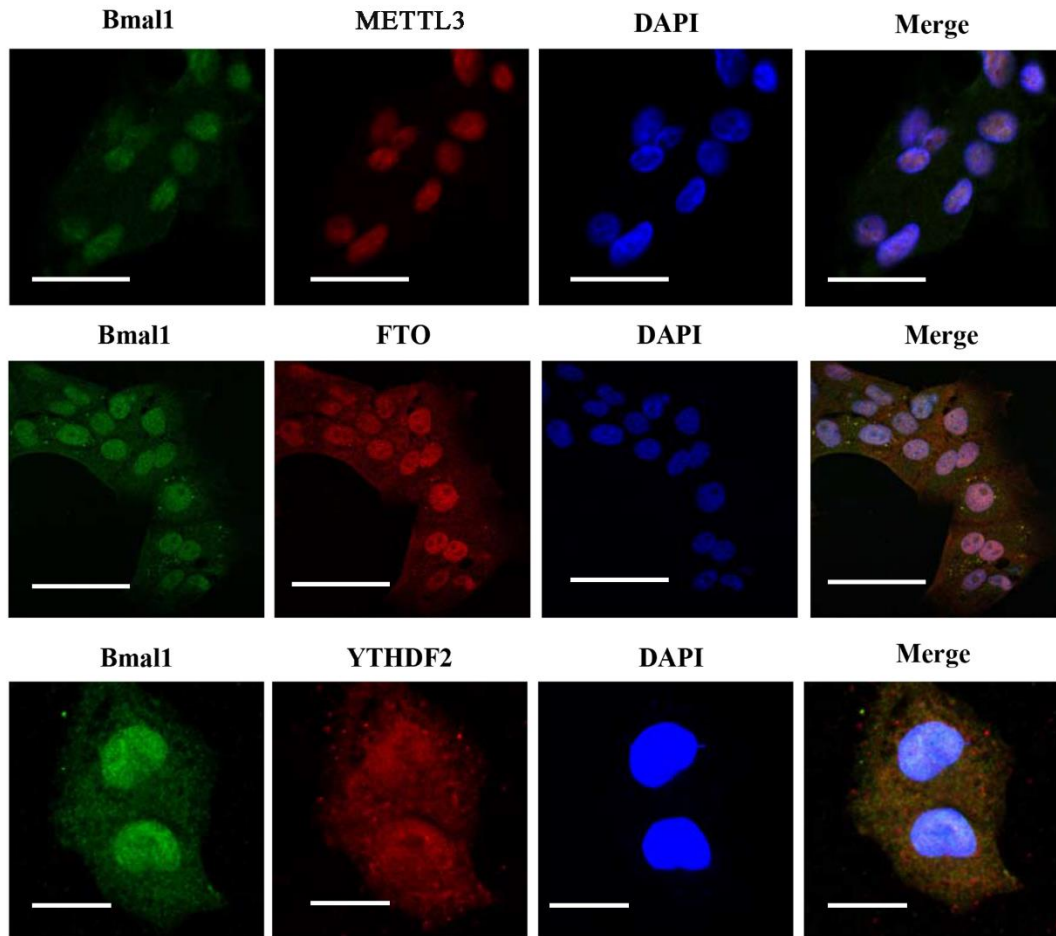


Figure S5. The distribution of METTL3, YTHDF2, FTO, and BMAL1 protein. Related to Figure 6.

The distribution of METTL3, YTHDF2, FTO, and BMAL1 proteins were examined by fluorescence immunostaining in HepG2 cells, indicating that METTL3, FTO, and BMAL1 were localized to the nucleus. Conversely, YTHDF2 was localized in both the nucleus and cytoplasm. Scale bar, 70 μm . All data are representative of three independent experiments.

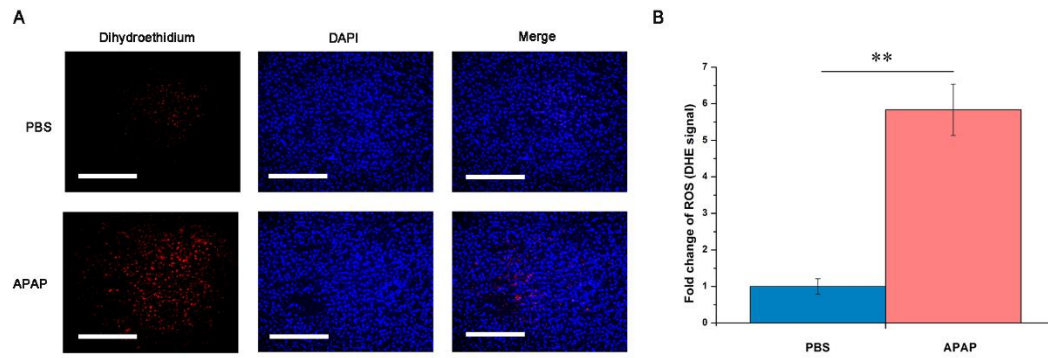


Figure S6. Detection of ROS in mouse liver by dihydroethidium. Related to Figure 6. **(A)** Representative images of ROS detection from dihydroethidium (DHE) stained liver sections from WT mice after acetaminophen (APAP) or PBS injection. Scale bar, 160 μm . **(B)** Quantification of ROS formation was achieved by measuring mean grey values of DHE signal per cell using ImageJ software, $n = 3$ per treatment. All data are representative of three independent experiments. Data are represented as mean \pm SEM. $*P < 0.05$, $**P < 0.01$ by two-tailed t -test.

Table S1. RNA methylation gene and m⁶A abundance CircWave cosinor statistical output. Related to Figure 1

Genotype	Gene	F-statistic	P-value	R ²
M⁶A writer, eraser, reader genes				
WT	<i>METTL3</i>	NA	NA	0
WT	<i>METTL14</i>	12.9429	0.00017	0.52952
WT	<i>FTO</i>	10.4670	0.00059	0.47649
WT	<i>ALKBH5</i>	7.55569	0.00300	0.39651
WT	<i>YTHDF2</i>	5.47771	0.01133	0.32264
WT	m ⁶ A	8.11851	0.00408	0.51980
<i>Bmal1</i> ^{-/-}	<i>METTL3</i>	3.47356	0.04545	0.20465
<i>Bmal1</i> ^{-/-}	<i>METTL14</i>	9.43082	0.00083	0.42044
<i>Bmal1</i> ^{-/-}	<i>FTO</i>	5.28065	0.01160	0.28118
<i>Bmal1</i> ^{-/-}	<i>ALKBH5</i>	NA	NA	0
<i>Bmal1</i> ^{-/-}	<i>YTHDF2</i>	NA	NA	0
<i>Bmal1</i> ^{-/-}	m ⁶ A	NA	NA	0
Lipid metabolism genes				
WT	<i>PPaRα</i>	26.92400	9X10 ⁻⁷	0.70071
WT	<i>PPARγ</i>	5.88272	0.00864	0.33842
WT	<i>SHP</i>	6.38064	0.00625	0.35685
WT	<i>RORα</i>	6.58642	0.00134	0.55645
WT	<i>ACC</i>	NA	NA	0
WT	<i>FAS</i>	NA	NA	0
WT	<i>FABP4</i>	NA	NA	0
WT	<i>FABP1</i>	5.98018	0.0081	0.34211
WT	<i>SREBP-1c</i>	5.18988	0.0138	0.31096
<i>Bmal1</i> ^{-/-}	<i>PPARα</i>	10.8668	0.00034	0.44597
<i>Bmal1</i> ^{-/-}	<i>PPARγ</i>	5.28796	0.01154	0.28145
<i>Bmal1</i> ^{-/-}	<i>SHP</i>	5.30759	0.01138	0.2822
<i>Bmal1</i> ^{-/-}	<i>RORα</i>	NA	NA	0
<i>Bmal1</i> ^{-/-}	<i>ACC</i>	3.6833	0.03851	0.21435
<i>Bmal1</i> ^{-/-}	<i>FAS</i>	18.9886	7.1X10 ⁻⁶	0.58447
<i>Bmal1</i> ^{-/-}	<i>FABP4</i>	NA	NA	0
<i>Bmal1</i> ^{-/-}	<i>FABP1</i>	NA	NA	0
<i>Bmal1</i> ^{-/-}	<i>SREBP-1c</i>	NA	NA	0

NA: Not available, No significant co-sinor pattern detected

Table S2. Primers for qRT-PCR. Related to KEY RESOURCES TABLE.

Gene	Forward	Reverse	Accession Number
Mouse			
<i>METTL3</i>	AGCAGAGCAAGAGACGAATTATC	GGTGGAAAGAGTCGATCAGCA	AF135789
<i>METTL14</i>	CTGAGAGTGCGGATAGCATTG	GAGCAGATGTATCATAGGAAGCC	NM_201638.2
<i>ALKBH5</i>	CGCGGTCATCAACGACTACC	ATGGGCTTGAAGTGAAGTTG	XM_021176347.1
<i>YTHDF2</i>	GAGCAGAGACCAAAAGGTCAAG	CTGTGGGCTCAAGTAAGTTTC	XM_021160169.1
<i>FTO</i>	TTCATGCTGGATGACCTCAATG	GCCAACTGACAGCGTTCTAAG	NM_011936.2
<i>PPARα</i>	TGCAAACCTTGGACTTGAACG	AGGAGGACAGCATCGTGAAG	XM_021183376.1
<i>PPARδ</i>	CAAGTGGGGTCAGTCATGGAA	GCTGGAAGGAAGCGTGTGTT	U01665
<i>PPARγ</i>	CTGACAGGACTGTGTGAC	TCTGTGTCAACCATGGTAAT	XM_021164280.1
<i>RORα</i>	ACGCCACCTACAACATCTC	TGCCCATCCATATAGGTGCT	XM_021171874.1
<i>SREBP-1c</i>	GGAGCCATGGATTGCACATT	GGCCCGGGAAGTCACTGT	XM_006532716.2
<i>ACC</i>	GCCTCCGTCAGCTCAGATAC	ATGTGAAAGGCCAAACCATC	XM_021178352.1
<i>FAS</i>	ATCCCAGCACTTCTTGATGG	CCGAAGCCAAATGAGTTGAT	XM_021177126.1
<i>FABP1</i>	CATCCAGAAAGGGAAGGACA	CTTCCCTTCATGCACGATTT	NM_017399.4
<i>FABP4</i>	CTTTGCCACAAGGAAAGTGG	TCCCCATTTACGCTGATGAT	XM_006530048.3
<i>FATP4</i>	ACTGTTCTCCAAGCTAGTGCT	GATGAAGACCCGGATGAAACG	NM_011989.4
<i>SHP</i>	TGGGTCCCAAGGAGTATGC	GCTCCAAGACTTCACACAGTG	NM_011850.3
<i>GAPDH</i>	GGCAAATTCAACGGCACAGT	AGATGGTGATGGGCTTCCC	NM_001289726.1
<i>HPRT1</i>	TCAGTCAACGGGGGACATAAA	GGGGCTGTACTGCTTAACCAG	NM_013556.2
Human			
<i>METTL3</i>	CAAGCTGCACTTCAGACGAA	GCTTGGCGTGTGGTCTTT	NM_019852.4
Continued			

<i>METTL14</i>	AGAAACTTGCAGGGCTTCCT	TCTTCTTCATATGGCAAATTTTCTT	NM_020961.3
<i>ALKBH5</i>	CGGCGAAGGCTACACTTACG	CCACCAGCTTTTGGATCACCA	NM_017758.3
<i>YTHDF1</i>	ACCTGTCCAGCTATTACCCG	TGGTGAGGTATGGAATCGGAG	NM_017798.3
<i>YTHDF2</i>	CCTTAGGTGGAGCCATGATTG	TCTGTGCTACCCAACTTCAGT	NM_001172828.1
<i>FTO</i>	ACTTGGCTCCCTTATCTGACC	GTGCAGTGTGAGAAAGGCTT	NM_001080432.2
<i>PPARα</i>	GGATGCTGGTAGCGTATGGA	GGACGATCTCCACAGCAAAT	NM_005036.4
<i>PPARδ</i>	TCTACAATGCCTACCTGAAAACTTC	ACAATGTCTCGATGTCGTGGATC	NM_001171820.1
<i>RORα</i>	ACTCCTGTCCTCGTCAGAAGA	CATCCCTACGGCAAGGCATTT	NM_002943.3
<i>SREBP-1c</i>	CGCCACCTGCCCCCTCTCCTTCC	TGCCCTGCCACCTATCCTCTCACG	NM_004599.3
<i>ACC</i>	ATGTCTGGCTTGCACCTAGTA	CCCCAAAGCGAGTAACAAATTCT	NM_198836.2
<i>FAS</i>	TCGTGGGCTACAGCATGGT	GCCCTCTGAAGTCGAAGAAGAA	NM_004104.4
<i>FABP4</i>	ACTGGGCCAGGAATTTGACG	CTCGTGGAAGTGACGCCTT	NM_001442.2
<i>FATP4</i>	GTGAAGGCAAAGGTGCGAC	CGGAAGGTCCAGTGGGTATC	NM_005094.3
<i>SHP</i>	GTGCCCAGCATACTCAAGAAG	TGGGGTCTGTCTGGCAGTT	NM_021969.2
<i>GAPDH</i>	CGACCACTTTGTCAAGCTCA	AGGGGAGATTCAGTGTGGTG	NM_001289746.1
<i>HPRT1</i>	TGACACTGGCAAAACAATGCA	GGTCCTTTTCACCAGCAAGCT	NM_000194.2