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Supplementary Materials for

Targeting the NF-KB signaling pathway in chronic tendon disease

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Fig. S1. Gene expression of cultured tendon fibroblasts in response to varying doses of IL-1 β . Fibroblasts were isolated from tail tendons from wildtype (WT, n = 10), tendon-specific IKK β knockout (IKK β KO^{Scx}, n = 6), and constitutively active IKK β (IKK β CA^{Scx}, n = 5) mice. Gene expression is normalized to *GAPDH*. Data are shown as mean \pm SD. Statistically significant differences were calculated using two-way ANOVA (genotype, dose) with Tukey's post-hoc test.



Fig. S2. Multiplex ELISA of cultured tendon fibroblasts in response to 10 ng IL-1 β over the 72-hour period. Fibroblasts were isolated from tail tendons from wildtype (WT, n = 5), tendon-specific IKK β knockout (IKK β KO^{Scx}, n = 5), and constitutively active IKK β (IKK β CA^{Scx}, n = 5) mice. Bars represent mean + SD. Statistically significant differences were calculated using two-way ANOVA (genotype, treatment) with Tukey's post-hoc test. Datasets without reported *P*-values were not compared due to undetectable baseline cytokine concentrations in WT controls.



Fig. S3. H&E- and Toluidine blue–stained section of tendons and tendon entheses of WT, IKK β KO^{Sex}, and IKK β CA^{Sex} mice. Black arrowheads: spindle shaped tendon fibroblasts indicated, white arrowheads: enthesis chondrocytes; metachromasia demonstrating fibrocartilage interface can be seen below the dashed line in Toluidine Blue-stained sections.



Fig. S4. μ CT results for treadmill overuse model. 10-week-old mice were subjected to a chronic overuse protocol with 1 week of progressive training followed by 4 weeks of downhill running. Control mice were permitted normal cage activity. The humeral head was scanned at an energy of 55 kVP, intensity of 145 μ A, and a resolution of 12.3 μ m. Data are shown as mean \pm SD with individual points representing biologically independent samples.



Fig. S5. μ CT results for acute injury and repair model. 10-week-old mice were subjected to a unilateral acute injury of the supraspinatus tendon and immediate repair followed by 2 weeks of recovery. Sham operations were performed on contralateral limbs. The humeral head was scanned at an energy of 55 kVP, intensity of 145 μ A, and a resolution of 12.3 μ m. Data are shown as mean \pm SD with individual points representing biologically independent samples. Statistically significant differences were calculated using One-way ANOVA (genotype, treatment) with Fisher's LSD post-hoc test ** - *P* < 0.01.



Fig. S6. Gene expression of cultured human tendon fibroblasts in response to IL-1 β and IKK β inhibitor. Data are shown as mean \pm SD with individual points representing biologically independent samples. Statistically significant differences were calculated using One-way ANOVA (treatment) with Fisher's LSD post-hoc test.



Fig. S7. Schematic of how IKKβ/NF-κB drives chronic tendinopathy. Schematic illustrating how injury-induced pro-inflammatory cytokines cause tendon fibroblasts to suppress tissue anabolism and increase matrix catabolism and cytokine production. NF-κB signaling within the tendon stromal and immune compartment increases during initial phases of healing. Constitutive activation of IKKβ (IKKβCA^{Scx}) chronically degrades the rotator cuff by synthesizing degenerative enzymes and pro-inflammatory cytokines. Fibroblasts without IKKβ (IKKβKO^{Scx}) remain agnostic to proinflammatory cytokines, maintain matrix production, and keep total NF-κB signaling lower. Dotted lines represent basal expression.

Table S1. Semi-quantitative histological assessment of supraspinatus tendons in WT, IKKβKO^{Scx}, IKKβCA^{Scx} mice. Mast Cells, PMN Acute Inflammatory Cells, Monocytes: # of samples where present.

	WT $(n = 8)$	IKKβKO ^{Sex} ($n = 4$)	IKKβCA ^{Sex} $(n = 4)$
Mast Cells	1 of 8	3 of 4	1 of 4
PMN	0 of 8	0 of 4	0 of 4
Monocytes	8 of 8	4 of 4	4 of 4
Total Inflammatory Cells	1.00 (1,1)	1.00 (1,1)	2.00 (2,2)
Total Cellularity	1.13 (1,2)	1.75 (1,3)	3.00 (3,3)