SUPPORTING INFORMATION

Table S1. Genes associated with hyperammonemia

Square brackets enclose protein function and disease, with the disease underlined.

Urea cycle ger	
ALDH18A1	aldehyde dehydrogenase 18 member 1 [ornithine, arginine, proline biosynthesis, <u>cutis laxa type IIIA]</u>
ARG1	arginase, liver [argininemia]
ASS1	argininosuccinate synthase 1 [citrullinemia type I]
ASL	argininosuccinate lyase [argininosuccinic acidemia]
CPS1	carbamovl phosphate synthase 1. mitochondrial
GLUI	glutamate-ammonia ligase [synthesis of glutamine from glutamate_congenital glutamine deficiency]
NAGS	N-acetylalutamate synthese
ORNT1	SI C25415 solute carrier family 25 (mitochondrial carrier: ornithine transporter) member 15
	buoranithinamia buoranmonomia bamoatrullinuria (HHI) audormal
	[hyperofiniting family 25 (mitchendrial corrier annihiling fragments) member 2
	SLC25A2, solute cameria family 25 (mitochondria) camer, omitime transporter) member 2
ORN 13	SLC25A29, solute camer family 25 (mitochondnai camitine/acyicamitine camer protein CACE) member 29
	ornithine transcarbamylase
SLC/A/	solute carrier family / (cationic amino acid transporter, y+ system) member / [Arg, Lys, ornithine transport
	in kidney and small intestine, lysinuric protein intolerance]
SLC25A13	solute carrier family 25 member 13 (citrin) [exchange of Asp for Glu across inner mitochondrial membrane,
	citrullinemia type II]
Fatty acid oxid	ation genes
ACÁDVL	acyl-CoA dehydrogenase, very long chain
ACADM	acvI-CoA dehvdrogenase. C-4 to C-12 straight chain
CPT1A	carnitine palmitovitransferase 1A (liver)
CPT2	carnitine palmitovitransferase 2
ETEA	electron-transfer-flavonrotein, alpha polypentide [dutaric acidemia []]
	election transfer flavoprotein, beta polyapetide [alutaric acidemia IIP]
	election-transferring flowen ten downer and a contraction downer and a
	budrownowi (CoA dobudrogonova) (CoA the logonova coa budrownia mc)
	hydroxyacyl-CoA denydrogenase/3-ketoacyl-CoA thiolase/enoyl-CoA hydratase, alpha subunit
HADHB	nydroxyacyi-CoA denydrogenase/3-ketoacyi-CoA thiolase/enoyi-CoA nydratase, beta subunit
HLCS	nolocarboxylase synthase (biotin- (propionyl-CoA-carboxylase (ATP-hydrolysing)) ligase)
	[gluconeogenesis, branched chain amino acid catabolism]
HMGCL	3-hydroxymethyl-3-methylglutaryl-CoA lyase [final step in leucine degradation]
IVD	isovaleryl-CoA dehydrogenase [valine, leucine, isoleucine degradation, isovaleric acidemia]
LMBRD1	LMBR1 domain containing 1 [cobalamin transporter, homocystinuria-megaloblastic anemia type F]
MCCC1	methylcrotonoyl-CoA carboxylase 1 (alpha) [leucine catabolism]
MCCC2	methylcrotonoyl-CoA carboxylase 2 (beta) [leucine catabolism]
MLYCD	malonyl-CoA-decarboxylase [stimulates fatty acid oxidation by converting malonyl-CoA to acetyl-CoA,
	[combined malonic and methylmalonic acidemia]
MMAA	methylmalonic acidemia (cobalamin deficiency) cblA type [methylmalonic acidemia]
MMAB	methylmalonic acidemia (cobalamin deficiency) cblB type [methylmalonic acidemia]
MMACHC	methylmalonic acidemia (cobalamin deficiency) cblC type with homocystinuria [methylmalonic acidemia]
MMADHC	methylmalonic acidemia (cobalamin deficiency) cblD type, with homocystinuria (methylmalonic acidemia)
MUT	methylmalonyl CoA mutase lisomerization of methylmalonyl-CoA to succinyl-CoA methylmalonic
	acidemial
PCCA	aciucinaj
PCCA	propional CoA carboxylase, alpha polypepide <u>[propionic acidemia]</u>
	propionty COA caliboxylase, beta polypeptide <u>[propionic acidemia]</u>
SLCZSAZU	solute carrier family 25 member 20 [Carrinne/acyclamine translocase denciency]
SLCZZAS	solute carrier family 22 (organic cation/carritine transporter) member 5 [carritine deficiency]
Other anaplero	<u>)sis genes</u>
DLAI	anyaronpoamide S-acetyltransterase [in mitochondrial complex that converts pyruvate to acetyl-CoA]
GLUD1	glutamate dehydrogenase 1 [hyperinsulinism-hyperammonemia syndrome]
PC	pyruvate carboxylase [mitochondrial pyruvate oxidation to oxaloacetate]
PDHA1	pyruvate dehydrogenase (lipoamide) alpha 1 [in mitochondrial complex that converts pyruvate to acetyl-
	CoA]
TUFM	Tu translation elongation factor, mitochondrial [protein translation in mitochondria, combined oxidative
	phosphorylation deficiency]

Table S2. Patient missense and splice site mutations

For the 43 hyperammonemia genes plus DPYD, we obtained allele frequencies and disease associations from the SNP database <u>https://www.ncbi.nlm.nih.gov/snp</u>. Abbreviations: NA, not applicable; NV, normal variant based on SIFT and PolyPhen2 predictions and high allele frequency; Ref DNA, reference DNA sequence; SD, splice donor site.

Gene	SNP ID	Ref DNA	Patient DNA	Mutation	SIFT	PolyPhen2	Allele freq	Conclusion
CPS1	rs1047891	С	C/A	T1406N	tolerated	benign	0.288	NV ^A
ORNT1	rs17849654	А	A/T	1254L	tolerated	benign	0.323	NV
ORNT2	rs3749780	G	G/A	V226I	tolerated	benign	0.128	NV
	rs10075302	G	G/T	G159C	damaging	possibly damaging	0.043	damaging ^B
MMAB	rs9593	Т	T/A	M239K	tolerated	benign	0.487	NV
LMBRD1	rs12648	А	A/T	D469E	tolerated	benign	0.385	NV
HLCS	rs61732502	G	G/A	V96I	tolerated	benign	0.058	NV
CPT2	rs1799821	G	G/A	V368I	tolerated	benign	0.413	NV
	rs776754218	С	C/T	P520L	tolerated	benign	0.00005	benign ^C
MUT	rs8589	А	A/G	I671V	tolerated	benign	0.451	NV
	rs2229385	G	G/A	A499T	tolerated	benign	0.086	benign ^D
ETFA	rs1801591	С	C/T	T171I	damaging	damaging	0.051	damaging ^E
ETFDH	rs11559290	С	C/T	T31I	tolerated	benign	0.327	NV
SLC7A7	rs8018462	А	G/G	SD-2	NA	NA	0.418	benign ^F
DPYD	rs1801159	А	A/G	1543V	tolerated	benign	0.185	NV^{G}
	rs1801265	Т	C/C	C29R	tolerated	benign	0.260	NV^{G}

Notes

A T1406N has no effect on plasma arginine concentrations¹.

B G159C shows decreased activity in cells transfected with a cDNA expression vector².

C P520L is predicted to preserve protein function and is not among the 64 mutations found in neonatal or severe infantile carnitine palmitoyltransferase II deficiency³.

D A499T confers normal enzymatic activity⁴.

E T171I affects thermal stability of the ETF enzyme and is over-represented among patients with very-long-chain acyl-CoA dehydrogenase deficiency⁵.

F The splice donor SD-2 polymorphism had no effect on RNA, as determined by RNA-sequence analysis of 12 acute myelogenous leukemias (Fig. 2E).

G Patient had normal enzymatic activity (Fig. 2B).

Table S3. Patient nonsense mutations

A nonsense mutation was deemed homozygous if we observed multiple homozygous reads at both the mutation site and adjacent SNPs (in parentheses). Other nonsense mutations were heterozygous.

Gene Mutation reads Function Function	
PDE4DIP p.R785X phosphodiesterase 4D interacting protein anchors cAMP PDE4D to carboxypeptidase N subunit 2	Golgi
SLC6A18 p.Y319X solute carrier family 6 member 18 Na-dependent neutral a.a. tra	insporter,
ZNF117 p.R428X zinc finger protein 117 possible transcriptional reg	gulator
FSCN3 p.R423X fascin-3 actin bundling protei	ר
LPL p.S474X lipoprotein lipase precursor triglyceride hydrolysi	S
VPS13B p.Y413X vacuolar protein sorting-assoc protein 13B vesicle-mediated trans	port
IFNE p.Q71X interferon epsilon precursor interferon	
CTBP2 p.Q445X C-terminal-binding protein 2 corepressor of transcriptional	regulators
OR1B1 p.R192X 20 (145) olfactory receptor 1B1 olfactory receptor	0
OR10X1 p.W66X 47 (118) olfactory receptor 10X1 olfactory receptor	
OR51Q1 p.R236X olfactory receptor 51Q1 olfactory receptor	
OR4X2 p.Y27X olfactory receptor 4X2 olfactory receptor	
OR4X1 p.Y273X 108 (71) olfactory receptor 4X1 olfactory receptor	
OR4C3 p.W174X olfactory receptor 4C3 olfactory receptor	
OR4A15 p.Q256X olfactory receptor 4A15 olfactory receptor	
OR4C16 p.Q17X olfactory receptor 4C16 olfactory receptor	
OR5AR1 p.Q19X olfactory receptor 5AR1 olfactory receptor	
OR10V1 p.Q123X olfactory receptor 10V1 olfactory receptor	
SI C22A10 p.W96X solute carrier family 22 member 10 organic anion transport	ter
KI RK1 p.W74X NKG2-D type II integral membrane protein receptor on NK cells	
HEBP1 p.Q147X heme-binding protein 1 removes free porphyring	nens
ACSM2A p R115X acvI-coA synthetase medium chain member 2A medium chain fatty acid:Co	A ligase
TM4SE5 p.S160X transmembrane 4 L6 family member 5 tetraspannin protein cell pro	liferation
MAP2K3 n Q73X dual specificity MAP kinase kinase 3 component of the MAP kinase	cascade
HAP1 n W104X huntingtin-assoc protein 1 isoform 3 associates with hunting	itin
CDC27 n R631X cell division cycle protein 27 homolog anaphase promoting complex	component
SERPINB7 n R266X sernin pentidase inhibit clade B member 7 inhibits lysine-specific pro-	eases
RHPN2 n Q378X rhophilin-2 organization of actin cytos	eleton
USP29 n Y913X 129 ubiquitin carboxyl-terminal hydrolase 29 de-ubiquitinating (DUB) er	zvme
WEDC8 n R5X WAP four-disulfide core dom protein 8 protease inhibitor	izyine
UBE2NI n I 89X nutative ubia-conjug enzyme E2 N-like 91% identical to UBE2N DN	IA renair
NEIL 1 n R325X endonuclease VIII-like 1 DNA glycoslylase in B	=R
ZACN n Q281X 55 (101) Zn-activated ligand-gated ion channel ?	
ZC3H3 n S880X 6 (21) Zn finger CCCH dom-containing protein 3 ?	
MAGEE2 n E120X 33 (106) melanoma-associated antigen E2 ?	
MAGEB16 n R272X melanoma-associated antigen R16 ?	
C5orf49 n R72X hypothetical protein LOC134121 ?	
UNC93A n W151X protein unc-93 homolog A isoform 2 ?	
CC2D2B n C320X protein CC2D2B isoform 2 ?	
PRAMEF2 n F211X PRAME family member 2 ?	
MOBKI 2C n R24X mns1 hinder kin activator-like 2C isoform ?	
SPATA8 n R34X spermatogenesis-associated protein 8 2	
TPTE n R211X nutative Tyr-protein phosphatase TPTE ?	
C1orf227 n R37X hypothetical protein LOC149643 ?	
V/WA3B n F219X von Willebrand factor A domain_containing 2	
SERH 2 n Q212X serine hydrolase-like protein 2 2	
FTHL17 p.E148X ferritin heavy polypeptide-like 17 ?	

Table S4. Patient invariant splice site mutations

The table shows genes with mutations in the splice donor site (SD) invariant GT, or splice acceptor site (SA) invariant AG. An invariant splice site mutation was deemed homozygous if we observed multiple homozygous reads at both the mutation site and adjacent SNPs (in parentheses). Other invariant splice site mutations were heterozygous.

Gene	Mutation	Homozygous reads	Protein	
TCTEX1D1	SD1		tctex1 domain-containing protein 1	
HTR3D	SD1	65 (65)	5-hydroxytryptamine receptor 3D isoform 3	
WDR67	SD1		WD repeat-containing protein 67 isoform 1	
C9orf43	SD1		hypothetical protein LOC257169	
NUCB2	SD1	74	nucleobindin-2 precursor	
GSTT2	SD1		glutathione S-transferase theta-2	
SAA1	SD2	3 (115)	serum amyloid A protein preproprotein	
AGL	SA-2		glycogen debranching enzyme isoform 1	
ZFP91	SA-2		zinc finger protein 91 homolog isoform 2	
GREB1	SA-1		protein GREB1 isoform c	
MUC7	SA-1		mucin-7 precursor	
XRCC4	SA-1	14 (24)	DNA repair protein XRCC4 isoform 2	
OAS1	SA-1	43 (86)	2'-5'-oligoadenylate synthase 1 isoform 1	
C13orf26	SA-1		hypothetical protein LOC122046	
C17orf57	SA-1		EF-hand dom-containing protein C17orf57	
LILRA2	SA-1		leukocyte Ig-like rec subfamily	

Table S5. Patient insertion/deletion mutations

The table shows insertions or deletions (indels) sequenced more than once. Indels for CLCA4, SMARCA2, and ATN1 occur in repeated amino acid sequences, and are therefore presumed to be polymorphisms.

Gene	Mutation	Reads	Protein	
CLCA4	2 a.a. del: (PT)5 > (PT)4	2	Calcium-activated chloride channel regulator 4 precursor	
OR7C2	frameshift	2	Olfactory receptor 7C2	
C21orf6	frameshift	3	Uncharacterized protein C21orf62	
GPATCH4	frameshift	3	G patch domain-containing protein 4	
SMARCA2	1 a.a. del: (Q)23 > (Q)22	4	Probable global transcription activator SNF2L2	
ATN1	5 a.a. del: (Q)19 > (Q)14	2	Atrophin-1 (Dentatorubral-pallidoluysian atrophy)	
ALMS1	L525_T527del/insP	2	Alstrom syndrome protein 1	

Table S6. Population deleterious SNPs among 43 hyperammonemia genes

We obtained data from the SNP database <u>https://www.ncbi.nlm.nih.gov/snp</u>. For each gene, the table shows the SNP with the highest allele frequency of the SNPs deemed pathogenic by the 1000 Genomes phase 3 genotype data from 2500 worldwide individuals, as released in the <u>May 2013</u> dataset. The table is restricted to the 16 genes with pathogenic SNPs with global mean allele frequencies of at least 0.0004, which corresponds to 2 observations in 1088 individuals (or 2176 chromosomes).

Gene	Chromosome	SNP id	Protein change	Allele frequency
ACADM	1	<u>rs147559466</u>	E43K	0.0020
ACADVL	17	<u>rs28934585</u>	P65L	0.0391
ASL	22	<u>rs28941471</u>	R190Q	0.0010
ASS1	9	rs35269064	R108L	0.0050
CPS1	2	rs200214298	G169R	0.0004
CPT1A	11	<u>rs2229738</u>	A275T	0.0238
CPT2	1	<u>rs1871748</u>	S565C	0.0016
IVD	15	<u>rs28940889</u>	A284V	0.0004
MCCC2	5	<u>rs119103219</u>	E99Q	0.0004
		<u>rs150591260</u>	V339M	0.0004
MMAB	12	<u>rs35648932</u>	A135T	0.0030
MMACHC	1	<u>rs398124292</u>	frameshift	0.0010
MUT	6	rs547709692	T297I	0.0022
ORNT1	13	<u>rs141028076</u>	intron variant	0.0010
OTC	Х	<u>rs1800328</u>	Q270R	0.0167
PDHA1	Х	<u>rs2229137</u>	M282L	0.0495
SLC22A5	5	<u>rs11568514</u>	Y473D	0.0016

Table S7. Population frequency of pathogenic SNPs

The sum of the allele frequencies for the 16 genes in Table S6 was 0.149. Five additional genes had pathogenic SNPs with global mean allele frequencies of 0.0002, providing a total contribution of 0.001. The remaining 21 genes did not have pathogenic SNPs observed in the 1000 Genomes phase 3 genotype data. Thus, the average number of deleterious SNPs in the global population was estimated to be 0.150. This number was used as the parameter λ for the Poisson distribution, which estimates the probability for n pathogenic SNPs, P(n) = ($\lambda^n/n!$) exp(– λ). Shown are the estimated fraction of the population carrying: zero, P(0); one or more, P(\geq 1); and two or more, P(\geq 2) deleterious SNPs.

Sum of allele frequencies	P(0)	P(≥1)	P(≥2)
0.150	0.861	0.139	0.010

Materials and Methods

DPYD analysis

DPYD enzymatic activity was measured in peripheral blood lymphocytes from the patient and an age-matched healthy control. Samples were harvested at the same time, and shipped on dry ice. The laboratory of Dr. Robert Diasio (Mayo Clinic, Rochester, MN) measured DPYD enzymatic activity and performed DPYD mutation analysis.

Exome sequencing

After obtaining informed consent from the Stanford University Administrative Panel for the Protection of Human Subjects, the patient's exome was analyzed using biotinylated RNA baits to capture exome DNA⁶, and then sequenced to 84-fold coverage, with a range of 6 to 498 reads (Hudson-Alpha Institute, Huntsville, AL). To determine if amino acid substitutions affected protein function, we applied two algorithms. SIFT (Sorting Tolerant From Intolerant) assumes that important positions in the amino acid sequence of a protein have been conserved during evolution, and predicts the effects of substitutions at each position in the amino acid sequence⁷. PolyPhen-2 (Polymorphism Phenotyping version 2) uses sequence-based and structure-based algorithms to predict the functional importance of an amino acid substitution⁸. Allele frequencies and other information for specific genes were obtained from GeneCards (http://www.genecards.org/).

RNA sequence analysis

To determine whether the SD-2 splice donor site SNP (A/C)**A**G|GUPuAGU > (A/C)**G**G|GUPuAGU at position 1083 affects RNA levels, we analyzed published RNAseq data from 12 acute myelogenous leukemias heterozygous for SNPs in at least one of five positions (Fig. 2E): SRR061899, SRR061823, SRR061886, SRR061900, SRR061757, SRR054844, SRR061824, SRR061898, SRR061897, SRR061758, SRR061885, SRR054845, respectively (http://o-www.ncbi.nlm.nih.gov.elis.tmu.edu.tw/Traces/sra/sra.cgi?view=search_obj).

Measurement of plasma ammonia levels

To measure plasma ammonia levels, Stanford Health Care Clinical Laboratory placed the blood sample on ice, and analyzed the sample within 15 minutes. Sample handling is important, because samples not placed on ice, or analyzed after a longer delay yield falsely elevated plasma ammonia levels^{9, 10}.

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