

1 **Supplemental Material**

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3 **Table S1.** Antibacterial specificity of PyS2-GN4 towards Gram-positive and Gram-negative
4 bacteria.

Gram-Positive			Gram-Negative		
Species	Strain	MIC ($\mu\text{g/ml}$)	Species	Strain	MIC ($\mu\text{g/ml}$)
<i>B. cereus</i>	RSVF1	> 256	<i>A. baumannii</i>	ATCC 17978	> 256
<i>E. faecium</i>	EFSK-2	> 256	<i>E. cloacae</i>	NR-50391	> 256
<i>S. aureus</i>	NR-45946	> 256	<i>E. coli</i>	ATCC 25922	> 256
<i>S. pyogenes</i>	D471	> 256	<i>K. pneumoniae</i>	NR-41916	> 256

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20 **Table S2.** List of bacterial strains used in this study.

Bacteria	Source
<i>A. baumannii</i> ATCC 17978	ATCC
<i>B. cereus</i> RSVF1	Vincent Fischetti, The Rockefeller University
<i>E. cloacae</i> NR-50391	BEI Resources, NIAID, NIH
<i>E. coli</i> ATCC 25922	ATCC
<i>E. faecium</i> EFSK-2	Alexander Tomasz, The Rockefeller University
<i>K. pneumoniae</i> NR-41916	BEI Resources, NIAID, NIH
<i>P. aeruginosa</i> ATCC 15692	ATCC
<i>P. aeruginosa</i> 442	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 443	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 445	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 446	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 448	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 449	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 450	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 451	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 452	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> 453	Lars Westblade, Weill Cornell Medical College
<i>P. aeruginosa</i> MDR-M-3	Daniel Green, Columbia University Medical Center
<i>S. aureus</i> NR-45946	BEI Resources, NIAID, NIH
<i>S. pyogenes</i> D471	The Rockefeller University Lancefield Collection

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22 ATCC, American Type Culture Collection; NIAID, National Institute of Allergy and Infectious
23 Disease; NIH, National Institutes of Health

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29 **Table S3.** List of primers used in this study.

Oligonucleotide	Nucleotide Sequence
GN4_F	5'-AACTTAAAGAAGGAGATATAATGCGCACCAGCCAGCGC-3'
GN4_R	5'-GTCGACGGAGCTGAATTGGATCCTTAGCTCAGCGGTTCCAGAACAGTCG-3'
PyS2_F	5'-AACTTAAAGAAGGAGATACCATGGCCGTGAACGATTATG-3'
PyS2-GN4_R	5'-TGGTGCATCGGGTCACGAAACATCAC-3'
PyS2-GN4_F	5'-TCGTGACCCGATGCGCACCAAGCCAGCGC-3'
PyS2-GN4ΔTBB_F	5'-[Phos]GTTCAGGGTGGTGGTCGTGACATTATCCAG
PyS2-GN4ΔTBB_R	5'-[Phos]GCTGCCGGCTATAATCGTCACGCCATG
PyS2-GN4_KO_1F	5'-[Phos]CGCCTATCAGGCTAGCGTGGGTGTGGACC-3'
PyS2-GN4_KO_1R	5'-[Phos]CTCAGGCGCAGGCCCTAAAGCTCTTAATC-3'
PyS2-GN4_KO_2F	5'-[Phos]GCGTGGGTGTGGGCCATTGGTTATGGTAC-3'
PyS2-GN4_KO_2R	5'-[Phos]TAGCCTGATAGGCCTCAGGCGCAGGCC-3'
FpvAI_F	5'-CGAAGGCCAGAACTACGAGA-3'
FpvAI_R	5'-TGTAGCTGGTAGAGGCTCAA-3'
FpvAII_F	5'-TACCTCGACGGCCTGCACAT-3'
FpvAII_R	5'-GAAGGTGAATGGCTTGCGTA-3'
FpvAIII_F	5'-ACTGGGACAAGATCCAAGAGAC-3'
FpvAIII_R	5'-CTGGTAGGACGAAATGCGAG-3'

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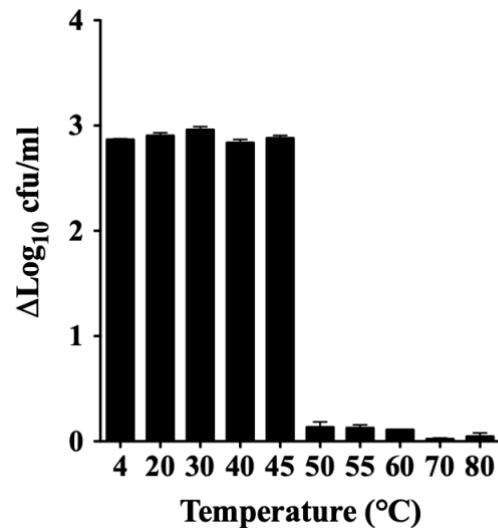
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42 **Figure S1.** Lysocin thermal stability. PyS2-GN4 was incubated in PBS for 30 min at
43 temperatures ranging from 4°C to 80°C. After cooling on ice, each sample at 50 µg/ml was
44 incubated statically with *P. aeruginosa* strain 453 for a total of 4 h at 37°C. The average residual
45 antipseudomonal activity of each sample was equated to the log₁₀ decrease in viable bacterial
46 cells when compared to the untreated control. All error bars correspond to ± SEM of triplicate
47 experiments.

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57 **SI Methods**58 ***Determining Thermal Stability***

59 Using an EchoTherm™ bench-top incubator (Torrey Pines Scientific), PyS2-GN4 was initially

60 incubated at 1 mg/ml in PBS, pH 7.4, for 30 min at temperatures ranging from 4°C to 80°C.

61 Following temperature treatment, the lysocin was immediately cooled on ice. Residual

62 antipseudomonal activity was determined by incubating each lysocin sample at 50 µg/ml with *P.*63 *aeruginosa* strain 453 at 10⁶ cfu/ml in CAA medium with EDDHA for a total of 4 h at 37°C. The

64 samples were plated on CAA agar in order to calculate viable cell counts. A control absent of

65 lysocin was included.

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